

Review



Exploring the Relevance of Senotherapeutics for the Current SARS-CoV-2 Emergency and Similar Future Global Health Threats

Marco Malavolta ^{1,*}^(D), Robertina Giacconi ¹^(D), Dario Brunetti ²^(D), Mauro Provinciali ¹^(D) and Fabrizio Maggi ³^(D)

- ¹ Advanced Technology Center for Aging Research, IRCCS INRCA, 60121 Ancona, Italy; r.giacconi@inrca.it (R.G.); m.provinciali@inrca.it (M.P.)
- ² Mitochondrial Medicine Laboratory, Department of Medical Biotechnology and Translational Medicine, University of Milan, 20133 Milan, Italy; dario.brunetti@unimi.it
- ³ Department of Translational Research, University of Pisa, 56126 Pisa, Italy; fabrizio.maggi63@gmail.com
- * Correspondence: m.malavolta@inrca.it; Tel.: +39-0718004116

Received: 21 March 2020; Accepted: 6 April 2020; Published: 8 April 2020



Abstract: The higher death rate caused by COVID-19 in older people, especially those with comorbidities, is a challenge for biomedical aging research. Here we explore the idea that an exacerbated inflammatory response, in particular that mediated by IL-6, may drive the deleterious consequences of the infection. Data shows that other RNA viruses, such as influenza virus, can display enhanced replication efficiency in senescent cells, suggesting that the accumulation of senescent cells with aging and age-related diseases may play a role in this phenomenon. However, at present, we are completely unaware of the response to SARS-CoV and SARS-COV-2 occurring in senescent cells. We deem that this is a priority area of research because it could lead to the development of several therapeutic strategies based on senotherapeutics or prevent unsuccessful attempts. Two of these senotherapeutics, azithromycin and ruxolitinib, are currently undergoing testing for their efficacy in treating COVID-19. The potential of these strategies is not only for ameliorating the consequences of the current emergence of SARS-CoV-2, but also for the future emergence of new viruses or mutated ones for which we are completely unprepared and for which no vaccines are available.

Keywords: cellular senescence; viral infection; inflammation; mitochondria; extracellular vesicles; senoptotics; senolytics; SASP inhibitors

1. Introduction

The global outbreak of severe acute respiratory syndrome coronavirus (SARS-CoV) in 2003 and the more recent coronavirus 2 (SARS-CoV-2) have made it clear that older people, especially those with comorbidities, die more easily from these infections than younger people [1–3]. The global spread of the novel coronavirus disease (COVID-19), and the emerging evidence of a sustained human-to-human transmission, suggest that we are dealing with a historic challenge to our capacity to protect the health of our elderly community.

This pandemic requires concerted action in all sectors of public health, but it also poses a challenge for biomedical aging research. Indeed, understanding the biological mechanisms that are the basis of the higher susceptibility to death in elderly people may be important to plan preventive and therapeutic strategies in the current pandemic, as well as for the future emergence of similar pandemic viruses for which vaccines are unavailable.

2. Mechanism Driving Inflammation during SARS-CoV-2 Infection

A reduced proportion of lymphocytes and an increased inflammatory response have been observed in the elderly with the new-type of SARS-CoV-2 pneumonia [4]. The novel coronavirus has been reported to use the same receptor, angiotensin-converting enzyme 2 (ACE2), used by SARS-CoV, and spreads mainly through the respiratory tract [5,6]. However, this seems not to be the only target used by coronavirus to attack human host cells. Glucose regulated protein 78 (GRP78) [7] and furin [8] have also been identified as membrane proteins that mediate the viral invasion. Once inside cells, the viral RNAs are detected by the pattern recognition receptors (PRRs) which trigger a downstream cascade of molecules leading to the activation of the transcription factor nuclear factor- κ B (NF- κ B) and interferon regulatory factor 3 (IRF3), with the subsequent production of type I interferons (IFN- α/β) and a series of pro-inflammatory cytokines [2,9] including interleukin (IL)-1 β and IL-6 [10]. The development of an uncontrolled inflammatory response can thus lead to potentially life-threatening damage to lung tissue.

For this reason, a clinical trial with a monoclonal antibody against the IL-6 receptor (tocilizumab, also known as atlizumab) has recently started in China in COVID-19 patients (ChiCTR2000029765). Tocilizumab is an immunosuppressive drug, mainly used for the treatment of rheumatoid arthritis (RA) and systemic juvenile idiopathic arthritis. Other drugs with similar targeting, such as TZLS-501 (Tiziana Life Science, London, UK) are also in course of development.

Hence, some of the proposed, putative therapies for COVID-19 to limit death in elderly (comorbid) patients are based on inflammation targets. However, increasing evidence points to the accumulation of senescent cells in different body tissues as the main source of inflammatory components during aging and age-related diseases [11]. Using network proximity analyses of drug targets and CoV-host interactions in the human interactome, additional anti-HCoV repurposable drugs have been identified [12]. Among them, rapamycin (sirolimus) is a specific inhibitor of mTOR that can promote autophagy and suppress the secretory phenotype of senescent cells [13]. Another proposed drug candidate is melatonin which was shown to display similar effects on the senescent secretory phenotype of lung fibroblasts [14]. A bioinformatic approach supported by artificial intelligence (AI) has identified ruxolitinib among the potential therapeutics [15]. This is a potent and selective JAK inhibitor approved for rheumatoid arthritis and myelofibrosis. It has been recently proposed as a potent suppressor of the secretory phenotype of senescent cells [16,17] and is currently being tested in at least one clinical trial for COVID-19 (NCT04331665). In contrast, chloroquine, an autophagy inhibitor, was shown to promote senescence in endothelial cells [18], and is currently undergoing testing in clinical trials for COVID-19 [19]. Chloroquine or its safer analogue hydroxychloroquine, is deemed to display enhanced effect when azithromycin is included in the treatment (NCT04322396). However, another pilot study found no evidence of antiviral clearance or clinical benefits with this combination [20]. The purpose of the combination of hydroxychloroquine with this antibiotic is mostly to prevent bacterial superinfection, but recent literature has identified azithromycin among the compounds (currently named senolytic drugs) which selectively kill senescent cells [21]. Importantly, it was reported that azithromycin presents more general antiviral effects, as shown for Zika virus [22] and influenza A(H1N1)pdm09 virus [23]; moreover it is recommended for the treatment of chronic airways diseases, such as cystic fibrosis [24] and chronic obstructive pulmonary disease [25], in which cellular senescence has been proposed as a key pathological mechanism [26,27]. Current therapies proposed for SARS-CoV-2 involving cellular senescence are reported in Table 1.

In light of this evidence from preliminary therapeutic findings, we deem that the interaction of senescent cells with coronaviruses deserves particular attention in the development of therapies for SARS-CoV, SARS-CoV-2 and future emerging viruses.

