

PERSPECTIVE

Receiver Operating Characteristic (ROC) Curve for Medical Researchers

RAJEEV KUMAR AND ABHAYA INDRAYAN

*From the Department of Biostatistics and Medical Informatics, University College of Medical Sciences, Delhi, India.
Correspondence to: Mr Rajeev Kumar, Department of Biostatistics and Medical Informatics, University College of Medical Sciences, Delhi 110 095. Rajeev.kumar.malhotra@gmail.com*

Sensitivity and specificity are two components that measure the inherent validity of a diagnostic test for dichotomous outcomes against a gold standard. Receiver operating characteristic (ROC) curve is the plot that depicts the trade-off between the sensitivity and (1-specificity) across a series of cut-off points when the diagnostic test is continuous or on ordinal scale (minimum 5 categories). This is an effective method for assessing the performance of a diagnostic test. The aim of this article is to provide basic conceptual framework and interpretation of ROC analysis to help medical researchers to use it effectively. ROC curve and its important components like area under the curve, sensitivity at specified specificity and *vice versa*, and partial area under the curve are discussed. Various other issues such as choice between parametric and non-parametric methods, biases that affect the performance of a diagnostic test, sample size for estimating the sensitivity, specificity, and area under ROC curve, and details of commonly used softwares in ROC analysis are also presented.

Key words: *Sensitivity, Specificity, Receiver operating characteristic curve, Sample size, Optimal cut-off point, Partial area under the curve.*

Diagnostic tests play a vital role in modern medicine not only for confirming the presence of disease but also to rule out the disease in individual patient. Diagnostic tests with two outcome categories such as a positive test (+) and negative test (–) are known as dichotomous, whereas those with more than two categories such as positive, indeterminate and negative are called polytomous tests. The validity of a dichotomous test compared with the gold standard is determined by sensitivity and specificity. These two are components that measure the inherent validity of a test.

A test is called continuous when it yields numeric values such as bilirubin level and nominal when it yields categories such as Mantoux test. Sensitivity and specificity can be calculated in both cases but ROC curve is applicable only for continuous or ordinal test.

When the response of a diagnostic test is continuous or on ordinal scale (minimum 5

categories), sensitivity and specificity can be computed across all possible threshold values. Sensitivity is inversely related with specificity in the sense that sensitivity increases as specificity decreases across various threshold. The receiver operating characteristic (ROC) curve is the plot that displays the full picture of trade-off between the sensitivity and (1-specificity) across a series of cut-off points. Area under the ROC curve is considered as an effective measure of inherent validity of a diagnostic test. This curve is useful in (i) finding optimal cut-off point to least misclassify diseased or non-diseased subjects, (ii) evaluating the discriminatory ability of a test to correctly pick diseased and non-diseased subjects; (iii) comparing the efficacy of two or more tests for assessing the same disease; and (iv) comparing two or more observers measuring the same test (inter-observer variability).

INTRODUCTION

This article provides simple introduction to measures of validity: sensitivity, specificity, and area under

ROC curve, with their meaning and interpretation. Some popular procedures to find optimal threshold point, possible bias that can affect the ROC analysis, sample size required for estimating sensitivity, specificity and area under ROC curve, and finally commonly used statistical softwares for ROC analysis and their specifications are also discussed.

PubMed search of pediatric journals reveals that ROC curve is extensively used for clinical decisions. For example, it was used for determining the validity of biomarkers such as serum creatine kinase muscle-brain fraction and lactate dehydrogenase (LDH) for diagnosis of the perinatal asphyxia in symptomatic neonates delivered non-institutionally where area under the ROC curve for serum creatine kinase muscle-brain fraction recorded at 8 hours was 0.82 (95% CI 0.69-0.94) and cut-off point above 92.6 U/L was found best to classify the subjects. The area under ROC curve for LDH at 72 hours was 0.99 (95% CI 0.99-1.00) and cut-off point above 580 U/L was found optimal for classifying the perinatal asphyxia in symptomatic neonates [1]. It has also been similarly used for parameters such as mid-arm circumference at birth for detection of low birth weight [2], and first day total serum bilirubin value to predict the subsequent hyperbilirubinemia [3]. It is also used for evaluating model accuracy and validation such as death and survival in children or neonates admitted in the PICU based on the child characteristics [4], and for comparing predictability of mortality in extreme preterm neonates by birth-weight with predictability by gestational age and with clinical risk index of babies score [5].

