

A conformational analysis of Walker motif A [GXXXXGKT (S)] in nucleotide-binding and other proteins

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The sequence GXXXXGKT/S, popularly known as Walker motif A, is widely believed to be the site for binding nucleotides in many proteins. Examination of the crystal structures in the Protein Data Bank showed that about half of the examples having these sequences do not bind or use nucleotides. Data analyses showed 92 different Walker sequences of the variable quartet (XXXX). Ramachandran angles in this segment revealed conformational similarity in the group of 45 proteins, known to bind or utilize nucleotides. The conformations of this segment in other proteins differ widely and it is not known whether they play any role in their functions. A flip of a peptide unit at different locations, with little change in the backbone conformation was noted in nine pairs of these proteins having same Walker sequence. An examination of the immediate neighborhood of the Walker sequence indicates that this region is preceded by a β-strand and followed by an α-helix, resulting in the motif β -W- α , an invariant feature amongst nucleotidebinding proteins.

Keywords: peptide flip/Ramachandran angles/β-turn/Walker motif

Introduction

The motif GXXXXGKT (X, any residue) as a common nucleotide binding fold in the α - and β -subunits of F_1 -ATPase, myosin and other ATP-requiring enzymes was first recognized in 1982 by Walker and colleagues (Walker et al., 1982). Since then, this sequence has been found in many proteins that bind nucleotides and thereby gained predictive value for nucleotide binding site in proteins. Crystal structure data of such proteins (Berchtold et al., 1993; Abrahams et al., 1994; Chattopadhyay et al., 2000) indicated that this motif is present in the shape of a loop around nucleotides and utilizes its highly conserved residues of lysine and threonine to bind to their phosphateoxygen atoms. This consensus sequence of GXXXXGKT (S), with serine substituting threonine in some cases, is more popularly known as Walker loop or P-loop (phosphate binding loop).

In view of growing interest in the proteins containing a segment with Walker sequence, the Brookhaven Protein Data Bank (Berman et al., 2000) was searched and 649 polypeptide chains were found to have such a sequence. Many of these proteins do not bind or use nucleotides in their reactions. Therefore, it appeared that the sequence of the variant quartet and the specific loop structure might have a role in nucleotide binding. To fill the lacunae of information, conformations of the backbone of the peptide fragments of GXXXXGKT (S) were examined using Ramachandran angles. The data analysis in this paper indicates that different foldings are possible for the Walker sequences and only in the nucleotide-binding proteins they have a distinctive loop structure.

Materials and methods

The Ramachandran angles (ϕ, ψ) (Ramachandran *et al.*, 1963; Ramachandran and Sasisekharan, 1968) were computed from the coordinates of atoms available in the Brookhaven Protein Data Bank (Berman et al., 2000). The segment structure similarity was obtained by evaluating the root mean square (r.m.s.) values of the Ramachandran angles. The package of RASMOL (Sayle and Milner-White, 1995) was used to draw the figures.

Results of data analysis

Proteins containing Walker sequences

Search for the sequence GXXXXGKT (S) in the Protein Data Bank (April 2001 release) revealed 649 entries having this sequence, occurring in 395 protein structures with a resolution of 4 Å or better. Out of the 20⁴ combinations of sequence possible for the variable region XXXX, only 92 were found to occur, of which 18 had only one entry. The present analysis is limited to these data

The Ramachandran angles of Walker sequence

Groups having more than one entry were examined from the structural viewpoint. The mean and r.m.s. values of the Ramachandran angles ϕ and ψ were computed at the eight residues of the segment. Should the same sequence give the same structure, as is widely believed, the r.m.s. values for a group would be small. Using a liberal upper limit of 40°, dissimilar structures were found to be present in 10 of these groups, as revealed by the high r.m.s. values for some of the Ramachandran angles. Using similarity of the Ramachandran angles as the criterion, these were divided into further sub-groups. The various sequences and location of the segment in the protein of the group thus obtained are given in Table I, along with the PDB code, chain identifier, resolution of the structure and r.m.s. for those groupings with more than one entry (the protein names are not included in Table I owing to the large number of examples; however, they are included in Table II, which gives the selected set). The sub-groups with same sequence are indicated by suffixes A, B and C, to the group number. It can be seen that the r.m.s. values are now reasonably small. Some sequences assume more than one conformation: two for six sequences (005 - GAGALGKT, 012 -GLRSDGKT, 016 - GLPAIGKT, 030 - GATGTGKT, 058 -GTAFEGKS and 077 - GLYRTGKS); three for three sequences (006 - GHVDHGKT, 033 - GPTGVGKT and

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 $\textbf{Table I.} \ Proteins \ containing \ the \ consensus \ sequence \ of \ GXXXXGKT(S): \ the \ location \ of \ the \ segment \ in \ the \ chain, \ PDB \ code \ and \ resolution \ of \ the \ crystal \ structure \ are given$

No.	Sequence Segment location	PDB code, (resolution in Å)) chain identifier	Residue r.m.s. (ϕ, ψ)	Residue r.m.s. (ϕ, ψ)
001	GLSGTGKT 248 255	1AQ2 (1.9)	1AYL (1.8)	G (1,1) L (1,1) S (6,1) G (2,1)	T (1,6) G (6,2) K (1,2) T (2,1)
002	GDRQTGKT 169 176	1BMF (2.8) (A,B,C) 1E1Q (2.6) (A,B,C) 1E79 (2.4) (A,B,C) 1MAB (2.8) (A)	1COW (3.1) (A,B,C) 1E1R (2.5) (A,B,C) 1EFR (3.1) (A,B,C) 1NBM (3.0) (A,B,C)	G (7,8) D (7,6) R (6,5) Q (7,11)	T (9,10) G (13,10) K (8,8) T (10,6)
003	GGAGVGKT 156 163	1BMF (2.8) (D,F) 1E1Q (2.6) (D,E,F) 1E79 (2.4) (D,E,F) 1NBM (3.0) (D,E,F)	1COW (3.1) (D,F) 1E1R (2.5) (D,F) 1EFR (3.1) (D,F)	G (23,11) G (7,19) A (19,18) G (13,17)	V (17,18) G (23,10) K (6,4) T (5,7)
003A	GGAGVGKT 156 163	1BMF (2.8) (E) 1EFR (3.1) (E)	1COW (3.1) (E)	G (1,2) G (2,0) A (1,1) G (2,2)	V (2,2) G (2,5) K (5,0) T (2,10)
003B	GGAGVGKT 156 163	1E1R (2.5) (E)			
003C	GGAGVGKT 156 163	1MAB (2.8) (B)			
004	GAHALGKT 173 180	1A2F (2.1) 1AA4 (2.1) 1AC8 (2.1) 1AED (2.1) 1AED (2.1) 1AEH (2.1) 1AEK (2.1) 1AEK (2.1) 1AEK (2.1) 1AEV (2.1) 1AEV (2.1) 1AEV (2.1) 1BES (2.0) 1BEY (2.2) 1BES (2.0) 1BVA (1.8) (A) 1CCB (2.1) 1CCI (2.4) 1CCK (2.1) 1CCP (2.2) 1CMQ (2.3) 1CMU (2.1) 1CPE (2.2) 1DPE (2.2) 1DPE (2.2) 1DCC (2.3) 1CCY (2.3)	1A2G (2.1) 1AC4 (2.1) 1AEB (2.1) 1AEB (2.1) 1AEG (2.1) 1AEG (2.1) 1AEJ (2.1) 1AEM (2.1) 1AEO (2.1) 1AES (2.1) 1AEU (2.1) 1BEJ (2.4) 1BEM (2.2) 1BEQ (2.1) 1BJ9 (2.2) 1CCA (1.8) 1CCC (2.0) 1CCJ (2.1) 1CCL (2.0) 1CMF (1.9) 1CMT (2.1) 1CPD (2.2) 1CPF (2.2) 1CYF (2.3) 1DJ1 (1.9) (A) 1RYC (1.8) 2CEP (2.2) 2PCB (2.8) (A,C) 3CCP (2.2) 4CCP (2.2) 5CCP (2.2) 5CCP (2.2)	G (5,8) A (5,4) H (5,7) A (8,5)	L (7,4) G (6,5) K (5,5) T (4,4)
005	GAGALGKT 173 180	1CCE (2.3)	1CCG (2.1)	G (5,9) A (5,10) G (11,3) A (1,3)	L (2,1) G (2,1) K (4,2) T (1,3)
005A	GAGALGKT 173 180	1DS4 (2.0) (A) 1DSG (2.5) (A) 1DSP (2.0) (A)	1DSE (2.0) (A) 1DSO (2.0) (A)	G (9,14) A (12,8) G (3,7) A (6,10)	L (13,2) G (4,6) K (3,6) T (5,3)
006	GHVDHGKT 18 25	1B23 (2.6) (P) 1D8T (2.3) (A,B) 1EFC (2.0) (A,B) 1EXM (1.7) A 1G7T (2.0) (A)	1D2E (1.9) (A–D) 1DG1 (2.5) (G,H) 1EFT (2.5) 1G7S (2.0) (A) 1TUI (2.7) (A,B,C)	G (9,4) H (5,6) V (6,5) D (6,7)	H (8,7) G (8,9) K (7,9) T (6,6)