Agent	Study Related to COVID-19 Therapy	Target/Pathways Related to Cellular Senescence	References
Tocilizumab (atlizumab)	Clinical trial (ChiCTR2000029765)	Blocker of IL-6R, IL-6 is among the most common SASP factor	[28,29]
Hydroxychloroquine and azithromycin	Clinical Trial (NCT04322396) and other pilot studies	Azithromycin exerts senolytic activity of human senescent lung fibroblasts; Hydroxychloroquine induces senescence in endothelial cells	[18,20,21]
Ruxolitinib	Bioinformatic study (supported by BenevolentAI) Clinical trial (NCT04331665)	JAK inhibitor, SASP suppressor	[15–17]
Rapamycin	Network-based drug repurposing methodology for HCoV	Inhibitor of mTOR, SASP suppressor	[12,13]
Melatonin	Network-based drug repurposing methodology for HCoV	SASP suppressor in lung fibroblasts	[12,14]

Table 1. Proposed therapies for COVID-19 targeting senescent cells or their signaling molecules.

HCoV: Human coronaviruses; IL-6R: Interleukin-6 receptor; mTOR: mammalian target of rapamycin; SASP: senescence-associated secretory phenotype; JAK: Janus kinases.

3. Cellular Senescence and Inflammatory Response

Cellular senescence entails a multitude of processes which culminate in a state of irreversible cell cycle arrest in which cells undergo distinctive phenotypic alterations, including profound chromatin and morphological changes, increased lysosomal activity and levels of tumor-suppressors, including p53, p16 (Cdkn2a), and p21 (Cdkn1a), as well as significant secretome changes [30]. Cellular senescence is considered to be an important anti-cancer mechanism as well as a physiological process for tissue regeneration. However, the excessive accumulation of senescent cells observed in aging and age-related diseases seems to contribute to chronic inflammation as well as to tissue and organ dysfunction.

Hence, it is reasonable to hypothesize the involvement of cellular senescence in the increased death rate of COVID-19 among elderly (comorbid) patients. Drugs targeting IL-6 have been included among the potential strategies to inhibit the deleterious consequences of the senescence-associated secretory phenotype (SASP), the secretome produced by senescent cells [31,32]. The SASP includes cytokines, chemokines, proteases, and growth factors, recently collected in a proteomic database [29], which functionally links the accumulation of senescent cells with its pathological processes [33,34]. However, the SASP appears to be beneficial or deleterious, depending on the biological context. For example, the SASP is also responsible for activating an immune surveillance response to remove senescent cells [35]. This response seems to involve macrophages, T-cells as well as NK-cells, and appears to differ in different tissues. In particular, an orthotopic transplantation model of senescence-inducible tumor cells in mice has provided evidence that, in contrast to the phenomenon observed in the liver [36], NK-cells may limit the efficient clearance of senescent cells in the lung [37]. Although this observation may suggest the hypothesis that senescent cells easily accumulate in aged lungs, understanding the extent and kind of senescent cell accumulation in aging lung tissue is still a challenge. A recent survey of senescent cell markers (presence of p16- and p21-positive cells) found an increased number of p21-positive and p16-positive cells with donor age in skin (epidermis), pancreas, and kidney, but not in lung [38]. However, senescent fibroblasts, smooth muscle cells, and/or alveolar epithelial cells have been implicated in the etiology or progression of several human diseases, including chronic obstructive pulmonary disease, idiopathic pulmonary fibrosis and emphysema [39]. In mice, aging does not contribute to the induction of cellular senescence by cigarette smoke in the lung of a mouse model of COPD/emphysema [40], but other studies using aged wild type mice display an increase of p16Ink4a mRNA and senescence-associated-β-galactosidase (SA-β-Gal) activity in the lung compared to young controls [41,42]. The stimuli that drive lung cells into senescence are still incompletely

understood and may include infection or inflammation due to infection. Notably, senescent cells are not only a source of inflammatory mediators, but they are also carriers of additional damage through a contagious spreading of senescence and inflammation in neighboring and even distant tissues [43]. The inflammatory components of the SASP are not the only player in this phenomenon. Most recent evidence has demonstrated that the SASP includes extracellular vesicles (EVs) through which senescent cells exert central effector functions in the local environment. Senescent cells secrete EVs with a distinctive, but still incompletely characterized, content of miRNA, proteins, and DNA that can spread senescence in surrounding and even distant tissues [44–47], thus promoting further inflammation and catastrophic consequences for the organism [48]. Interestingly, it has been documented that exosomes are crucial components in the pathogenesis of virus infection, but specific studies for SARS-CoV or SARS-CoV are still lacking [49].

4. Cellular Senescence and Response to Viruses

Importantly, cellular senescence has been also described as an anti-viral mechanism [50]. Indeed, the cell cycle arrest and the release of proinflammatory cytokines and chemokines that characterize cellular senescence are strikingly similar to the features observed by many cells during an antiviral response. Senescence can also be induced by prolonged signaling of cytokines, such as β -interferon, in response to viral infection [51] and the SASP can induce senescence in neighboring cells [52] which have a high probability of being infected. Moreover, many viruses encode inhibitors of programmed cell death to subvert the host responses during infection [53], suggesting that chronic viral infection may contribute to the resistance to apoptosis hypothesized for some senescent cells that accumulate with aging [30]. This hypothesis was also confirmed by the observation that the replication of vesicular stomatitis virus is impaired both in primary and tumor senescent cells in comparison to non-senescent cells [54]. Many studies have described a higher prevalence or increased circulating viral load of some viruses, such as cytomegalovirus (CMV) [55] and Torquetenovirus (TTV) [56,57], with aging. The increased grade of inflammation associated with this phenomenon and the association with mortality of TTV and CMV in the elderly highlights the possible involvement of cellular senescence. In this context, TTV viremia can predict CMV reactivation [58] and a mechanistic link between CMV and accumulating senescent cells has already been proposed [59]. The CMV serostatus plays also a prominent role in determining the magnitude of response to influenza vaccination [60].

Some viruses have been shown to induce cellular senescence, such as human immunodeficiency virus-1 [61], Epstein–Barr virus [62], measles virus [63] and dengue virus [64]. Similar results have been obtained using the virulence factor, NS1 protein, of the influenza A virus (IFV-A) [65]. Other viruses have developed mechanisms to overcome senescence and to evade anti-viral response, as reported for simian virus 40 [66] and human papillomavirus [67]. More recently, it has been shown that IFV and Varicella Zoster Virus display enhanced replication efficiency in senescent human bronchial epithelial cells as well as in senescent human dermal fibroblasts compared to non-senescent cells [68].