SENSITIVITY AND SPECIFICITY

Two popular indicators of inherent statistical validity of a medical test are the probabilities of detecting

correct diagnosis by test among the true diseased subjects (D+) and true non-diseased subjects (D-). For dichotomous response, the results in terms of test positive (T+) or test negative (T-) can be summarized in a 2x2 contingency table (**Table I**). The columns represent the dichotomous categories of true diseased status and rows represent the test results. True status is assessed by gold standard. This standard may be another but more expensive diagnostic method or a combination of tests or may be available from the clinical follow-up, surgical verification, biopsy, autopsy, or by panel of experts. Sensitivity or true positive rate (TPR) is conditional probability of correctly identifying the diseased

subjects by test: $S_N = P(T+/D+) = \frac{TP}{TP+FN}$; and specificity or true negative rate (TNR) is conditional probability of correctly identifying the non-disease subjects by test: $S_p = P(T-/D-) = \frac{TN}{TN+FP}$. False positive rate (FPR) and false negative rate (FNR) are the two other common terms, which are conditional probability of positive test in non-diseased subjects: $P(T+/D-) = \frac{FP}{FP+TN}$; and conditional probability of negative test in diseased subjects: $P(T-/D+) = \frac{FN}{TP+FN}$, respectively.

Calculation of sensitivity and specificity of various values of mid-arm circumference (cm) for detecting low birth weight on the basis of a hypothetical data are given in **Table II** as an illustration. The same data have been used later to draw a ROC curve.

ROC CURVE

ROC curve is graphical display of sensitivity (TPR) on y-axis and (1 – specificity) (FPR) on x-axis for varying cut-off points of test values. This is

TABLE I DIAGNOSTIC TEST RESULTS IN RELATION TO TRUE DISEASE STATUS IN A 2x2 TABLE

Diagnostic test result	Disease status		Total
	Present	Absent	
Present	True positive (TP)	False positive (FP)	All test positive (T+)
Absent	False negative (FN)	True negative (TN)	All test negative (T-)
Total	Total with disease (D+)	Total without disease (D-)	Total sample size

TABLE II HYPOTHETICAL DATA SHOWING THE SENSITIVITY AND SPECIFICITY AT VARIOUS CUT-OFF POINTS OF MID-ARM CIRCUMFERENCE TO DETECT LOW BIRTH WEIGHT

Mid-arm circumference (cm)	Low birthweight (<2500 grams) (n=130)		Normal birth weight (≥2500 grams) (n=870)		Sensitivity = TP/(TP+FN)	Specificity = TN/(TN+FP)
	True positive (TP)	False negative (FN)	False positive (FP)	True negative (TN)		
≤8.3	13	117	3	867	0.1000	0.9966
≤8.4	24	106	26	844	0.1846	0.9701
≤8.5	73	57	44	826	0.5615	0.9494
≤8.6	90	40	70	800	0.6923	0.9195
≤8.7	113	17	87	783	0.8692	0.9000
≤8.8	119	11	135	735	0.9154	0.8448
≤8.9	121	09	244	626	0.9308	0.7195
≤9.0	125	05	365	505	0.9615	0.5805
≤9.1	127	03	435	435	0.9769	0.5000
≤9.2 & above	130	00	870	0	1.0000	0.0000

generally depicted in a square box for convenience and its both axes are from 0 to 1. **Figure 1** depicts a ROC curve and its important components as explained later. The area under the curve (AUC) is an effective and combined measure of sensitivity and specificity for assessing inherent validity of a diagnostic test. Maximum AUC = 1 and it means diagnostic test is perfect in differentiating diseased with non-diseased subjects. This implies both sensitivity and specificity are one and both errors—false positive and false negative—are zero. This can happen when the distribution of diseased and non-diseased test values do not overlap. This is extremely unlikely to happen in practice. The AUC closer to 1 indicates better performance of the test.

The diagonal joining the point (0, 0) to (1,1) divides the square in two equal parts and each has an area equal to 0.5. When ROC is this line, overall there is 50-50 chances that test will correctly discriminate the diseased and non-diseased subjects. The minimum value of AUC should be considered 0.5 instead of 0 because AUC = 0 means test incorrectly classified all subjects with disease as negative and all non-disease subjects as positive. If the test results are reversed then area = 0 is transformed to area = 1; thus a perfectly inaccurate test can be transformed into a perfectly accurate test!

ADVANTAGES OF THE ROC CURVE

ROC curve has following advantages compared with single value of sensitivity and specificity at a particular cut-off.

1. The ROC curve displays all possible cut-off points, and one can read the optimal cut-off for correctly identifying diseased or non-diseased subjects as per the procedure given later.

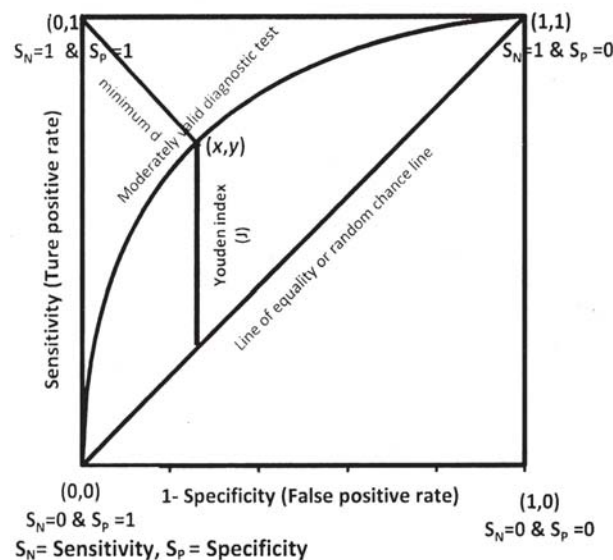


FIG. 1 ROC curve and its components.

