

Quantifying the source-sink balance and carbohydrate content in three tomato cultivars

Tao Li, Ep Heuvelink and Leo Marcelis

Journal Name:	Frontiers in Plant Science
ISSN:	1664-462X
Article type:	Original Research Article
Received on:	23 Jan 2015
Accepted on:	23 May 2015
Provisional PDF published on:	23 May 2015
Frontiers website link:	www.frontiersin.org
Citation:	Li T, Heuvelink E and Marcelis L(2015) Quantifying the source-sink balance and carbohydrate content in three tomato cultivars. <i>Front. Plant Sci.</i> 6:416. doi:10.3389/fpls.2015.00416
Copyright statement:	© 2015 Li, Heuvelink and Marcelis. This is an open-access article distributed under the terms of the Creative Commons Attribution License (CC BY) . The use, distribution and reproduction in other forums is permitted, provided the original author(s) or licensor are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.

This Provisional PDF corresponds to the article as it appeared upon acceptance, after rigorous peer-review. Fully formatted PDF and full text (HTML) versions will be made available soon.

1
2
3 **Quantifying the source-sink balance and carbohydrate content in**
4 **three tomato cultivars**
5
6
7
8
9
10
11
12

13
14 T. Li^{1,2}, E. Heuvelink¹, L.F.M. Marcelis*¹
15

16 ¹Horticulture and Product Physiology Group, Wageningen University , P.O. Box 630,
17 6700AP Wageningen, the Netherlands

18 ²Institute of Environment and Sustainable Development in Agriculture, Chinese Academy of
19 Agriculture Science, 100081, Beijing, China
20

21
22
23 * Correspondence:

24 Prof. dr. L.F.M. Marcelis, Horticulture and Product Physiology Group, Wageningen
25 University, P.O. Box 16, 6700AA Wageningen, the Netherlands

26 Tel.:+31 (0)317 483678

27 E-mail address: Leo.Marcelis@wur.nl
28
29
30
31
32
33
34
35
36
37
38
39
40
41

42 Number of words: 5440 (excluding figure captions, table, abstract, and references)

43
44 Number of figures: 8
45
46
47

48 **ABSTRACT**

49
50 Supplementary lighting is frequently applied in the winter season for crop production in
51 greenhouses. The effect of supplementary lighting on plant growth depends on the balance
52 between assimilate production in source leaves and the overall capacity of the plants to use
53 assimilates. This study aims at quantifying the source-sink balance and carbohydrate content
54 of three tomato cultivars differing in fruit size, and to investigate to what extent the
55 source/sink ratio correlates with the potential fruit size. Cultivars Komeett (large size),
56 Capricia (medium size) and Sunstream (small size, cherry tomato) were grown from 16 Aug
57 to 21 Nov, at similar crop management as in commercial practice. Supplementary lighting
58 (High Pressure Sodium lamps, photosynthetic active radiation at 1 m below lamps was 162
59 $\mu\text{mol photons m}^{-2} \text{ s}^{-1}$; maximum 10 hours per day depending on solar irradiance level) was
60 applied from 19 Sep onwards. Source strength was estimated from total plant growth rate
61 using periodic destructive plant harvests in combination with the crop growth model
62 TOMSIM. Sink strength was estimated from potential fruit growth rate which was determined
63 from non-destructively measuring the fruit growth rate at non-limiting assimilate supply,
64 growing only one fruit on each truss. Carbohydrate content in leaves and stems were
65 periodically determined. During the early growth stage, 'Komeett' and 'Capricia' showed sink
66 limitation and 'Sunstream' was close to sink limitation. During this stage reproductive organs
67 had hardly formed or were still small and natural irradiance was high (early Sep.) compared to
68 winter months. Subsequently, during the fully fruiting stage all three cultivars were strongly
69 source-limited as indicated by the low source/sink ratio (average source/sink ratio from 50
70 days after planting onwards was 0.17, 0.22 and 0.33 for 'Komeett', 'Capricia' and
71 'Sunstream', respectively). This was further confirmed by the fact that pruning half of the
72 fruits hardly influenced net leaf photosynthesis rates. Carbohydrate content in leaves and
73 stems increased linearly with the source/sink ratio. We conclude that during the early growth
74 stage under high irradiance, tomato plants are sink-limited and that the level of sink limitation
75 differs between cultivars but is not correlated with their potential fruit size. During the fully
76 fruiting stage tomato plants are source-limited and the extent of source limitation of a cultivar
77 is positively correlated with its potential fruit size.

78
79
80
81 **KEY WORDS:** Source-sink balance, plant development stage, carbohydrate content,
82 quantification, tomato cultivars, *Solanum lycopersicum*

98 **INTRODUCTION**

99

100 Plant growth is closely correlated with source and sink strength and the balance between them
101 (Gifford and Evans, 1981; Smith and Stitt, 2007; Wardlaw, 1990). Source strength of a plant
102 is defined as the rate at which the plant produces assimilates (photosynthesis rate). The sink
103 strength of a plant is composed of sink strengths of all individual organs. Sink strength of an
104 organ is the competitive ability of an organ to attract assimilates and can be quantified by its
105 potential growth rate (Marcelis, 1996). Although fruits are the major sink organs in crops like
106 tomato, also leaves, stems and roots utilize assimilates and have a sink strength; hence leaves
107 are not only source organ but also sink organ.

108

109 Source-sink balance regulates carbon status in plants (Osorio et al., 2014). Differences in
110 source-sink balance are expected to result in differences in carbohydrate content in plants
111 (Dingkuhn et al., 2007; Paul and Foyer, 2001; Patrick and Colyvas, 2014). In a source-limited
112 situation, carbohydrate content in the plants might be low as plants have sufficient sinks to
113 utilize the produced assimilates. However, in a sink-limited situation plant growth cannot
114 keep pace with assimilate production. When assimilate production exceeds its utilisation
115 carbohydrates (starch and soluble sugars) are usually stored in leaves (Yelle et al., 1989) as
116 well as stems (Hocking and Steer, 1994; Scofield et al., 2009). Limited sink demand could
117 result in feedback regulation of photosynthesis as it may down-regulate the net photosynthetic
118 activity through carbohydrate accumulation in source leaves (Franck et al., 2006; Iglesias et
119 al., 2002; McCormick et al., 2006; Velez-Ramirez et al., 2014).

120

121 Manipulating source and sink organs (e.g. fruit and leaf pruning) are often applied to
122 investigate plant source-sink balance (Cockshull and Ho, 1995; Iglesias et al., 2002; Matsuda
123 et al., 2011). Crop growth models can be used to quantify the source and sink strength (De
124 Koning, 1994; Heuvelink, 1996b; Wubs et al., 2009, 2012). In these models the sink strength
125 of a growing organ is determined by its potential growth rate (i.e. growth under non-limiting
126 assimilate supply) (Marcelis, 1996), which depends on its developmental stage (Marcelis and
127 Baan Hofman-Eijer, 1995). Cumulating the sink strength of each organ on the plant results in
128 total plant sink strength. The plant source strength is calculated as the supply of assimilates
129 during a day, which is estimated by the crop growth rate ($\text{g dry mass plant}^{-1} \text{day}^{-1}$) (Heuvelink,
130 1995).

131

132 The growth environment plays a pivotal role in determining the source-sink balance.
133 Under non-stressing conditions, irradiance becomes particularly important as it is the driving
134 force for photosynthesis. Supplementary lighting is commonly applied in greenhouses in
135 order to improve crop photosynthesis and thus production (Heuvelink et al., 2006; Moe et al.,
136 2005). The beneficial effect of supplementary lighting is determined by the balance between
137 assimilate production in source leaves and the overall capacity of the plants to use these
138 assimilates. This implies that it is important to identify the plant source-sink balance in order
139 to efficiently utilize supplementary lighting.

140

141 The source-sink balance of a plant varies significantly during its life span because of the
142 continuous organ initiation and development which affects both the sink and source strength
143 (Wardlaw, 1990). During the early growth stage, tomato plants might be prone to sink
144 limitation as there might be insufficient sinks to utilize all the produced assimilates. This
145 might occur especially under high irradiance. During the reproductive stage, tomato plants
146 generally bear many fruits, and assimilate supply might not meet the sink demand. This has
147 been suggested in studies where fruit pruning increased fruit size of the remaining fruits

148 without influencing the total plant biomass production (Cockshull and Ho, 1995; Heuvelink,
149 1996b; Matsuda et al., 2011). Tomato source-sink balance could also differ between cultivars
150 which often differ in fruit load and potential fruit growth rate, suggesting differences in sink
151 strength (Heuvelink and Marcelis, 1989; Marcelis, 1996). Cultivars may also differ in source
152 strength as leaf photosynthetic properties, leaf area and plant architecture may differ. Dueck et
153 al. (2010) observed that under commercial crop management effects of supplementary
154 lighting were small in cherry tomato compared with cultivars with large-sized fruits. They
155 argued that cherry tomato had less sink demand although it bears more fruits. A detailed
156 analysis of the source-sink balance from early growth stage to fully fruiting stage for cultivars
157 with different potential fruit size has not performed so far.

158
159 The objectives of this study are to provide a detailed quantitative analysis of source-sink
160 balance as well as carbohydrate content of tomato plants with standard fruit load during their
161 development; and to investigate to what extent the source/sink ratio of a cultivar depends on
162 the potential fruit size. Our hypotheses are 1) tomato plants are sink-limited during their early
163 growth stage when grown under high irradiance; 2) tomato plants are source-limited during
164 the fully fruiting stage, and the source-sink ratio negatively correlates with the potential fruit
165 size (when comparing cultivars at their commercial fruit load). To test these hypotheses, three
166 types of tomato cultivars with different potential fruit size were grown under conditions
167 comparable to commercial crop management from mid-August until end of November. The
168 source/sink ratio and carbohydrate content were examined during this period through
169 experimental observation combined with model estimation.