Coronaviruses are similar to IFVs, in that they both are RNA viruses, but we are completely unaware of the types of interaction and type of response that SARS-CoV or SARS-CoV-2 can display in senescent cells. Regarding the molecular targets of SARS-CoV-2 virus, there is no evidence that these can be upregulated in senescent cells, and some studies may eventually suggest that ACE2 [69] as well as for GRP78 [70] are downregulated by senescence. However, in few months of studies of COVID-19 pathogenesis, at least four routes of entry have been identified and it is unlikely that the reduced expression of one or two of these targets may prevent infection of senescent cells. Conversely, it should be urgently addressed if the results observed for normal IFVs (i.e., that they can induce senescence and can use senescence to increase their replication rate) can be extended also to SARS-CoV and SARS-CoV-2. If this is proven to be true, it could be reasonable to hypothesize that the massive presence of senescent cells in elderly and comorbid patients may exacerbate the mortality rate. The evidence that patients with cancer, hypertension or with smoking habits (conditions associated with a pathological role of cellular senescence) experienced worse outcomes from COVID-19 [71], further supports the

hypothesis that an accumulation of senescent cells may favor the development of severe events during SARS-CoV-2 infection.

Spreading of senescent cells through the organism is limited by immune surveillance [72–74]. Impaired immune surveillance of senescent cells may contribute to their accumulation during aging [75] and to the development of cancer [74]. Some component of the SASP contributes to attract and activate immune cells, thus suggesting that alteration of this process could be particularly important in the case of viral infection. Age-associated changes in CD4 T-cell functionality have been linked to chronic inflammation and decreased immunity [76] as well as to senescence immune surveillance [74,77]. Depletion of CD4 T cells resulted in an enhanced immune-mediated interstitial pneumonitis and delayed clearance of SARS-CoV from the lungs [78].

There are also mechanisms that some viruses have evolved to overcome immune surveillance of infected cells, for example by targeting mitochondria.

Mitochondria dysfunction in senescent cells can drive important and even specific alterations of the SASP [79]. When the mitochondrial quality control system is impaired, mitochondria could release into extracellular space various components (including formyl peptides, mitochondrial DNA, TFAM, cardiolipin, ATP, succinate and cytochrome C) that could promote or exacerbate SASP; this phenomenon is named mitochondrial damage-associated molecular patterns (DAMPs) [80]. Many models of senescent cells are associated with a shift of mitochondria toward more hyperfusion events, resulting in the presence of abnormally elongated mitochondria [81]. It was also demonstrated that the reduction in mitochondrial content by mTOR inhibition prevents senescence and attenuates the SASP [82].

A mechanism based on mitochondria alterations was proposed to explain how SARS-CoV can escape innate immune surveillance. A protein encoded by SARS-CoV designated as open reading frame-9b (ORF-9b) localizes to the mitochondria, where it promotes proteasomal degradation of Drp1, a protein involved in mitochondrial fission [83], leading to mitochondrial hyperfusion [84], which is a common phenotype of senescent cells and a mechanism involved in resistance to apoptosis [85,86]. The SARS-CoV ORF-9b provides a receptive intracellular environment for viral replication by targeting the mitochondrial antiviral signaling protein (MAVS) signalosome [84]. During the SARS-CoV infection, in the presence of ORF-9b, MAVS undergo degradation and this process is accompanied by the loss of TNF Receptor Associated Factor 3 and 6, two other key signaling intermediaries in antiviral defenses. This leads to an impairment of the host cell IFN responses [84]. However, silencing of MAVS induces a general repressive action on NF-KB with the subsequent suppression of IL-6 expression in senescent cells [87]. This may eventually favor viral replication in the initial phases of the infection without alerting the immune system. In agreement, a significantly enhanced production of IL-6 is not detected until day 4 after infection of human lung epithelial cells with SARS-CoV [88]. This could be consistent with an initial suppression of the mitochondrial antiviral response followed by the later development of a senescent-like phenotype (Figure 1). Hence, it should be important to verify if SARS-CoV and SARS-CoV-2 may induce alteration of the SASP in senescent cells through a similar process.

Mitochondrial fusion is generally required for intracellular proliferation of the viruses and evasion of the antiviral innate immune signaling, as demonstrated for the dengue virus infection mechanism [89]. In the case of dengue virus infection, cellular senescence seems to exert an anti-viral function [64] but other viruses, such as IFV, have evolved mechanisms to increase replication in senescent cells [68]. These observations illustrate that the behavior of different viruses in response to senescence can be different and that specific studies for each virus are needed.



Figure 1. Modulation of mitochondrial dynamics upon CoV Infection. ORF-9b, a virulence factor of severe acute respiratory syndrome coronavirus (SARS-CoV), localizes to mitochondria and causes mitochondrial elongation by triggering ubiquitination and proteasomal degradation of dynamin-like protein (DRP1). Further, acting on mitochondria ORF-9b targets the mitochondrial-associated adaptor molecule MAVS signalosome by usurping poly(C)-binding protein 2 (PCBP2) and the HECT domain E3 ligase (AID4E3) to trigger the degradation of MAVS. This reduces host cell interferon responses and antiviral signaling. However, these changes may later trigger cellular senescence and contribute to enhance the inflammatory response via the senescence associated secretory phenotype (SASP).

5. Therapeutic Perspectives

Clarifying the role of cellular senescence in SARS-CoV infection may additionally provide a strong rationale for the use of senotherapeutics, such as SASP inhibitors, in the management of elderly patients affected by COVID-19. Ongoing clinical trials with SASP inhibitors targeting IL-6 as well as with ruxolitinib (NCT04331665) may provide useful information in this field. This research could eventually pave the way to exploration of whether other senotherapeutics, such as senolytics (compounds that selectively kill senescent cells) [90,91], or compounds able to promote clearance of senescent cells by the immune system [16,92], may eventually provide an additional advantage in the COVID-19 therapy.

Regarding this last aspect, T-cells engineered to express the NKG2D chimeric antigen receptor (CAR), which recognizes NKG2D ligands on the surface of SCs, may be used to target senescent cells [16,35]. In turn, the recent discovery of dipeptidyl peptidase 4 (DPP4) as a selective membrane marker of cellular senescence may provide a target for the development of immune-mediated clearance of senescent cells [16,93]. However, senescence immune surveillance in the lung may work through specific and different mechanisms than those reported for other tissues [37]. Hence, addressing this strategy in the case of infections with a lung target may be very challenging.