2. The ROC curve is independent of prevalence of disease since it is based on sensitivity and specificity which are known to be independent of prevalence of disease [6-7].
3. Two or more diagnostic tests can be visually compared simultaneously in one figure.
4. Sometimes sensitivity is more important than specificity or vice versa, ROC curve helps in finding the required value of sensitivity at fixed value of specificity.
5. Empirical area under the ROC curve (explained later) is invariant with respect to the addition or subtraction of a constant or transformation like log or square root [8]. Log or square root transformation condition is not applicable for binormal ROC curve. Binormal ROC is also shortly explained.
6. Useful summary of measures can be obtained for determining the validity of diagnostic test such as AUC and partial area under the curve.

NON-PARAMETRIC AND PARAMETRIC METHODS TO OBTAIN AREA UNDER THE ROC CURVE

Statistical softwares provide non-parametric and parametric methods for obtaining the area under ROC curve. The user has to make a choice. The following details may help.

Non-parametric Approach

This does not require any distribution pattern of test values and the resulting area under the ROC curve is called empirical. First such method uses trapezoidal rule. It calculates the area by just joining the points $(1-S_p, S_M)$ at each interval of the observed values of continuous test and draws a straight line joining the x-axis. This forms several trapezoids (**Fig 2**) and their area can be easily calculated and summed. **Figure 2** is drawn for the mid-arm circumference and low birth weight data in **Table II**. Another non-parametric method uses Mann-Whitney statistics, also known as Wilcoxon rank-sum statistic and the c-index for calculating area. Both these methods of estimating AUC estimate have been found equivalent [7].

Standard errors (SE) are needed to construct a confidence interval. Three methods have been suggested for estimating the SE of empirical area under ROC curve [7,9-10]. These have been found similar when sample size is greater than 30 in each group provided test value is on continuous scale [11]. For small sample size it is difficult to recommend any one method. For discrete ordinal outcome, Bamber method [9] and Delong method [10] give equally good results and better than Hanley and McNeil method [7].

Parametric Methods

These are used when the statistical distribution of diagnostic test values in diseased and non-diseased is known. Binormal distribution is commonly used for this purpose. This is applicable when test values in both diseased and non-diseased subjects follow normal distribution. If data are actually binormal or a transformation such as log, square or Box-Cox [12] makes the data binormal then the relevant parameters can be easily estimated by means and variances of test values in diseased and non-diseased subjects. Details are available elsewhere [13].

Another parametric approach is to transform the test results into an unknown monotone form when both the diseased and non-diseased populations

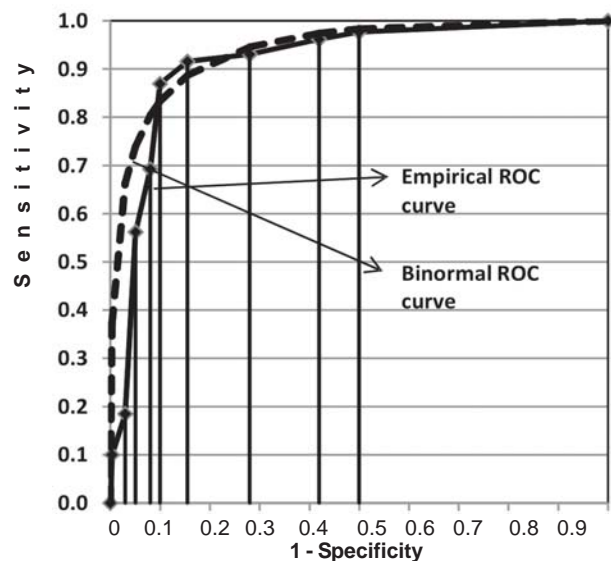


FIG. 2 Comparison of empirical and binormal ROC curves for hypothetical neonatal data in **Table II**.

follow binormal distribution [14]. This first discretizes the continuous data into a maximum 20 categories, then uses maximum likelihood method to estimate the parameters of the binormal distribution and calculates the AUC and standard error of AUC. ROCKIT package containing ROCFIT method uses this approach to draw the ROC curve, to estimate the AUC, for comparison between two tests, and to calculate partial area [15].

The choice of method to calculate AUC for continuous test values essentially depends upon availability of statistical software. Binormal method and ROCFIT method produce results similar to non-parametric method when distribution is binormal [16]. In unimodal skewed distribution situation, Box-Cox transformation that makes test value binormal and ROCFIT method perform give results similar to non-parametric method but former two approaches have additional useful property for providing smooth curve [16,17]. When software for both parametric and non-parametric methods is available, conclusion should be based on the method which yields greater precision to estimate the AUC. However, for bimodal distribution (having two peaks), which is rarely found in medical practice, Mann-Whitney gives more accurate estimates compare to parametric methods [16]. Parametric method gives small bias for discrete test value compared to non-parametric method [13].