170 171 **MATERIALS AND METHODS**

172 173 **Plant materials and growth conditions**

174
175 Tomato (*Solanum lycopersicum*) plants were planted in a Venlo-type glasshouse compartment
176 on 16 August and grown until 21 November 2013. The greenhouse compartment had an area
177 of 150 m² with a gutter height of 5 m, and was located in Wageningen, the Netherlands (52 °N,
178 5 ° E). Eight growth gutters were evenly arranged in the compartment in the East to West
179 direction with a distance of 150 cm between gutters. Plants on each gutter were alternatively
180 trained to two high wires which were 30 cm to the right and left of the growth gutter. 45
181 plants were grown on each gutter at an inter-plant distance of 20 cm. All plants were grown
182 with single shoot. Plant density was initially 3.3 plants m⁻² and gradually decreased to 2.2
183 plants m⁻² at the end of the experiment due to periodical destructive harvests. Plants were
184 grown on Rockwool with drip irrigation according to the commercial practice. From 43 days
185 after planting onwards, leaves below the 2nd lowest truss were regularly removed. Fruits were
186 picked when they turned red-ripe.

187
188 Solar radiation was continuously measured outside the greenhouse throughout the
189 experimental period. Photosynthetic active radiation (PAR) was estimated from solar
190 radiation, assuming half the global radiation is PAR (Jacovides et al., 2003). Greenhouse
191 transmissivity of PAR was 62 %. Supplementary lighting (High Pressure Sodium lamps,
192 HortiluxSchreder, HPS600W/400V) was applied from 19 September until the end of the
193 experiment. PAR of the supplementary lighting was 162 ± 9 μmol photons m⁻² s⁻¹ at 1 m
194 below the lamps. The lamps were turned on when global radiation was below 200 W m⁻² and
195 turned off when it exceeded 300 W m⁻² between 6:00 to 16:00 hours. A standard greenhouse
196 computer (Hoogendoorn-Economic, Hoogendoorn, Vlaardingen, The Netherlands) was used
197 to control the greenhouse climate as well as supplementary lighting. Sunrise to sunset at start

198 of the experiment was from 6:30 to 21:00, it was from 8:00 to 16:40 at end of the experiment.
199 During the experiment, average daily outside global radiation was $9 \text{ MJ m}^{-2} \text{ d}^{-1}$; inside the
200 greenhouse, average day/night temperature was 24/18 °C, air humidity was 77 % and day
201 time CO_2 concentration was $577 \text{ } \mu\text{mol mol}^{-1}$. Daily PAR integral inside the greenhouse is
202 presented in Figure. 1.

203

204 **Treatments**

205

206 Three tomato cultivars with different potential fruit size and with standard fruit load were
207 grown on eight gutters (double rows) in the same greenhouse in order to compare their
208 source-sink balance during plant development: cv. Komeett (large size, 5 fruits per truss),
209 Capricia (medium size, 6 fruits per truss), and Sunstream (small size, 10 fruits per truss).
210 Additionally, a set of plants of these cultivars were pruned to one fruit per truss, in order to
211 determine the potential growth rate of a single fruit which is an estimate of sink strength of a
212 single fruit (Marcelis, 1996). Furthermore, another set of plants of all cultivars were pruned to
213 half fruit load: cv. Komeett (2 fruits per truss), Capricia (3 fruits per truss), Sunstream (5
214 fruits per truss), in order to determine the effect of reduced sink strength on total biomass and
215 net leaf photosynthesis.

216

217 The greenhouse was divided into 3 equal parts, perpendicular on the gutters: at the West
218 side the tallest cultivar (Sunstream) was grown, at the East side the smallest cultivar (Capricia)
219 was grown and in the middle cultivar Komeett was grown. For each of the six central gutters,
220 six plants were grown with standard fruit load and one with half fruit load for each cultivar.
221 The number of plants with standard fruit load was larger than those at half fruit load as for
222 standard fruit load destructive measurements were taken at 6 moments while for half fruit
223 load these measurements were only performed at the end of the experiment. Each plant with
224 standard and half fruit load was surrounded on both sides by an internal border plant. All
225 plants on the two outer gutters as well as the internal border plants were pruned to one fruit
226 per truss. Fruit pruning was done immediately after fruit set for each truss.

227

228 **Plant development registration**

229

230 Observations on flowering and fruit age were taken three times a week. Flowering was
231 defined as three fully open flowers on a truss, which indicates fruit age 0. For the treatment
232 with standard fruit load, 12 plants of each cultivar which were used for the last two
233 destructive harvests were investigated. This observation was used for estimating the sink
234 strength of the plant with standard fruit load. Due to more plants were available for the
235 treatment with one fruit per truss, observations on flowering and fruit age of this treatment
236 were taken on 15-20 plants of each cultivar. Furthermore, the maximum fruit length and
237 diameter of the fruits from the treatment with one fruit per truss were measured with caliper
238 three times a week since fruit set in order to obtain fruit volume over time, number of
239 measured fruits ranged from 34 to 48 fruits per cultivar, these fruits were from the first three
240 trusses which developed in September. The observation of fruit volume and fruit age of the
241 treatment with one fruit per truss was used for estimating the potential growth rate of a single
242 fruit. Total formed truss number was 11, 11, and 14 for Komeett, Capricia, and Sunstream,
243 respectively, until the end of the experiment. Plant development registration was not
244 performed in the treatment with half fruit load due to sink strength of this treatment was not
245 addressed.

246

247 Fruit set started between 20-30 days after planting for the three cultivars. Therefore, the

248 first 30 days after planting was defined as early growth stage, since 30 days after planting
249 onwards was defined as fully fruiting stage.

250

251 **Destructive measurements**

252

253 Six plants per cultivar were destructively measured before planting (on 15 August) to
254 determine their initial total biomass and leaf area. For the plants with standard fruit load six
255 plants of each cultivar (one from each gutter) were harvested on 18, 33, 47, 61, 81, 97 days
256 after planting. For plants with half fruit load six plants (one from each gutter) were harvested
257 on 97 days after planting. Fresh and dry weight of leaves, stems and fruit trusses were
258 determined. Plant organs were dried for at least 48 h at 105°C in a ventilated oven. Leaf area
259 was measured with a leaf area meter (LI-3100C, Li-Corinc., Lincoln, USA). Specific leaf area
260 (SLA) was calculated by dividing leaf area by leaf dry weight. The regularly removed leaves
261 and harvested fruits were dried and dry weight was added to obtain the cumulative dry
262 weights per plant; area of the regularly removed leaves was also determined for estimating
263 total LAI at different moments which was needed as model input.

264

265 For each cultivar, 97 to 148 fruits from the plants with one fruit per truss were randomly
266 sampled during the experiment, the samples were taken once per week, and fruit diameter,
267 length, age, fresh and dry weight were recorded. These observations were used to get two
268 relationships: a relationship between fruit volume and fresh weight; and a relationship
269 between fruit age and fruit dry matter content.

270

271 **Sample collection and carbohydrates analysis**

272

273 Leaf and stem samples for carbohydrate analyses were taken from plants with standard fruit
274 load. Leaf samples were taken at the beginning of the day (6:00-7:00 AM) at one day before
275 each destructive harvest. The samples were taken at every other leaf from leaf number 5
276 (uppermost fully expanded leaf; leaf number 1 was the uppermost leaf longer than 5 cm)
277 downward to the bottom of the canopy. In each selected leaf, one leaflet adjacent to the
278 terminal leaflet was collected. The collected leaflets from one plant were pooled together to
279 represent one canopy leaf sample. Stem samples were taken on the day of destructive harvest.
280 Stem sections (0.5 cm length) were taken from top to the bottom where the leaf samples were
281 taken, these sections were pooled together to represent one stem sample. Six replicates were
282 taken for each type of sample at each time. Fresh weight of all collected samples was
283 determined and added to the total plant weight.

284

285 Samples were inserted in vials and flash-frozen in liquid nitrogen. They were transferred
286 to a freezer (-80 °C) for storage. Starch and soluble sugar content were analysed with a HPLC
287 Dionex system (GS 50 pump and PED 2 electrochemical detector) as described by Savvides
288 et al. (2014); the soluble sugars that were monitored were fructose, glucose and sucrose.

289

290 **Net photosynthesis measurements**

291

292 Net photosynthesis rates were measured with a portable gas exchange device equipped with a
293 leaf chamber fluorometer (Li-6400; LI-COR) at leaf number 6 from top of the canopy. In the
294 measurement chamber, PAR (10% blue, 90% red) was 1000 $\mu\text{mol m}^{-2} \text{s}^{-1}$, CO₂ concentration
295 was 500 $\mu\text{mol mol}^{-1}$, air temperature was 23 °C and vapor pressure deficit (VPD) was
296 between 0.5-1 kPa. The measurements were performed on plants with standard fruit load as
297 well as plants with half fruit load on 20, 28, 39, 54-55, 64-65 and 75-76 days after planting

298 (plants with half fruit load only from 54 days onwards). For each cultivar each time 6
299 measurements were taken before noon (between 8:30 and 12:00) and 6 were taken after noon
300 (between 12:30 and 16:00).

301

302 **Plant source/sink ratio determination**

303

304 Source/sink ratio was estimated based on source strength of the plant divided by the sum of
305 the vegetative sink strength and total fruit sink strength.

306

307 Plant growth rate (g dry mass plant⁻¹ day⁻¹) was used as an estimate of source strength.
308 Daily plant growth rate was estimated by the crop growth model TOMSIM (Heuvelink, 1996b)
309 with measured SLA (from planting date to first destructive harvest date), measured LAI (from
310 first destructive harvest date onwards), dry matter partitioning among plant organs (leaves,
311 fruits, stems, roots), and the climate data (global radiation, intensity and timing of the
312 supplementary lighting, greenhouse temperature and CO₂) were input to the model. The
313 fraction dry matter partitioned to roots was set to 13% at planting; and 4% from first fruit
314 harvest onwards; in between this fraction was estimated by linear interpolation (Heuvelink,
315 1995). Estimated daily plant growth rate was multiplied by a correction factor such that
316 estimated cumulative plant weights corresponded to the measured cumulative plant weights.
317 This factor was estimated by minimizing the sum of squares of the residuals between
318 measured and estimated total dry weight at each destructive harvest (one factor for each
319 cultivar).