In the case of senolytics, it should be also addressed if the mechanism used to kill senescent cells can favor the release of intact viral particles in the environment. Preliminary results indicating the efficacy of the senolytic azithromycin in COVID-19 deserve particular attention. In the original paper describing the senolytic activity of this drug, an effect on mitochondria and mitochondrial oxygen consumption rate was reported [21], which are well known metabolic targets to induce apoptosis in

senescent cells [94,95]. Azithromycin can also induce apoptosis by down-regulating Bcl-xL [96], which is a target of various senolytic compounds including ABT-263, ABT-737 and B1331852 [16]. So far, there has not been much attention paid to the mechanism of action of senolytics, as the goal of most research in this area is aimed at removing senescent cells independently from the mechanism of action. Necrosis, necroptosis, apoptosis and other subcategories of processes leading to cell death play an important and differential role in the host response to different viral infections [97]. This is an additional challenge, as it should be carefully evaluated if the mechanism inducing senescent cell death could eventually or not—the spreading of the virus into neighboring cells. In this context, it might be useful to favoremploy the recently proposed general term of 'senocidals' [90], to identify the pharmaceutical class of drugs that selectively kills senescent cells, and to discriminate between compounds that selectively eliminate senescent cells by apoptosis ('senoptotics') or by non-apoptotic means ('senolytics'). In the case of elderly (comorbid) patients, a clean elimination of infected senescent cells may be critical for the termination of viral infection without side effects. While this area of research is completely unexplored, we deem that addressing these aspects could lead to the development of a wide range of therapeutic strategies to ameliorate the consequences associated with various viral infections.

Another aspect that deserves particular attention is that senotherapeutic molecules have off-target effects [98] and this is especially true for elderly or comorbid patients, where senotherapy side-effects due to systemic dosing could actually exacerbate the virally induced disease. Systemic delivery is currently the only means available for these compounds, for which we have convincing preclinical data from rodent studies [16]. However, there is a need to develop more-targeted delivery approaches so that senotherapeutic compounds can reach the senescent cells in the intended tissue site. The recent development of delivery systems based on encapsulation of diagnostic or therapeutic agents with $\beta(1,4)$ -galacto-oligosaccharides and their preferential delivery to lysosomes of senescent cells is a step forward towards this intent [99]. Notably, this approach was effective in-vivo against pulmonary fibrosis.

The potential therapeutic strategies and the key points that need to be addressed to pave the way to these strategies are shown in Figure 2. This field of research could be important, not only to cope with the current SARS-CoV-2 emergency, but also for future emergencies related to new or mutated viruses for which we are not fully prepared and for which there are no vaccines available.





Funding: This study was supported by Ricerca Corrente funding from Ministero della Salute to IRCCS INRCA. **Conflicts of Interest:** The authors declare no conflict of interest.

References

- 1. Chan-Yeung, M.; Xu, R.H. SARS: Epidemiology. *Respirology* 2003, *8*, S9–S14. [CrossRef] [PubMed]
- 2. Yang, X.; Yu, Y.; Xu, J.; Shu, H.; Xia, J.; Liu, H.; Wu, Y.; Zhang, L.; Yu, Z.; Fang, M.; et al. Clinical course and outcomes of critically ill patients with SARS-CoV-2 pneumonia in Wuhan, China: A single-centered, retrospective, observational study. *Lancet Respir. Med.* **2020**. [CrossRef]
- Kobayashi, T.; Jung, S.; Linton, N.M.; Kinoshita, R.; Hayashi, K.; Miyama, T.; Anzai, A.; Yang, Y.; Yuan, B.; Akhmetzhanov, A.R.; et al. Communicating the Risk of Death from Novel Coronavirus Disease (COVID-19). *J. Clin. Med.* 2020, *9*, 580. [CrossRef] [PubMed]
- 4. Liu, K.; Chen, Y.; Lin, R.; Han, K. Clinical feature of COVID-19 in elderly patients: A comparison with young and middle-aged patients. *J. Infect.* **2020**. [CrossRef]
- 5. Yan, R.; Zhang, Y.; Li, Y.; Xia, L.; Guo, Y.; Zhou, Q. Structural basis for the recognition of the SARS-CoV-2 by full-length human ACE2. *Science* 2020. [CrossRef]
- 6. Guo, Y.-R.; Cao, Q.-D.; Hong, Z.-S.; Tan, Y.-Y.; Chen, S.-D.; Jin, H.-J.; Tan, K.-S.; Wang, D.-Y.; Yan, Y. The origin, transmission and clinical therapies on coronavirus disease 2019 (COVID-19) outbreak an update on the status. *Mil. Med. Res.* **2020**, *7*, 11. [CrossRef]
- Ibrahim, I.M.; Abdelmalek, D.H.; Elshahat, M.E.; Elfiky, A.A. COVID-19 spike-host cell receptor GRP78 binding site prediction. *J. Infect.* 2020. pii: S0163-4453(20)30107-9. [CrossRef]
- Coutard, B.; Valle, C.; de Lamballerie, X.; Canard, B.; Seidah, N.G.; Decroly, E. The spike glycoprotein of the new coronavirus 2019-nCoV contains a furin-like cleavage site absent in CoV of the same clade. *Antiviral Res.* 2020, 176. [CrossRef]
- 9. Kawai, T.; Akira, S. The role of pattern-recognition receptors in innate immunity: Update on toll-like receptors. *Nat. Immunol.* **2010**, *11*, 373–384. [CrossRef]
- 10. Conti, P.; Ronconi, G.; Caraffa, A.; Gallenga, C.; Ross, R.; Frydas, I.; Kritas, S. Induction of pro-inflammatory cytokines (IL-1 and IL-6) and lung inflammation by COVID-19: Anti-inflammatory strategies. *J. Biol. Regul. Homeost. Agents* **2020**, *34*.
- Zampino, M.; Ferrucci, L.; Semba, R.D. Biomarkers in the path from cellular senescence to frailty. *Exp. Gerontol.* 2020, 129, 110750. [CrossRef] [PubMed]
- 12. Zhou, Y.; Hou, Y.; Shen, J.; Huang, Y.; Martin, W.; Cheng, F. Network-based drug repurposing for novel coronavirus 2019-nCoV/SARS-CoV-2. *Cell Discov.* **2020**, *6*, 14. [CrossRef] [PubMed]
- Wang, R.; Yu, Z.; Sunchu, B.; Shoaf, J.; Dang, I.; Zhao, S.; Caples, K.; Bradley, L.; Beaver, L.M.; Ho, E.; et al. Rapamycin inhibits the secretory phenotype of senescent cells by a Nrf2-independent mechanism. *Aging Cell* 2017, *16*, 564–574. [CrossRef] [PubMed]
- Yu, S.; Wang, X.; Geng, P.; Tang, X.; Xiang, L.; Lu, X.; Li, J.; Ruan, Z.; Chen, J.; Xie, G.; et al. Melatonin regulates PARP1 to control the senescence-associated secretory phenotype (SASP) in human fetal lung fibroblast cells. *J. Pineal Res.* 2017, 63. [CrossRef]
- 15. Stebbing, J.; Phelan, A.; Griffin, I.; Tucker, C.; Oechsle, O.; Smith, D.; Richardson, P. COVID-19: Combining antiviral and anti-inflammatory treatments. *Lancet Infect. Dis.* **2020**, *20*, 400–402. [CrossRef]
- Kim, E.C.; Kim, J.R. Senotherapeutics: Emerging strategy for healthy aging and age-related disease. *BMB Rep.* 2019, 52, 47–55. [CrossRef]
- 17. Xu, M.; Palmer, A.K.; Ding, H.; Weivoda, M.M.; Pirtskhalava, T.; White, T.A.; Sepe, A.; Johnson, K.O.; Stout, M.B.; Giorgadze, N.; et al. Targeting senescent cells enhances adipogenesis and metabolic function in old age. *Elife* **2015**, *4*, e12997. [CrossRef]
- Lin, J.-R.; Shen, W.-L.; Yan, C.; Gao, P.-J. Downregulation of dynamin-related protein 1 contributes to impaired autophagic flux and angiogenic function in senescent endothelial cells. *Arterioscler. Thromb. Vasc. Biol.* 2015, 35, 1413–1422. [CrossRef]
- 19. Cortegiani, A.; Ingoglia, G.; Ippolito, M.; Giarratano, A.; Einav, S. A systematic review on the efficacy and safety of chloroquine for the treatment of COVID-19. *J. Crit. Care* **2020**. [CrossRef]