The area under the curve by trapezoidal rule and Mann-Whitney U are 0.9142 and 0.9144, respectively, of mid-arm circumference for indicating low birth weight in our data. The SE also is nearly equal by three methods in these data: Delong SE = 0.0128, Bamber SE = 0.0128, and Hanley and McNeil SE = 0.0130. For parametric method, smooth ROC curve was obtained assuming binormal assumption (**Fig 2**) and the area under the curve is calculated by using means and standard deviations of mid-arm circumference in normal and low birth weight neonates which is 0.9427 and its SE is 0.0148 in this example. Binormal method showed higher area compared to area by non-parametric method which might be due to violation of binormal assumption in this case. Binormal ROC curve is initially above the empirical curve (**Fig 2**) suggesting higher sensitivity compared to empirical

values in this range. When (1-specificity) lies between 0.1 to 0.2, the binormal curve is below the empirical curve, suggesting comparatively low sensitivity compared to empirical values. When values of (1-specificity) are greater than 0.2, the curves are almost overlapping suggesting both methods giving the similar sensitivity. The AUC by using ROCFIT methods is 0.9161 and its standard error is 0.0100. This AUC is similar to the non-parametric method; however standard error is little less compared to standard error by non-parametric method. The data in our example has unimodal skewed distribution and results agree with previous simulation study [16, 17] on such data. All calculations were done using MS Excel and STATA statistical software for this example.

Interpretation of ROC Curve

Total area under ROC curve is a single index for measuring the performance a test. The larger the AUC, the better is overall performance of diagnostic test to correctly pick up diseased and non-diseased subjects. Equal AUCs of two tests represents similar overall performance of medical tests but this does not necessarily mean that both the curves are identical. They may cross each other. Three common interpretations of area under the ROC curve are: (i) the average value of sensitivity for all possible values of specificity, (ii) the average value of specificity for all possible values of sensitivity [13]; and (iii) the probability that a randomly selected patient with disease has positive test result that indicates greater suspicion than a randomly selected patient without disease [10] when higher values of the test are associated with disease and lower values are associated with non-disease. This interpretation is based on non-parametric Mann-Whitney U statistic for calculating the AUC.

Figure 3 depicts three different ROC curves. Considering the area under the curve, test A is better than both B and C, and the curve is closer to the perfect discrimination. Test B has good validity and test C has moderate.

Hypothetical ROC curves of three diagnostic tests A, B, and C applied on the same subjects to

classify the same disease are shown in **Fig 4**. Test B (AUC=0.686) and C (AUC=0.679) have nearly equal area but cross each other whereas test A (AUC=0.805) has higher AUC value than curves B and C. The overall performance of test A is better than test B as well as test C at all the threshold points. Test C performed better than test B where high sensitivity is required, and test B performed better than C when high specificity is needed.

Sensitivity at Fixed Point and Partial Area Under the ROC Curve

The choice of fixed specificity or range of specificity depends upon clinical setting. For example, to diagnose serious disease such as cancer in a high risk group, test that has higher sensitivity is preferred even if the false positive rate is high because test giving more false negative subjects is more dangerous. On the other hand, in screening a low risk group, high specificity is required for the diagnostic test for which subsequent confirmatory test is invasive and costly so that false positive rate should be low and patient does not unnecessarily suffers pain and pays price. The cut-off point should be decided accordingly. Sensitivity at fixed point of specificity or vice versa and partial area under the

curve are more suitable in determining the validity of a diagnostic test in above mentioned clinical setting and also for the comparison of two diagnostic tests when applied to same or independent patients when ROC curves cross each other.

Partial area is defined as area between range of false positive rate (FPR) or between the two sensitivities. Both parametric (binormal assumption) and non-parametric methods are available in the literature [13,18] but most statistical softwares do not have option to calculate the partial area under ROC curve (**Table III**). STATA software has all the features for ROC analysis.

Standardization of the partial area by dividing it with maximum possible area (equal to the width of the interval of selected ranged of FPRs or sensitivities) has been recommended [19]. It can be interpreted as average sensitivity for the range of selected specificities. This standardization makes partial area more interpretable and its maximum value will be 1. **Figure 5** shows that the partial area under the curve for FPR from 0.3 to 0.5 for test A is 0.132, whereas for test B is 0.140. After standardization, they it would be 0.660 and 0.700,