Table I. Co	ommed				
No.	Sequence Segment location	PDB code, (resolution in Å)	chain identifier	Residue r.m.s. (φ,ψ)	Residue r.m.s. (φ,ψ)
006A	GHVDHGKT 18 25	1AIP (3.0) (A,B,E,F)	1EFU (2.5) (A,C)	G (18,5) H (5,4) V (9,4) D (11,7)	H (2,4) G (5,14) K (10,5) T (3,5)
006B	GHVDHGKT 18 25	1ETU (2.9)		D (11,7)	1 (3,3)
007	GYLVNGKT 1264 271	10MH (2.5) (A) HMY (2.5) 2HMY (2.6) (B) 4MHT (2.7) (A) 6MHT (2.0) (A) 8MHT (2.7) (A)	1FJX (2.2) (A) 1MHT (2.8) (A) 3MHT (2.7) (A) 5MHT (2.7) (A) 7MHT (2.8) (A) 9MHT (2.3) (A)	G (7,4) Y (5,4) L (3,7) V (8,6)	N (9,6) G (9,7) K (10,7) T (9,8)
008	GLDAAGKT 24 31	1EOS (2.2) (A) 1HUR (2.0) (A,B) 1RRG (2.4) A,B	1HFV (2.8) (A,B) 1RRF (3.0)	G (4,5) L (9,6) D (7,8) A (3,7)	A (7,9) G (13,9) K (9,9) T (7,6)
009	GPHGMGKT 56 63	1E2H (1.9) (A,B) 1E2J (2.5) (A,B) 1E2L (2.4) (A,B) 1KI3 (2.3) (A,B) 1KI6 (2.3) (A,B) 1KI8 (2.2) (A,B) 1KIN (2.0) (A,B) 1VTK (2.7) 2VTK (2.8)	1E2I (1.9) (A,B) 1E2K (1.7) (A,B) 1KI2 (2.2) (A,B) 1KI4 (2.3) (A,B) 1KI7 (2.2) (A,B) 1KIM (2.1) (A,B) 1QHI (1.9) (A,B) 2KI5 (1.9) (A,B) 3VTK (3.0)	G (14,4) P (5,6) H (5,5) G (7,12)	M (9,14) G (14,10) K (8,9) T (7,8)
010	GVRSDGKT 487 494	1MHY (2.) (D)	1MHZ (2.7) (D)	G (5,9) V (11,2) R (8,3) S (9,8)	D (2,3) G (9,0) K (14,10) T (1,2)
011	GESGAGKT 179 186	1B7T (2.5) (A) 1BR2 (2.9) (A–F) 1D0X (2.0) (A) 1D0Z (2.0) (A) 1D1B (2.0) (A) 1DFK (4.2) (A) 1FMV (2.1) (A) 1G8X (2.8) (A,B) 1MMA (2.1) 1MMG (1.9) 1MND (2.6) 1VOM (1.9)	1BR1 (3.5) (A,C,E,G) 1BR4 (3.6) (A,C,E,G) 1D0Y (2.0) (A) 1D1A (2.0) (A) 1D1C (2.3) (A) 1DFL (4.2) (A,B) 1FMW (2.1) (A) 1LVK (1.9) 1MMD (2.0) 1MMN (2.1) 1MNE (2.7) 2MYS (2.8) (A)	G (26,15) E (7,6) S (10,28) G (28,12)	A (10,17) G (20,18) K (18,6) T (6,12)
012	GLRSDGKT 487 494	1FYZ (2.1) (A,B) 1FZ1 (1.9) (A,B) 1FZ3 (2.0) (A,B) 1FZ5 (2.4) (A,B) 1MMO (2.2) (E)	1FZ0 (2.0) (A,B) 1FZ2 (2.1) (A,B) 1FZ4 (2.3) (A,B) 1FZ7 (1.9) (A,B) 1MTY (1.7) (D,E)	G (2,5) L (4,4) R (4,4) S (8,10)	D (5,4) G (5,5) K (9,8) T (3,2)
012A	GLRSDGKT 487 494	1MMO (2.2) (D)	11111 (111) (23,2)		
013	GLSGSGKT 248 255	10EN (1.9)		6.02	D (2.4)
014	GTAFPGKT 212 219	1QPA (1.8) (A,B)		G (3,3) T (3,1) A (2,2) F (1,1)	P (2,4) G (4,0) K (2,3) T (1,5)
015	GKVTGGKT 102 109	1STE (2.0)		- (-,-)	1 (1,0)
016	GLPAIGKT 499 506	1BGX (2.3) (T) 1TAQ (2.4)	1CMW (2.6) (A)	G (4,12) L (16,5) P (2,20) A (12,11)	I (8,13) G (19,13) K (22,15) T (2,10)
016A	GLPAIGKT 499 506	1QSS (2.3) (A) 1QTM (2.3) (A) 3KTQ (2.3) (A)	1QSY (2.3) (A) 2KTQ (2.3) (A) 4KTQ (2.5) (A)	G (13,10) L (11,2) P (3,8) A (5,20)	I (15,8) G (4,4) K (2,4) T (5,7)

