RESEARCH ARTICLE

REVISED The dipeptidyl peptidase IV inhibitors vildagliptin and K-579 inhibit a phospholipase C: a case of promiscuous scaffolds in proteins [version 3; referees: 2 approved]

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v3 **First published:** 27 Dec 2013, **2**:286 (doi: 10.12688/f1000research.2-286.v1)
Second version: 14 Jan 2015, **2**:286 (doi: 10.12688/f1000research.2-286.v2)
Latest published: 29 Jan 2015, **2**:286 (doi: 10.12688/f1000research.2-286.v3)

Abstract

The long term side effects of any newly introduced drug is a subject of intense research, and often raging controversies. One such example is the dipeptidyl peptidase-IV (DPP4) inhibitor used for treating type 2 diabetes, which is inconclusively implicated in increased susceptibility to acute pancreatitis. Previously, based on a computational analysis of the spatial and electrostatic properties of active site residues, we have demonstrated that phosphoinositide-specific phospholipase C (PI-PLC) from *Bacillus cereus* is a prolyl peptidase using *in vivo* experiments. In the current work, we first report the inhibition of the native activity of PI-PLC by two DPP4 inhibitors - vildagliptin (LAF-237) and K-579. While vildagliptin inhibited PI-PLC at micromolar concentrations, K-579 was a potent inhibitor even at nanomolar concentrations. Subsequently, we queried a comprehensive, non-redundant set of 5000 human proteins (50% similarity cutoff) with known structures using serine protease (SPASE) motifs derived from trypsin and DPP4. A pancreatic lipase and a gastric lipase are among the proteins that are identified as proteins having promiscuous SPASE scaffolds that could interact with DPP4 inhibitors. The presence of such scaffolds in human lipases is expected since they share the same catalytic mechanism with PI-PLC. However our methodology also detects other proteins, often with a completely different enzymatic mechanism, that have significantly congruent domains with the SPASE motifs. The reported elevated levels of serum lipase, although contested, could be rationalized by inhibition of lipases reported here. In an effort to further our understanding of the spatial and electrostatic basis of DPP4 inhibitors, we have also done a

comprehensive analysis of all 76 known DPP4 structures liganded to inhibitors till date. Also, the methodology presented here can be easily adopted for other drugs, and provide the first line of filtering in the identification of pathways that might be inadvertently affected due to promiscuous scaffolds in proteins.

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How to cite this article: Chakraborty S, Rendón-Ramírez A, Ásgeirsson B *et al.* **The dipeptidyl peptidase IV inhibitors vildagliptin and K-579 inhibit a phospholipase C: a case of promiscuous scaffolds in proteins [version 3; referees: 2 approved]** *F1000Research* 2015, **2**:286 (doi: 10.12688/f1000research.2-286.v3)

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Grant information: FMG thanks the Spanish Ministerio de Ciencia e Innovacion for grant No. BFU 2012-36241, and the University of the Basque Country for grant No. IT 849-13. BJ and RV acknowledge financial support from Tata Institute of Fundamental Research (Department of Atomic Energy). Additionally, BJR is thankful to the Department of Science and Technology for the JC Bose Award Grant. BA extends gratitude to the University of Iceland Research Found for supporting the project financially. AMD wishes to acknowledge grant #12-0130-SA from California Department of Food and Agriculture CDFA PD/GWSS Board. MO was supported in part by a Grant-in-aid for Scientific Research from the Ministry of Education, Culture, Sports, Science and Technology, Japan; (grant No. 21790431). The work in Liege was supported by an IUAP program funded by the Belgian federal government.

Competing interests: No competing interests were disclosed.

First published: 27 Dec 2013, **2**:286 (doi: 10.12688/f1000research.2-286.v1)

REVISED **Amendments from Version 2**

In the current version, we have changed the title, and cited previous research (ref 41 and 54) based on referee suggestions.

We have also included some minor corrections as suggested by a co-author.

See referee reports

Introduction

Oral glucose elicits a greater insulin response than intravenous glucose infusion, a phenomenon known as the incretin effect[1]. This effect is mostly attributed to the intestinally derived hormones glucagon-like peptide-1 (GLP-1) and gastric inhibitory polypeptide (GIP)[2]. These hormones have a very short half-life as they are rapidly inactivated by the ubiquitous enzyme dipeptidyl peptidase-IV (DPP4)[3]. The finding that the incretin effect is impaired in subjects with type 2 diabetes[4] led to two major types of GLP-1 based therapies[5] - intravenously or sub-cutaneously administered GLP-1 mimetics that are resistant to DPP4 (exenatide, liraglutide, etc.)[6], and the orally administered gliptins that prolong the physiological actions of incretin hormones by inhibiting DPP4 (sitagliptin, vildagliptin, etc.)[7–9]. Due to the multifarious roles played by the DPP4 enzyme[10–12], the possible side effects of these drugs (acute pancreatitis, pancreatic cancer, etc.[13–15]) are strongly contested by researchers who argue that current statistics are insufficient[16,17] to conclusively attribute these side effects to the otherwise beneficial GLP-1 drugs[18]. Compound promiscuity is another phenomenon that might play a crucial role in determining the side effects of these therapies, although this aspect has rarely been pursued intensively[19].

Previous work by our group has established the spatial and electrostatic congruence in cognate residue pairs of the active site in proteins with the same functionality (CLASP)[20,21]. CLASP analysis indicated that the phosphoinositide-specific phospholipase C (PI-PLC) from *Bacillus cereus* has spatial and electrostatic congruence with a serine protease motif[22]. This was validated by protease assays, mass spectrometry and by inhibition of the native phospholipase activity of PI-PLC by the well-known serine protease inhibitor AEBSF (IC_{50} = 0.018 mM). The specificity of the protease activity was for a proline in the amino terminal, suggesting that PI-PLC is a prolyl peptidase, similar to the DPP4 enzyme. This finding led us to believe that the gliptins would have similar inhibitory effect on PI-PLC. In the current work, we have confirmed the inhibition of the native phospholipase activity of PI-PLC using two gliptins - vildagliptin[23] (at μ-molar concentrations) and K579[24] (at nano-molar concentrations).

Subsequently, we used a motif derived from a DPP4 protein[25], in addition to the trypsin motif used previously[22], to query a comprehensive and non-redundant (50% sequence identity) list of ~5000 human proteins with known structures using CLASP, intending to identify other proteins that might be inhibited by the gliptins. From the set of proteins with significant congruent matches with these two motifs, we identified a pancreatic lipase[26] and a gastric lipase[27], keeping the context of lipases, acute pancreatitis and GLP-1 based therapies in mind. Our findings rationalize the elevated levels of serum lipase found in patients undergoing DPP4 inhibitor based therapies[28,29], although these reports are in disagreement with other findings[30,31]. While it is logical and expected to find scaffolds that are congruent to trypsin and DPP4 active sites in lipases based on the current results and our previous findings[22], we also show the presence of the serine catalytic triad in close proximity to the active site residues of proteins which have a completely different enzymatic mechanism (for example, in glutaminyl cyclase which is a transferase[32]). This corroborates the current belief that convergent evolution occurs more frequently than previously believed[33]. Thus, we propose a rational method to identify proteins that might have unintended and undesirable interactions with newly introduced compounds, and substantiate our claims by demonstrating the inhibition of the native phospholipase activity of PI-PLC from *B. cereus* using gliptins that are used in type 2 diabetes therapy.

Results

The active site motifs

The active sites of serine proteases differ in their specificities owing to residues other than the conserved catalytic triad. Thus, in addition to the trypsin motif used previously (Asp102, Ser195 and His57 - PDBid 1A0J)[22] (Motif1), we choose another motif from a DPP4 enzyme (Asp708, Ser630 and His740 - PDBid:1N1M) (Motif2) (Table 1). Apart from the catalytic triad, we chose another non-polar residue in order to increase the specificity of the matches (Ala56 in Motif1 and Val711 in Motif2). This fourth residue is chosen as the closest residue to any one of the catalytic triad residues. Using the ability of CLASP to include stereochemically equivalent residues, this last residue could be matched by another non-polar residue - one of Gly, Ala, Val, Leu, Ile or Met. Further, it has been seen that the second (ac) and fifth (bd) (Table 1) pairwise electrostatic potential differences (EPD) are not discriminatory - thus, this pair is not used to score the EPD difference (although it is included in the distance deviation score).

Inhibition of phosphoinositide-specific phospholipase C (PI-PLC) using dipeptidyl peptidase-IV (DPP4) inhibitors. DPP4 (EC 3.4.14.5), a serine protease that is expressed in many tissues (kidney, liver, lung, intestinal membranes, lymphocytes and endothelial cells), cleaves peptides with Pro or Ala residues in the second amino terminal position. Previously, we have experimentally demonstrated the existence of the serine protease domain in PI-PLC from *Bacillus cereus* - both by virtue of its proteolytic activity, and the inhibition of its native activity on phospholipids in the presence of serine protease inhibitors[22]. Furthermore, the specificity of the proteolytic activity indicated that it was a prolyl peptidase - thus, leading us to believe that DPP4 inhibitors should have a similar inhibitory effect on the PI-PLC enzyme. Table 1 shows the presence of a congruent motif in the PI-PLC protein with both Motif1 and Motif2. His32 and Asp67 are known to be a part of the active site scaffold in PI-PLC[22]. These proteins have completely different folds, and thus a superimposition (using both MUSTANG[34] and DECAAF[35]) does not show any detectable similarity in their structures (Supplementary Figure 1). Figure 1 shows the active sites of these proteins, and the superimposition of these proteins

Table 1. Potential and spatial congruence of the active site residues in proteins queried using two motifs - Motif1 from Trypsin and Motif2 from DPP4. Rmsd1 and Rmsd2 are the root mean square deviation of the scaffold with respect to Motif1 and Motif2. DPP4 - dipeptidyl peptidase-IV, PI-PLC - phosphoinositide-specific phospholipase C, PLASE - human pancreatic lipase-Related Protein 2, GPASE - human gastric lipase, QC - glutaminyl cyclase. D = Pairwise distance in Å. PD = Pairwise potential difference. APBS writes out the electrostatic potential in dimensionless units of kT/e where k is Boltzmann's constant, T is the temperature in K and e is the charge of an electron.