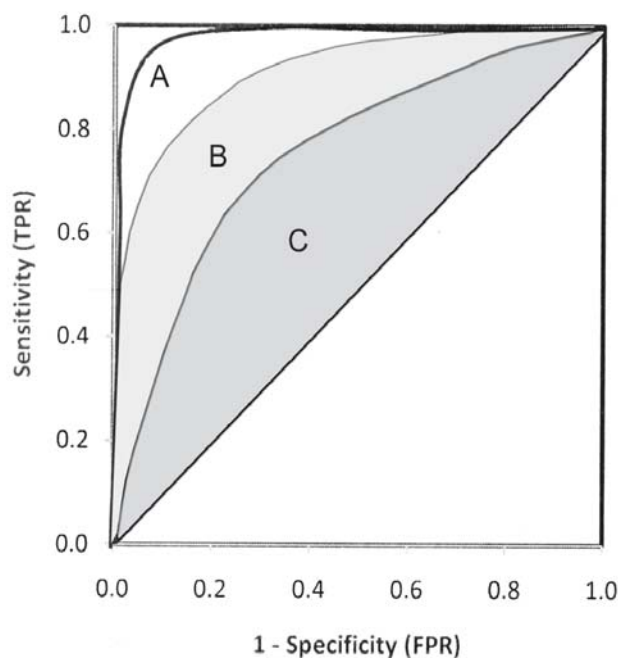


FIG. 3 Comparison of three smooth ROC curves with different areas.

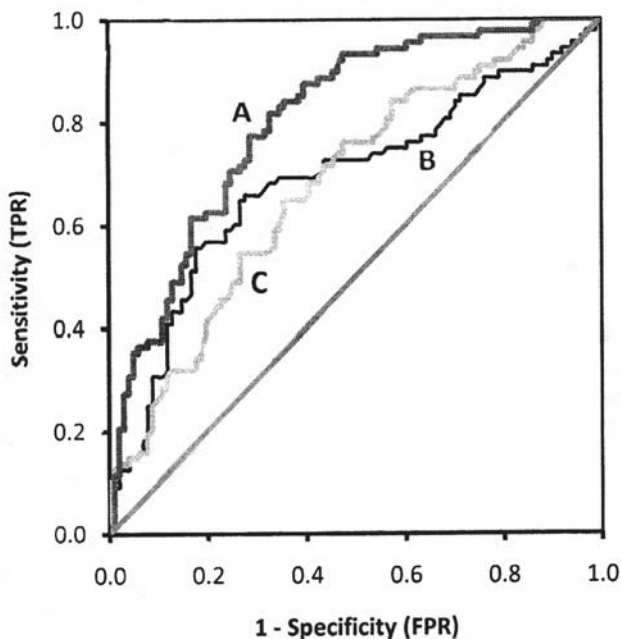


FIG. 4 Three empirical ROC curves. Curves for B and C cross each other but have nearly equal areas, curve A has bigger area.

TABLE III SOME POPULAR ROC ANALYSIS SOFTWARES AND THEIR IMPORTANT FEATURES

Name of ROC analysis software	Methods used to estimate AUC of ROC curve and its variance	Comparison of two or more ROC curves	Partial area		Important by-products
			Calculation	Comparison	
Medcalc software version 11.3; Commercial software trial version available at: www.medcalc.be	Non-parametric	Available	Not available	Not available	Sensitivity, specificity, LR+, LR- with 95% CI at each possible cut-point
SPSS version 17.0; Commercial software	Non-parametric	Not available	Not available	Not available	Sensitivity and specificity at each cut-off point - no 95% CI
STATA version 11; Commercial software	Non-parametric Parametric (Metz et al.)	Available for paired and unpaired subjects with Bonferroni adjustment when 3 or more curves to be compared	Available specificity at specified range and sensitivity range	Only two partial AUCs can be compared	Sensitivity, specificity, LR+, LR- at each cut-off point but no 95% CI
ROCKIT (Beta version) Free software; Available at www.radiology.uchicago.edu	Parametric (Metz et al.)	Available for both paired and unpaired subjects	Available	Available	Sensitivity and specificity at each cut-point
Analyse - it Commercial software; add on in the MS Excel; trial version available at www.analyse-it.com	Non-parametric	Available – no option for paired and unpaired subjects	Not available	Not available	Sensitivity, specificity, LR+, LR- with 95% CI of each possible cut-point; Various decision plots
XLstat2010 Commercial software; add on in the MS Excel; trial version available at: www.xlstat.com	Non-parametric	Available for both paired and unpaired subjects	Not available	Not available	Sensitivity, specificity, LR+, LR- with 95% CI of each possible cut-Various - decision plots
Sigma plot Commercial software; add on in the MS Excel; trial version available at: www.sigmaplot.com	Non-parametric	Available for both paired and unpaired subjects	Not available	Not available	Sensitivity, specificity, LR+, LR- with 95% CI of each possible cut-point: Various- decision plots

*LR+ = Positive likelihood ratio, LR- = Negative likelihood ratio, Paired subjects means both diagnostic tests (test and gold) applied to same subjects and unpaired subjects means diagnostic tests applied to different subjects, CI = Confidence interval, AUC=Area under curve, ROC=Receiver operating characteristic.

respectively. The portion of partial area will depend on the range of interest of FPRs selected by researcher. It may lie on one side of intersecting point or may be on both sides of intersecting point of ROC curves. In Figure 5, $S_N(A)$ and $S_N(B)$ are sensitivities at specific value of FPR. For example, sensitivity at FPR=0.3 is 0.545 for test A and 0.659 for test B. Similarly sensitivity at fixed FPR=0.5 for test A is 0.76 and 0.72 for test B. All these calculations were done by STATA (version 11) statistical software using *comproc* command with option *pcvmeth(empirical)*.