- 20. Molina, J.M.; Delaugerre, C.; Goff, J.L.; Mela-Lima, B.; Ponscarme, D.; Goldwirt, L.; de Castro, N. No Evidence of Rapid Antiviral Clearance or Clinical Benefit with the Combination of Hydroxychloroquine and Azithromycin in Patients with Severe COVID-19 Infection. *Médecine Mal. Infect.* **2020**. [CrossRef]
- 21. Ozsvari, B.; Nuttall, J.R.; Sotgia, F.; Lisanti, M.P. Azithromycin and Roxithromycin define a new family of "senolytic" drugs that target senescent human fibroblasts. *Aging (Albany. NY).* **2018**, *10*, 3294–3307. [CrossRef] [PubMed]
- 22. Li, C.; Zu, S.; Deng, Y.Q.; Li, D.; Parvatiyar, K.; Quanquin, N.; Shang, J.; Sun, N.; Su, J.; Liu, Z.; et al. Azithromycin protects against Zika virus infection by upregulating virus-induced type I and III interferon responses. *Antimicrob. Agents Chemother.* **2019**, 63. [CrossRef] [PubMed]
- 23. Tran, D.H.; Sugamata, R.; Hirose, T.; Suzuki, S.; Noguchi, Y.; Sugawara, A.; Ito, F.; Yamamoto, T.; Kawachi, S.; Akagawa, K.S.; et al. Azithromycin, a 15-membered macrolide antibiotic, inhibits influenza A(H1N1)pdm09 virus infection by interfering with virus internalization process. *J. Antibiot. (Tokyo).* **2019**, *72*, 759–768. [CrossRef] [PubMed]
- 24. Principi, N.; Blasi, F.; Esposito, S. Azithromycin use in patients with cystic fibrosis. *Eur. J. Clin. Microbiol. Infect. Dis.* **2015**, *34*, 1071–1079. [CrossRef]
- 25. Menzel, M.; Akbarshahi, H.; Bjermer, L.; Uller, L. Azithromycin induces anti-viral effects in cultured bronchial epithelial cells from COPD patients. *Sci. Rep.* **2016**, *6*, 28698. [CrossRef]
- 26. Bezzerri, V.; Piacenza, F.; Caporelli, N.; Malavolta, M.; Provinciali, M.; Cipolli, M. Is cellular senescence involved in cystic fibrosis? *Respir. Res.* **2019**, *20*, 32. [CrossRef]
- 27. Kumar, M.; Seeger, W.; Voswinckel, R. Senescence-associated secretory phenotype and its possible role in chronic obstructive pulmonary disease. *Am. J. Respir. Cell Mol. Biol.* **2014**. [CrossRef]
- 28. Zhang, C.; Wu, Z.; Li, J.-W.; Zhao, H.; Wang, G.-Q. The cytokine release syndrome (CRS) of severe COVID-19 and Interleukin-6 receptor (IL-6R) antagonist Tocilizumab may be the key to reduce the mortality. *Int. J. Antimicrob. Agents* **2020**, 105954. [CrossRef]
- 29. Basisty, N.; Kale, A.; Jeon, O.H.; Kuehnemann, C.; Payne, T.; Rao, C.; Holtz, A.; Shah, S.; Sharma, V.; Ferrucci, L.; et al. A proteomic atlas of senescence-associated secretomes for aging biomarker development. *PLoS Biol.* **2020**, *18*, e3000599. [CrossRef]
- 30. van Deursen, J.M. The role of senescent cells in ageing. Nature 2014, 509, 439–446. [CrossRef]
- 31. Watanabe, S.; Kawamoto, S.; Ohtani, N.; Hara, E. Impact of senescence-associated secretory phenotype and its potential as a therapeutic target for senescence-associated diseases. *Cancer Sci.* **2017**, *108*, 563–569. [CrossRef]
- 32. Saleh, T.; Tyutynuk-Massey, L.; Cudjoe, E.K.; Idowu, M.O.; Landry, J.W.; Gewirtz, D.A. Non-cell autonomous effects of the senescence-associated secretory phenotype in cancer therapy. *Front. Oncol.* **2018**, *8*. [CrossRef] [PubMed]
- 33. Coppé, J.-P.; Desprez, P.-Y.; Krtolica, A.; Campisi, J. The senescence-associated secretory phenotype: The dark side of tumor suppression. *Annu. Rev. Pathol.* **2010**, *5*, 99–118. [CrossRef] [PubMed]
- 34. Bojko, A.; Czarnecka-Herok, J.; Charzynska, A.; Dabrowski, M.; Sikora, E. Diversity of the Senescence Phenotype of Cancer Cells Treated with Chemotherapeutic Agents. *Cells* **2019**, *8*, 1501. [CrossRef] [PubMed]
- 35. Burton, D.G.A.; Stolzing, A. Cellular senescence: Immunosurveillance and future immunotherapy. *Ageing Res. Rev.* **2018**, *43*, 17–25. [CrossRef] [PubMed]
- 36. Krizhanovsky, V.; Yon, M.; Dickins, R.A.; Hearn, S.; Simon, J.; Miething, C.; Yee, H.; Zender, L.; Lowe, S.W. Senescence of Activated Stellate Cells Limits Liver Fibrosis. *Cell* **2008**. [CrossRef]
- Stokes, K.L.; Cortez-Retamozo, V.; Acosta, J.; Lauderback, B.; Robles-Oteiza, C.; Cicchini, M.; Pittet, M.J.; Feldser, D.M. Natural killer cells limit the clearance of senescent lung adenocarcinoma cells. *Oncogenesis* 2019, *8*, 24. [CrossRef] [PubMed]
- 38. Idda, M.L.; McClusky, W.G.; Lodde, V.; Munk, R.; Abdelmohsen, K.; Rossi, M.; Gorospe, M. Survey of senescent-cell markers with age in human tissues. *Aging (Albany. NY)* **2020**, *12*, 4052–4066. [CrossRef]
- 39. Campisi, J. Cellular senescence and lung function during aging: Yin and Yang. *Ann. Am. Thorac. Soc.* **2016**, *13*, S402–S406. [CrossRef]
- 40. Rashid, K.; Sundar, I.K.; Gerloff, J.; Li, D.; Rahman, I. Lung cellular senescence is independent of aging in a mouse model of COPD/emphysema. *Sci. Rep.* **2018**, *8*, 9023. [CrossRef]
- 41. Perez-Lanzon, M.; Zitvogel, L.; Kroemer, G. Failure of immunosurveillance accelerates aging. *Oncoimmunology* **2019**, *8*, e1575117. [CrossRef] [PubMed]