PDB	Active site atoms (a,b,c,d)		ab	ac	ad	bc	bd	cd	Rmsd1	Rmsd2
TRYPSIN (1A0J)	D102,S195 H57,A56	D PD	7.8 -144.1	5.6 -39.2	2.9 -248.3	3.3 104.8	9.0 -104.3	6.9 -209.1	0	0.5
DPP4 (1N1M)	D708,S630 H740,V711	D PD	7.6 -154.4	5.4 124.4	2.6 -148.8	2.6 278.8	6.8 5.6	5.4 -273.2	0.5	0
PI-PLC (1PTD)	D67,S234 H32,I68	D PD	8.2 -93.7	6.2 39.7	4.1 -245.2	3.8 133.4	11.5 -151.5	9.2 -284.8	0.6	1.1
PLASE (2OXE)	D195,S171 H282,G235	D PD	7.7 -150.2	6.4 26.7	4.4 -132.1	3.0 176.9	6.7 18.2	5.8 -158.8	0.5	0.4
GPASE (1HLG) Motif1	D324,S153, H353,L326	D PD	7.5 -202.6	5.0 -15.0	2.9 -272.3	2.7 187.6	8.4 -69.7	6.2 -257.3	0.2	0.3
GPASE (1HLG) Motif2	D324,S153 H353,A327	D PD	7.5 -202.6	5.0 -15.0	2.6 -207.1	2.7 187.6	7.1 -4.5	5.3 -192.1	0.4	0.1
QC (3PB4)	D170,S187, H168,G224	D PD	7.5 -92.8	4.8 -16.5	3.4 -214.0	3.3 76.3	10.7 -121.2	8.0 -197.5	0.4	0.8

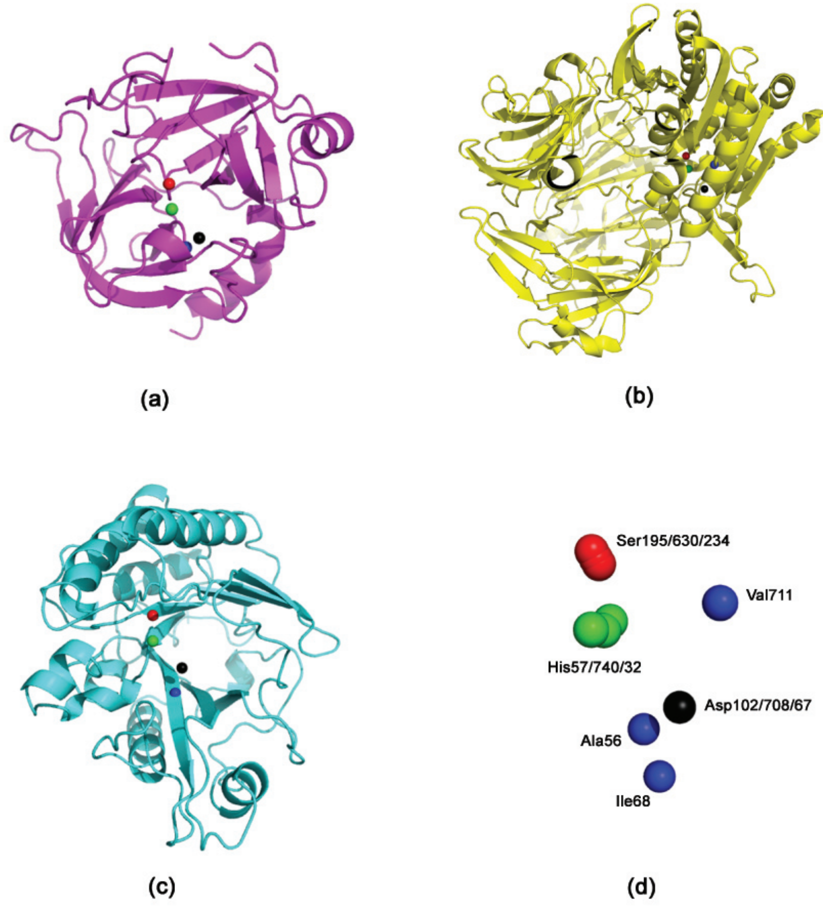


Figure 1. The active site residues in Trypsin, DPP4 and PI-PLC. (**a**) Trypsin (PDBid:1A0J) (**b**) DPP4 (PDBid:1N1M); (**c**) PI-PLC (PDBid:1PTD) (**d**) Superimposing the active site residues using DE- CAAF[35]. The superimposition can be viewed in Superimposeproteins.p1m in Dataset 1.

based on their catalytic residues[35]. It can be seen that the closest non-polar residue to the catalytictriad in trypsin and PI-PLC (Ala56 in PDBid:1A0J, Ile68 in PDBid:1PTD) is differently placed from Val711 in DPP4 (PDBid:1N1M). This is also indicated by the greater RMSD (root mean square deviation) of the scaffold in PI-PLC to Motif2 as compared to Motif1. The differences in the position of peripheral residues is the source of the diverse specificities exhibited by these proteases. Figure 2 shows the inhibition of PI-PLC using two gliptins - vildagliptin (LAF-237)[23] and K579[24]. PI-PLC catalyzes hydrolysis of phospholipids to yield diacylglycerol and a phosphoryl alcohol. In the absence of inhibitors enzyme addition to the vesicle suspension causes an increase in turbidity due to vesicle aggregation (Figure 2 a,c). Aggregation in turn occurs as a result of formation of the enzyme endproduct diacylglycerol[36,37]. A steady-state is reached under our conditions after 6–8 min. Addition of either LAF-237 (vildagliptin) or K579 leads to an obvious inhibition of the enzyme activity. Dose-response curves for the inhibitors are shown in Figure 2 (b,d). K579 is two orders of magnitude more potent than LAF-237 as a PI-PLC inhibitor, with half-maximal inhibitory concentrations IC_{50} respectively of 1 μM and 100 μM.

Phosphoinositide-specific phospholipase C inhibition data using the dipeptidyl peptidase-IV inhibitors K-579 and LAF-237

12 Data Files

http://dx.doi.org/10.6084/m9.figshare.880620

Querying a non-redundant set of human proteins using Motif1 and Motif2. Currently, the PDB database has about 25,000 human proteins. Using a identity cutoff of 50%, we chose a set of ~5000 proteins (Supplementary Table 1) as the target proteins.

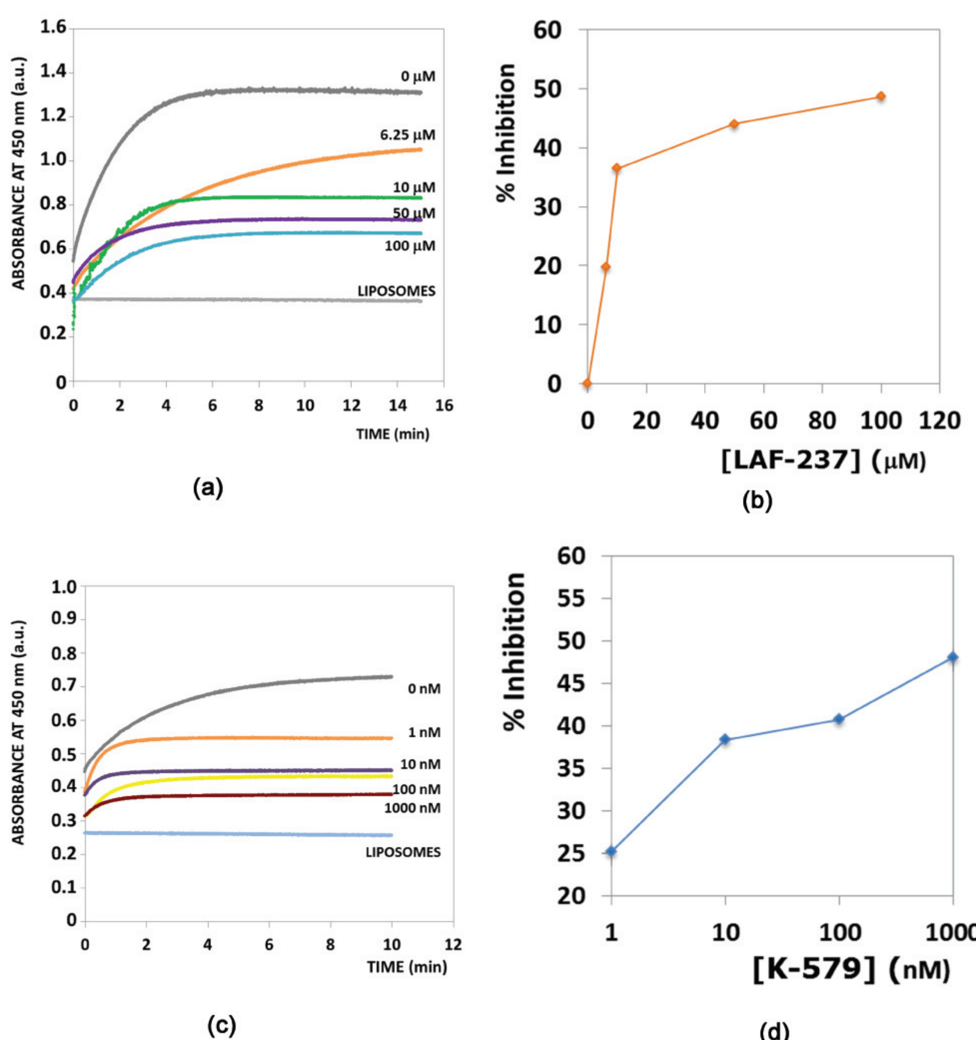


Figure 2. PI-PLC inhibition using DPP4 inhibitors. (**a**,**c**) Time courses of enzyme activity in the presence of varying amounts of inhibitors, respectively LAF-237 and K579. The trace marked LIPOSOMES corresponds to a control in the absence of PI-PLC. (**b**,**d**) Dose-response effect of inhibitors on PI-PLC activity. Activity was computed as the extent of vesicle aggregation after 10 min enzyme activity.

Table 2 shows ten proteins which have signicant matches with Motif1 and Motif2. Given the context of lipases, acute pancreatitis and GLP-1 based therapies, we picked two proteins - the human pancreatic lipase-related protein 2 (PDBid:2OXE)[26] and a human gastric lipase (PDBid:1HLG)[27] - to demonstrate the distinct possibility that these proteins might be inhibited by DPP4 inhibitors. Table 1 shows the congruence of the DPP4 motif to these proteins using Motif1 and Motif2. It is interesting to note that the gastric lipase (PDBid:1HLG) has a good match with both motifs - Leu326 in PDBid:1HLG is congruent to Ala56 in PDBid:1A0J, and Ala237 (PDBid:1HLG) is congruent to Val711 (PDBid:1N1M).

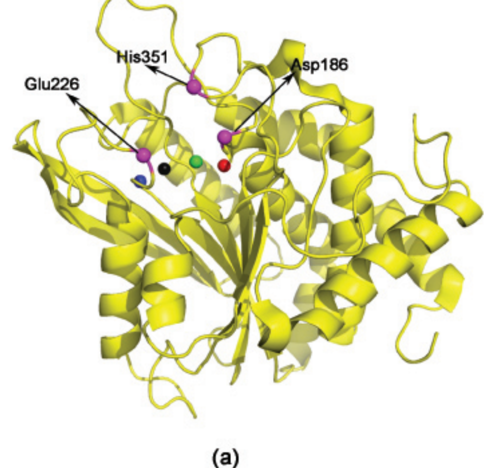
Since both these proteins are lipases (hydrolases), this congruence to Motif1 and Motif2 is expected based on our previous results with PI-PLC[22]. However, our methodology also detects other proteins, often with a completely different enzymatic mechanism from hydrolases. A glutaminyl cyclase (PDBid:3PB4[32]), a transferase, has a significantly congruent domain with Motif1 (lesser congruence with Motif2, as indicated by the RMSD) (Table 1). Figure 3 shows the proximity of the promiscuous scaffold to the active site of the cyclase, and also the congruence of the scaffold to Motif1.