No.	Sequence Segment location	PDB code, (resolution in Å) chain identifier	Residue r.m.s. (ϕ, ψ)	Residue r.m.s. (ϕ, ψ)
017	GSQAGGKT 47 54	1WGT (1.9) (A,B)		G (4,1) S (1,1) Q (2,7) A (3,0)	G (9,4) G (5,7) K (4,1) T (2,0)
018	GPESSGKT 66 73	1G18 (3.8) (A) 2REB (2.3)	1G19 (3.0) (A)	G (5,6) P (6,1) E (4,18) S (20,22)	S (22,12) G (27,28) K (33,30) T (12,9)
019	GDVACGKT 12 19	1A2B (2.4) 1DPF (2.0) (A)	1CXZ (2.2) (A)	G (6,5) D (2,6) V (6,2) A (6,10)	C (6,0) G (1,7) K (9,1) T (1,4)
020	GDGGTGKT 7 24	1A2K (2.5) (C,D,E) 1IBR (2.3) (A,C) 1QG2 (2.5) (A) 1RRP (2.9) (A,C)	1BYU (2.1) (A,B) 1QBK (3.0) (C) 1QG4 (2.5) (A,B) 3RAN (2.1) (A-D)	G (12,8 D (6,10) G (8,5) G (4,13)	T (14,5) G (9,13) K (11,4) T (4,6)
)21	GDVAVGKT 210 217	1A4R (2.5) (A,B)		G (0,1) D (1,0) V (1,0) A (1,3)	V (4,1) G (3,3) K (3,3) T (2,0)
)22	GDGAVGKT 10 17	1AM4 (2.7) (D,E,F) 1DOA (2.6) (A) 1E96 (2.4) (A) 1G4U (2.3) (R) 1HE1 (2.0) (C,D)	1AN0 (2.8) (A,B) 1DS6 (2.3) (A) 1FOE (2.8) (B,D,F,H) 1GRN (2.1) (A) 1MH1 (1.3)	G (12,7) D (11,12) G (12,11) A (6,27)	V (26,5) G (6,12) K (11,6) T (4,5)
)23	GQTSSGKT 86 93	2NGR (1.9) (A) 1BG2 (1.8) 3KIN (3.1) (A,C)	2KIN (1.9) (A)	G (13,11) Q (15,6) T (7,4) S (6,14)	S (14,12) G (12,12) K (8,4) T (2,3)
024	GLPARGKT 45 52	1BIF (2.0) 3BIF (2.3) (A)	2BIF (2.4) (A,B)	G (4,3) L (2,2) P (1,5) A (3,4)	R (2,7) G (3,8) K (4,3) T (6,3)
025	GMDLKGKT 206 213	1BVU (2.5) (A–F)		G (16,25) M (26,9) D (12,6) L (4,12)	K (7,5) G (7,11) K (8,9) T (7,7)
026	GDGACGKT 12 19	1CC0 (5.0) (A,C)	1FTN (2.1) 1TX4 (1.6) (B)	G (1,4) D (3,9) G (3,1) A (7,8)	C (3,1) G (2,1) K (1,1) T (1,0)
)27	GLHAMGKT 24 31	1CP2 (1.9) (A,B)		G (3,2) L (1,4) H (2,3) A (3,5)	M (3,7) G (8,1) K (3,3) T (1,1)
)28	GAPANGKT 513 520	1CWV (2.3) (A)			
)29	GQTGSGKT 474 481	1CZ7 (2.9) (A–D) 3KAR (2.3)	2NCD (2.5) (A)	G (5,4) Q (4,4) T (3,13) G (11,11)	S (11,16) G (19,13) K (11,9) T (6,4)
030	GATGTGKT 39 46	1D2M (1.9) (A)	1D9Z (3.1) (A)	G (8,3) A (3,5) T (5,6) G (14,7)	T (3,2) G (2,15) K (17,13) T (13,1)
030A	GATGTGKT 39 46	1D9X (2.6) (A)			
031	GPPHSGKT 543 550	1D2N (1.7) (A)	1NSF (1.9)	G (1,1) P (0,1) P (1,1) H (1,2)	S (1,1) G (1,2) K (2,0) T (1,2)
0032	GEQAVGKT 18 25	1D5C (2.3) (A)		H (1,2)	T (1,2)

Table I. Co					
No.	Sequence Segment location	PDB code, (resolution in Å)	chain identifier	Residue r.m.s. (φ,ψ)	Residue r.m.s. (ϕ, ψ)
033	GPTGVGKT 57 64	1DO0 (3.0) (A–F) 1E94 (2.8) (E,F) 1G41 (2.3) (A) 1G4B (7.0) (E,F,K,L)	1DO2 (4.0) (A,C) 1G3I (3.4) (A–F) 1G4A (3.0) (E,F)	G (8,8) P (9,10) T (10,11) G (17,24)	V (19,19) G (13,20) K (18,11) T (13,10)
033A	GPTGVGKT 57 64	1DO2 (4.0) (B,D)		G (2,1) P (1,3) T (3,1) G (2,1)	V (1,1) G (1,2) K (1,2) T (1,1)
)33B	GPTGVGKT 57 64	1G31 (3.4) (S,T,U,V,W)		G (1,1) P (1,1) T (0,1) G (0,0)	V (0,1) G (1,1) K (0,0) T (0,0)
034	GTEFEGKT 44 51	1DT0 (2.1) (A,B,C)		G (0,1) T (2,1) E (1,1) F (1,2)	E (2,1) G (1,2) K (1,2) T (2,2)
035	GKGGVGKT 15 22	1F48 (2.3) (A)			
036	GLQGSGKT 105 112	1FFH (2.0) 2FFH (3.2) (A,B,C) 3NG1 (2.3) A,B	1NG1 (2.0) 2NG1 (2.0)	G (5,7) L (6,4) Q (4,5) G (10,19)	S (19,7) G (9,7) K (3,4) T (4,5)
037	GRPGTGKT 50 57	1FNN (2.0) (A,B)		G (2,1) R (2,4) P (2,2) G (1,7)	T (4,4) G (2,2) K (1,2) T (2,1)
038	GAPVDGKT 116 123	1FS7 (1.6) (A) 1FS9 (2.0) (A)	1FS8 (1.6) (A)	G (2,2) A (3,1) P (2,1) V (1,3)	D (1,2) G (1,0) K (2,0) T (2,2)
039	GVNGVGKT 300 307	1FTS (2.2)			
040	GLDNAGKT 24 31	1FZQ (1.7) (A)			
041	GPSGCGKT 36 43	1G29 (1.9) (1,2)		G (3,2) P (2,1) S (1,3) G (1,2)	C (1,1) G (5,5) K (1,1) T (2,0)
042	GGTGSGKT 178 185	1G6O (2.5) (A,B)		G (1,4) G (8,3) T (2,5) G (4,1)	S (2,2) G (1,3) K (3,1) T (0,3)
043	GPPGLGKT 45 52	1HQC (3.2) (A,B)		G (1,9) P (9,6) P (2,15) G (25,13)	L (5,8) G (8,11) K (16,4) T (0,2)
)44	GKGGTGKT 10 17	1HYQ (2.6) (A)		(23,13)	1 (0,2)
045	GKVTSGKT 102 109	1JCK (3.5) (B,D)		G (0,0) K (0,0) V (0,0) T (0,0)	S (0,0) G (0,0) K (0,0) T (0,0)
046	GARGCGKT 9 16	1SHK (1.9) (A,B)	2SHK (2.6) (A,B)	G (5,5) A (4,1) R (4,2) G (3,4)	C (7,2) G (2,8) K (7,3) T (4,1)
047	GLDRTGKT 12 19	1TMK (2.1) (A,B) 3TMK (2.0) (A-H)	2TMK (2.4) (A,B)	G (5,1) L (4,9) D (5,14) R (9,8)	T (9,9) G (13,10) K (7,4) T (2,5)