METHOD TO FIND THE 'OPTIMAL' THRESHOLD POINT

Optimal threshold is the point that gives maximum correct classification. Three criteria are used to find optimal threshold point from ROC curve. First two methods give equal weight to sensitivity and specificity and impose no ethical, cost, and no prevalence constraints. The third criterion considers cost which mainly includes financial cost for correct and false diagnosis, cost of discomfort to person caused by treatment, and cost of further investigation when needed. This method is rarely used in medical literature because it is difficult to implement. These three criteria are known as points on curve closest to the (0, 1), Youden index, and minimize cost criterion, respectively.

The distance between the point (0, 1) and any point on the ROC curve is $d^2 = [(1 - S_N)^2 + (1 - S_p)^2]$. To obtain the optimal cut-off point to discriminate the disease with non-disease subject, calculate this distance for each observed cut-off point, and locate the point where the distance is minimum. Most of the ROC analysis softwares (**Table III**) calculate the sensitivity and specificity at all the observed cut-off points allowing you to do this exercise.

The second is Youden index [20] that maximizes the vertical distance from line of equality to the point $[x, y]$ as shown in **Fig 1**. The x -axis represents (1 - specificity) and y -axis represents sensitivity. In other words, the Youden index J is the point on the ROC curve which is farthest from line of equality (diagonal line). The main aim of Youden index is to maximize the difference between TPR (S_N) and

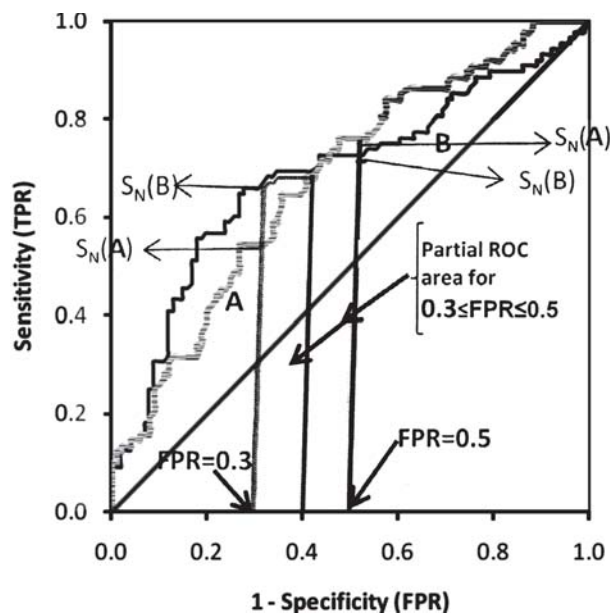


FIG. 5 The partial area under the curve and sensitivity at fixed point of specificity (see text).

FPR ($1 - S_p$) and little algebra yields $J = \max[S_N + S_p]$. The value of J for continuous test can be located by doing a search of plausible values where sum of sensitivity and specificity can be maximum. Youden index is more commonly used criterion because this index reflects the intention to maximize the correct classification rate and is easy to calculate. Many authors advocate this criterion [21]. Third method that considers cost is rarely used in medical literature and is described in [13].

BIASES THAT CAN AFFECT ROC CURVE RESULTS

We describe more prevalent biases in this section that affect the sensitivity, specificity and consequently may affect the area under the ROC curve. Interested researcher can find detailed description of these and other biases such as withdrawal bias, lost to follow-up bias, spectrum bias, and population bias, elsewhere [22,23].

1. **Gold standard:** Validity of gold standard is important—ideally it should be error free and the diagnostic test under review should be independent of the gold standard as this can increase the area under the curve spuriously. The gold standard can be clinical follow-up, surgical verification, biopsy or autopsy or in some cases

opinion of panel of experts. When gold standard is imperfect, such as peripheral smear for malaria parasites [24], sensitivity and specificity of the test are under estimated [22].

2. *Verification bias*: This occurs when all disease subjects do not receive the same gold standard for some reason such as economic constraints and clinical considerations. For example, in evaluating the breast bone density as screening test for diagnosis of breast cancer and only those women who have higher value of breast bone density are referred for biopsy, and those with lower value but suspected are followed clinically. In this case, verification bias would overestimate the sensitivity of breast bone density test.
3. *Selection bias*: Selection of right patients with and without diseased is important because some tests produce perfect results in severely diseased group but fail to detect mild disease.
4. *Test review bias*: The clinician should be blind to the actual diagnosis while evaluating a test. A known positive disease subject or known non-disease subject may influence the test result.