320

321 Sink strength of a single fruit, quantified by the potential fruit growth rates, was obtained
322 by non-destructive measurements on potentially growing fruits (i.e. one fruit per truss). On
323 the basis of the lengths and diameters of the potentially growing fruits, their volume was
324 calculated assuming a deformed sphere

$$325 \quad v = \frac{4}{3}\pi\left(\frac{d}{2}\right)^2 \frac{h}{2} \quad (1)$$

326 where v is fruit volume (cm³), d is fruit diameter (cm), h is fruit length (cm).

327

328 Fruit volume was subsequently converted into fresh weight, using a cultivar specific
329 linear regression between fruit volume and fruit fresh weight ($r^2 = 0.97-0.99$ for three
330 cultivars). A Gompertz function was fitted through fresh weight over time

$$331 \quad w(t) = w_{max}e^{-e^{-k(t-t_m)}} \quad (2)$$

332 where $w(t)$ is the weight at age t (d after anthesis), w_{max} is upper asymptote of fruit weight (g),
333 k represents the weighted mean relative growth rate and t_m the age (d) at maximum growth
334 rate.

335

336 The Gompertz function was fitted through the data with non-linear mixed modelling.
337 Non-linear mixed models take into account that the measurements on one fruit are grouped. A
338 lower variation is assumed between the measurements of one fruit than between the
339 measurements of different fruits. The three parameter means (w_{max} , t_m , k) were estimated to
340 describe fruit growth (Wubs et al., 2009).

341

342 A fourth-order polynomial function was fitted for the destructively determined fruit dry
343 matter content as a function of fruit age according to Wubs et al. (2012). The potential growth

344 rate in g dry matter per day of an individual fruit (representing the sink strength of a single
345 fruit) was calculated as the product of the derivative of the Gompertz function for fruit fresh
346 weight and this fourth-order polynomial function. The daily total fruit sink strength of a plant
347 was calculated by accumulating the sink strength of all fruits which were present on the plant
348 that day.

349
350 Vegetative sink strength was estimated as the integral of sink strengths of each vegetative
351 unit (De Koning,1994; Heuvelink,1996b).

$$352 \quad PVGR = ae^{-0.168(T-19)}PFGR \quad (3)$$

353 where *PVGR* is the potential growth rate for a vegetative unit (g d^{-1}) and *PFGR* is the
354 potential fruit growth rate (g d^{-1}) for a single fruit. *a* is a specific factor between potential fruit
355 growth rate and potential growth rate of a vegetative unit, which was estimated by minimizing
356 the sum of squares of the residuals between measured and estimated dry matter partitioning to
357 fruits, the latter was calculated as estimated fruit dry matter divided by cumulative plant dry
358 matter; this factor is cultivar dependent. *T* is the average greenhouse diel temperature during
359 the experiment period ($^{\circ}\text{C}$).

360
361 Before anthesis of the first truss, vegetative growth is an input. Usually about three
362 vegetative units precede the first truss (Dieleman and Heuvelink, 1992), which was also
363 observed in this experiment. The sink strengths of these three units were estimated by using
364 *PVGR* multiplied by three specific factors [0.6, 0.75, 0.9, respectively, from the first to the
365 third unit, these factors were derived based on Heuvelink (1996a)], this is because the first
366 few units are relatively small and hence have a low sink strength. The daily total vegetative
367 sink strength of a plant was calculated by accumulating the vegetative sink strength of all
368 units which were present that day. A more detailed description see De Koning (1994) and
369 Heuvelink (1996a).

370 371 **Statistical analysis**

372
373 Destructive measurements and carbohydrate determination were based on 6 replicate plants;
374 net leaf photosynthesis was based on 12 replicates (two leaves per plant, 6 replicate
375 plants).The effects of cultivars, days after planting, and fruit pruning treatments on measured
376 plant parameters were evaluated by ANOVA followed by Fisher's protected least significant
377 difference test (l.s.d) at 95% confidence, using GenStat16th edition.

378 379 **RESULTS**

380 381 **Plant growth**

382
383 Maximum growth rate and growth duration of single fruit were highest in 'Komeett'; while
384 these parameters were lowest in 'Sunstream' (Figure. 2). These differences together resulted
385 in the largest potential fruit size in 'Komeett' and smallest in 'Sunstream'. Potential fresh fruit
386 weight was 180 g for 'Komeett', 137 g for 'Capricia' and 20 g for 'Sunstream' as determined
387 in this study.

388
389 'Sunstream' had highest LAI during a large part of the growing period (Figure. 3A), and
390 highest total dry weight except for the initial period after planting (Figure. 3B); while these
391 parameters were similar between 'Komeett' and 'Capricia' (Figure. 3). For all cultivars, plant

392 total dry weight was not affected by the half fruit load treatments (Table 1). However, half
393 fruit load treatments resulted in significantly higher fraction of dry mass partitioned to leaves
394 and stems, and lower partitioning to fruits (Table 1).

395

396 **Carbohydrate content and net photosynthesis rate**

397

398 In tomato stems, starch content was negligible compared to sugar content which was
399 apparently the main carbohydrate in stems (Figure. 4A, B). For all cultivars, soluble sugar
400 content was at a high level until 33 days after planting. Thereafter, it decreased gradually until
401 the end of the experiment (Figure. 4A). This phenomenon was not observed for starch content
402 which reached a peak at 33 days after planting for ‘Capricia’ and ‘Sunstream’, and remained
403 relatively constant from 60 days after planting onwards for all three cultivars (Figure. 4B).
404 ‘Sunstream’ had higher sugar content than the other two cultivars ($P < 0.001$) except for at 18
405 days after planting (Fig. 4A); it also had highest starch content ($P < 0.001$) (Figure. 4B).

406

407 In leaves, soluble sugar content was relatively constant during the growing period
408 compared to starch content (Figure. 4C, D). For all cultivars, starch content was initially (18
409 days after planting) high and decreased gradually until 60 days after planting. Surprisingly,
410 starch content at 80 days after planting suddenly increased and reached a level as high as that
411 observed at 18 days after planting in ‘Komeett’. At the end of the experiment, starch content
412 increased in ‘Capricia’ and ‘Sunstream’ (Figure. 4D).

413

414 For all cultivars, the highest net photosynthesis rates were observed at 28 days after
415 planting; thereafter it decreased gradually until the end of the experiment (Figure. 5).
416 Interestingly, net photosynthesis rates at 20 days after planting were tended to be lower than
417 at 28 days after planting, although this difference was only significant in ‘Capricia’ (Figure.
418 5). Furthermore, ‘Capricia’ had higher net photosynthesis rates than the other two cultivars
419 ($P < 0.001$). Half fruit pruning treatments had no effect on net photosynthesis rates in all three
420 cultivars (data not shown).

421

422 **Source-sink balance and its relationship with plant carbohydrate content**

423

424 The vegetative sink strength differed between cultivars and was highest for ‘Sunstream’ and
425 lowest for ‘Capricia’ (Figure. 6A). The total fruit sink strength was highest for ‘Komeett’ and
426 lowest for ‘Sunstream’ (Figure. 6B). Furthermore, the total fruit sink strength was initially
427 low and soon increased to a plateau and kept constant onwards. ‘Sunstream’ had highest total
428 plant sink strength before 25 days after planting; thereafter, ‘Komeett’ had highest and
429 ‘Sunstream’ had lowest total plant sink strength (Figure. 6C).

430

431 Source strength (crop growth rate) was initially low and increased drastically until about
432 30 days after planting (Figure. 7A); it was decreasing from 45 days after planting onwards
433 until the end of the experiment. ‘Sunstream’ had higher source strength than the other two
434 cultivars during a large part of the growing period (Figure. 7A).

435

436 Plant source/sink ratio was initially low (below 1) for all three cultivars, and it soon
437 exceeded 1 in ‘Komeett’ and ‘Capricia’, and came close to 1 in ‘Sunstream’ (Figure. 7B).
438 ‘Komeett’ had shorter duration of sink limitation than ‘Capricia’, the source/sink ratio in
439 ‘Komeett’ was also lower than in ‘Capricia’. During the fully-fruited stage, source/sink ratio
440 was lower than 1 for all three cultivars, ‘Sunstream’ had the highest and ‘Komeett’ had lowest

441 source/sink ratio during this stage. Total carbohydrate content in stems and leaves over the
442 three cultivars increased linearly with the source/sink ratio (Figure. 8).

443

444 **DISCUSSION**

445

446 **Tomato plants are sink-limited during their early growth stage in greenhouses under** 447 **high irradiance**

448

449 Young plants are likely to be sink-limited (Ark and Drake, 1991). Indeed, we found in our
450 study that three types of tomato cultivars experienced a period of sink limitation or came close
451 to sink limitation during their early growth stage (Figure. 7B). Sink limitation during the early
452 growth stage was caused by the low total plant sink strength (Figure. 6C) combined with a
453 fast increase in source strength (Figure. 7A). This increase in source strength resulted from a
454 fast increase in LAI. In addition, irradiance might also have played an important role, because
455 sink limitation was observed during a period (early September) that plants received high
456 natural irradiance to maintain a high rate of net photosynthesis compared to late autumn and
457 winter months (Figure. 1). The combination of the high irradiance and fast increase in LAI
458 with limited reproductive organs during the early growth stage, resulted in plants not being
459 able to use the extra assimilates, so that the high sugar content in stems was observed during
460 this stage (Figure. 4A). Tomato stems have been reported as an important storage organ for
461 assimilates (Hocking and Steer, 1994), this is in line with our study that carbohydrate content
462 in stems was higher than in leaves. Starch is predominantly utilized for diurnal carbon storage
463 in leaves, it degrades to soluble sugar at night for mobilization and utilization (Smith and Stitt,
464 2007; Osorio et al., 2014), so that in stems sugar content was significantly higher than starch
465 content (Figure. 4A). In leaves the highest starch content was observed at 18 days after
466 planting which was during the period of sink limitation (Figure. 4B). Similarly, Nakano et al.
467 (2000) and Plaut et al. (1987) also reported starch accumulation in leaves when sink limitation
468 occurs.