Docking vildagliptin to the PIPLC structure. Since there are no DPP4 structures solved which ligand K-579, a DPP4 protein structure in complex with vildagliptin (PDBid:3W2TA)[38] was used to dock vildagliptin to the PIPLC structure complexed with myo-inositol (PDBid:1PTG[39]) using DOCLASP[40] (Figure 4). The Pymol script for visualizing the docking (SupplementaryPymol.p1m) is provided as Supplementary information.

Statistics of atoms making contact with inhibitors. There are 76 unique DPP4 inhibitors, defined by three letter codes, for which the

Table 2. Best matches in the set of ~5000 human proteins. (a) Motif1 (Asp102, Ser195, His57, Ala56) from Trypsin (b) Motif2 (Asp708, Ser630, His740, Val711) from DPP4.

Motif	PDB	Description	CLASP Score
1	2ANY	Plasma kallikrein, light chain	0.028
1	2OQ5	Transmembrane protease, serine 11E	0.037
1	3U0V	Lysophospholipase-like protein 1	0.041
1	2ODP	Complement C2	0.060
1	1IMJ	CCG1-interacting factor B	0.065
1	3F6U	Vitamin K-dependent protein C heavy chain	0.065
1	1ELV	Complement C1S component	0.068
1	1MD8	C1R complement serine protease	0.068
1	1ORF	Granzyme A	0.070
1	1FJ2	Acyl protein thioesterase 1	0.071
2	1HLG	Gastric lipase	0.042
2	1SPJ	Kallikrein 1	0.114
2	2F83	Coagulation factor XI	0.120
2	1ZJK	Mannan-binding lectin serine protease 2	0.131
2	3QLP	Thrombin light chain	0.145
2	2QXI	Kallikrein-7	0.146
2	2XU7	Histone-binding protein RBBP4	0.174
2	2W2N	Proprotein convertase subtilisin/kexin type 9	0.180
2	2HEH	KIF2C protein	0.195
2	2ANY	Plasma kallikrein, light chain	0.197



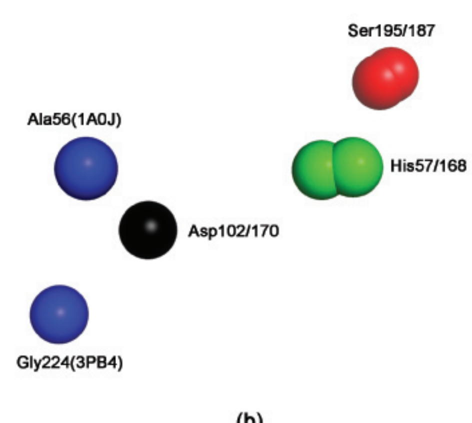


Figure 3. A scaffold congruent to the active site of Trypsin (PDBid:1A0J) in a glutaminyl cyclase (PDBid:3PB4). (**a**) The active site residues are marked in magenta. They are seen to be proximal to the identified scaffold. (**b**) Superimposition of Motif1 and the scaffold in glutaminyl cyclase. The exact pairwise interatomic distance and electrostatic potential differences are specified in Table 1.

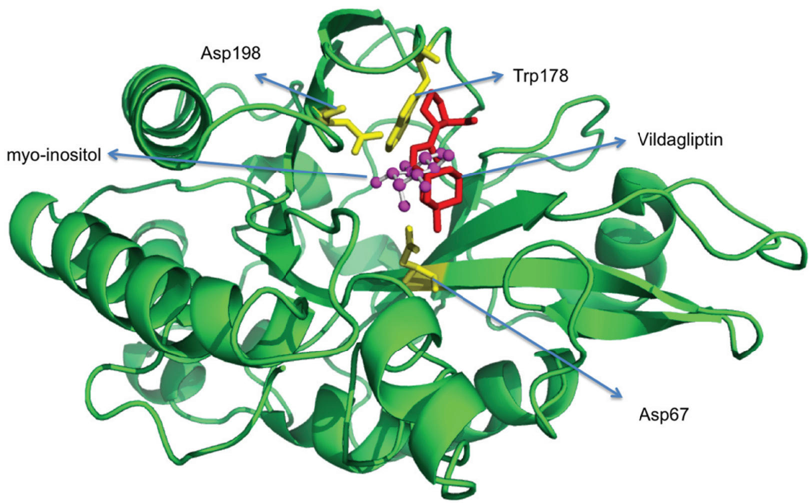


Figure 4. Docking vildagliptin to the PI-PLC structure in complex with myo-inositol (PDBid:1PTGA). Docking done using DOCLASP[40]. The Pymol script for visualizing the docking (SupplementaryPymol.p1m) is provided as Supplementary information.

ligand-DPP4 structure is solved (Supplementary Table 2). For uniformity, we chose the first four closest atoms from the protein that make contacts to the ligand, excluding hydrophobic interactions. Table 3 shows the number of times each residue in DPP4 makes contact to the ligand. Three residues are ubiquitous in making contacts in all these ligands: Glu205, Glu206 and Tyr662 made contacts in 71, 68 and 63 ligands, respectively. Interestingly, Glu205 and Glu206 have been implicated as critical residues for the enzymatic activity of DPP4 through point mutations[41]. Note, that since only the first four residues were considered, these counts are conservative (and might be more). A recent study has found that inhibitors that bind to residues beyond the extensive subsite (defined as Val207, Ser209, Phe357 and Arg358) increases DPP4 inhibition, as compared to those inhibitors that form a covalent bond with Ser630[38]. Table 3 shows that very few inhibitors make such contacts. We created a library of motifs from these structures that can be used to query any protein using CLASP to determine the possibility that DPP4 inhibitors might bind to it (Supplementary Table 3), after removing equivalent ones to eliminate redundancy. This table shows the final list of 39 motifs (pruned from the initial 76): this is a comprehensive set of motifs that encapsulates the current knowledge about protein ligand interactions for the DPP4 enzyme. A facet of ligand binding that needs to be accounted for while choosing a motif is the spatial and electrostatic changes that can be induced by ligand binding. Thus, we obtain the residues involved in binding from the holo enzyme, but extract the motif values (pairwise distance and EPD) from the apo enzyme.

Discussion

The controversy regarding the side effects of the dpp4 inhibitors, particularly with respect to acute pancreatitis and pancreatic cancer, continues unabated. While some researchers feel that it is not acceptable to assume that ‘absence of evidence is evidence of absence’[42,43], others believe that current data are not conclusive and the ‘benefits by far outweigh the potential risks’[16]. Adding to the uncertainties are conflicting reports presented by different groups[28–31]. Notwithstanding the antagonistic views on the subject, it is unanimously accepted that current data are insufficient to establish a causal pathogenic effect of these drugs on such side effects[44].

Table 3. Number of times residues from the DPP4 enzyme ligand an inhibitor. Three residues - Glu205, Glu206 and Tyr662 - make contacts in 71, 68 and 63 ligands, respectively. Note, that since we only choose the first four residues based on proximity of the atoms closest to the ligand, these counts are conservative (and might be actually more).

Residue	Number of ligands
ARG125	11
GLU205	71
GLU206	68
VAL207	1
SER209	3
ARG358	6
TYR547	18
GLN553	1
TYR585	1
TRP629	1
SER630	10
TYR631	12
TYR662	63
ASN710	15

Various database studies have been undertaken in order to ascertain the effects of the GLP-1 therapies. Some studies 'did not find an association between the use of exenatide or sitagliptin and acute pancreatitis' with the caveat that the 'limitations of this observational claims-based analysis cannot exclude the possibility of an increased risk'[45]. On the other hand, other studies have shown that the use of 'sitagliptin or exenatide increased the odds ratio for reported pancreatitis 6-fold as compared with other therapies'[14]. Further, they reported that 'pancreatic cancer was more commonly reported among patients who took sitagliptin or exenatide as compared with other therapies'[14]. Although these studies concern the usage of both GLP-1 mimetics and the orally administered gliptins, and our study exclusively focusses on gliptins, and is not concerned with the GLP-1 mimetics data. The close relationship between chronic pancreatitis and pancreatic cancer is also a subject of intense research[46]. Another administrative database study of US adults with type 2 diabetes reported increased odds of hospitalization for acute pancreatitis for patients undergoing GLP-1 based therapies sitagliptin[13]. Once again, such correlation of GLP-1 based therapies to acute pancreatitis is contested by other studies[47].

Our findings rationalize the elevated levels of serum lipase found in patients undergoing DPP4 inhibitor based therapies[28,29], keeping in mind that other studies contradict these reports[30,31]. While several studies have reported that the GLP-1 mimetics do not induce pancreatitis in rats, mouse and/or monkey[48–50], these studies did not include DPP4 inhibitors, which are the compounds that might be responsible for interactions with pancreatic proteins according to our study. It is to be noted however that these mimetics may have other physiological effects and 'the long-term consequences of sustained GLP-1 receptor activation in the human thyroid remain unknown and merit further investigation'[51]. Once again, the previous study[51] has been challenged by another group who note that 'findings previously reported in rodents may not apply to humans'[52].

The orally administered gliptins differ in many aspects such as potency, excretion mechanism, target selectivity, half-life, metabolism and possible drug-drug interactions[9,53,54]. This difference is also highlighted in the different concentrations of vildagliptin and K579 that inhibit PI-PLC. A recent study has also noted the differential off-target inhibition of enzymes by vildagliptin and sitagliptin using a high-throughput, multiplexed assay[55]. Interestingly, the PI-PLC scaffold has a better match with the trypsin motif than with the DPP4 motif (Table 1). In order to be able to model these differences in our *in silico* search, it is important to be able to provide flexibility in the scoring mechanism.

To summarize, it has been noted in the case of GLP-1 based therapies that as 'evidence of harm accumulates, but is vigorously discounted' the 'burden of proof now rests with those who wish to convince us of their safety'[43]. Surveillance programs, real-life cohort studies and case-control studies can be supplemented by rational investigations of relevant proteins based on anecdotal reports[56]. The methodology proposed in the current work, which specifically demonstrates the effects of the DPP4 inhibitors, also presents a rational way of determining the inadvertent interactions of newly designed compounds with proteins, and thus prevent the recurrence of drug induced diseases being detected after considerable damage has already been inflicted on humans subjected to these drugs[57].