No.	Sequence Segment location	PDB code, (resolution in Å	chain identifier	Residue r.m.s. (ϕ, ψ)	Residue r.m.s. (ϕ, ψ)
048	GNSSVGKT 29 36	1ZBD (2.6) (A)	3RAB (2.0) (A)	G (5,2) N (1,2) S (3,3) S (2,3)	V (3,2) G (1,5) K (1,5) T (4,2)
)49	GLEGAGKT 10 17	4TMK (1.9) (A)	5TMP (1.9) (A)	G (3,1) L (2,2) E (3,3) G (2,9)	A (1,9) G (15,7) K (9,9) T (2,3)
050	GAGGVGKS 10 17	121P (1.5) 1CTQ (1.2) (A) 1GNQ (2.5) 1LFD (2.1) (B,D) 1QRA (1.6) (A) 221P (2.3) 5P21 (1.3) 6Q21 (1.9) (A–D)	1BKD (2.8) (R) 1GNP (2.7) 1GNR (1.8) 1Q21 (2.2) 1WQ1 (2.5) (R) 4Q21 (2.0) 621P (2.4) 721P (2.0)	G (6,10) A (13,7) G (9,11) G (10,10)	V (14,13) G (15,9) K (7,11) S (7,7)
051	GADGVGKS 10 17	1AGP (2.3)			
052	GAGESGKS 36 43	1AGR (2.8) (A,D) 1AZT (2.3) (A,B) 1BOF (2.2) 1CJK (3.0) (C) 1CJU (2.8) (C) 1CS4 (2.5) (C) 1FQJ (2.0) (A,D) 1GDD (2.2) 1GG2 (2.4) (A) 1GIL (2.3) 1GOT (2.0) (A) 1TAD (1.7) (A,B,C) 1TND (2.2) (A,B,C)	1AZS (2.3) (C) 1BH2 (2.1) 1CIP (1.5)(A) 1CJT (2.8) (C) 1CJV (3.0) (C) 1CUL (2.4) (C) 1FQK (2.3) (A,C) 1GFI (2.2) 1GIA (2.0) 1GIT (2.6) 1GP2 (2.3) (A) 1TAG (1.8)	G (14,5) A (7,5) G (7,7) E (8,8)	S (9,10) G (10,9) K (7,5) S (6,5)
)53	GIVSYGKS 211 218	1AU8 (1.9) (A)	1CGH (1.8) (A)	G (3,1) I (0,0) V (1,3) S (5,2)	Y (1,1) G (1,3) K (6,1) S (2,3)
054	GDGTGGKS 78 85	1CYN (1.8) (A)			
)55	GPSGTGKS 8 15	1EX6 (2.3) (A,B) 1GKY (2.0)	1EX7 (1.9) (A)	G (6,5) P (3,5) S (10,6) G (4,6)	T (7,19) G (27,7) K (5,3) S (4,9)
056	GSGGVGKS 10 17	1C1Y (1.9) (A) 1KAO (1.7) 3RAP (2.2) R,S	1GUA (2.0) (A) 2RAP (2.6)	G (5,6) S (5,4) G (5,7) G (7,10)	V (10,3) G (4,4) K (3,4) S (6,6)
057	GDTSDGKS 183 189	1HYL (1.8) (A,B)		G (1,5) D (3,2) T (2,2) S (5,6)	D (5,3) G (2,3) K (4,7) S (6,2)
)58	GTAFEGKS 44 51	1ISA (1.8) (A) 1ISC (1.8) (A)	1ISB (1.8) (A)	G (1,2) T (0,0) A (1,1) F (1,0)	E (1,0) G (1,4) K (2,0) S (0,1)
)58A	GTAFEGKS 44 51	1ISA (1.8) (B) 1ISC (1.8) (B)	1ISB (1.8) (B)	G (1,2) T (1,1) A (2,2) F (1,2)	E (2,1) G (2,2) K (3,2) S (3,1)
)59	GKGGIGKS 9 16	1CP2 (1.9) (A,B) 1FP6 (2.1) (A–D) 1G21 (3.0) (E–H) 1N2C (3.0) (E–H)	1DE0 (2.4) (A,B) 1G1M (2.2) (A,B) 1G5P (2.2) (A,B) 1NIP (2.9) (A,B)	G (17,11) K (14,17) G (16,19) G (12,7)	I (24,21) G (23,13) K (13,11) S (10,12)
059A	GKGGIGKS 9 16	2NIP (2.2) (A,B) 1G20 (2.2) (E,F)		G (8,9) K (13,6) G (8,1) G (2,7)	I (6,14) G (16,2) K (1,4) S (3,5)