- 42. Yousefzadeh, M.J.; Zhao, J.; Bukata, C.; Wade, E.A.; McGowan, S.J.; Angelini, L.A.; Bank, M.P.; Gurkar, A.U.; McGuckian, C.A.; Calubag, M.F.; et al. Tissue specificity of senescent cell accumulation during physiologic and accelerated aging of mice. *Aging Cell* **2020**. [CrossRef] [PubMed]
- 43. Ogrodnik, M.; Salmonowicz, H.; Gladyshev, V.N. Integrating cellular senescence with the concept of damage accumulation in aging: Relevance for clearance of senescent cells. *Aging Cell* **2019**, *18*. [CrossRef] [PubMed]
- 44. Takasugi, M. Emerging roles of extracellular vesicles in cellular senescence and aging. *Aging Cell* **2018**, *17*, e12734. [CrossRef]
- 45. Alibhai, F.J.; Lim, F.; Yeganeh, A.; DiStefano, P.V.; Binesh-Marvasti, T.; Belfiore, A.; Wlodarek, L.; Gustafson, D.; Millar, S.; Li, S.-H.; et al. Cellular senescence contributes to age-dependent changes in circulating extracellular vesicle cargo and function. *Aging Cell* **2020**, e13103. [CrossRef]
- 46. Jeon, O.H.; Wilson, D.R.; Clement, C.C.; Rathod, S.; Cherry, C.; Powell, B.; Lee, Z.; Khalil, A.M.; Green, J.J.; Campisi, J.; et al. Senescence cell–associated extracellular vesicles serve as osteoarthritis disease and therapeutic markers. *JCI Insight* **2019**, *4*. [CrossRef]
- 47. Mensà, E.; Guescini, M.; Giuliani, A.; Bacalini, M.G.; Ramini, D.; Corleone, G.; Ferracin, M.; Fulgenzi, G.; Graciotti, L.; Prattichizzo, F.; et al. Small extracellular vesicles deliver miR-21 and miR-217 as pro-senescence effectors to endothelial cells. *J. Extracell. Vesicles* **2020**, *9*, 1725285. [CrossRef]
- Xu, M.; Pirtskhalava, T.; Farr, J.N.; Weigand, B.M.; Palmer, A.K.; Weivoda, M.M.; Inman, C.L.; Ogrodnik, M.B.; Hachfeld, C.M.; Fraser, D.G.; et al. Senolytics improve physical function and increase lifespan in old age. *Nat. Med.* 2018, 24, 1246–1256. [CrossRef]
- 49. Crenshaw, B.J.; Gu, L.; Sims, B.; Matthews, Q.L. Exosome Biogenesis and Biological Function in Response to Viral Infections. *Open Virol. J.* **2018**, *12*, 134–148. [CrossRef]
- 50. Reddel, R.R. Senescence: An antiviral defense that is tumor suppressive? *Carcinogenesis* **2010**, *31*, 19–26. [CrossRef]
- 51. Moiseeva, O.; Mallette, F.A.; Mukhopadhyay, U.K.; Moores, A.; Ferbeyre, G. DNA damage signaling and p53-dependent senescence after prolonged beta-interferon stimulation. *Mol. Biol. Cell* **2006**, *17*, 1583–1592. [CrossRef] [PubMed]
- 52. Nelson, G.; Kucheryavenko, O.; Wordsworth, J.; von Zglinicki, T. The senescent bystander effect is caused by ROS-activated NF-κB signalling. *Mech. Ageing Dev.* **2018**, 170, 30–36. [CrossRef] [PubMed]
- 53. Upton, J.W.; Chan, F.K.M. Staying alive: Cell death in antiviral immunity. *Mol. Cell* **2014**, *54*, 273–280. [CrossRef] [PubMed]
- 54. Baz-Martínez, M.; Da Silva-Álvarez, S.; Rodríguez, E.; Guerra, J.; El Motiam, A.; Vidal, A.; Garciá-Caballero, T.; González-Barcia, M.; Sánchez, L.; Munõz-Fontela, C.; et al. Cell senescence is an antiviral defense mechanism. *Sci. Rep.* **2016**, *6*. [CrossRef]
- 55. Spyridopoulos, I.; Martin-Ruiz, C.; Hilkens, C.; Yadegarfar, M.E.; Isaacs, J.; Jagger, C.; Kirkwood, T.; von Zglinicki, T. CMV seropositivity and T-cell senescence predict increased cardiovascular mortality in octogenarians: Results from the Newcastle 85+ study. *Aging Cell* **2016**, *15*, 389–392. [CrossRef]
- 56. Giacconi, R.; Maggi, F.; Macera, L.; Pistello, M.; Provinciali, M.; Giannecchini, S.; Martelli, F.; Spezia, P.G.; Mariani, E.; Galeazzi, R.; et al. Torquetenovirus (TTV) load is associated with mortality in Italian elderly subjects. *Exp. Gerontol.* **2018**, *112*, 103–111. [CrossRef]
- 57. Giacconi, R.; Maggi, F.; Macera, L.; Spezia, P.G.; Pistello, M.; Provinciali, M.; Piacenza, F.; Basso, A.; Bürkle, A.; Moreno-Villanueva, M.; et al. Prevalence and loads of torquetenovirus (TTV) in the European MARK-AGE Study population. *J. Gerontol. A. Biol. Sci. Med. Sci.* **2019**. [CrossRef]
- Maggi, F.; Focosi, D.; Statzu, M.; Bianco, G.; Costa, C.; Macera, L.; Spezia, P.G.; Medici, C.; Albert, E.; Navarro, D.; et al. Early Post-Transplant Torquetenovirus Viremia Predicts Cytomegalovirus Reactivations In Solid Organ Transplant Recipients. *Sci. Rep.* 2018, *8*, 15490. [CrossRef]
- 59. Heath, J.J.; Grant, M.D. The Immune Response Against Human Cytomegalovirus Links Cellular to Systemic Senescence. *Cells* **2020**, *9*, 766. [CrossRef]
- 60. McElhaney, J.E.; Garneau, H.; Camous, X.; Dupuis, G.; Pawelec, G.; Baehl, S.; Tessier, D.; Frost, E.H.; Frasca, D.; Larbi, A.; et al. Predictors of the antibody response to influenza vaccination in older adults with type 2 diabetes. *BMJ Open Diabetes Res. Care* **2015**, *3*, e000140. [CrossRef]
- 61. Chen, N.C.; Partridge, A.T.; Tuzer, F.; Cohen, J.; Nacarelli, T.; Navas-Martín, S.; Sell, C.; Torres, C.; Martín-García, J. Induction of a senescence-like phenotype in cultured human fetal microglia during HIV-1 infection. *J. Gerontol. Ser. A Biol. Sci. Med. Sci.* **2018**, *73*, 1187–1196. [CrossRef] [PubMed]