469

470 Photosynthetic capacity often correlates with the source-sink balance (Iglesias et al., 2002;
471 McCormick et al., 2006). In this study, net photosynthesis rates at 20 days after planting
472 tended to be lower than at 28 days after planting when measured at the same conditions,
473 although this was only significant for ‘Capricia’ (Figure. 5). Sink limitation around 20 days
474 after planting in combination with the high starch content in leaves (Figure. 4D) might have
475 led to a slight down-regulation of net photosynthesis (Iglesias et al., 2002; Nakano et al.,
476 2000; Paul and Foyer, 2001). Irradiance induced acclimation could not play a role because the
477 daily light sum was similar during this period (Figure. 1). When young tomato plants not yet
478 producing fruits were grown under elevated CO₂, this resulted in photosynthetic acclimation
479 (Besford, 1993; Yelle et al., 1989), which was probably caused by an imbalance in the
480 supply and demand of assimilates. These studies further indicate that tomato plants are likely
481 sink-limited during the early growth stage.

482

483 Source-sink balance is cultivar specific (Figure. 7B). During the early growth stage
484 cultivar differences in source/sink ratio were mainly due to differences in vegetative sink
485 strength, as reproductive organs had hardly been formed or were still small and source
486 strength was similar for the different cultivars (Figure. 7A). ‘Sunstream’ had the highest
487 vegetative sink strength (Figure. 6A), and hence the lowest source/sink ratio during this
488 period (Figure. 7B). Wubs et al. (2009) also reported that cultivars with the smallest potential
489 fruit size had the highest vegetative sink strength in sweet pepper. ‘Capricia’ had the lowest

490 vegetative sink strength and consequently the highest source/sink ratio during the early
491 growth stage (Figure. 7).

492

493 **Fruiting tomato plants are source-limited and source/sink ratio negatively correlates**
494 **with the potential fruit size when standard fruit load is maintained**

495

496 A major change in plant development is the switch from vegetative growth to generative
497 growth. This change was also followed by a marked change in source-sink balance in the
498 current experiment (Figure. 7B). For all three cultivars, source/sink ratio was below 1 during
499 the fully fruiting stage (Figure. 7B), suggesting source limitation. This is also supported by
500 the observation that half fruit load treatment did not influence the total plant dry weight
501 (Table 1). This result is in agreement with many previous studies that fruiting tomato plants
502 grown in greenhouses are source-limited (Cockshull and Ho, 1995; De Koning, 1994;
503 Heuvelink and Buiskool, 1995; Matsuda et al., 2011; Qian et al., 2012). Our results
504 contradicts those of Dueck et al. (2010) who estimated that cherry tomato is most likely sink-
505 limited. The source/sink ratio of fruiting tomato plants in this study (average source/sink ratio
506 was 0.17-0.33 from 50 days after planting onwards for all three cultivars) was lower than the
507 value (about 0.5) which has been reported by De Koning (1994) and Heuvelink (1996b). This
508 is mainly attributed to the low irradiance level during the fully fruiting stage (Fig. 1).
509 Furthermore, De Koning (1994) reported that tomato potential fruit growth rate positively
510 correlates with the irradiance level. In this study, the potential fruit growth rate used for sink
511 strength estimation was mainly determined from those fruits that developed under relatively
512 high irradiance level (in September and early October). This might have slightly
513 overestimated the sink strength of the plants during the low irradiance period. Additionally,
514 fruit position within a truss also plays a role, i.e. potential growth rate of the first six fruits
515 was higher than the other fruits within a truss (De Koning, 1994). In this study, the potential
516 growth rate of a single fruit was estimated from the first three fruits within a truss, therefore,
517 the sink strength of ‘Sunstream’ (10 fruits per truss) might have been overestimated. Although
518 there were several pitfalls for the estimation of sink strength in this study, the average fresh
519 weight of harvest-ripe fruits from the plants with half fruit load was 1.4, 2.2 and 2.3 times
520 higher than the fruits from plants with standard fruit load in ‘Sunstream’, ‘Capricia’ and
521 ‘Komeett’, respectively. This clearly indicates that fruiting tomato plants were source-limited
522 for all three cultivars.

523

524 During the fully fruiting stage, total fruit sink strength played a pivotal role in
525 determining the source/sink ratio, because differences in source strength and vegetative sink
526 strength between cultivars were small (Figure. 6). ‘Sunstream’ (cherry tomato) showed the
527 lowest total fruit sink strength, while ‘Komeett’ (large-sized fruits) showed the highest total
528 fruit sink strength (Figure. 6B). Hence, a negative correlation between potential fruit size and
529 source/sink ratio during the fully fruiting stage was observed when standard fruit load was
530 maintained (Figure. 7B).

531

532 Plant carbohydrate content is positively correlated with the source-sink balance (Iglesias
533 et al., 2002; Li et al., 2002; Schnyder, 1993). In line with these results a linear relationship
534 between plant source/sink ratio and total carbohydrate content in stems (Figure. 8A) as well as
535 in leaves (Figure. 8B) was observed, which relationship was independent of cultivar.
536 Carbohydrate content (i.e. sugar content in stems and starch content in leaves) during the fully
537 fruiting stage was generally lower than during the early growth stage (Figure. 4). Among the
538 three cultivars, ‘Sunstream’ showed the highest source/sink ratio and consequently the highest
539 sugar content in stems during the fully fruiting stage, while ‘Komeett’ showed the lowest

540 source/sink ratio and sugar content in stems (Figure. 4A). The positive correlation between
541 carbohydrate content in stems and source/sink ratio was also observed by Ho et al. (1983) and
542 Hall and Milthorpe (1978). In leaves, the sudden increase in starch content at 80 days after
543 planting in ‘Komeett’ and to a lesser extent at 97 days after planting in the other two cultivars
544 was unexpected as source/sink ratio was very low during this period (Figure. 7B); this
545 remains unexplained.

546

547 **IMPLICATIONS**

548

549 Fruiting tomato plants were strongly source-limited even for cherry tomato (‘Sunstream’) as
550 indicated by the low source/sink ratio (average source/sink ratio from 50 days after planting
551 onwards was 0.17-0.33 for three tomato cultivars). Despite the application of supplementary
552 lighting (162 $\mu\text{mol m}^{-2} \text{s}^{-1}$ PAR; maximum 10 hours per day), irradiance in the greenhouse
553 declined due to decreased natural irradiance towards the winter. Therefore, extending the
554 duration or increasing the PAR intensity of supplementary lighting in combination with
555 maintaining lower fruit load could be considered to better balance source and sink strength.
556 Early growth stage tomato plants showed sink limitation as indicated by a source/sink ratio
557 exceeding 1. For sink-limited plants, giving more light will not increase plant growth as
558 surplus assimilates in leaves could down-regulate leaf photosynthesis.

559

560 **CONCLUSION**

561

562 Our conclusions are: (1) tomato plants are sink-limited during the early growth stage under
563 high irradiance; (2) under commercial crop management fully fruiting tomato plants are
564 source-limited, this is even the case for small fruited cherry tomato; (3) during the fully
565 fruiting stage of tomato cultivars, the source/sink ratio is negatively correlated with the
566 potential fruit size when standard fruit load is maintained; and (4) carbohydrate content in
567 tomato stems and leaves increases linearly with the plant source/sink ratio.

568

569 **AUTHOR CONTRIBUTIONS**

570 T.L. carried out the measurements, data analysis, and drafted the manuscript. L.M and E.H
571 made substantial contributions to conception and experiment design, and critically revised the
572 manuscript.

573

574 **ABBREVIATIONS**

575

576 PAR, photosynthetic active radiation; LAI, leaf area index; SLA, specific leaf area.

577

578 **ACKNOWLEDGEMENT**

579

580 The authors would like to thank China Scholarship Council for awarding a scholarship to T.Li;
581 X. Zhao and L.F. Peng for conducting the destructive measurements, carbohydrate analysis
582 and plant development registration. We also thank Maaïke Wubs for fitting the potential fruit
583 growth curve; Menno Bakker for TOMSIM model adaptation; and Arjen van de Peppel for
584 instructing the carbohydrate analysis.

585

586 **REFERENCES**

587

588 Arp WJ, Drake BG. (1991). Increased photosynthetic capacity of *Scirpus olneyi* after 4 years
589 of exposure to elevated CO₂. *Plant, Cell & Environment*, 14: 1003-1006.