		DDD 1 (1 1 1 2)			
No.	Sequence Segment location	PDB code, (resolution in Å) chain identifier	Residue r.m.s. (φ,ψ)	Residue r.m.s. (φ,ψ)
059B	GKGGIGKS 9 16	1G20 (2.2) (G,H)		G (12,7) K (9,4) G (19,30) G (1,22)	I (6,9) G (18,9) K (2,12) S (4,5)
060	GAVGVGKS 10 17	1HE8 (3.0) (B) 2Q21 (2.2)	1RVD (1.9) (A) 521P (2.6)	G (16,2) A (4,6) V (13,3) G (9,15)	V (13,9) G (8,10) K (10,10) S (8,9)
061	GMVVEGKS 190 197	1DJO (2.0) (A,B) 3PGA (2.) (1 – 4)	1DJP (1.9) (A,B) 4PGA (1.7) (A,B)	G (5,3) M (3,4) V (5,2) V (2,5)	E (9,6) G (10,12) K (6,4) S (5,3)
062	GARGVGKS 10 17	421P (2.2)		()-/	(2)2)
063	GIIAPGKS 539 546	1AHU (2.7) (A,B) 1AHZ (3.3) (A,B) 1E0Y (2.7) (A,B) 1E8G (2.1) (A,B) 1QLT (2.2) (A,B) 1VAO (2.5) (A,B)	1AHV (3.1) (A,B) 1DZN (2.8) (A,B) 1E8F (2.9) (A,B) 1E8H (2.6) (A,B) 1QLU (2.4) (A,B) 2VAO (2.8) (A,B)	G (8,13) I (14,12) I (6,5) A (7,11)	P (8,6) G (6,7) K (5,6) S (5,6)
064	GAVESGKS 40 47	1AS0 (2.0) 1AS3 (2.4)	1AS2 (2.8)	G (3,1) A (1,4) V (7,3) E (2,3)	S (4,3) G (6,3) K (2,3) S (4,3)
065	GSSGSGKS 39 46	1B0U (1.5) (A)		L (2,3)	3 (4,5)
066	GVRFPGKS 313 320	1BAG (2.5)			
067	GGARSGKS 406 413	1C9K (2.2) (A,B,C)	1CBU (2.3) (A,B,C)	G (7,9) G (7,10) A (8,7) R (6,5)	S (5,6) G (5,7) K (7,3) S (6,10)
068	GFAKTGKS 195 202	1CA1 (1.9) 1QMD (2.2) (A,B)	1QM6 (2.5) (A,B)	G (1,3) F (2,1) A (2,3) K (1,1)	T (2,1) G (2,3) K (2,1) S (1,2)
069	GLLEAGKS 174 181	1CD1 (2.6) (A,C)		G (9,4) L (6,3) L (5,5) E (6,12)	A (13,4) G (10,21) K (16,3) S (3,6)
070	GAPGVGKS 10 17	1CLU (1.7) (A) 1JAI (1.8)	1JAH (1.8) 821P (1.5)	G (5,2) A (4,3) P (5,2) G (8,3)	V (2,2) G (4,3) K (3,2) S (3,3)
071	GGSCTGKS 434 441	1CLV (2.0) (A) 1TMQ (2.5) (A)	1JAE (1.6) 1VIW (3.0) (A)	G (5,13) G (7,8) S (5,7) C (11,2)	T (5,8) G (8,28) K (21,16) S (14,32)
072	GRSGRGKS 138 145	1HJB (3.0) (C,F) 1IO4 (3.0) (C)	1HJC (2.6) (A,D)	G (8,4) R (11,9) S (12,12) G (14,5)	R (3,6) G (5,11) K (11,3) S (4,3)
073	GPLYLGKS 587 594	1CMX (2.2) (A,C)		G (0,1) P (1,1) L (0,2) Y (2,2)	L (2,3) G (3,1) K (1,2) S (3,0)
074	GLSASGKS 32 39	1D6J (2.0) (A,B)		G (5,2) L (1,2) S (2,1) A (4,7)	S (5,4) G (1,1) K (2,3) S (4,2)
075	GQAMPGKS 105 112	1DDZ (2.2) (A,B)		G (1,1) Q (0,0) A (1,0) M (1,0)	P (1,1) G (1,1) K (0,1) S (1,1)

No.	Sequence Segment location	PDB code, (resolution in Å) chain identifier	Residue r.m.s. (φ,ψ)	Residue r.m.s. (φ,ψ)
076	GVVQPGKS 510 517	1DEE (2.7) (B,D,F)		G (1,1) V (1,1) V (0,2)	P (2,1) G (1,2) K (1,0)
077	GLYRTGKS	1DG3 (1.8) (A)		Q (5,1)	S (2,3)
)77A	45 52 GLYRTGKS 45 52	1F5N (1.7) (A)			
078	GENGIGKS 42 49	1DYW (1.8) (A)	1E3B (1.8) (A)	G (2,2) E (2,1) N (3,2) G (3,3)	I (1,1) G (0,2) K (2,2) S (4,2)
79	GRPNVGKS 15 22	1EGA (2.4) (A,B)		G (2,0) R (2,3) P (2,1) N (1,3)	V (4,3) G (4,1) K (2,1) S (1,1)
80	GAAAAGKS 893 900	1EJ6 (3.6) (A)			
081	GEAAVGKS 14 21	1EK0 (1.4) (A)			
082	GSVAVGKS 95 102	1ESM (2.5) (A–D)	1ESN (2.6) (A–D)	G (3,5) S (13,14) V (4,2) A (13,10)	V (3,2) G (8,3) K (6,17) S (8,10)
183	GAEAAGKS 188 195	1F07 (2.0) (A–D)		G (1,3) A (4,2) E (3,4) A (4,2)	A (4,4) G (4,2) K (2,4) S (5,3)
84	GQNGSGKS 30 37	1F2T (1.6) (A)	1F2U (1.6) (A,C)	G (13,3) Q (3,12) N (8,4) G (16,14)	S (13,8) G (7,3) K (7,13) S (2,6)
85	GHVDSGKS 14 21	1F60 (1.6) (A)	1G7C (2.0) (A)	G (0,1) H (5,4) V (3,4) D (0,2)	S (4,6) G (5,3) K (1,1) S (1,3)
86	GDSGVGKS 27 34	1G16 (1.8) (A–D)	1G17 (2.0) (A,B)	G (3,3) D (3,3) S (1,5) G (4,4)	V (4,2) G (2,3) K (3,2) S (2,2)
87	GLVSPGKS 951 958	1HQM (3.3) (C)		,	,
88	GESAVGKS 28 35	1HUQ (1.8) (A)			
89	GIPGVGKS 8 15	1NKS (2.5) (A–F)		G (7,2) I (4,3) P (4,7) G (12,11)	V (10,8) G (12,9) K (6,3) S (5,8)
90	GDVSPGKS 457 464	1QHB (2.3) (A–F)		G (2,2) D (2,2) V (1,2) S (1,1)	P (2,1) G (3,2) K (3,3) S (4,3)
91	GGNGAGKS 34 41	1QHL (2.2) (A)		- () - /	- (- ,-)
092	GGSSAGKS 10 17	1QHN (2.7) (A) 1QHX (2.5) (A)	1QHS (2.8) (A) 1QHY (2.6) (A)	G (3,2) G (4,3) S (3,2) S (2,1)	A (1,2) G (4,3) K (3,1) S (4,5)

For those groups which have more than one entry, structural similarity is brought out by the small r.m.s. values of the Ramachandran angles (ϕ, ψ) , given in the last column.

059 – GKGGIGKS); and four for one sequence (003 – GGAGVGKT). These data implied that highly localized conformational variants are possible in these segments retaining overall structural similarity.