Materials and methods

In silico analysis

A comprehensive, non-redundant set of ~5000 human proteins (50% identity cutoff) was obtained from the PDB database[58]. The CLASP package (http://www.sanchak.com/clasp) used for querying these proteins using motifs from trypsin and DPP4 is written in Perl on Ubuntu[20]. Hardware requirements are modest - all results here are from a simple workstation (8GB ram), and runtimes for analyzing the ~5000 proteins was about 24 hours. Adaptive Poisson-Boltzmann Solver (APBS) and PDB2PQR packages were used to calculate the potential difference between the reactive atoms of the corresponding proteins[59,60]. The APBS parameters and electrostatic potential units were set as described previously in Chakraborty *et al.*[20]. All protein structures were rendered by PyMol (http://www.pymol.org/). Protein structures have been superimposed using MUSTANG[34] and DECAAF[35].

Protein, substrate and reagents

PI-PLC was purchased from Sigma. Vildagliptin (LAF-237) was obtained from Selleckchem, and K579 was obtained from Santa Cruz.

PI-PLC assay and inhibition using DPP4 inhibitors

Vesicle preparation and characterization. The appropriate lipids were mixed in organic solution, and the solvent was evaporated to dryness under N_2. Solvent traces were removed by evacuating the lipids for at least 2 hours. The lipids were then swollen in 10 mM Hepes, 150 mM NaCl, pH 7.5 buffer. Large unilamellar vesicles (LUV) were prepared from the swollen lipids by extrusion and sized by using 0.1 μm poresize Nuclepore filters, as described by Ahyayauch *et al.*[36]. LUV composition was egg phosphatidylcholine: egg phosphatidylethanolamine: cholesterol at a 2:1:1 mole ratio. The average size of LUV was measured by quasi-elastic light scattering, using a Malvern Zeta-sizer instrument. Lipid concentration, determined by phosphate analysis, was 0.3 mM in all experiments.

Aggregation Assay. Enzyme activity was assayed measuring enzyme-induced vesicle aggregation. All assays were carried out at 39°C with continuous stirring, in 10 mM Hepes, 150 mM NaCl buffer (pH 7.5), in the presence of 0.1% BSA for optimum catalytic activity. Enzyme concentration was 0.16 U/mL, and liposomal concentration was 0.3 mM. Lipid aggregation was monitored in a Cary Varian UV-vesicle spectrometer as an increase in turbidity (absorbance at 450 nm) of the sample, as described by Villar *et al.*[37]. The data are average values of two closely similar experiments.

Analyzing known DPP4 inhibitors with solved structures. In order to obtain all known structures of DPP4 with inhibitors bound to the

active site, we did a search for the keyword dipeptidyl-peptidase on the PDB database, and choose proteins with DPP4 inhibitors as ligands. There are 76 such unique compounds (defined by three letter codes) that are reported to date (May 2014). We docked the DPP4 inhibitor to the PIPLC active site using DOCLASP[40].

Data availability

figshare: Phosphoinositide-specific phospholipase C inhibition data using the dipeptidyl peptidase-IV inhibitors K-579 and LAF-237, http://dx.doi.org/10.6084/m9.figshare.880620

Author contributions

SC, ARR and BA performed the experiments. All authors analyzed the data, and contributed equally to the writing and subsequent refinement of the manuscript.

Competing interests

No competing interests were disclosed.

Grant information

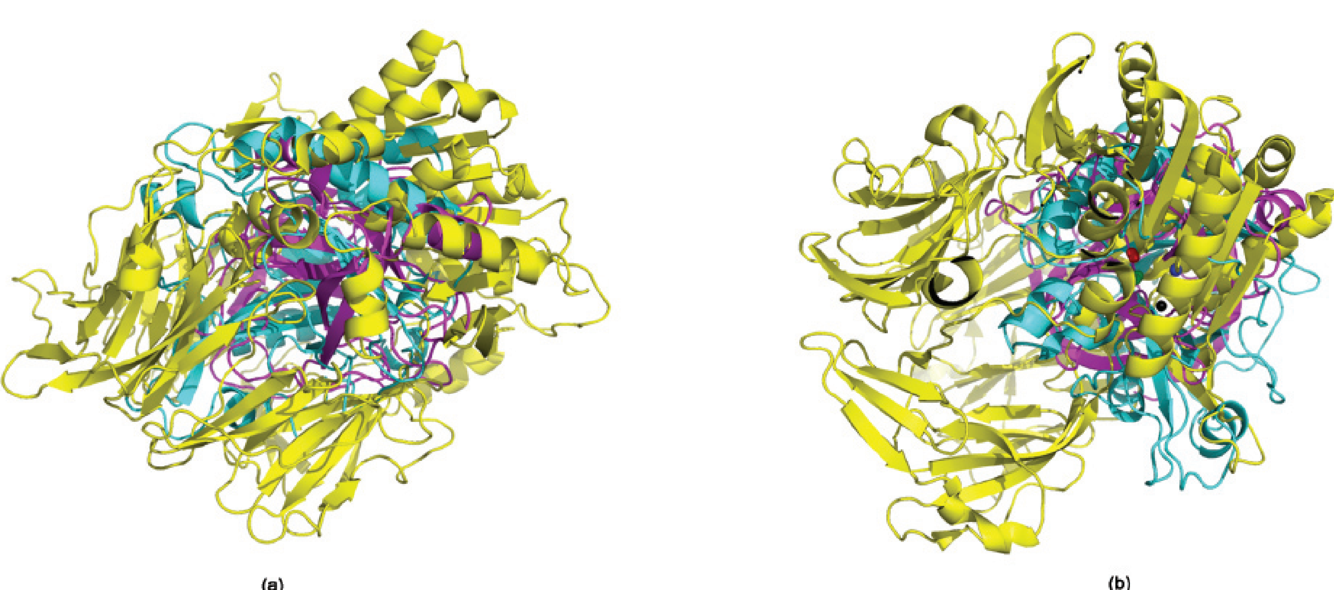
FMG thanks the Spanish Ministerio de Ciencia e Innovacion for grant No. BFU 2012-36241, and the University of the Basque Country for grant No. IT 849-13. BJ and RV acknowledge financial support from Tata Institute of Fundamental Research (Department of Atomic Energy). Additionally, BJR is thankful to the Department of Science and Technology for the JC Bose Award Grant. BA extends gratitude to the University of Iceland Research Found for supporting the project financially. AMD wishes to acknowledge grant #12-0130-SA from California Department of Food and Agriculture CDFA PD/GWSS Board. MO was supported in part by a Grant-in-aid for Scientific Research from the Ministry of Education, Culture, Sports, Science and Technology, Japan; (grant No. 21790431). The work in Liege was supported by an IUAP program funded by the Belgian federal government.

Acknowledgements

We are grateful to Jean-Marie Frère, Centre for Protein Engineering, Universite de Liège, Institut de Chimie B6, Sart Tilman, B-4000 Liège, Belgium for critical inputs.

Supplementary information

Supplementary Pymol scripts. Click here to access the files. http://dx.doi.org/10.5256/f1000research.3002.s40929



Supplementary Figure 1. Superimposition of trypsin (PDBid:1A0J - magenta), dipeptidyl peptidase-IV (PDBid:1N1M - yellow) and phosphoinositide-specific phospholipase C (PDBid:1PTD - cyan). It is seen that there is no structural similarity in the two proteins. (**a**) Using MUSTANG[34]. (**b**) Using DECAAF[35].

Supplementary Table 1. PDB IDs of ~5000 human proteins analyzed in this study.