- 590 Besford RT. (1993). Photosynthetic acclimation in tomato plants grown in high CO₂. *In: CO₂*
591 *Biosphere, 104/405: 441-448. Cambridge.*
- 592 Cockshull K, Ho L. (1995). Regulation of tomato fruit size by plant density and truss thinning.
593 *Journal of Horticultural Science, 70: 395-407.*
- 594 De Koning ANM. (1994). Development and dry matter distribution in glasshouse tomato: a
595 quantitative approach. PhD thesis, Wageningen University, Wageningen, the Netherlands.
- 596 Dieleman JA, Heuvelink E. (1992). Factors affecting the number of leaves preceding the first
597 inflorescence in the tomato. *Journal of Horticultural Science, 67: 1-10.*
- 598 Dingkuhn M, Luquet D, Clément-Vidal A, Tambour L, Kim HK, Song YH. (2007). Is plant
599 growth driven by sink regulation? Implications for crop models, phenotyping approaches
600 and ideotypes. *Frontis, 21: 155-168.*
- 601 Dueck, TA., Janse, J, Schapendonk, AHCM, Kempkes, FLK, Eveleens-Clark, BA, Scheffers,
602 CP, Pot, S, Trouwborst, G, Nederhoff, EM, Marcelis, LFM. (2010b). Lichtbenutting van
603 tomaat onder LED en SON-T belichting. Wageningen: Wageningen UR
604 Glastuinbouw/Plant Dynamics BV, Rapporten GTB 1040.
- 605 Franck N, Vaast P, G énard M, Dauzat J. (2006). Soluble sugars mediate sink feedback down-
606 regulation of leaf photosynthesis in field-grown *Coffea arabica*. *Tree physiology, 26: 517-*
607 *525.*
- 608 Gifford RM, Evans LT. (1981). Photosynthesis, carbon partitioning, and yield. *Annual Review*
609 *of Plant Physiology, 32: 485-509.*
- 610 Hall AJ, Milthorpe FL. (1978). Assimilate source-sink relationships in *Capsicum annum* L.
611 III. The effects of fruit excision on photosynthesis and leaf and stem carbohydrates.
612 *Functional Plant Biology, 5: 1-13.*
- 613 Heuvelink E. (1995). Dry matter production in a tomato crop: measurements and simulation.
614 *Annals of Botany, 75: 369-379.*
- 615 Heuvelink E. (1996a). Dry matter partitioning in tomato: validation of a dynamic simulation
616 model. *Annals of Botany, 77: 71-80.*
- 617 Heuvelink E. (1996b). Tomato growth and yield: quantitative analysis and synthesis, PhD
618 thesis, Wageningen University, Wageningen, the Netherlands.
- 619 Heuvelink E, Bakker MJ, Hogendonk L, Janse J, Kaarsemaker R, Maaswinkel R. (2006).
620 Horticultural lighting in the Netherlands: new developments. *Acta Horticulturae, 711: 25-*
621 *33.*
- 622 Heuvelink E, Buiskool RPM. (1995). Influence of sink-source interaction on dry matter
623 production in tomato. *Annals of Botany, 75: 381-389.*
- 624 Heuvelink E, Marcelis LFM. (1989). Dry matter distribution in tomato and cucumber. *Acta*
625 *Horticulturae, 260: 149-179.*
- 626 Ho LC, Shaw AF, Hammond JBW, Burton KS. (1983). Source-sink relationships and carbon
627 metabolism in tomato leaves I. ¹⁴C assimilate compartmentation. *Annals of Botany, 52:*
628 *365-372.*
- 629 Hocking PJ, Steer BT. (1994). The distribution and identity of assimilates in tomato with
630 special reference to stem reserves. *Annals of Botany, 73: 315-325.*
- 631 Iglesias DJ, Lliso I, Tadeo FR, Talon M. (2002). Regulation of photosynthesis through source:
632 sink imbalance in citrus is mediated by carbohydrate content in leaves. *Physiologia*
633 *plantarum, 116: 563-572.*
- 634 Jacovides CP, Tymvios FS, Asimakopoulos DN, Theofilou KM, Pashiardes S. (2003). Global
635 photosynthetically active radiation and its relationship with global solar radiation in the
636 eastern mediterranean basin. *Theoretical and Applied Climatology, 74: 227-233.*
- 637 Li MH, Hoch G, Körner C. (2002). Source/sink removal affects mobile carbohydrates in
638 *Pinus cembra* at the Swiss treeline. *Trees, 16: 331-337.*

639 Marcelis LFM. (1996). Sink strength as a determinant of dry matter partitioning in the whole
640 plant. *Journal of Experimental Botany*,47: 1281-1291.

641 Marcelis LFM, Baan Hofman-Eijer LR. (1995). Growth analysis of sweet pepper fruits
642 (*Capsicum Annuum* L.). *Acta Horticulturae*, 412: 470-478.

643 Matsuda R, Suzuki K, Nakano A, Higashide T, Takaichi M. (2011). Responses of leaf
644 photosynthesis and plant growth to altered source–sink balance in a Japanese and a Dutch
645 tomato cultivar. *Scientia Horticulturae*,127: 520-527.

646 McCormick AJ, Cramer MD, Watt DA. (2006). Sink strength regulates photosynthesis in
647 sugarcane. *New Phytologist*,171: 759-770.

648 Moe R, Grimstad SO, Gislerod HR. (2006). The use of artificial light in year round
649 production of greenhouse crops in Norway. *Acta Horticulturae*,711:35-42.

650 Nakano H, Muramatsu S, Makino A, Mae T. (2000). Relationship between the suppression of
651 photosynthesis and starch accumulation in the pod-removed bean. *Functional Plant
652 Biology*,27: 167-173.

653 Osorio S, Ruan YL, Fernie AR. (2014). An update on source-to-sink carbon partitioning in
654 tomato. *Frontiers in Plant Science*, 5: 516.

655 Patrick JW, Colyvas K. (2014). Crop yield components -photoassimilate supply- or utilisation
656 limited-organ development? *Functional Plant Biology*, 41: 893-913.

657 Paul MJ, Foyer CH. (2001). Sink regulation of photosynthesis. *Journal of Experimental
658 Botany*,52: 1383-1400.

659 Plaut Z, Mayoral ML, Reinhold L. (1987). Effect of altered sink: source ratio on
660 photosynthetic metabolism of source leaves. *Plant Physiology*,85: 786-791.

661 Qian T, Dieleman JA, Elings A, Marcelis LFM. (2012). Leaf photosynthetic and
662 morphological responses to elevated CO₂ concentration and altered fruit number in the
663 semi-closed greenhouse. *Scientia Horticulturae*,145: 1-9.

664 Savvides A, Ntagkas N, van Ieperen W, Dieleman JA, Marcelis LFM. (2014). Impact of light
665 on leaf initiation: a matter of photosynthate availability in the apical bud? *Functional
666 Plant Biology*, 41: 547-556.

667 Schnyder H. (1993). The role of carbohydrate storage and redistribution in the source-sink
668 relations of wheat and barley during grain filling—a review. *New Phytologist*,123: 233-
669 245.

670 Scofield GN, Ruuska SA, Aoki N, Lewis DC, Tabe LM, Jenkins CLD. (2009). Starch storage
671 in the stems of wheat plants: localization and temporal changes. *Annals of Botany*,103:
672 859-868.

673 Smith AM, Stitt M. (2007). Coordination of carbon supply and plant growth. *Plant, Cell &
674 Environment*,30: 1126-1149.

675 Velez-Ramirez AI, van Ieperen W, Vreugdenhil D, van Poppel PM, Heuvelink E, Millenaar
676 FF. (2014). A single locus confers tolerance to continuous light and allows substantial
677 yield increase in tomato. *Nature communications*,5:doi:10.1038/ncomms5549.

678 Wardlaw IF. (1990). The control of carbon partitioning in plants. *New Phytologist*,116: 341-
679 381.

680 Wubs AM, Ma YT, Heuvelink E, Marcelis LFM. (2009). Genetic differences in fruit-set
681 patterns are determined by differences in fruit sink strength and a source: sink threshold
682 for fruit set. *Annals of Botany*,104: 957-964.

683 Wubs AM, Ma YT, Heuvelink E, Hemerik L, Marcelis LFM. (2012). Model selection for
684 nondestructive quantification of fruit growth in pepper. *Journal of the American Society
685 for Horticultural Science*,137: 71-79.

686 Yelle S, Beeson RC, Trudel MJ, Gosselin A. (1989). Acclimation of two tomato species to
687 high atmospheric CO₂. I. Sugar and starch concentrations. *Plant Physiology*,90: 1465-
688 1472.

689 **FIGURE LEGENDS**

690

691 Figure. 1. Daily photosynthetic active radiation (PAR) integral inside the greenhouse (sum of
692 natural irradiance and supplementary lighting) during the experiment. Line represents moving
693 average over five days.

694

695 Figure. 2. Potential growth rate of individual fruits for three tomato cultivars. Curves end at
696 the average growth duration (time from anthesis until harvest ripe) of each cultivar. Number
697 of measured fruits ranged from 34 to 48 fruits per cultivar. Potential growth was created by
698 maintaining only one fruit per truss.

699

700 Figure. 3. Measured (symbols) and estimated (lines) leaf area index (LAI) (A) and total dry
701 weight (B) over time for three tomato cultivars with standard fruit load. Error bars through
702 data points show \pm s.e. ($n = 6$). The result of two-way ANOVA with cultivar (Cv.) and days
703 after planting (D.) as independent variables and their interaction (Cv. \times D.) for each dependent
704 variable is shown in each panel. The value in the bracket indicates the least significant
705 difference at $P = 0.05$ (l.s.d). Arrow in X-axis indicates 30 days after planting. Fruit set
706 started between 20-30 days after planting for the three cultivars. Therefore, the left side of
707 arrow was defined as early growth stage, the right side of arrow was defined as fully fruiting
708 stage.

709

710 Figure. 4. Time course of the soluble sugar (A, C) and starch (B, D) concentration in the
711 stems (A, B) and leaves (C, D) of three tomato cultivars with standard fruit load. Soluble
712 sugar is the sum of glucose, fructose and sucrose. Error bars through data points show \pm s.e. (n
713 $= 6$). The result of two-way ANOVA with cultivar (Cv.) and days after planting (D.) as
714 independent variables and their interaction (Cv. \times D.) for each dependent variable is shown in
715 each panel. The value in the bracket indicates the least significant difference at $P = 0.05$ (l.s.d).
716 Arrow in X-axis indicates 30 days after planting. Fruit set started between 20-30 days after
717 planting for the three cultivars. Therefore, the left side of arrow was defined as early growth
718 stage, the right side of arrow was defined as fully fruiting stage.

719

720 Figure. 5. Time course of the net photosynthesis rate of leaf number six from top of the
721 canopy in the three tomato cultivars with standard fruit load. In the measurement chamber,
722 light intensity, CO₂ concentration, air temperature and VPD were maintained at 1000 $\mu\text{mol m}^{-2}$
723 s^{-1} , 500 $\mu\text{mol mol}^{-1}$, 23 °C and between 0.5-1 kPa. Error bars through data points show \pm s.e.
724 ($n = 12$). The result of two-way ANOVA with cultivar (Cv.) and days after planting (D.) as
725 independent variables and their interaction (Cv. \times D.) for each dependent variable is shown in
726 the figure. The value in the bracket indicates the least significant difference at $P = 0.05$ (l.s.d).
727 Arrow in X-axis indicates 30 days after planting. Fruit set started between 20-30 days after
728 planting for the three cultivars. Therefore, the left side of arrow was defined as early growth
729 stage, the right side of arrow was defined as fully fruiting stage.