Conformational variants of segments of Walker sequences The next step was the grouping of the conformations irrespective of the sequence of the variable region of the Walker segment. This was done as follows: (1) for those

Table II. The entries selected from Table I regrouped based on their structural similarity [the examples in set VII do not possess any structural similarity, as analyzed using the Ramachandran angles (ϕ, ψ)]

2 G D R O T G K T 169-176	the protein
2 G D R Q T C R T 169-176 1E79 (A) F, ATPas 3 G P E S S G K T 66-73 2REB REC A p 4 G L P A R G K T 45-52 1BH 6-Phosph 5 G Q T G S G K T 474-481 3KAR Kinesin-16 G G A T G T G K T 36-43 1D2M (A) UvrB pro 7 G P P H S G K T 55-15-58 1D2M (A) UvrB pro 7 G P P H S G K T 55-15-58 1D2M (A) W-Ethylm 8 G P T G V G K T 55-15-58 1D2M (A) Heat shot 9 G L Q G S G K T 105-112 1FFH Signal rec 10 G R P C T G K T 50-57 1FNN (A) CD-C pro 11 G L D M A G K T 24-31 1F2Q (A) ADP-ribo 12 G C R P C T G K T 24-31 1F2Q (A) ADP-ribo 14 G C R P C T G K T 24-31 1F2Q (A) ADP-ribo 14 G C R P G T G K T 105-112 1FFH Signal rec 10 G C R P G T G K T 105-112 1FFH Signal rec 10 G C R P G T G K T 105-112 1FFH Signal rec 10 G C R P G T G K T 105-117 1FXQ (A) ADP-ribo 14 G C G K T 105-117 1FXQ (A) ADP-ribo 15 G F S G T G K S 8-15 1EXT (A) Guanylate 15 G C G K T 105-17 1FXQ (A) Thymicly 15 G G F S G T G K S 8-15 1EXT (A) Guanylate 15 G G S G G G K S 105-17 1KAQ (A) Nitrogena 15 G G G G G G G G G G G G G G G G G G	
Q P E S S G K T	nol pyruvate kinase
G L P A R G K T	e α subunit
G Q T G S G K T G G T G T G K T G G T G T G K T G G T G T G K T G G P T G K G K T G G P T G K G K T G G P T G K G K T G P T G K G K T G P T G K G K T G P T G K G K T G P T G K G K T G P T G K G K T G P T G K G K T G R P G T G K S G R S G G K S G G G G K T G R S G G K S G G G G K S G G G G K S G G G G K S G G G G K S G G G G K S G G G G K S G G G G K S G G G G K S G G G G K S G G G G K S G G G G K S G G G G K S G G G G K S G G G G K S G G G G K S G G G G K S G G G G K S G G G G G K S G G G G K S G G G G K S G G G G K S G G G G K S G G G G K S G G G G K S G G G G K S G G G G K S G G G G K S G G G G K S G G G G K S G G K S G G G G K S G G K S G G G G K S G G K S G G G G K S G G K S G G G K S G G K S G G G K S G	rotein.
G A T G T G K T G F P H S G K T G F P H S G K T G F P G Y G K T G F P G Y G K T G F P G Y G K T G F P G Y G K T G F P G Y G K T G F C G G G K T G F C G K T G F C G K T G F C G K T G F C G K T G F C G K T G F C G K T G F C G K T G F C G K T G F C G K T G F C G G K T G F C G C G K T G F C G C G K T G F C G C G K T G F C G C G K T G F C G G G K T G F C G G G K T G F C G G G K T G F C G G G K T G F C G G G K T G F C G G G K T G F C G G G K T G F C G G G K T G F C G G G K T G F C G G G G K T G F C G G G G K T G F C G G G G K T G F C G G G G K T G F C G G G G K T G F C G G G G K S G F C G G K S G F C G G F C G C G K S G F C G G F C G C G C C C C C C C C C C	ofructo-2-kinase/fructose-2,6-bisphosphatase
GATGTGKT GPPHSGKT 551-558 ID2N (A) GPPHSGKT 551-558 ID2N (A) GPPTGVGKT GPPGGKT 551-558 ID2N (A) GPTGVGKT GLQGSGKT ID5-112 IFFH Signal rec GRPGTGKT SO-57 IFNN (A) CDCGp GLDNAGKT 24-31 IFZQ (A) APP-rbo GPSGCGKT GLDNAGKT GLDNAGKT GLDRTGKT GLDRTGKT GLDRTGKT ID-19 3TMK (A) Thymidyl GLEGAGKT ID-17 IKAO SMIGG GSGGVGKS ID-17 IKAO SMIGG GSGGVGKS ID-17 IKAO SMIGG GSGGVGKS GSGGGGGGGGGGGGGGGGGGGGGGGGGG	ke protein KAR3
GPPHSGRT S51-558 ID2N (A) M-Elnylm	*
G P T G V G K T G L Q G S G K T G L Q G S G K T G L Q G S G K T G L Q G S G K T G L Q G S G K T G L D N A G K T G L D N A G K T G L D N A G K T G L D N A G K T G L D N A G K T G L D N A G K T G L D N A G K T G L D N A G K T G L D N A G K T G L D N A G K T G L D N A G K T G L D N A G K T G L D R T G C T G L D R T G C T G L D R T G C	aleimide-sensitive fusion protein
GLQGSGKT 105-112 1FFH Signal recommendation of the commendation of	ek protein HslU
GRPGTGKT 50-57 GLDNAGKT 24-31 GEDNAGKT 24-31 GPQ (A) GPSGCGKT 36-43 GPSGCGKT 12-19 GLDRTGKT 12-19 GLDRTGKT 12-19 GLDRTGKT 12-17 GLEGAGKT 10-17 GLEGAGKT 10-17 HKAO 5mall GP GKGGTGKS 10-17 IKAO 5mall GP GKGGTGKS 8-15 GKGGTGKS 10-17 IKAO 5mall GP GKGGTGKS 8-15 GKGGTGKS 10-17 IKAO 5mall GP GKGGTGKS 45-52 IFSN (A) Guanylate GKGGTGKS 45-15 GKAO Adenosyl GLYRTGKS 45-52 IFSN (A) GAGNOSY GLYRTGKS 45-52 IFSN (A) GAGNOSY GRANGKGKS 30-37 IFT (A)	cognition protein FFH
GLDNAGKT 24-31 IFZQ (A) ADP-fibe GPSGCGKT 36-43 IG29 (I) Mal K GLDRTGKT 12-19 3TMK (A) Thymidyl GLEGAGKT 10-17 4TMK (A) Thymidyl GPSGCGKS 8-15 IEXT (A) Guanylate GSGGYGKS 10-17 IKAO Small G- GKGGTGKS 8-15 ICCP (A) Nitrogens GSSGSGKS 39-46 IBOU (A) Histidine GGARSGKS 6-13 ICCW (A) Adenoylat GLYRTGKS 45-52 IFSN (A) Guanylate GLYRTGKS 45-52 IFSN (A) Guanylate GCARSGKS 14-21 IEKO (A) YPT51 GDSGCKS 27-34 IGI6 (A) Sect GGNGSGKS 30-37 IFZT (A) Rad50 Al GGSSAGKS 27-34 IGI6 (A) Sect GFSAVGKS 28-35 IHUQ (A) Rab5c GIPGYGKS 8-15 INKS (A) Adenoylate GGSSAGKS 10-17 IQHX (A) Chlorang GHYDHGKT 18-25 IEXM (A) EF-Tu GPHGMGKT 56-63 IEZK (A) Thymidia GGSGCGCKT 179-186 IMMG Myosin n GDGGTGKT 17-24 IBYU (A) RAN-GT GDGAYGKT 12-19 ITX4 (B) Rho A GGGAYGKT 12-19 ITX4 (B) Rho A GGGAGGCKT 12-19 ITX4 (B) Rho A GGGAGGCKT 12-19 ITX4 (B) Rho A GGGAGGCKT 15-22 IFAB (A) Arsenite 1 GGGAGGCKT 15-22 IFAB (A) Arsenite 1 GGAGGCKT 178-185 IGGO (A) Traffic A' GAGGGCKT 19-16 ISHK (A) Shikimate GGAGGCKT 19-17 ICTQ (A) P21 Ras GGAGGCKT 19-16 ISHK (A) Shikimate GGAGGCKT 19-17 ICTQ (A) P21 Ras GGAGGCKT 10-17 ICTQ (A) P21 Ras GGAGGCKT 10-17 ICTQ (A) P21 Ras GGAGGCKT 15-22 IEGM (A) GTAFIC A' GGAGGCKT 15-22 IEGM (A) GTAFIC A' GCAGGCKT 15-22 IEGM (A) GTAFIC A' GCAGGCKT 15-22 IEGM (A) GTAFIC A' GCAGGCKT 15-22 IEGM (A) GTAFIC A' GCAGGCCKT 15-24 IEGM (A) GTAFIC A' GCAGGCCCKT 1	ogmuon protein 1111