- 62. Hafez, A.Y.; Luftig, M.A. Characterization of the EBV-induced persistent DNA damage response. *Viruses* **2017**, *9*.
- Chuprin, A.; Gal, H.; Biron-Shental, T.; Biran, A.; Amiel, A.; Rozenblatt, S.; Krizhanovsky, V. Cell fusion induced by ERVWE1 or measles virus causes cellular senescence. *Genes Dev.* 2013, 27, 2356–2366. [CrossRef] [PubMed]
- 64. Abubakar, S.; Shu, M.H.; Johari, J.; Wong, P.F. Senescence affects endothelial cells susceptibility to dengue virus infection. *Int. J. Med. Sci.* 2014, *11*, 538–544. [CrossRef] [PubMed]
- Yan, Y.; Du, Y.; Zheng, H.; Wang, G.; Li, R.; Chen, J.; Li, K. NS1 of H7N9 Influenza A Virus Induces NO-Mediated Cellular Senescence in Neuro2a Cells. *Cell. Physiol. Biochem.* 2017, 43, 1369–1380. [CrossRef] [PubMed]
- 66. Bryan, T.M.; Reddel, R.R. SV40-induced immortalization of human cells. *Crit. Rev. Oncog.* **1994**, *5*, 331–357. [CrossRef] [PubMed]
- 67. DiPaolo, J.A.; Popescu, N.C.; Alvarez, L.; Woodworth, C.D. Cellular and molecular alterations in human epithelial cells transformed by recombinant human papillomavirus DNA. *Crit. Rev. Oncog.* **1993**, *4*, 337–360.
- Kim, J.A.; Seong, R.K.; Shin, O.S. Enhanced viral replication by cellular replicative senescence. *Immune Netw.* 2016, 16, 286–295. [CrossRef]
- Takeshita, H.; Yamamoto, K.; Nozato, S.; Takeda, M.; Fukada, S.; Inagaki, T.; Tsuchimochi, H.; Shirai, M.; Nozato, Y.; Fujimoto, T.; et al. Angiotensin-converting enzyme 2 deficiency accelerates and angiotensin 1-7 restores age-related muscle weakness in mice. *J. Cachexia. Sarcopenia Muscle* 2018, *9*, 975–986. [CrossRef]
- 70. Li, W.; Wang, W.; Li, Y.; Wang, W.; Wang, T.; Li, L.; Han, Z.; Wang, S.; Ma, D.; Wang, H. Proteomics analysis of normal and senescent NG108-15 cells: GRP78 plays a negative role in cisplatin-induced senescence in the NG108-15 cell line. *PLoS ONE* **2014**, *9*, e90114. [CrossRef]
- 71. Liang, W.; Guan, W.; Chen, R.; Wang, W.; Li, J.; Xu, K.; Li, C.; Ai, Q.; Lu, W.; Liang, H.; et al. Cancer patients in SARS-CoV-2 infection: A nationwide analysis in China. *Lancet. Oncol.* **2020**, *21*, 335–337. [CrossRef]
- 72. Xue, W.; Zender, L.; Miething, C.; Dickins, R.A.; Hernando, E.; Krizhanovsky, V.; Cordon-Cardo, C.; Lowe, S.W. Senescence and tumour clearance is triggered by p53 restoration in murine liver carcinomas. *Nature* **2007**, *445*, 656–660. [CrossRef] [PubMed]
- 73. Sagiv, A.; Krizhanovsky, V. Immunosurveillance of senescent cells: The bright side of the senescence program. *Biogerontology* **2013**, *14*, 617–628. [CrossRef] [PubMed]
- Kang, T.-W.; Yevsa, T.; Woller, N.; Hoenicke, L.; Wuestefeld, T.; Dauch, D.; Hohmeyer, A.; Gereke, M.; Rudalska, R.; Potapova, A.; et al. Senescence surveillance of pre-malignant hepatocytes limits liver cancer development. *Nature* 2011, 479, 547–551. [CrossRef] [PubMed]
- 75. Ovadya, Y.; Landsberger, T.; Leins, H.; Vadai, E.; Gal, H.; Biran, A.; Yosef, R.; Sagiv, A.; Agrawal, A.; Shapira, A.; et al. Impaired immune surveillance accelerates accumulation of senescent cells and aging. *Nat. Commun.* **2018**, *9*. [CrossRef] [PubMed]
- 76. Elyahu, Y.; Hekselman, I.; Eizenberg-Magar, I.; Berner, O.; Strominger, I.; Schiller, M.; Mittal, K.; Nemirovsky, A.; Eremenko, E.; Vital, A.; et al. Aging promotes reorganization of the CD4 T cell landscape toward extreme regulatory and effector phenotypes. *Sci. Adv.* 2019, *5*, eaaw8330. [CrossRef]
- 77. Prata, L.G.P.L.; Ovsyannikova, I.G.; Tchkonia, T.; Kirkland, J.L. Senescent cell clearance by the immune system: Emerging therapeutic opportunities. *Semin. Immunol.* **2018**, *40*, 101275. [CrossRef]
- Chen, J.; Lau, Y.F.; Lamirande, E.W.; Paddock, C.D.; Bartlett, J.H.; Zaki, S.R.; Subbarao, K. Cellular Immune Responses to Severe Acute Respiratory Syndrome Coronavirus (SARS-CoV) Infection in Senescent BALB/c Mice: CD4+ T Cells Are Important in Control of SARS-CoV Infection. J. Virol. 2010, 84, 1289–1301. [CrossRef]
- 79. Wiley, C.D.; Velarde, M.C.; Lecot, P.; Liu, S.; Sarnoski, E.A.; Freund, A.; Shirakawa, K.; Lim, H.W.; Davis, S.S.; Ramanathan, A.; et al. Mitochondrial dysfunction induces senescence with a distinct secretory phenotype. *Cell Metab.* **2016**, *23*, 303–314. [CrossRef]
- 80. Grazioli, S.; Pugin, J. Mitochondrial damage-associated molecular patterns: From inflammatory signaling to human diseases. *Front. Immunol.* **2018**, *9*, 832. [CrossRef]
- 81. Ziegler, D.V.; Wiley, C.D.; Velarde, M.C. Mitochondrial effectors of cellular senescence: Beyond the free radical theory of aging. *Aging Cell* **2015**, *14*, 1–7. [CrossRef] [PubMed]
- 82. Correia-Melo, C.; Marques, F.D.; Anderson, R.; Hewitt, G.; Hewitt, R.; Cole, J.; Carroll, B.M.; Miwa, S.; Birch, J.; Merz, A.; et al. Mitochondria are required for pro-ageing features of the senescent phenotype. *EMBO J.* **2016**, *35*, 724–742. [CrossRef] [PubMed]

