

Draft Genome Sequence and Gene Annotation of *Stemphylium lycopersici* Strain CIDEFI-216

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***Stemphylium lycopersici* is a plant-pathogenic fungus that is widely distributed throughout the world. In tomatoes, it is one of the etiological agents of gray leaf spot disease. Here, we report the first draft genome sequence of *S. lycopersici*, including its gene structure and functional annotation.**

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Dematiaceous fungi belonging to the genus *Stemphylium* Wallr., a taxonomical name established in 1833, produce single-cell conidia (often dictyoconidia) in a percurrent conidiogenous cell. This genus is the anamorphic stage of the teleomorph *Pleospora* (*Pleosporaceae*, *Pleosporales*, *Dothideomycetes*, ascomycetes) (1, 2). Currently, the number of described *Stemphylium* species varies from 30 to 150 (2, 3). Fungi within this genus are widely distributed and have a broad plant host range, establishing a pathogenic, saprotrophic, or entophytic relationship with their hosts (4–7). Pathogenic forms cause severe yield reduction and economic losses in horticultural and fruit tree crops (2, 8). Since *Stemphylium lycopersici* was first described on tomatoes (*Solanum lycopersicum* L.) in 1931 (9, 10), it has been considered the causal agent of leaf spot in >30 host genera worldwide (8, 11). Here, we report the first whole-genome shotgun sequence of the plant-pathogenic fungus *S. lycopersici* strain CIDEFI-216.

S. lycopersici strain CIDEFI-216 was isolated from a tomato plant affected by gray leaf spot disease collected at the tomato-growing area of Bella Vista, Corrientes, Argentina, in 2010. Total genomic DNA was isolated from a monospore culture using the DNeasy plant minikit (Qiagen). The quality of the DNA was assessed by gel electrophoresis, and the quality and quantity were determined by the NanoDrop 2000 UV-Vis spectrophotometer (Thermo Scientific). Libraries were prepared with the TruSeq Nano DNA library preparation kit, 2 × 100-bp paired-end sequencing was performed using an Illumina HiSeq 2000 sequencing system, and reads were assembled with the SOAPdenovo2 assembler software (12) at Macrogen Co. (Seoul, South Korea). *Ab initio* gene prediction was performed using Fgenesh 2.6 (13), trained on an algorithm optimized for *Alternaria brassicicola*. tRNAs and rRNAs were predicted using the tRNAscan-SE (14) and hmmsrch 3.0 (15–18) tools from the WebMGA server (19). A homology-based automatic annotation of the predicted open reading frames (ORFs) was carried out using Blast2GO Basic (20)

(BLASTp against the NCBI nonredundant protein database with an e-value cutoff of 1E-6 and a maximum of 20 BLAST hits) and InterProScan (21); gene names, Gene Ontology (GO) terms, Enzyme Commission (EC) numbers, and InterPro codes were assigned.

The shotgun sequencing yielded 3.14 Gbp of raw data, which represents 60-fold genome coverage. The genome was assembled into 414 scaffolds (>1,000 bp; N_{50} , 498,048 bp), with a total length of 35.18 Mbp and an overall G+C content of 50.8%. A total of 8,999 protein-coding genes were predicted; among them, 6,885 were assigned with 26,524 GO terms (42.8% GO function, 40.7% GO process, and 16.5% GO component), 1,526 with 1,723 EC numbers, and 8,169 with 99,069 InterPro codes from all databases that make up the InterPro Consortium. Additionally, 94 tRNAs and 16 rRNAs were found.

The *S. lycopersici* draft genome sequence generated in this study represents a new source of knowledge that will be useful to further understand the molecular mechanisms of its pathogenesis and other lifestyles. It will also be valuable in comparative genomic studies to shed light into the taxonomy of this fungus.

Nucleotide sequence accession numbers. This whole-genome shotgun project has been deposited at DDBJ/EMBL/GenBank under the accession no. [LGLR000000000](https://www.ncbi.nlm.nih.gov/nuclink/LGLR000000000). The version described in this paper is version LGLR01000000.