G P S G C G K T G L D R T G K T G L D R T G K T G L D R T G K T G L D R T G K T G L D R T G K T G L D R T G K T G L D R T G K T G L E G A G K T G L E G A G K T G P S G T G K S 8-15 G S G G G V G K S 10-17 1 KAO Small G G G K G G I G K S 8-15 I LCP2 (A) Nitrogene G S G G G K S G S G G G K S G S G G G K S G G A R S G K S G L Y R T G K S G G A R S G K S G G A R S G K S G L Y R T G K S G S G G G K S G D Y R T G K S G D Y D H G K T G D G G T G K T G D G G T G K T G D G G T G K T G D G G T G K T G D G G T G K T G D G G T G K T G D G G T G K T G D G G T G K T G D G G T G K T G D G G T G K T G D G G T G	sylation-like factor
C L D R T G K T	sylution like factor
G L E G A G K T G P S G T G K S G S G G V G K S G S G G V G K S G S G G V G K S G S G G V G K S G S G G V G K S G S G G V G K S G S G G G K S G G G G G K S G G G G G K S G G G G G K S G G G G K S G G G G K S G G G G K S G G G G K S G G G G K S G G G K S G G A R S G K S G G S G K S G G S G K S G G S G K S G G S G K S G G S G K S G G S G K S G G S G K S G G S G K S G G S G K S G G G S G K S G G G G G G K S G G G G G G G G G G G G G G G G G G G	ate kinase (S.cerevisiae)
G P S G T G K S G S G G V G K S G S G G V G K S G S G G V G K S G S G G V G K S G S G G K S G S G G K S G S G G K S G S G G K S G G G G G K S G G G G K S G G G G K S G G G G K S G G G G G K S G G G G G K S G G G G G K S G G G G G K S G G G G G K S G G G A R S G K S G G G A R S G K S G G A R S G K S G G L Y R T G K S G E A A V G K S G E A A V G K S G E A A V G K S G E A A V G K S G E A S G K S G E A S G K S G E A S G K S G E A S G K S G E A S G K S G E A S G K S G E A S G K S G E A S G K S G E S A V G K S	
G S G G V G K S	
C K G G I G K S S-15 ICP2 (A) Nitrogena G S S G S G K S 39-46 1B0U (A) Histidine G G A R S G K S 6-13 IC9K (A) Adenosyl G L Y R T G K S 45-52 IF5N (A) Guanylate G E A A V G K S 14-21 IEKO (A) YPT51 G O N G S G K S 30-37 IF2T (A) Rad50 (A) G D S G V G K S 27-34 IGIG (A) Sec4 G E S A V G K S 28-35 IHUQ (A) Rad50 (A) G G S G K S G E S A V G K S 28-35 IHUQ (A) Rad50 (A) G G S G K S G F Y G V G K S 8-15 INKS (A) Adenylate G G S S A G K S 10-17 IQHX (A) Chloramp G H V D H G K T 18-25 IEXM (A) IEF-Tu G P H G M G K T 179-186 IMMG Myosin n G D G G T G K T 179-186 IMMG Myosin n G D G G T G K T 179-186 IMMG Myosin n G D G G T G K T 17-24 IB YU (A) RAN-GT G D G A C G K T 12-19 ITX4 (B) Rho A G G G G V G K T 18-25 ID5C (A) Rab56 + G G G V G K T 178-185 IG6O (A) Traffic A' G R G G V G K T 300-307 IFTS Signal red G T G G G G G G G G G G G G G G G G G	
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G N S S V G K T G N S S V G K T G A G G V G K S G A G G V G K S G A G E S G K S G A G E S G K S G R P N V G K S G S V A V G K S G G G G G V G K T G G S V A V G K S G G G G G V G K T G G G G G V G K T G G G G G V G K T G G G G G V G K T G G G G G G V G K T G G G G G G V G K T G G G G G G V G K T G G G G G G G G G G G G G G G G G G G	
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G S V A V G K S G G A G V G K T G G A G V G K T G L D A A G K T G L D A G T G L D A G T G L D A G T G L D A G T G L D A G T G L D A G T G L D A G T G L D A G T G	
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G K G G I G K S G L Y R T G K S G L Y R T G K S G L Y R T G K S Hhal met G M V V E G K S G A E A A G K S G A E A A G K S G L L E A G K S G A T G T G K T G A P P G L G K T G A P P G L G K T G A F A G K T G A F A G K S G A F A G	
G L Y R T G K S 45–52 1DG3 (A) Guanylate t III G Y L V N G K T G M V V E G K S 190–197 4PGA (A) Glutamina t IV G L H A M G K T G A E A A G K S 188–195 1F07 (A) Tetrahydr t V G F A K T G K S 195–202 1 CA1 G L E A G K S 174–181 1 CD1 (A) CD1 t VI G A T G T G K T G P P G L G K T 45–52 1DG3 (A) Guanylate 6MHT (A) Hhal met 6MHT (A) Hhal met 6MHT (A) Hhal met 6 Glutamina 1 CP2 (A) Nitrogena 1 Tetrahydr 1 Tetrahydr 1 UVI G A T G T G K T 39–46 G P P G L G K T 45–52 1 HQC (A) RuvB	
GYLVNGKT 264–271 6MHT (A) HhaI met GMVVEGKS 190–197 4PGA (A) Glutamin: t IV GLHAMGKT 24–31 1CP2 (A) Nitrogena GAEAAGKS 188–195 1F07 (A) Tetrahydr t V GFAKTGKS 195–202 1CA1 Alpha-tox GLLEAGKS 174–181 1CD1 (A) CD1 t VI GATGTGKT 39–46 1D9X (A) UvrB GPPGLGKT 45–52 1HQC (A) RuvB	
GYLVNGKT GMVVEGKS 190–197 4PGA (A) Glutamina t IV GLHAMGKT GAEAAGKS 188–195 1F07 (A) Tetrahydr t V GFAKTGKS 195–202 1CA1 GLLEAGKS 174–181 1CD1 (A) CD1 t VI GATGTGKT 39–46 GPPGLGKT 45–52 1HQC (A) Hhal met APGA (A) Hhal met APGA (A) Glutamina Tetrahydr	e binding protein-1
GMVVEGKS 190–197 4PGA (A) Glutamina t IV GLHAMGKT 24–31 1CP2 (A) Nitrogena GAEAAGKS 188–195 1F07 (A) Tetrahydr t V GFAKTGKS 195–202 1CA1 Alpha-tox GLLEAGKS 174–181 1CD1 (A) CD1 t VI GATGTGKT 39–46 1D9X (A) UvrB GPPGLGKT 45–52 1HQC (A) RuvB	
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### ### ##############################	ase–asparaginase
G L H A M G K T G A E A A G K S 188–195 1F07 (A) Tetrahydr V G F A K T G K S G L L E A G K S 195–202 1CA1 Alpha-tox G L L E A G K S 174–181 1CD1 (A) CD1 VV G A T G T G K T G P P G L G K T 45–52 1HQC (A) Nitrogena Tetrahydr 1EVA 1	ase asparaginase
GAEAAGKS 188–195 1F07 (A) Tetrahydr GFAKTGKS 195–202 1CA1 Alpha-tox GLLEAGKS 174–181 1CD1 (A) CD1 t VI GATGTGKT 39–46 1D9X (A) UvrB GPPGLGKT 45–52 1HQC (A) RuvB	
GFAKTGKS 195-202 1CA1 Alpha-tox GLLEAGKS 174-181 1CD1 (A) CD1 t VI GATGTGKT 39-46 1D9X (A) UvrB GPPGLGKT 45-52 1HQC (A) RuvB	se iron protein
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GATGTGKT 39-46 1D9X (A) UvrB GPPGLGKT 45-52 1HQC (A) RuvB	
G A T G T G K T 39–46 1D9X (A) UvrB G P P G L G K T 45–52 1HQC (A) RuvB	
G P P G L G K T 45–52 1HQC (A) RuvB	
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	- 0
	e β subunit (bovine) e β subunit (bovine)