1A02 1A12 1A17 1A1W 1A1X 1A27 1A31 1A35 1A36 1A3Q 1A4I 1A66 1A6Q 1A6Y 1A7S 1A9N 1AAP
1AB2 1AD6 1ADX 1ADZ 1AIE 1AJJ 1ALE 1ALF 1ALU 1ALY 1AM9 1AN4 1AOA 1APJ 1APQ 1APY
1AQC 1ATZ 1AU1 1AUD 1AV1 1AVO 1AW0 1AWE 1AWW 1AX8 1AYE 1B1C 1B22 1B34 1B4F 1B4G
1B4Q 1B4R 1B64 1B6A 1B6E 1B6U 1B72 1B9O 1BA5 1BAK 1BBO 1BC7 1BC8 1BCI 1BD8 1BF5 1BH7
1BH9 1BHD 1BHI 1BIK 1BJ8 1BKR 1BL1 1BNK 1BNL 1BO9 1BOE 1BOR 1BPV 1BPX 1BPY 1BPZ
1BQU 1BTK 1BTR 1BTS 1BUO 1BV8 1BW6 1BX4 1BY1 1BY2 1BY4 1BYG 1BYW 1BZ4 1BZG 1C07
1C7U 1C9B 1CB0 1CCZ 1CDW 1CF7 1CI4 1CJY 1CK7 1CKS 1CMZ 1CNT 1CQ0 1CQT 1CR8 1CS8
1CSK 1CSY 1CVJ 1CXW 1CY5 1CZA 1CZT 1CZY 1D2J 1D2L 1D2S 1D3B 1D4A 1D4B 1D4T 1D4U
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1H3O 1H4A 1H4R 1H5P 1H6G 1H6H 1H6I 1H6K 1H6P 1H7C 1H7S 1H8U 1H95 1H9D 1H9E 1H9F 1H9O
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3MQI 3MQL 3MQM 3MQP 3MR2 3MR3 3MR5 3MR6 3MSH 3MT5 3MTC 3MTR 3MTS 3MTT 3MU6
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3N01 3N2Z 3N3F 3N50 3N5N 3N6S 3N7Q 3N7S 3N8E 3N8I 3N9Y 3NA3 3NAF 3NAR 3NAU 3NBI 3NBN 3NCE 3NCL 3NCU 3NDD 3NDQ 3NER 3NEY 3NF1 3NGD 3NGO 3NGQ 3NH6 3NHC 3NHD 3NHE 3NHN 3NKB 3NKS 3NMD 3NMR 3NMW 3NMZ 3NNA 3NNC 3NNH 3NO8 3NOI 3NQJ 3NR1 3NR5 3NRX 3NSL 3NSZ 3NV1 3NVF 3NW0 3NWH 3NWN 3NWV 3NXA 3NXB 3NXP 3NXU 3NY3 3NY5 3NZL 3O0Z 3O10 3O22 3O2G 3O2T 3O36 3O3I 3O46 3O47 3O4R 3O5Q 3O6E 3O70 3O7V 3OA6 3OAD 3OB9 3OBQ 3OCP 3OD5 3OD8 3ODA 3ODC 3ODE 3ODW 3ODX 3OES 3OG6 3OG7 3OG8 3OGU 3OHU 3OLC 3OLJ 3OLL 3OMZ 3OOI 3OP3 3OP5 3OP8 3OPE 3ORH 3OSE 3OSK 3OSN 3OTC 3OU5 3OUI 3OV1 3OV6 3OVP 3OW8 3OX6 3P0C 3P0F 3P0L 3P0U 3P1A 3P1F 3P1J 3P1X 3P23 3P2T 3P3Y 3P49 3P4L 3P57 3P6D 3P6Y 3P7G 3P8C 3P8D 3PA6 3PB6 3PBH 3PC7 3PCV 3PD7 3PDF 3PDY 3PE0 3PE6 3PFF 3PFN 3PFS 3PFY 3PG6 3PG7 3PGW 3PH9 3PKI 3PLZ 3PM0 3PMI 3PML 3PMN 3PNC 3POW 3POZ 3PP2 3PPD 3PQ1 3PRY 3PS5 3PT3 3PTA 3PUA 3PUC 3PUF 3PV7 3PYC 3PZ7 3PZP 3Q01 3Q05 3Q06 3Q0H 3Q0L 3Q0M 3Q0N 3Q0O 3Q0P 3Q0Q 3Q0R 3Q0S 3Q13 3Q18 3Q1D 3Q1I 3Q2C 3Q2E 3Q2T 3Q2U 3Q6L 3Q6M 3Q6O 3Q6S 3Q6Z 3Q71 3Q72 3Q8K 3Q8L 3Q8M 3Q91 3Q93 3QCR 3QE2 3QE9 3QEA 3QEB 3QF2 3QFT 3QH9 3QI3 3QI5 3QII 3QIJ 3QIK 3QIR 3QIS 3QJ4 3QK3 3QKG 3QL9 3QLP 3QMB 3QMC 3QMD 3QMG 3QMH 3QMI 3QNT 3QO4 3QOW 3QP3 3QQN 3QRF 3QTE 3QU6 3QVE 3QWE 3QWL 3QWM 3QWP 3QWQ 3QX1 3QX3 3QXL 3QXR 3QXY 3QYE 3QYM 3QYN 3R0N 3R1H 3R1L 3R27 3R2P 3R3I 3R62 3R6B 3R6N 3R7G 3R8J 3R8Q 3R90 3R9A 3R9M 3RAU 3RAY 3RBG 3RBN 3RBS 3RC3 3RC8 3RCO 3RCP 3RCQ 3RCW 3RD2 3RDV 3RFE 3RGH 3RGK 3RH4 3RH5 3RH6 3RI4 3RIP 3RIY 3RJD 3RJE 3RJF 3RJG 3RJH 3RJI 3RJJ 3RJK 3RJO 3RK6 3RKQ 3RLE 3RLO 3RMU 3RN2 3RN5 3RNJ 3RNU 3RPP 3RPX 3RQ4 3RRQ 3RRU 3RSN 3RW6 3RY4 3RZ3 3RZG 3RZH 3RZJ 3RZK 3RZL 3RZM 3RZN 3RZV 3S24 3S4Y 3S57 3S58 3S59 3S5A 3S5J 3S5O 3S6W 3S79 3S7R 3S84 3S8I 3S8S 3S8W 3S92 3S93 3S94 3S95 3S98 3S9D 3S9G 3SAF 3SAK 3SC0 3SD6 3SEI 3SEN 3SF4 3SFJ 3SGM 3SGN 3SGO 3SGP 3SGR 3SGS 3SH4 3SHU 3SHW 3SI8 3SIU 3SIV 3SJM 3SKP 3SL9 3SM9 3SMJ 3SMQ 3SMT 3SMZ 3SNH 3SNV 3SOA 3SOC 3SOE 3SOM 3SOO 3SOV 3SP7 3SP8 3SPA 3SQD 3SR4 3SSU 3SW0 3SWK 3SWR 3SWY 3SWZ 3SYX 3SZA 3SZR 3T0H 3T0O 3T1I 3T1W 3T30 3T3L 3T5O 3T5X 3T6A 3T6P 3T7A 3T7L 3T92 3TBD 3TBG 3TC5 3TDC 3TDU 3TE3 3TEG 3TEQ 3TFR 3TFS 3TG4 3TGX 3THC 3THT 3THW 3THX 3THY 3THZ 3TIW 3TJM 3TJO 3TJQ 3TKU 3TKZ 3TLP 3TMI 3TMM 3TN2 3TNU 3TO8 3TOJ 3TOP 3TOW 3TQ1 3TQ6 3TRT 3TS8 3TSV 3TSZ 3TT0 3TT9 3TTJ 3TUO 3TV0 3TWR 3TYY 3TZD 3TZM 3U0R 3U0V 3U10 3U12 3U1K 3U1N 3U1U 3U21 3U23 3U2P 3U2U 3U3P 3U3Z 3U5L 3U5S 3U83 3U8I 3U9H 3U9J 3U9Q 3U9W 3UB2 3UBY 3UCG 3UCU 3UCW 3UCZ 3UD1 3UD3 3UD4 3UE2 3UEM 3UF1 3UFJ 3UFN 3UGC 3UI4 3UK3 3UKM 3ULH 3ULL 3UMH 3UN9 3UNN 3UO7 3UO9 3UOA 3UOB 3UOM 3UP1 3UPQ 3UQ0 3UQ2 3URF 3URO 3US0 3US1 3US2 3UUN 3UV2 3UV4 3UV5 3UVT 3UW5 3UWT 3UX2 3V2A 3V2B 3V30 3V33 3V34 3V3E 3V3L 3V42 3V43 3V4K 3V4O 3V4Q 3V53 3V56 3V70 3V79 3V8D 3V8S 3V98 3V9H 3VAF 3VAG 3VAH 3VAI 3VAJ 3VAL 3VAM 3VBB 3VD0 3VD1 3VD2 3VF3 3VFD 3VG7 3VHE 3VHS 3VHV 3VI6 3VJ9 3VKE 3VN9 3VNN 3VO3 3VOQ 3VOW 3VOY 3VPP 3VTU 3VTV 3VTW 3VVV 3VW9 3VYX 3VYY 3VZB 3W1B 3W3J 3W9Y 3ZCW 3ZD2 3ZDK 3ZI1 3ZIM 3ZJC 3ZJE 3ZN0 3ZNF 3ZNN 3ZNV 3ZON 3ZQK 3ZQS 3ZR0 3ZRH 3ZRT 3ZSJ 3ZTG 3ZVZ 3ZW5 3ZWF 3ZWT 3ZXF 3ZY0 3ZYQ 3ZYW 4A04 4A0D 4A0P 4A14 4A1G 4A1N 4A24 4A27 4A35 4A3N 4A3P 4A4F 4A4I 4A5S 4A5X 4A5Z 4A64 4A6D 4A7U 4A82 4A9C 4A9Z 4AA6 4AAA 4ABL 4ABM 4ACQ 4ACR 4AD9 4AE2 4AE7 4AE8 4AFL 4AGU 4AH6 4AIF 4AIW 4AJ5 4AJY 4AK8 4AKM 4AKV 4AL0 4ALG 4AMT 4ANK 4AOH 4AOW 4AP5 4AP8 4APO 4AQB 4AQL 4AS4 4ASC 4ASZ 4ATM 4AUV 4AVP 4AVS 4AW0 4AW6 4AWL 4AWN 4AY2 4AYA 4AYT 4AZ3 4AZ9 4B0F 4B2R 4B2S 4B3F 4B3G 4B4C 4B4O 4B53 4B5O 4B6D 4B6H 4B7L 4B7Y 4B87 4B91 4B94 4B9D 4BB9 4BBQ 4BC3 4BD2 4BDV 4BDX 4BEJ 4BGJ 4BGQ 4BHX 4BK0 4BKJ 4BKW 4BL1 4BN4 4BPB 4BQA 4BQY 4BSP 4D86 4D8K 4D8O 4D90 4DA1 4DA5 4DB1 4DBG 4DD8 4DDJ 4DDP 4DEQ 4DGJ 4DHX 4DIP 4DJC 4DK9 4DKC 4DKK 4DKX 4DL2 4DL3 4DL4 4DL5 4DL6 4DL7 4DLO 4DND 4DNL 4DO4 4DO9 4DOA 4DOB 4DOC 4DOH 4DON 4DOU 4DPZ 4DQY 4DRI 4DUR 4DVQ 4DWF 4DXT 4DY0 4DYL 4DYO 4DZO 4E1H 4E1I 4E1O 4E34 4E45 4E4H 4E54 4E5Y 4E5Z 4E6R 4E74 4E82 4E9E 4E9F 4E9G 4E9M 4EA4 4EA5 4EAR 4EBB 4EBC 4EBD 4EBE 4ECQ 4ECR 4ECS 4ECT 4ECU 4ECV 4ECW 4ECX 4ECY 4ECZ 4ED0 4ED1 4ED2 4ED3 4ED5 4ED6 4ED7 4ED8 4EEW 4EEY 4EF0 4EFO 4EGL 4EGX 4EHD 4EI1 4EI3 4EIH 4EJN 4EJQ 4EKU 4EKZ 4ELJ 4ELL 4EMO 4EMT 4ENZ 4EO7 4EOT 4EOZ 4EPU 4ERC 4ERN 4ERV 4ERY 4ES7 4ESR 4EUT 4EUU 4EUW 4EWE 4EWI 4EYH 4EYI 4EZF 4F02 4F0D 4F11 4F14 4F25 4F2J 4F3J 4F3T 4F6M 4F6N 4F6U 4F7H 4F7O 4F80 4F92 4F9C 4F9K 4F9Z 4FBN 4FC7 4FCJ 4FDI 4FGL 4FH0 4FHQ 4FIE 4FKA 4FKL 4FL3 4FLA 4FLB 4FMU 4FMW 4FNC 4FO0 4FO6 4FO9 4FOM 4FQG 4FQN 4FQP 4FR4 4FRW 4FTG 4FU3 4FU6 4FVQ 4FWW 4FXM 4FXV 4FXW 4FYO 4FYT 4FZV 4G0F 4G1M 4G1T 4G31 4G3O 4G82 4G83 4G84 4G85 4G8K 4G9A 4GA0 4GA7 4GBA 4GDK 4GDV 4GE6 4GEH 4GEI 4GGA 4GGC 4GGF 4GIF 4GIW 4GJZ 4GL2 4GLM 4GLP 4GMJ 4GMV 4GNE 4GO6 4GOF 4GOS 4GQ4 4GQB 4GQR 4GRZ 4GS4 4GT4 4GUT 4GV1 4GV2 4GWG 4GWM 4GXL 4GYW 4GYX 4H10 4H22 4H27 4H2D 4H2G 4H6Y 4H75 4H7W 4H7Y 4H87 4H9N 4HAE 4HAN 4HAS 4HBD 4HBQ 4HC4 4HC7 4HC9 4HCA 4HCK 4HCU 4HCZ 4HFX 4HL4 4HLH 4HOQ 4HOR 4HOS 4HOT 4HOU 4HPF 4HPM 4HQA 4HQU 4HQX 4HR9 4HRG 4HT2 4HTJ 4HTM 4HTP 4HVC 4HW4 4HWK 4HWN 4HXH 4HY4 4HZH 4HZR 4HZS 4I1F 4I4E 4I5I 4I5J 4I6O 4I6X 4I79 4IAX 4IC3 4IC7 4IDO 4IDT 4IE5 4IEJ 4IF8 4IG8 4IGD 4IGG 4IGZ 4II1 4IIM 4IJD 4IJX 4IKD 4IKP 4IM0 4IN0 4INC 4IQR 4IQY 4IR5 4IS1 4ITJ 4IU6 4IUL 4IVE 4IYP 4J0W 4J15 4J19 4J1Y 4J37 4J3M 4J5R 4J6G 4J8S 4J9K 4J9L 4J9M 4J9N 4J9O 4J9P 4J9Q 4J9R 4J9S 4JA8 4JGC 4JGT 4JHN 4JHS 4JIF 4JJ7 4JJH 4JK8 4JNC 4JNK 4JOI 4JOL 4JON 4JQF 4JSN 4JUY 4JV8 4JVH 4JWM 4JWN 4JXO 4K6J 4K92 4KA4 4KBL 4KFO 4KM5 4KNV 4KRF 4KSY 4KT1 4L0N 4L58 4L6E 4SKN 5ZNF 6PAX 6RLX 7ICE 7ICF 7ICG 7ICH 7ICI 7ICJ 7ICK 7ICL 7ICM 7ICN 7ICO 7ICP 7ICQ 7ICR 7ICS 7ICT 7ICU 7ICV 8ICA 8ICB 8ICC 8ICE 8ICF 8ICG 8ICH 8ICI 8ICJ 8ICK 8ICL 8ICM 8ICN 8ICO 8ICP 8ICQ 8ICR 8ICS 8ICT 8ICU 8ICV 8ICW 8ICX 8ICY 8ICZ 9ICA 9ICB 9ICC 9ICE 9ICF 9ICG 9ICH 9ICI 9ICJ 9ICK 9ICL 9ICM 9ICN 9ICO 9ICP 9ICQ 9ICR 9ICS 9ICT 9ICU 9ICV 9ICW 9ICX 9ICY