- 83. Tilokani, L.; Nagashima, S.; Paupe, V.; Prudent, J. Mitochondrial dynamics: Overview of molecular mechanisms. *Essays Biochem.* **2018**, *62*, 341–360. [PubMed]
- Shi, C.-S.; Qi, H.-Y.; Boularan, C.; Huang, N.-N.; Abu-Asab, M.; Shelhamer, J.H.; Kehrl, J.H. SARS-Coronavirus Open Reading Frame-9b Suppresses Innate Immunity by Targeting Mitochondria and the MAVS/TRAF3/TRAF6 Signalosome. *J. Immunol.* 2014, 193, 3080–3089. [CrossRef]
- Frank, S.; Gaume, B.; Bergmann-Leitner, E.S.; Leitner, W.W.; Robert, E.G.; Catez, F.; Smith, C.L.; Youle, R.J. The Role of Dynamin-Related Protein 1, a Mediator of Mitochondrial Fission, in Apoptosis. *Dev. Cell* 2001, 1, 515–525. [CrossRef]
- Karbowski, M.; Lee, Y.J.; Gaume, B.; Jeong, S.Y.; Frank, S.; Nechushtan, A.; Santel, A.; Fuller, M.; Smith, C.L.; Youle, R.J. Spatial and temporal association of Bax with mitochondrial fission sites, Drp1, and Mfn2 during apoptosis. J. Cell Biol. 2002, 159, 931–938. [CrossRef]
- 87. Liu, F.; Wu, S.; Ren, H.; Gu, J. Klotho suppresses RIG-I-mediated senescence-associated inflammation. *Nat. Cell Biol.* **2011**, *13*, 254–262. [CrossRef]
- Yoshikawa, T.; Hill, T.; Li, K.; Peters, C.J.; Tseng, C.-T.K. Severe Acute Respiratory Syndrome (SARS) Coronavirus-Induced Lung Epithelial Cytokines Exacerbate SARS Pathogenesis by Modulating Intrinsic Functions of Monocyte-Derived Macrophages and Dendritic Cells. J. Virol. 2009, 83, 3039–3048. [CrossRef]
- 89. Barbier, V.; Lang, D.; Valois, S.; Rothman, A.L.; Medin, C.L. Dengue virus induces mitochondrial elongation through impairment of Drp1-triggered mitochondrial fission. *Virology* **2017**, *500*, 149–160. [CrossRef]
- Myrianthopoulos, V.; Evangelou, K.; Vasileiou, P.V.S.; Cooks, T.; Vassilakopoulos, T.P.; Pangalis, G.A.; Kouloukoussa, M.; Kittas, C.; Georgakilas, A.G.; Gorgoulis, V.G. Senescence and senotherapeutics: A new field in cancer therapy. *Pharmacol. Ther.* 2019, 193, 31–49. [CrossRef]
- 91. Myrianthopoulos, V. The emerging field of senotherapeutic drugs. *Future Med. Chem.* **2018**, *10*, 2369–2372. [CrossRef] [PubMed]
- 92. Malavolta, M.; Costarelli, L.; Giacconi, R.; Piacenza, F.; Basso, A.; Pierpaoli, E.; Marchegiani, F.; Cardelli, M.; Provinciali, M.; Mocchegiani, E. Modulators of cellular senescence: Mechanisms, promises, and challenges from in vitro studies with dietary bioactive compounds. *Nutr. Res.* 2014, 34, 1017–1035. [CrossRef] [PubMed]
- Kim, K.M.; Noh, J.H.; Bodogai, M.; Martindale, J.L.; Yang, X.; Indig, F.E.; Basu, S.K.; Ohnuma, K.; Morimoto, C.; Johnson, P.F.; et al. Identification of senescent cell surface targetable protein DPP4. *Genes Dev.* 2017, 31, 1529–1534. [CrossRef] [PubMed]
- 94. Hubackova, S.; Davidova, E.; Rohlenova, K.; Stursa, J.; Werner, L.; Andera, L.; Dong, L.F.; Terp, M.G.; Hodny, Z.; Ditzel, H.J.; et al. Selective elimination of senescent cells by mitochondrial targeting is regulated by ANT2. *Cell Death Differ.* **2019**, *26*, 276–290. [CrossRef]
- Dörr, J.R.; Yu, Y.; Milanovic, M.; Beuster, G.; Zasada, C.; Däbritz, J.H.M.; Lisec, J.; Lenze, D.; Gerhardt, A.; Schleicher, K.; et al. Synthetic lethal metabolic targeting of cellular senescence in cancer therapy. *Nature* 2013, 501, 421–425. [CrossRef]
- Mizunoe, S.; Kadota, J.I.; Tokimatsu, I.; Kishi, K.; Nagai, H.; Nasu, M. Clarithromycin and azithromycin induce apoptosis of activated lymphocytes via down-regulation of Bcl-xL. *Int. Immunopharmacol.* 2004, *4*, 1201–1207. [CrossRef]
- 97. Orzalli, M.H.; Kagan, J.C. Apoptosis and Necroptosis as Host Defense Strategies to Prevent Viral Infection. *Trends Cell Biol.* **2017**, *27*, 800–809. [CrossRef]
- 98. Childs, B.G.; Durik, M.; Baker, D.J.; Van Deursen, J.M. Cellular senescence in aging and age-related disease: From mechanisms to therapy. *Nat. Med.* **2015**, *21*, 1424–1435. [CrossRef]
- Muñoz-Espín, D.; Rovira, M.; Galiana, I.; Giménez, C.; Lozano-Torres, B.; Paez-Ribes, M.; Llanos, S.; Chaib, S.; Muñoz-Martín, M.; Ucero, A.C.; et al. A versatile drug delivery system targeting senescent cells. *EMBO Mol. Med.* 2018, 10. [CrossRef]



© 2020 by the authors. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license (http://creativecommons.org/licenses/by/4.0/).