730

731 Figure. 6. Estimated vegetative (A), total fruit (B), and total plant (C) sink strength over time
732 for the three tomato cultivars with standard fruit load. Lines are moving averages over five
733 days. Vegetative sink strength is the sum of the sink strengths of all the vegetative units of a
734 plant; total fruit sink strength is the sum of the sink strengths of all fruits which are present on
735 the plant; total plant sink strength is the sum of vegetative and total fruit sink strength. Arrow
736 in X-axis indicates 30 days after planting. Fruit set started between 20-30 days after planting

737 for the three cultivars. Therefore, the left side of arrow was defined as early growth stage, the
 738 right side of arrow was defined as fully fruiting stage.

739

740 Figure. 7. Estimated source strength (crop growth rate) (A) and source/sink ratio (B) over
 741 time for the three tomato cultivars with standard fruit load. Lines are moving averages over
 742 five days. Dashed horizontal line in B represents a source/sink ratio of 1. Arrow in X-axis
 743 indicates 30 days after planting. Fruit set started between 20-30 days after planting for the
 744 three cultivars. Therefore, the left side of arrow was defined as early growth stage, the right
 745 side of arrow was defined as fully fruiting stage.

746

747 Figure. 8. The relationship between total carbohydrate content (sum of soluble sugar and
 748 starch content) and plant source/sink ratio in stems (A) and leaves (B) for three tomato
 749 cultivars with standard fruit load. Lines represent linear regression line. In B, carbohydrate
 750 content determined at 81 and 97 days after planting (Fig. 4D) were not included as these data
 751 were unexpected and remain unexplained.

752

753

Table 1. Plant total dry mass and fraction of dry mass partitioned to leaves, stems and fruits of three tomato cultivars in response to fruit pruning treatment (data are collected at the end of the experiment, n = 6).

Treatment	Total dry weight (g plant ⁻¹)	Dry mass partitioning (%)		
		Leaves	Stems	Fruits
<i>'Komeett'</i>				
Standard fruit load	271.5 (±11) a	37.9 (±1.4)a	16.3 (±0.4)a	45.8 (±1.6)b
Half fruit load	275.1 (±10) a	42.3 (±0.7)b	20.2 (±0.5)b	37.5 (±1.0)a
<i>'Capricia'</i>				
Standard fruit load	278.2 (±5) a	36.3 (±1.0)a	17.3 (±0.6)a	46.4 (±1.4)b
Half fruit load	277.0 (±16) a	41.0 (±0.9)b	19.5 (±0.5)b	39.5 (±0.7)a
<i>'Sunstream'</i>				
Standard fruit load	317.3 (±10) b	45.2 (±0.5)a	20.1 (±0.4)a	34.7 (±0.8)b
Half fruit load	316.4 (±17) b	52.7 (±0.3)b	25.1 (±0.5)b	22.2 (±0.6)a

Means followed by different letters within one column of each cultivar differ significantly as established by the least significant difference (l.s.d) test at $P = 0.05$.