5. *Inter-observer bias*: In the studies where observer abilities are important in diagnosis, such as for bone density assessment through MRI, experienced radiologist and junior radiologist may differ. If both are used in the same study, the observer bias is apparent.
6. *Co-morbidity bias*: Sometimes patients have other types of known or unknown diseases which may affect the positivity or negativity of test. For example, NESTROFT (Naked eye single tube red cell osmotic fragility test), used for screening of thalassaemia in children, shows good sensitivity in patients without any other hemoglobin disorders but also produces positive results when other hemoglobin disorders are present [25].
7. *Uninterpretable test results*: This bias occurs when test provides results which can not be interpreted and clinician excludes these subjects from the analysis. This results in over estimation of validity of the test.

It is difficult to rule out all the biases but researcher should be aware and try to minimize them.

TABLE IV SAMPLE SIZE FORMULA FOR ESTIMATING SENSITIVITY AND SPECIFICITY AND AREA UNDER THE ROC CURVE

Problem	Formula	Description of symbol used
Estimating the sensitivity of test	$\frac{Z^2_{1-\alpha/2} S_N (1-S_N)}{\epsilon^2 \times \text{Prev}}$	<p>S_N = Anticipated sensitivity</p> <p>Prev = Prevalence of disease in population can be obtained from previous literature or pilot study</p> <p>ϵ = required absolute precision on either side of the sensitivity</p>
Estimating the specificity of test	$\frac{Z^2_{1-\alpha/2} S_p (1-S_p)}{\epsilon^2 \times (1-\text{Prev})}$	<p>S_N = Anticipated specificity</p> <p>Prev = Prevalence of disease in population can be obtained from previous literature or pilot study</p> <p>ϵ = required absolute precision on either side of the specificity.</p>
Estimating the area under the ROC curve	$n_D = \frac{Z^2_{\alpha/2} \times V(\text{AUC})}{\epsilon^2}$	<p>V(AUC) = Anticipated variance anticipated area under ROC curve</p> <p>ϵ = required absolute precision on either side of the area under the curve.</p>
	<p>n_D = number of diseased subjects</p> <p>$n = n_D(1+k)$, k is ratio of prevalence of non-disease to disease subjects</p>	

$Z_{1-\alpha/2}$ is a standard normal value and α is the confidence level. $Z_{1-\alpha/2} = 1.645$ for $\alpha=0.10$ and $Z_{1-\alpha/2} = 1.96$ for $\alpha=0.05$.

KEY MESSAGES

- For sensitivity and specificity to be valid indicators, the gold standard should be nearly perfect.
- For a continuous test, optimal cut-off point to correctly pick-up a disease and non-diseased cases is the point where sum of specificity and sensitivity is maximum, when equal weight is given to both.
- ROC curve is obtained for a test on continuous scale. Area under the ROC curves is an index of overall inherent validity of test as well as used for comparing sensitivity and specificity at particular cut-offs of interest.
- Area under the curve is not an accurate index of comparing two tests when their ROC curves cross each other.
- Adequate sample size and proper designing is required to yield valid and unbiased results.

SAMPLE SIZE

Adequate power of the study depends upon the sample size. Power is probability that a statistical test will indicate significant difference where certain pre-specified difference is actually present. In a survey of eight leading journals, only two out of 43 studies reported a prior calculation of sample size in diagnostic studies [26]. In estimation set-up, adequate sample size ensures the study will yield the estimate with desired precision. Small sample size produces imprecise or inaccurate estimate, while large sample size is wastage of resources especially when a test is expensive. The sample size formula depends upon whether interest is in estimation or in testing of the hypothesis. **Table IV** provides the required formula for estimation of sensitivity, specificity and AUC. These are based on the normal distribution or asymptotic assumption (large sample theory) which is generally used for sample size calculation.

Variance of AUC, required in formula 3 (**Table IV**), can be obtained by using either parametric or non-parametric method. This may also be available in literature on previous studies. If no previous study is available, a pilot study is done to get some workable estimates to calculate sample size. For pilot study data, appropriate statistical software can provide estimate of this variance.

Formulas of sample size to test hypothesis on sensitivity-specificity or the AUC with a pre-specified value and for comparison on the same subjects or different subjects are complex. Refer [13] for details. A Nomogram was devised to read the sample size for anticipated sensitivity and

specificity at 90%, 95%, 99% confidence level [27].

There are many more topics for interested reader to explore such as combining the multiple ROC curve for meta-analysis, ROC analysis to predict more than one alternative, ROC analysis in the clustered environment, and for tests for repeated over the time. For these see [13,28]. For predictivity based ROC, see [6].

Contributors: Both authors contributed to concept, review of literature, drafting the paper and approved the final version.

Funding: None.

Competing interests: None stated.