Supplementary Table 2. Residues of DPP4 closest to the bound ligand with possible hydrogen bonds. Interactions sorted based on the distance. N: Number of atoms in the ligand, R/A/LA/D: Residue number/Atom of the residue/Atom of ligand/distance between the interacting atoms (in Å). For example, 'E205/OE1/N25/2.7' means that the atom OE1 from Glu205 is at 2.7 Å from the N25 atom of W94 in PDBid:3VJLA. For uniformity, we choose the first four closest atoms. This might result in choosing some atoms which are unlikely to form a hydrogen bond (for example, in PDBid:4J3JA S209/OG is at 4.8 Å from NAQ).

PDB	HET	N	R/A/LA/D	R/A/LA/D	R/A/LA/D	R/A/LA/D
3VJLA	W94	33	E205/OE1/N25/2.7	E206/OE1/N25/2.8	N710/ND2/O33/2.9	Y662/OH/O33/3
2AJ8A	SC3	26	E205/OE2/N13/2.7	E206/OE1/N13/3	Y631/N/O23/3.1	Y547/OH/N7/3.4
2RGUA	356	35	E205/OE2/N27/3	Y662/OH/N27/3.1	Y631/N/O10/3.1	E206/OE2/N27/3.1
4A5SA	N7F	37	E205/OE2/N18/2.7	E206/OE2/N18/2.7	Y662/OH/N18/2.8	Y631/N/O26/3
2QTBA	474	32	E205/OE2/N6/2.7	N710/ND2/O7/2.8	E206/OE1/N6/2.8	R358/NE/N56/2.8
2OGZA	U1N	24	Y631/N/O25/3	E206/OE1/N12/3.1	R125/NH2/O15/3.1	E205/OE2/O15/3.3
2JIDA	GVB	24	E206/OE2/N20/2.8	E205/OE2/N20/2.9	Y662/OH/N20/3	R125/NH1/O25/3.8
2I78B	KIQ	31	Y662/OH/N/3	E205/O/O/4.1	E206/OE1/O/4.2	R669/NH2/O/4.4
3H0CA	PS4	32	E205/OE2/N21/2.7	Y662/OH/N21/2.9	E206/OE2/N21/2.9	Q553/N/O3/3
2AJLI	JNH	24	S630/OG/N3/2.3	E206/OE2/N2/2.5	Y547/OH/N3/2.6	E205/OE2/N2/2.7
2BUBA	FPB	28	E206/OE2/N18/2.5	E205/OE2/N18/2.9	Y662/OH/N18/3	Y547/OH/O16/4.3
4DSAA	D1C	29	E206/OE2/NAY/2.6	E205/O/OBC/3.1	Y662/OH/NAY/3.1	Y585/OH/NAI/3.9
2OPHA	277	23	E205/OE2/N33/2.7	N710/ND2/O32/2.8	Y662/OH/N33/3.1	E206/OE2/N33/3.1
1RWQA	5AP	27	Y662/OH/N21/2.5	E206/OE2/N21/2.8	E205/OE2/N23/2.9	R125/NH2/N1/3.4
2QJRA	PZF	29	E205/OE2/N20/2.6	R358/NE/O18/2.8	E206/OE2/N20/2.9	Y662/OH/N20/3
2FJPA	S14	31	E205/OE2/N30/2.7	N710/ND2/O32/2.8	E206/OE2/N30/2.8	Y547/OH/O33/2.8
2OAEA	AIL	21	E203/OE2/N2/2.8	E204/OE2/N2/2.8	N711/ND2/O8/3.1	Y663/OH/N9/3.1
3G0CA	RUF	27	E205/OE1/N9/3	E206/OE1/N9/3.2	Y631/N/O23/3.4	Y547/OH/N12/3.6
3C43A	315	31	E205/OE2/N6/2.8	Y662/OH/N6/3	N710/ND2/O5/3	E206/OE2/N6/3
3BJMA	BJM	23	S630/OG/N23/2.4	E205/OE2/N7/2.7	E206/OE2/N7/2.7	Y547/OH/O15/2.8
3O95A	01T	26	E206/OE2/N13/2.5	E205/OE1/N13/2.8	Y662/OH/N13/2.8	R125/NH1/O19/3
3G0GA	RUM	24	E205/OE1/N24/2.9	E206/OE1/N24/3.1	Y631/N/O8/3.2	R125/NH2/N17/3.3
2G5PA	ADF	29	S630/OG/N22/2.4	Y662/OH/N8/3.1	Y547/OH/N22/3.1	E206/OE2/N7/3.1
2BUCA	008	26	E206/OE2/N10/2.7	Y662/OH/N10/2.8	E205/OE2/N10/3	Y547/OH/O13/4.5
2QOEA	448	29	E206/OE2/N20/2.7	E205/OE2/N20/2.9	Y662/OH/N20/2.9	Y547/OH/O22/4.6
2OLEA	KR2	30	E206/OE2/NAM/2.7	Y662/OH/NAM/3.6	E205/OE2/NAM/4	Y547/OH/OAP/4.5
3KWFA	B1Q	27	E205/OE2/N21/2.7	N710/ND2/O19/2.7	Y662/OH/N21/3	R125/NH2/O19/3
3SX4A	KXA	58	Y662/OH/N25/2.7	E206/OE2/N25/2.7	E205/OE2/N25/2.8	R125/NH1/O26/3.1
2ONCA	SY1	27	E205/OE1/N1/2.6	Y631/N/O17/3.1	Y547/OH/N18/3.2	E206/OE1/N1/3.4
2I03B	AXD	29	S630/OG/N14/2.4	E206/OE1/N1/2.8	Y662/OH/O16/2.9	Y547/OH/N14/3
3KWJA	23Q	27	E205/OE2/N17/2.6	Y662/OH/N17/2.8	E206/OE2/N17/2.8	S209/OG/O19/3.3
3CCCA	7AC	21	E205/OE1/N20/2.5	Y662/OH/N20/2.7	E206/OE2/N20/3.2	Y631/N/N9/3.3
3SWWA	KXB	25	E205/OE2/N21/2.7	Y662/OH/N21/2.8	E206/OE2/N21/2.9	R125/NH2/N19/3.5
4G1FA	0WG	24	E206/OE2/N9/2.8	Y662/OH/N9/2.9	Y547/OH/N2/3.1	Y631/N/O20/3.1
3C45A	317	30	E205/OE2/N6/2.8	E206/OE2/N6/2.8	Y662/OH/N6/3	Y547/OH/N29/3.7