754

755

Figure 1.JPEG

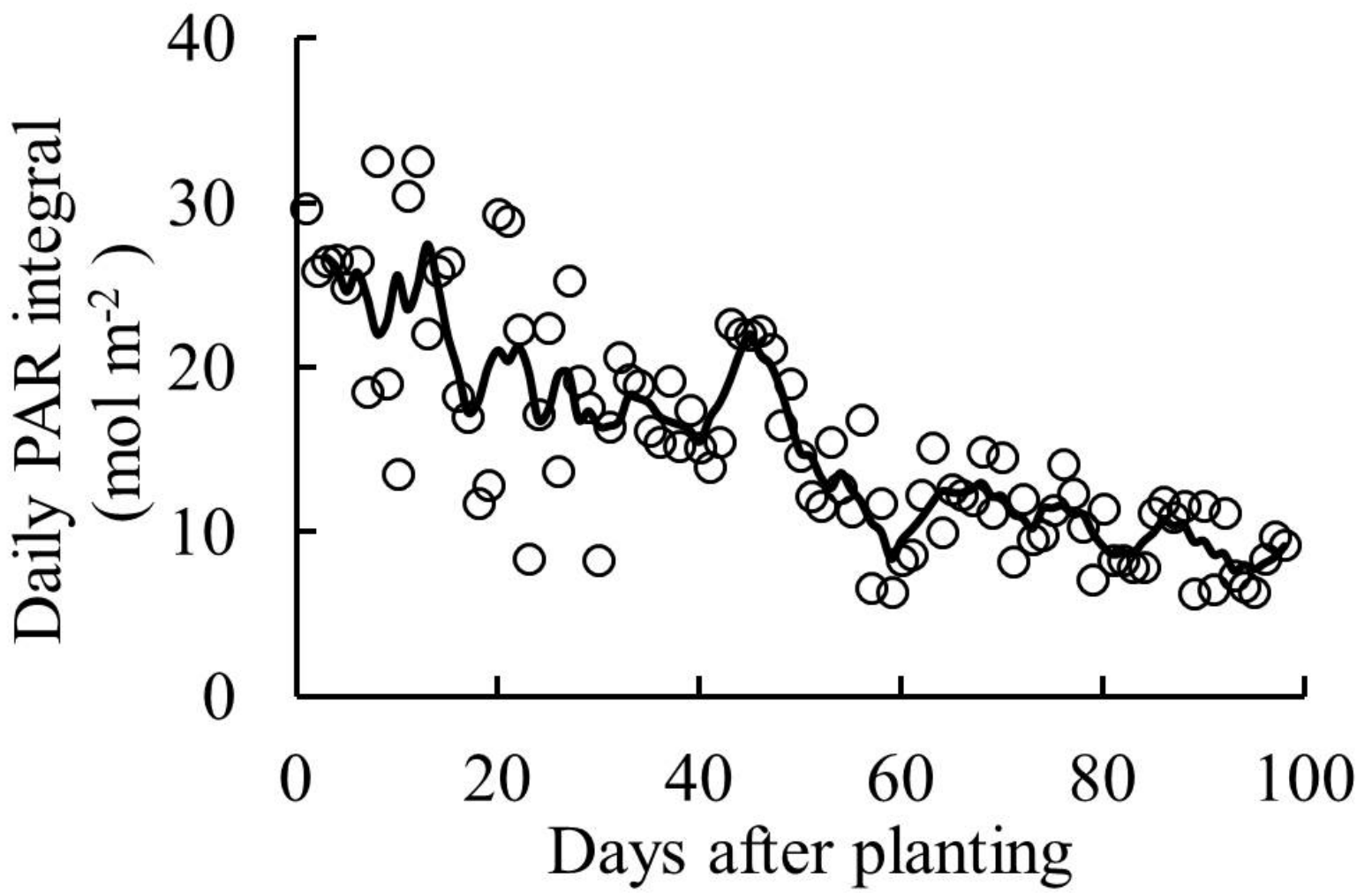


Figure 2.JPEG

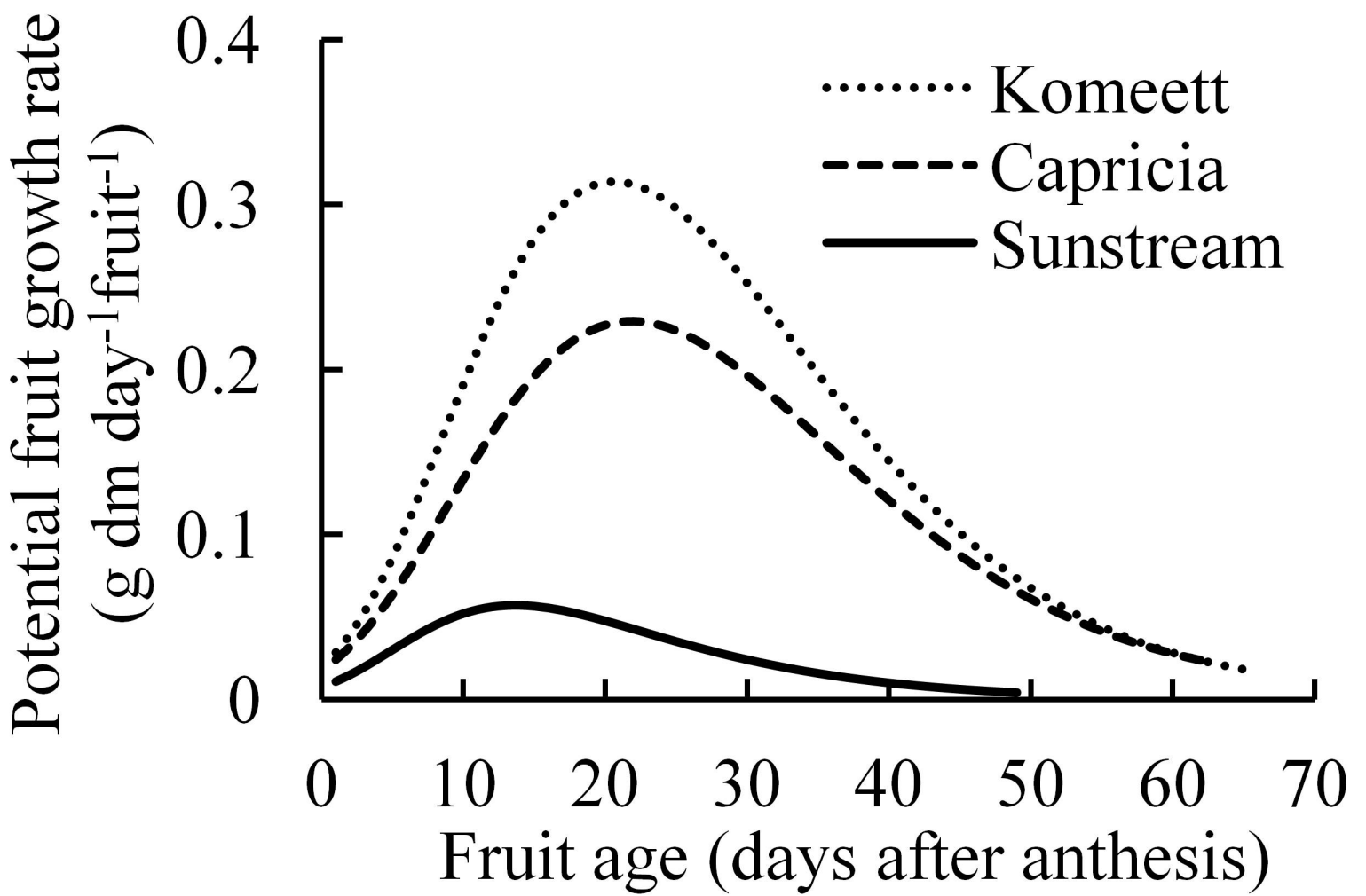


Figure 3.JPEG

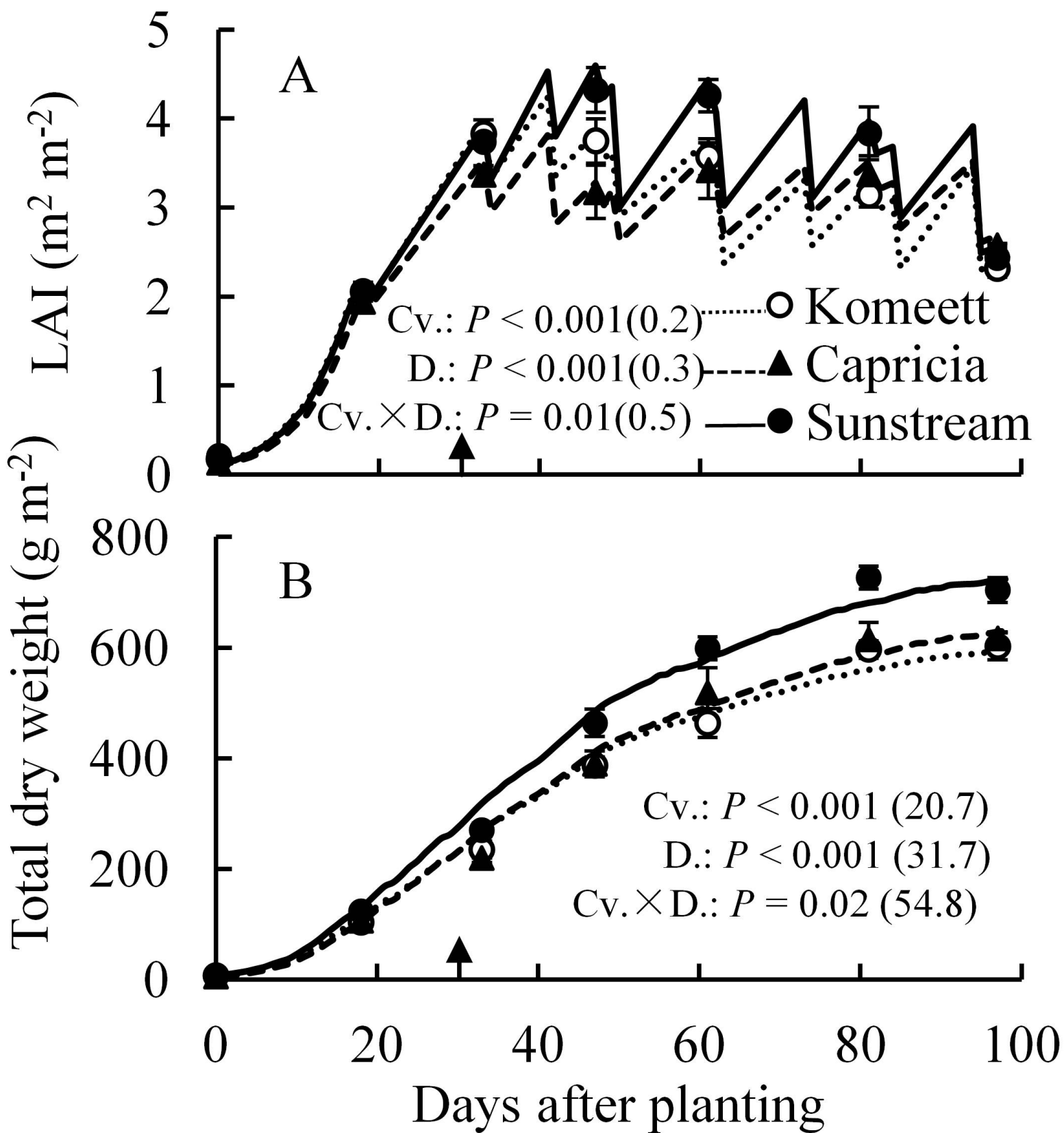


Figure 4.JPEG