REFERENCES

1. Reddy S, Dutta S, Narang A. Evaluation of lactate dehydrogenase, creatine kinase and hepatic enzymes for the retrospective diagnosis of perinatal asphyxia among sick neonates. *Indian Pediatr.* 2008;45:144-7.
2. Sood SL, Saiprasad GS, Wilson CG. Mid-arm circumference at birth: A screening method for detection of low birth weight. *Indian Pediatr.* 2002;39:838-42.
3. Randev S, Grover N. Predicting neonatal hyperbilirubinemia using first day serum bilirubin levels. *Indian J Pediatr.* 2010; 77:147-50.
4. Vasudevan A, Malhotra A, Lodha R, Kabra SK. Profile of neonates admitted in pediatric ICU and validation of score for neonatal acute physiology (SNAP). *Indian Pediatr.* 2006;43:344-8.
5. Khanna R, Taneja V, Singh SK, Kumar N, Sreenivas V, Puliyl JM. The clinical risk index of babies (CRIB) score in India. *Indian J Pediatr.* 2002;69:957-60.
6. Indrayan A. *Medical Biostatistics (Second Edition)*. Boca Raton: Chapman & Hall/ CRC Press; 2008. p. 263-7.
7. Hanley JA, McNeil BJ. The meaning and use of the area under a receiver operating characteristic (ROC) curve. *Radiology.* 1982;143:29-36.
8. Campbell G. Advances in statistical methodology for evaluation of diagnostic and laboratory tests. *Stat Med.* 1994;13:499-508.

9. Bamber D. The area above the ordinal dominance graph and area below the receiver operating characteristic graph. *J Math Psychol.* 1975;12:387-415.
10. DeLong ER, DeLong DM, Clarke-Pearson DL. Comparing the area under two or more correlated receiver operating characteristic curves: a nonparametric approach. *Biometrics.* 1988;44:837-45.
11. Cleves MA. Comparative assessment of three common algorithms for estimating the variance of the area under the nonparametric receiver operating characteristic curve. *Stata J.* 2002;3:280-9.
12. Box GEP, Cox DR. An analysis of transformation. *J Royal Statistical Society, Series B.* 1964;26:211-52.
13. Zhou Xh, Obuchowski NA, McClish DK. *Statistical Methods in Diagnostic Medicine.* New York: John Wiley and Sons, Inc; 2002.
14. Metz CE, Herman BA, Shen JH. Maximum likelihood estimation of receiver operating characteristic (ROC) curve from continuously distributed data. *Stat Med.* 1998;17:1033-53.
15. ROCKIT [Computer program]. Chicago: University of Chicago. Available from: www-radiology.uchicago.edu/krl/KRL_ROC/software_index6.htm. Accessed on February 27, 2007.
16. Faraggi D, Reiser B. Estimating of area under the ROC curve. *Stat Med.* 2002; 21:3093-3106.
17. Hajian Tilaki KO, Hanley JA, Joseph L, Collet JP. A comparison of parametric and nonparametric approaches to ROC analysis of quantitative diagnosis tests. *Med Decis Making.* 1997;17:94-102.
18. Pepe M, Longton G, Janes H. Estimation and comparison of receiver operating characteristic curves. *Stata J.* 2009;9:1.
19. Jiang Y, Metz CE, Nishikawa RM. A receiver operating characteristic partial area index for highly sensitive diagnostic test. *Radiology.* 1996;201:745-50.
20. Youden WJ. An index for rating diagnostic test. *Cancer.* 1950;3:32-5.
21. Perkins NJ, Schisterman EF. The inconsistency of 'optimal' cut points obtained using two criteria based on the receiver operating characteristic curve. *Am J Epidemiol.* 2006;163:670-5.
22. Whiting P, Ruljes AW, Reitsma JB, Glas AS, Bossuyt PM, Kleijnen J. Sources of variation and bias in studies of diagnostic accuracy – a systematic review. *Ann Intern Med.* 2004;140:189-202.
23. Kelly S, Berry E, Proderick P, Harris KM, Cullingworth J, Gathercale L, *et al.* The identification of bias in studies of the diagnostic performance of imaging modalities. *Br J Radiol.* 1997;70:1028-35.
24. Malaria Site. Peripheral smear study for malaria parasites – Available from: www.malariasite.com/malaria/DiagnosisOfMalaria.htm. Accessed on July 05, 2010.
25. Thomas S, Srivastava A, Jeyaseelan L, Dennison D, Chandy M. NESTROFT as screening test for the detection of thalassaemia & common haemoglobinopathies – an evaluation against a high performance liquid chromatographic method. *Indian J Med Res.* 1996;104:194-7.
26. Bachmann LM, Puhan MA, ter Riet G, Bossuyt PM. Sample sizes of studies on diagnostic accuracy: literature survey. *BMJ.* 2006;332:1127-9.
27. Malhotra RK, Indrayan A. A simple nomogram for sample size for estimating the sensitivity and specificity of medical tests. *Indian J Ophthalmol.* 2010;58:519-22.
28. Kester AD, Buntinx F. Meta analysis of curves. *Med Decis Making.* 2000;20:430-9.