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REFERENCES

1. Wallroth FG. 1833. Flora Cryptogamica Germaniae; pars posterior, continens Algas et Fungos. Nuremberg: J.L. Schrag.

2. Câmara MP, O'Neill NR, van Berkum P. 2002. Phylogeny of *Stemphylium* spp. based on ITS and glyceraldehyde-3-phosphate dehydrogenase gene sequences. *Mycologia* 94:660–672. <http://dx.doi.org/10.2307/3761717>.
3. Wang Y, Zhang XG. 2006. Three new species of *Stemphylium* from China. *Mycotaxon* 96:77–81.
4. Simmons EG. 1969. Perfect states of *Stemphylium*. *Mycologia* 61:1–26. <http://dx.doi.org/10.2307/3757341>.
5. Ellis MB. 1971. Dematiaceous hyphomycetes, p 608. Commonwealth Mycological Institute, Kew, Surrey, United Kingdom.
6. Farr DF, Bills GF, Chamuris GP, Rossman AY. 1989. Fungi on plants and plant products in the United States. APS Publishing, St. Paul, MN.
7. Debbab A, Aly AH, Edrada-Ebel R, Wray V, Müller WE, Totzke F, Zirrgiebel U, Schächtele C, Kubbutat MH, Lin WH, Mosaddak M, Hakiki A, Proksch P, Ebel R. 2009. Bioactive metabolites from the endophytic fungus *Stemphylium globuliferum* isolated from *Mentha pulegium*. *J Nat Prod* 72:626–663. <http://dx.doi.org/10.1021/np8004997>.
8. Nasehi A, Kadir J, Nasr-Esfahani M, Abed-Ashtiani F, Wong M, Rambe S, Golkhandan E. 2014. Analysis of genetic and virulence variability of *Stemphylium lycopersici* associated with leaf spot of vegetable crops. *Eur J Plant Pathol* 140:261–273. <http://dx.doi.org/10.1007/s10658-014-0460-3>.
9. Enjoji S. 1931. Two diseases of tomato. *J Plant Protect* 18:48–53. (In Japanese.)
10. Ellis MB, Gibson IAS. 1975. *Stemphylium lycopersici*. *CMI Descr Pathog Fungi Bact* 471:1–2.
11. Farr DF, Rossman AY. Fungal databases—quick search. United States Department of Agriculture, Agricultural Research Service, Washington, DC. <http://nt.ars-grin.gov/fungaldatabases/>.
12. Luo R, Liu B, Xie Y, Li Z, Huang W, Yuan J, He G, Chen Y, Pan Q, Liu Y, Tang J, Wu G, Zhang H, Shi Y, Liu Y, Yu C, Wang B, Lu Y, Han C, Cheung DW, Yiu SM, Peng S, Xiaoqian Z, Liu G, Liao X, Li Y, Yang H, Wang J, Lam TW, Wang J. 2012. SOAPdenovo2: an empirically improved memory-efficient short-read *de novo* assembler. *GigaScience* 1:18. <http://dx.doi.org/10.1186/2047-217X-1-18>.
13. Solovyev V, Kosarev P, Seledsov I, Vorobyev D. 2006. Automatic annotation of eukaryotic genes, pseudogenes and promoters. *Genome Biol* 7(Suppl 1):. <http://dx.doi.org/10.1186/gb-2006-7-s1-s10>.
14. Lowe TM, Eddy SR. 1997. tRNAscan-SE: a program for improved detection of transfer RNA genes in genomic sequence. *Nucleic Acids Res* 25:955–964. <http://dx.doi.org/10.1093/nar/25.5.0955>.
15. Eddy SR. 1998. Profile hidden Markov models. *Bioinformatics* 14:755–763. <http://dx.doi.org/10.1093/bioinformatics/14.9.755>.
16. Huang Y, Gilna P, Li W. 2009. Identification of ribosomal RNA genes in metagenomic fragments. *Bioinformatics* 25:1338–1340. <http://dx.doi.org/10.1093/bioinformatics/btp161>.
17. Szymanski M, Barciszewska MZ, Erdmann VA, Barciszewski J. 2002. 5S Ribosomal RNA database. *Nucleic Acids Res* 30:176–178. <http://dx.doi.org/10.1093/nar/30.1.176>.
18. Wuyts J, Perrière G, Van de Peer Y. 2004. The European ribosomal RNA database. *Nucleic Acids Res* 32:D101–D103. <http://dx.doi.org/10.1093/nar/gkh065>.
19. Wu S, Zhu Z, Fu L, Niu B, Li W. 2011. WebMGA: a customizable Web server for fast metagenomic sequence analysis. *BMC Genomics* 12:444. <http://dx.doi.org/10.1186/1471-2164-12-444>.
20. Conesa A, Götz S, Garcia-Gomez JM, Terol J, Talon M, Robles M. 2005. Blast2GO: a universal tool for annotation, visualization and analysis in functional genomics research. *Bioinformatics* 21:3674–3676. <http://dx.doi.org/10.1093/bioinformatics/bti610>.
21. Quevillon E, Silventoinen V, Pillai S, Harte N, Mulder N, Apweiler R, Lopez R. 2005. InterProScan: protein domains identifier. *Nucleic Acids Res* 33:W116–W120. <http://dx.doi.org/10.1093/nar/gki442>.