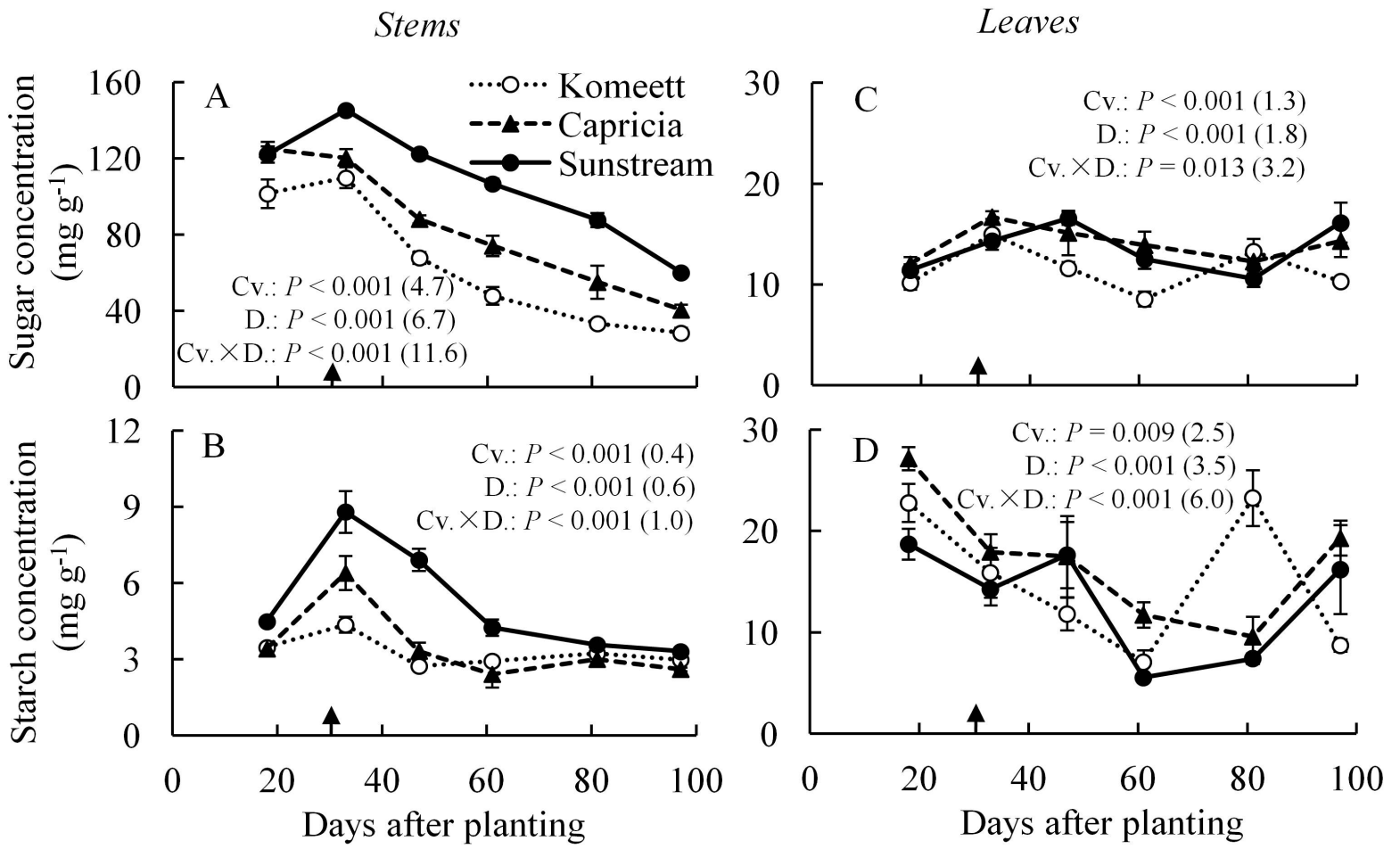


Figure 5.JPEG

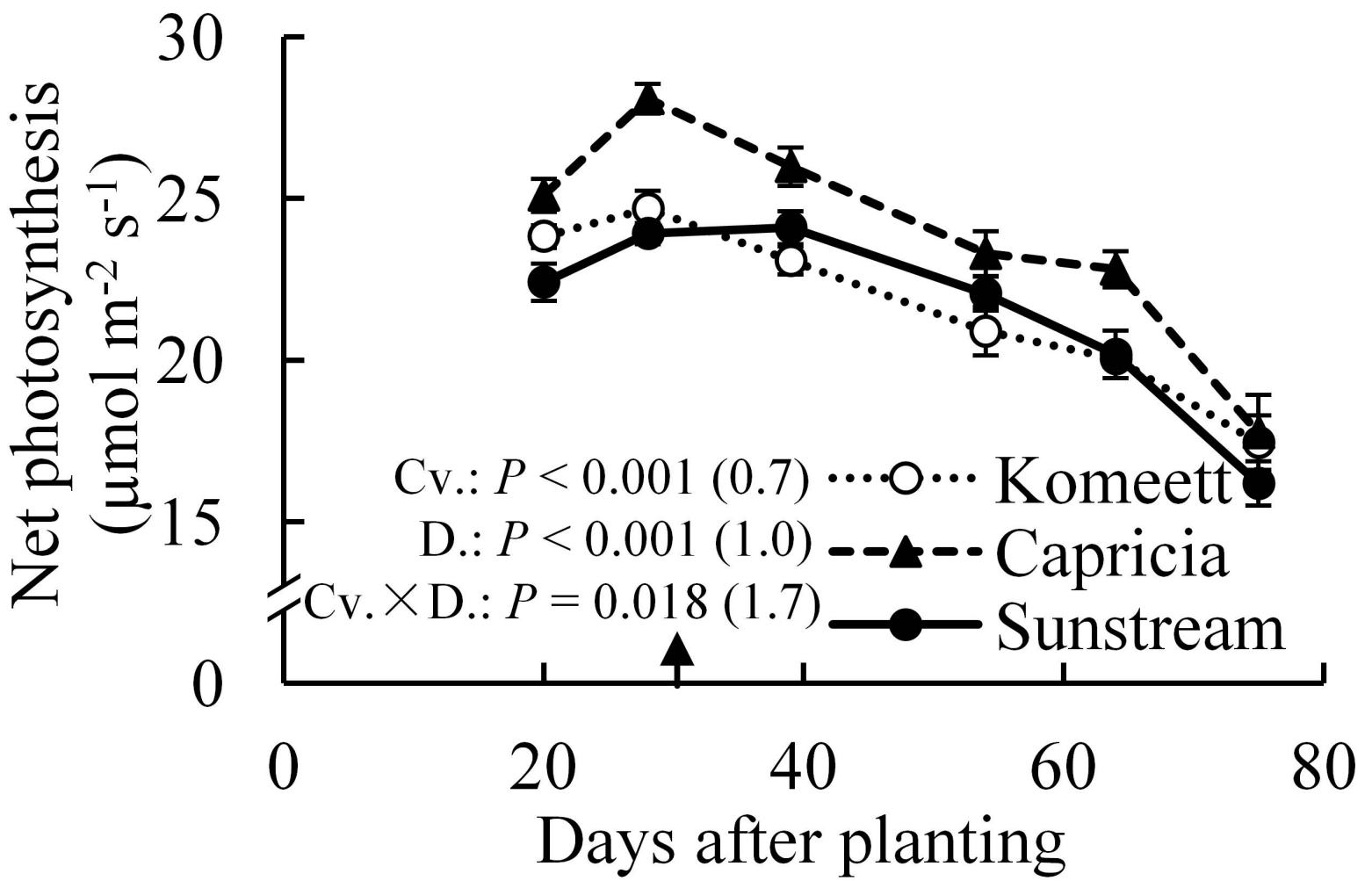


Figure 6.JPEG

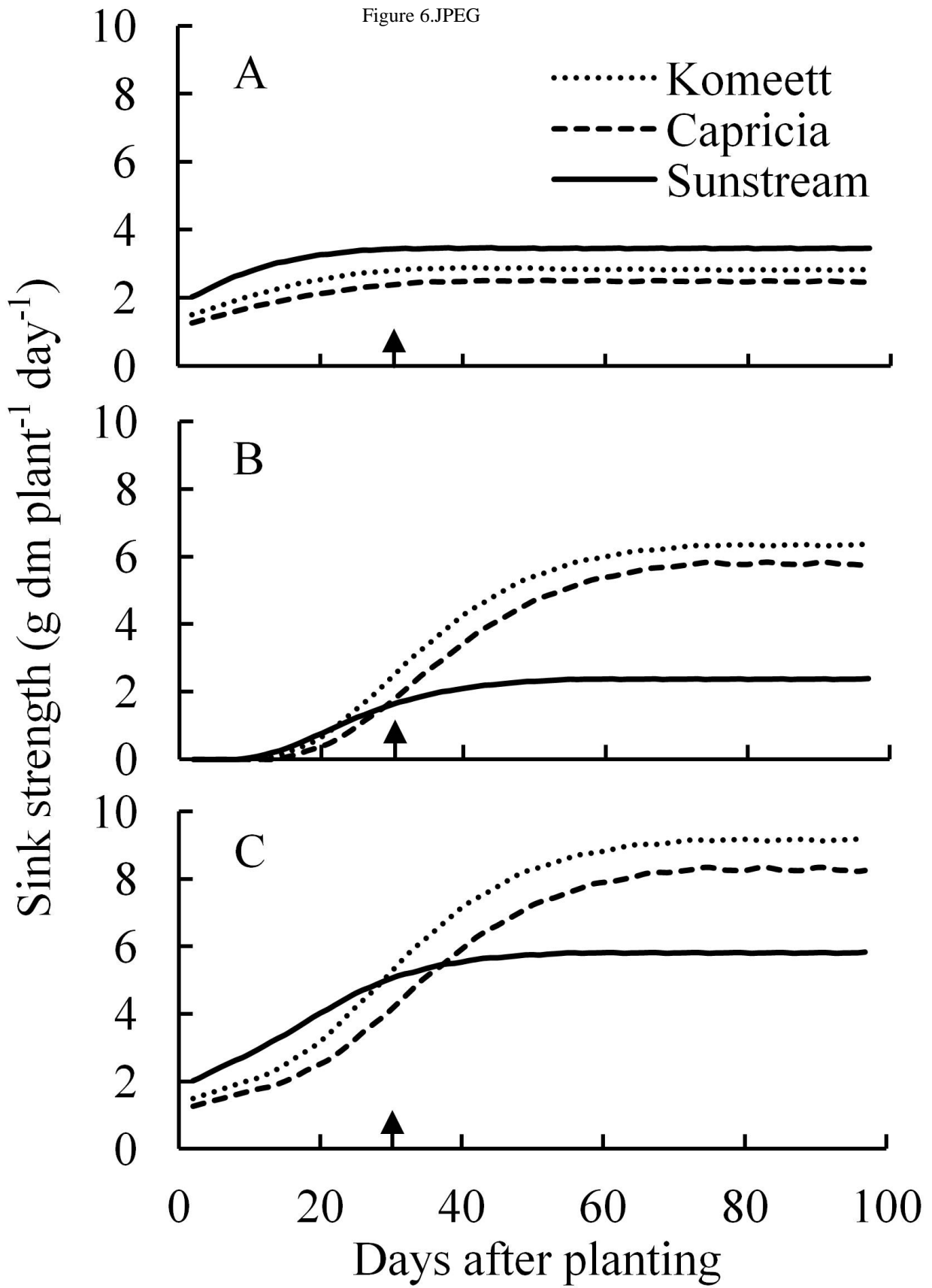


Figure 7.JPEG

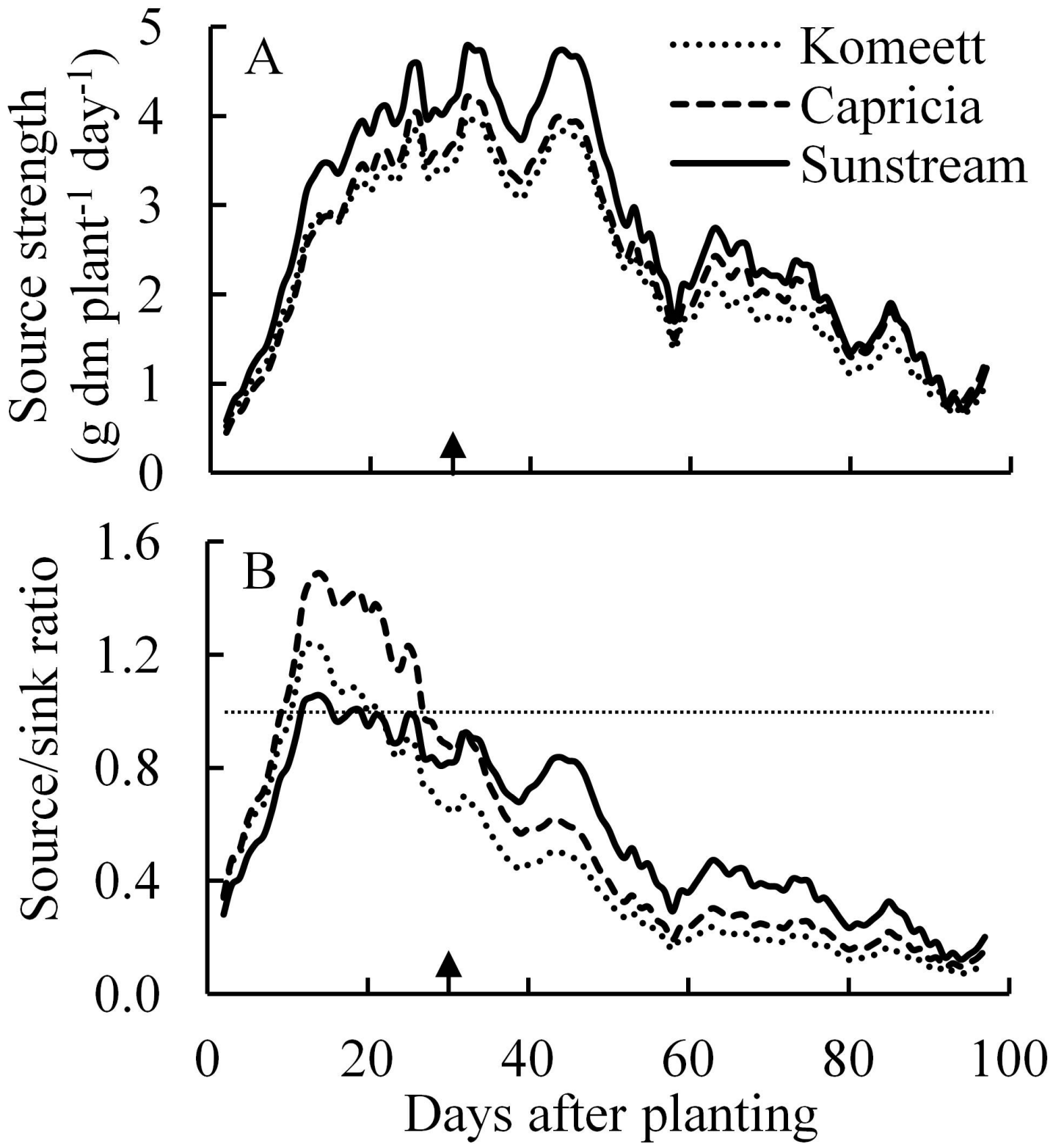


Figure 8.JPEG