PDB	HET	N	R/A/LA/D	R/A/LA/D	R/A/LA/D	R/A/LA/D
2G63B	AAF	29	S630/OG/N18/2.4	E205/OE2/N7/2.6	Y662/OH/N8/3.1	Y547/OH/N18/3.1
1X70A	715	28	E206/OE2/N20/2.7	E205/OE2/N20/2.8	Y662/OH/N20/2.8	S209/OG/N27/3.9
2GBIA	XIH	29	E204/OE2/N14/2.3	Y632/N/O/2.8	E203/OE2/N14/3	Y663/OH/N14/3.1
3G0BA	T22	25	E205/OE1/N13/2.5	R125/NH2/N24/3.1	Y631/N/O26/3.2	E206/OE1/N13/3.3
2IITA	872	28	E205/OE2/N20/2.7	Y662/OH/N20/2.8	E206/OE2/N20/2.9	N710/OD1/N20/4.5
4JH0A	1MD	27	Y662/OH/N16/2.7	E206/OE2/N16/2.7	Y547/OH/O1/2.8	E205/OE2/N16/2.9
4LKOA	1WH	25	Y662/OH/N/2.7	E206/OE2/N/2.8	E205/OE2/N/2.9	Y547/OH/O2/3
2RIPA	34Q	25	N710/ND2/O1/2.6	E205/OE2/N3/2.8	Y662/OH/N3/2.8	E206/OE1/N3/2.8
3QBJA	NXZ	25	Y662/OH/N18/2.7	E206/OE2/N18/2.7	N710/ND2/O25/2.8	E205/OE2/N18/2.9
3HACA	361	23	Y662/OH/N23/2.7	E205/OE2/N23/2.8	E206/OE1/N12/4.2	N710/OD1/N23/4.3
3VJMA	W61	32	E205/OE1/N28/2.7	E206/OE1/N28/2.7	Y662/OH/O57/2.9	N710/ND2/O57/2.9
3O9VA	10T	23	Y547/OH/O15/2.5	E206/OE2/N19/2.6	Y662/OH/N19/2.7	E205/OE1/N19/2.8
4DSZA	DC3	26	E206/OE2/NAM/2.8	E205/OE2/NAM/3.1	Y662/OH/NAM/3.1	S209/OG/NAR/4.7
4J3JA	D3C	30	E206/OE2/NAM/2.8	E205/OE2/NAM/3.1	Y662/OH/NAM/3.3	S209/OG/NAQ/4.8
4DTCA	D5C	33	E206/OE2/NAM/2.7	E205/OE2/NAM/3.1	Y662/OH/NAM/3.3	R669/NH2/OAQ/4.2
3OPMA	LUI	28	E205/OE1/N18/2.7	Y662/OH/N18/2.8	E206/OE2/N18/2.9	W629/O/N27/3
2OAGB	DLI	31	E205/OE2/N22/2.5	Y662/OH/N22/2.6	E206/OE2/N22/2.9	R358/NE/O1/3.2
2GBGA	1AD	19	S631/OG/N12/2.4	E203/OE2/N14/2.7	Y548/OH/N12/3	E204/OE2/N14/3.1
2HHAA	3TP	26	E205/OE2/N6/2.7	E206/OE2/N6/2.7	Y662/OH/N6/2.9	N710/ND2/O5/2.9
2QT9A	524	31	E205/OE2/N19/2.6	E206/OE2/N19/2.8	Y662/OH/N19/2.9	N710/ND2/O20/2.9
2OQVA	MA9	32	E206/OE2/N27/2.7	Y662/OH/N27/3	R358/NE/O4/3.1	E205/OE2/N27/3.3
3F8SA	PF2	26	E205/OE2/N3/2.5	Y662/OH/O7/2.8	N710/OD1/O7/3	E206/OE2/N3/3.2
2AJBA	0QG	24	S630/OG/N2/2.4	E205/OE2/N/2.7	HIS740/NE2/O2/2.9	Y662/OH/O/3
2G5TA	ACF	26	S630/OG/N22/2.4	E205/OE2/N7/2.6	Y662/OH/O3/3	N710/ND2/O3/3
3NOXA	6A5	28	E205/OE2/N16/2.4	E206/OE2/N16/2.7	Y662/OH/N16/3	R125/NH2/N4/3.7
3W2TA	LF7	22	S630/OG/N2/2.4	E205/OE1/N12/2.8	Y662/OH/O20/3	E206/OE2/N12/3
3D4LA	605	26	E205/OE2/N15/2.6	R358/NE/O42/2.8	Y662/OH/N15/2.9	V207/O/N41/2.9
2QKYA	13Z	26	S630/OG/O2/2.1	E205/O/O4/2.5	Y547/OH/O2/2.6	E206/OE1/O4/2.8
3Q8WA	AZV	38	R125/NH1/O/2.5	E206/OE2/NAG/2.6	Y662/OH/NAG/2.8	E205/OE2/NAG/2.9
3EIOA	AJH	33	Y585/OH/OBD/2.6	E205/OE2/NBG/2.7	Y662/OH/NBG/2.9	E206/OE2/NBG/2.9
3Q0TA	LGE	26	Y662/OH/N21/2.6	E205/OE2/N21/2.7	E206/OE2/N21/2.9	R125/NH1/O22/3.4
2P8SA	417	58	E205/OE2/N38/2.8	E206/OE2/N38/2.9	Y662/OH/N38/3.2	S209/OG/N34/3.4
2OQIB	GGO	28	E205/OE2/N/2.4	Y662/OH/N/2.7	E206/OE2/N/2.9	R358/NE/O/3.2
4PNZA	2VH	28	E205/OE2/N/2.7	E206/OE2/N/2.8	Y662/OH/N/2.9	Y547/OH/O/3.2
3VJKA	M51	30	E205/OE1/N21/2.9	Y662/OH/O30/2.9	E206/OE1/N21/2.9	N710/ND2/O30/3
3OC0A	B2Q	23	E205/OE2/NS/2.8	S209/OG/OB/2.9	Y662/OH/NS/3.4	E206/OE2/NS/3.5
4N8DA	2KS	24	E206/OE2/N10/2.7	E205/OE2/N10/2.8	Y662/OH/N10/2.8	N710/OD1/N10/4.5
4N8EA	2KV	22	E205/OE2/N15/2.7	E206/OE2/N15/2.7	Y662/OH/N15/2.8	N710/OD1/N15/4.3
2IIVA	565	24	E205/OE2/N20/2.7	Y662/OH/N20/2.8	E206/OE2/N20/2.9	N710/ND2/N20/4.4
3HABA	677	27	E205/OE2/N23/2.7	Y662/OH/N23/2.7	E206/OE1/N12/4.3	N710/OD1/N23/4.4
2I3ZA	LIR	27	E203/OE2/N18/2.5	Y632/N/O9/2.7	Y548/OH/N6/3.4	E204/OE2/N18/3.4

Supplementary Table 3. Library of non-redundant motifs. This library of motifs can be used to query any protein using CLASP to determine the possibility that DPP4 inhibitors might bind to it.

PDB	Motif Name	Motif
3VJLA	2OQVA1	GLU205/OE1 GLU206/OE1 TYR662/OH ASN710/ND2
2AJ8A	2OQVA2	GLU205/OE2 GLU206/OE1 TYR547/OH TYR631/N
2RGUA	2OQVA3	GLU205/OE2 GLU206/OE2 TYR631/N TYR662/OH
2QTBA	2OQVA4	GLU205/OE2 GLU206/OE1 ARG358/NE ASN710/ND2
2OGZA	2OQVA5	ARG125/NH2 GLU205/OE2 GLU206/OE1 TYR631/N
2JIDA	2OQVA6	ARG125/NH1 GLU205/OE2 GLU206/OE2 TYR662/OH
2I78B	2OQVA7	GLU205/O GLU206/OE1 TYR662/OH ARG669/NH2
3H0CA	2OQVA8	GLU205/OE2 GLU206/OE2 GLN553/N TYR662/OH
2AJLI	2OQVA9	GLU205/OE2 GLU206/OE2 TYR547/OH SER630/OG
2BUBA	2OQVA10	GLU205/OE2 GLU206/OE2 TYR547/OH TYR662/OH
4DSAA	2OQVA11	GLU205/O GLU206/OE2 TYR585/OH TYR662/OH
2OPHA	2OQVA12	GLU205/OE2 GLU206/OE2 TYR662/OH ASN710/ND2
1RWQA	2OQVA13	ARG125/NH2 GLU205/OE2 GLU206/OE2 TYR662/OH
2QJRA	2OQVA14	GLU205/OE2 GLU206/OE2 ARG358/NE TYR662/OH
2FJPA	2OQVA15	GLU205/OE2 GLU206/OE2 TYR547/OH ASN710/ND2
2OAEA	2OQVA16	GLU205/OE2 GLU206/OE2 TYR662/OH ASN711/ND2
3G0CA	2OQVA17	GLU205/OE1 GLU206/OE1 TYR547/OH TYR631/N
3O95A	2OQVA18	ARG125/NH1 GLU205/OE1 GLU206/OE2 TYR662/OH
3G0GA	2OQVA19	ARG125/NH2 GLU205/OE1 GLU206/OE1 TYR631/N
2G5PA	2OQVA20	GLU206/OE2 TYR547/OH SER630/OG TYR662/OH
3KWFA	2OQVA21	ARG125/NH2 GLU205/OE2 TYR662/OH ASN710/ND2
2I03B	2OQVA22	GLU206/OE1 TYR547/OH SER630/OG TYR662/OH
3KWJA	2OQVA23	GLU205/OE2 GLU206/OE2 SER209/OG TYR662/OH
3CCCA	2OQVA24	GLU205/OE1 GLU206/OE2 TYR631/N TYR662/OH
4G1FA	2OQVA25	GLU206/OE2 TYR547/OH TYR631/N TYR662/OH
2G63B	2OQVA26	GLU205/OE2 TYR547/OH SER630/OG TYR662/OH
2IITA	2OQVA27	GLU205/OE2 GLU206/OE2 TYR662/OH ASN710/OD1
2RIPA	2OQVA28	GLU205/OE2 GLU206/OE1 TYR662/OH ASN710/ND2
3HACA	2OQVA29	GLU205/OE2 GLU206/OE1 TYR662/OH ASN710/OD1
3O9VA	2OQVA30	GLU205/OE1 GLU206/OE2 TYR547/OH TYR662/OH
4DTCA	2OQVA31	GLU205/OE2 GLU206/OE2 TYR662/OH ARG669/NH2
3OPMA	2OQVA32	GLU205/OE1 GLU206/OE2 TRP629/O TYR662/OH
2AJBA	2OQVA33	GLU205/OE2 SER630/OG TYR662/OH HIS740/NE2
2G5TA	2OQVA34	GLU205/OE2 SER630/OG TYR662/OH ASN710/ND2
3W2TA	2OQVA35	GLU205/OE1 GLU206/OE2 SER630/OG TYR662/OH
3D4LA	2OQVA36	GLU205/OE2 VAL207/O ARG358/NE TYR662/OH
2QKYA	2OQVA37	GLU205/O GLU206/OE1 TYR547/OH SER630/OG
3EIOA	2OQVA38	GLU205/OE2 GLU206/OE2 TYR585/OH TYR662/OH
2I3ZA	2OQVA39	GLU205/OE2 GLU206/OE2 TYR547/OH TYR631/N

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Open Peer Review

Current Referee Status:

Version 2

Referee Report 20 January 2015

doi:10.5256/f1000research.6371.r7383

Mark D Gorrell
Molecular Hepatology, Centenary Institute, Newtown, NSW, Australia

Thank you to the authors for developing this paper.

I have some further comments.

1. The primary issue now is the speculation in the title.

   The title seeks to extrapolate the obtained data on two compounds to suggest that it is applicable to all DPP-IV inhibitors.

   That is, the speculation of this paper is that the presented data is relevant to an entire drug class. The comments and the title should be restricted to one or two of the compounds that were studied in this paper. Moreover, K-579 is not a diabetes drug. In this context, the title needs changing to avoid ambiguity.

   I suggest:

   "The dipeptidyl peptidase IV inhibitor vildagliptin used in type 2 diabetes inhibits a phospholipase C: a case of promiscuous scaffolds in proteins."

   or

   "The dipeptidyl peptidase IV inhibitors vildagliptin and K-579 inhibit a phospholipase C: a case of promiscuous scaffolds in proteins."

2. This study complements the much broader work using focused, direct technology for measuring and detecting off-target inhibition. That paper is published in Nature Chemical Biology in 2014 (Bachovchin *et al.* 2014). That study similarly showed that vildagliptin inhibits enzymes other than DPP-IV. That study showed that DPP4 inhibitors differ, such that sitagliptin does not inhibit other enzymes.

   The authors need to comment and restrict their conclusions to the compounds that they studied rather than imply that DPP-IV inhibiting compounds that they did not study, such as sitagliptin, have similar characteristics to the compounds that they did study.

3. The data of this study is biochemical yet 16 of the cited references concern the safety of DPP-IV inhibition. The manuscript now carefully does not draw a link to drug safety; the title needs to do the same.

4. As the paper is focused upon DPP-IV structure and function, more papers on this topic could be cited and linked with the data. For example, the author's amendment mentions contacts in DPP-IV at Glu205, Glu206 and Tyr662. The authors could state that Glu205 and Glu206 have been shown to be essential for catalysis by DPP-IV and cite the paper Abbott *et al.* (1999).

I have read this submission. I believe that I have an appropriate level of expertise to confirm that it is of an acceptable scientific standard.

Competing Interests: This reviewer recently received a speaker honorarium from Boehringer Ingelheim, which manufactures linagliptin.

Author Response 28 Jan 2015

Sandeep Chakraborty, Tata Institute of Fundamental Research, India

We would like to thank you for your positive comments, and your informative suggestions.

We agree with your suggested change in the title. In the latest version, we have also cited the research you have brought to our attention.

Competing Interests: No competing interests were disclosed.

Version 1

Referee Report 26 March 2014

doi:10.5256/f1000research.3236.r4249

Mark D Gorrell
Molecular Hepatology, Centenary Institute, Newtown, NSW, Australia

The successful targeting of DPP4 using small molecule compounds to treat type 2 diabetes has attracted a great deal of attention towards the study of this protease.

The authors applied sophisticated techniques that they have developed in order to discover that two DPP4 inhibitors, including one that is in limited clinical use, can to some extent inhibit the activity of a bacterial lipase (PI-PLC). Many lipases and esterases and hydrolases including DPP4 and related enzymes use the *alpha/beta* hydrolase fold and the authors show how this related protein topology can place the residues in positions that are sufficiently similar to interact with an inhibitor.

The major difficulty with this paper is that it attempts to connect these data with possible clinical outcomes. No evidence for such a link is presented. Therefore, the title and much of the conclusions need to be modified so that they reflect the data without speculation.

Two inhibitors of DPP4, LAF237 and K-579, were studied. K-579 is not in clinical use. LAF237 is licensed in Europe and is known to exhibit some inhibition of the DPP4-related proteases DPP8 and DPP9. The extent of inhibition of DPP8 and DPP9 by LAF237 is believed not to have physiological effects in humans. The IC50 of LAF237 on DPP9 is less than 0.01 mM. The IC50 of LAF237 on bacterial PI-PLC is 0.1mM, which is close to the lower limit of detection of inhibition of an enzyme. No mammalian homolog of PI-PLC was examined.

The literature that the authors cite to suggest that DPP4 inhibition might be detrimental for human health, particularly the pancreas, is data on sitagliptin or exenatide. Exenatide is not a DPP4 inhibitor and sitagliptin is quite different to LAF237, both in protease specificity and in chemical structure. The contact points of LAF237 and sitagliptin in the catalytic site of DPP4 differ considerably. The authors present no data on sitagliptin or any other DPP4 inhibitor (other than LAF237) that in is the clinic.

The images of overlaid catalytic triads of various enzymes presented in Fig 1 and Fig 3 need to be depicted in 3D in order to evaluate how close they are in 3D. Intermolecular distances should be shown on thee figures. To convince the reader that LAF237 sits into and makes contacts with enzymes other than DPP4, we need to see the compound docked into the structure of each enzyme of interest.

I have read this submission. I believe that I have an appropriate level of expertise to confirm that it is of an acceptable scientific standard, however I have significant reservations, as outlined above.

Competing Interests: No competing interests were disclosed.

Author Response 10 Dec 2014

Sandeep Chakraborty, Tata Institute of Fundamental Research, India

We would like to thank you for taking the time to review this paper, and also for your insightful comments. We also apologize for the inordinate time taken to respond to your comments. A lot of this time was spent in understanding docking methods, instead of blindly applying this to the problem at hand. A by-product of this learning process was the implementation of a new method (DOCLASP) for docking molecules to proteins[1]. We have docked vildagliptin to the PI-PLC structure complexed with myo-inositol using DOCLASP. Based on your suggestion, we have also done a comprehensive analysis of all 76 known DPP4 structures liganded to inhibitors till date.

Please find out detailed responses to your comments below.

- *The successful targeting of DPP4 using small molecule compounds to treat type 2 diabetes has attracted a great deal of attention towards the study of this protease. The authors applied sophisticated techniques that they have developed in order to discover that two DPP4 inhibitors, including one that is in limited clinical use, can to some extent inhibit the activity of a bacterial lipase (PI-PLC). Many lipases and esterases and hydrolases including DPP4 and related enzymes use the alpha/beta hydrolase fold and the authors show how this related protein topology can place the residues in positions that are sufficiently similar to interact with an inhibitor. The major difficulty with this paper is that it attempts to connect these data with possible clinical outcomes. No evidence for such a link is presented. Therefore, the title and much of the conclusions need to be modified so that they reflect the data without speculation.*

We have tried to keep away from taking sides on the clinical outcomes, since that is not our forte. Also, we believe our title is innocuous in that context - it just speaks of promiscuous scaffolds. We only highlight that if (and only if) our data of PIPLC inhibition holds true for human lipases, then it might provide some arguing points for those worried about the side effects of these drugs.

For example, we say ‘The reported elevated levels of serum lipase, although contested, could be rationalized by inhibition of lipases reported here’. If you could kindly point out specifically any speculations that is unwarranted, we will modify those.

- *Two inhibitors of DPP4, LAF237 and K-579, were studied. K-579 is not in clinical use. LAF237 is licensed in Europe and is known to exhibit some inhibition of the DPP4-related proteases DPP8 and DPP9. The extent of inhibition of DPP8 and DPP9 by LAF237 is believed not to have physiological effects in humans.*

  Since this study does not emphasize on the clinical relevance of the inhibitions (but on the methodology of finding such interactions), and we are not a group specializing in diabetes, we believe the choice of the inhibitors would not alter our reasoning our conclusions.

- *The IC50 of LAF237 on DPP9 is less than 0.01 mM. The IC50 of LAF237 on bacterial PI-PLC is 0.1mM, which is close to the lower limit of detection of inhibition of an enzyme.*

  We agree to this point. However, K-579 was inhibiting even at nanomolar concentrations.

- *No mammalian homolog of PI-PLC was examined.*

  We are currently evaluating that possibility.

- *The literature that the authors cite to suggest that DPP4 inhibition might be detrimental for human health, particularly the pancreas, is data on sitagliptin or exenatide. Exenatide is not a DPP4 inhibitor and sitagliptin is quite different to LAF237, both in protease specificity and in chemical structure.*

  We were referring to the inhibitor part of the data, but that point needs to be made explicit as you have correctly pointed out. Also, we agree that the possible difference of sitagliptin with LAF237 needs to be stated. We have modified the text to include these criticisms. Once again, we reiterate we intend not to comment on clinical outcomes or debates, but to suggest a rational methodology to act as a guide for tests that look for possible interactions.

- *The contact points of LAF237 and sitagliptin in the catalytic site of DPP4 differ considerably. The authors present no data on sitagliptin or any other DPP4 inhibitor (other than LAF237) that in is the clinic.*

  We have included a comprehensive study on the contact points of various inhibitors. Once again, this does not negate any of our conclusions.

- *The images of overlaid catalytic triads of various enzymes presented in Fig 1 and Fig 3 need to be depicted in 3D in order to evaluate how close they are in 3D.*

The 3D images of the superimposition of these enzymes are not pleasing to the eye, since they lack structural homology. However, we have added a PyMol script in case someone wishes to do that (Superimposeproteins.p1m). The script specifies the color coding of the residues.

- *Intermolecular distances should be shown on thee figures.*

  Once again, we think that the intermolecular distances clutter the figure. The superimposition gives an approximate idea of the congruence. The exact values are specified in Table 1. We have modified the legend of Fig.3 to specify that.

- *To convince the reader that LAF237 sits into and makes contacts with enzymes other than DPP4, we need to see the compound docked into the structure of each enzyme of interest.*

  As mentioned previously, we have docked sitagliptin to PI-PLC using DOCLASP[1]. We have provided the Pymol script as supplementary data to help visualize the docking. There is no solved structure where LAF237 inhibits DPP4.

Once again, we are thankful for the comments. We hope that we have addressed your concerns by the changes that we have made, and that the manuscript will be found suitable in the modified form.

References

1. Chakraborty S. *DOCLASP* - Docking ligands to target proteins using spatial and electrostatic congruence extracted from a known holoenzyme and applying simple geometrical transformations [v2; ref status: awaiting peer review, http://f1000r.es/4pb] *F1000Research* 2014, **3**:262 (doi: 10.12688/f1000research.5145.2)

Competing Interests: No competing interests were disclosed.

Referee Report 06 March 2014

doi:10.5256/f1000research.3236.r3818

Rodney Rouse

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Disclaimer: I lack the protein chemistry expertise to comment on the assumptions and protein chemistry used in the computational method described in this article.

The title and abstract are appropriate.The overall experimental design is simple but strong and well suited for this project. The methods were generally well described. The conclusions are not overstated and any implications are justified based on the presented data. The article is very well written.

This is a very interesting study that uses a previously defined computational method, CataLytic Active Site Prediction (CLASP), that compares structural and charge similarities of catalytic sites to identify

functionally similar proteins. This methodology was used to assess the potential for adverse events based on off target effects of the inhibitors of DPP-IV. Using CLASP, the authors had previously indentified a Bacillus cerus phosphoinositide specific phospholipase-C (PI-PLC) as similar in active catalytic site to the enzyme, DPP-IV. They used laboratory techniques to verify this finding.

In the present study, the authors demonstrated the ability of two separate DPP-IV inhibitors to significantly reduce the activity of this PI-PLC in the lab. Subsequent to this experimentation, the authors returned CLASP to identify catalytic sites in other proteins that might also be inhibited by DPP-IV inhibitors thereby yielding unforeseen inhibition and biological effects. As applied to the case of DPP-IV inhibitors, which are not extremely specific, the authors identify a number of other proteins that could be promiscuously impacted by DPP-IV inhibitors thereby providing mechanisms for unexpected adverse events. Although the significance of DPP-IV inhibitor related adverse events has yet to be determined, the fact that changes have been reported non-clinically and clinically are undeniable. Eventually, the benefit of these molecules may far outweigh their associated risks, but the authors provide a potential path forward for investigation of unexpected events with this class of drug. If contradictory reports persist, this path may require further illumination.

The approach is theoretically similar to using structural similarities to identify off target receptor binding and consequent biological effects, an expanding approach in safety assessment and in identification of mechanisms for adverse events in the pharmaceutical lifecycle. Similarly, this method could be predictive for off target effects and suggest what those effects might be. However, whether this is a method that can be generally applicable to other molecules is beyond my ability to comment and the scope of this work.

Comments/Suggestions:

1) Were the inhibition experiments done in duplicate, triplicate, etc? Some slight expansion of the protocols would help with attempts to replicate.

I have read this submission. I believe that I have an appropriate level of expertise to confirm that it is of an acceptable scientific standard.

Competing Interests: No competing interests were disclosed.

Author Response 10 Mar 2014

Adela Rendón-Ramirez, Unidad de Biofisica CSIC UPV, Spain

Dear Dr Rouse,

We would like to thank you for taking the time and reviewing our paper. Your positive comments encourage us to further our research in this area.

We concur with your statement - *"Eventually, the benefit of these molecules may far outweigh their associated risks"*. And it is our endeavor to improve the accuracy and generality of our method through different compounds. We would specifically like to highlight another case of antagonist binding identified through CLASP, although in this case most alkaline phosphatases were not affected - Chakraborty *et al.*(2012)

The data for PI-PLC inhibition using DPP4 inhibitors, as shown in Figure 2, are average values of two closely similar experiments. We will revise the manuscript to include this point when we hear

from another referee.

Best regards,

Sandeep Chakraborty and Adela Rendón-Ramirez

Competing Interests: No competing interests were disclosed.