Table II. Continued

Sr. No.	Sequence of Walker motif	Segment location	PDB code (chain)	Name of the protein
61	GGAGVGKT	156–163	1MAB (B)	F ₁ -ATPase β subunit (rat)
62	GAHALGKT	173-180	2CYP	Cytochrome c peroxidase
63	GAGALGKT	173-180	1DS4 (A)	Cytochrome c peroxidase
64	GHVDHGKT	18-25	1ETU	EF-Tu domain 1
65	GTAFPGKT	212-219	1QPA (A)	Lignin peroxidase
66	GLRSDGKT	487-494	1MTY (D)	Methane monoxygenase (Mc) ^a
67	GLRSDGKT	487-494	1MMO (D)	Methane monoxygenase (Mt) ^a
68	GKVTGGKT	102-109	1STE	Sec2 superantigen
69	GLPAIGKT	499-506	1BGX (T)	Taq polymerase
70	GLPAIGKT	499-506	1QSS (A)	Taq Klenow fragment
71	GSQAGGKT	47–54	1WGT (A)	Wheat germ agglutinin
72	GMDLKGKT	206-213	1BVU (A)	Glutamate dehydrogenase
73	GAPANGKT	513-520	1CWV (A)	Invasin
74	GPTGVGKT	57–64	1DO2 (B)	HslU
75	GPTGVGKT	57–64	1G3I (S)	HslU protease
76	GAPVDGKT	116-123	1FS7 (A)	Cytochrome <i>c</i> nitrite reductase
77	GKGGTGKT	10–17	1HYQ (A)	MinD-1
78	GKVTSGKT	102-109	1JCK (B)	Sec3 superantigen
79	GIVSYGKS	211-218	1CGH (A)	Cathepsin G
80	GDGTGGKS	78–85	1CYN (A)	Cyclophilin B
81	GDTSDGKS	183-189	1HYL (A)	Collagenase
82	GTAFEGKS	44-51	1ISA (A)	Superoxide dismutase
83	GTAFEGKS	44-51	1ISA (B)	Superoxide dismutase
84	GKGGIGKS	9–16	1G20 (E)	Nitrogenase iron protein
85	GIIAPGKS	539-546	1E8G (A)	Vanillyl-alchohol oxidase
86	GVRFPGKS	313-320	1BAG	α-Amylase (B.subtilis)
87	GGSCTGKS	434-441	1JAE	α-Amylase (yellow mealworm)
88	GRSGRGKS	138-145	1HJC (A)	RUNT-related transcription factor-1
89	GPLYLGKS	187-194	1CMX (A)	Ubiquitin
90	GQAMPGKS	105-112	1DDZ (A)	Carbonic anhydrase
91	GVVQPGKS	510-517	1DEE (B)	IgM heavy chain
92	GENGIGKS	42-49	1DYW (A)	Cylophilin 3
93	GAAAGKS	893-900	1EJ6 (A)	Reovirus core protein λ2
94	GLVSPGKS	951-958	1HQM (C)	Bacterial RNA polymerase β subunit
95	GDVSPGKS	457-464	1QHB (A)	Vanadium bromo-peroxidase
96	GGNGAGKS	34-41	1QHL (A)	MukB N-terminal domain

 $^{{}^}a\!M\!c,\, \textit{Methylococcus capsulatus};\, Mt,\, \textit{Methylosinus trichosporum}$

Table III. Mean and r.m.s. (φ,ψ) values $(^\circ)$ for the first six sets given in Table II

	Position No.									
	1	2	3	4	5	6	7	8		
Set I [45] Mean (φ,ψ) R.m.s.(φ,ψ)	(159,166) (18,11)	(-67,162) (10,10)	(-63,141) (8,11)	(79,12) (14,15)	(-87,-11) (17,14)	(104,20) (16,11)	(-60,-46) (7,10)	(-66,-40) (5,5)		
Set II [5] Mean (ϕ, ψ) R.m.s. (ϕ, ψ)	(158,171) (11,7)	(-66,157) (7,8)	(-60,-36) (13,7)	(-82,6) (19,15)	(-86,-5) (20,14)	(73,29) (12,13)	(-63,-32) (5,9)	(-66,-43) (4,8)		
Set III [2] Mean (ϕ, ψ) R.m.s. (ϕ, ψ)	(-138,176) (35,25)	(-138,154) (2,6)	(-106,114) (17,8)	(-127,125) (8,4)	(41,54) (1,3)	(84,–11) (3,5)	(-115,149) (10,13)	(-104,134) (7,7)		
Set IV [2] Mean (ϕ, ψ) R.m.s. (ϕ, ψ)	(-63,-45) (4,4)	(-62,-44) (9,7)	(-65,-38) (1,3)	(-62,-21) (3,9)	(-97,7) (7,8)	(82,16) (2,2)	(-99,163) (12,19)	(-114,141) (13,16)		
Set V [2] Mean (φ,ψ) R.m.s.(φ,ψ)	(-52,-47) (8,2)	(-67,-45) (4,6)	(-61,-28) (5,7)	(-76,-44) (7,5)	(-73,-36) (8,2)	(-71,-34) (4,12)	(-44,-38) (17,3)	(-63,-49) (1,1)		
Set VI [2] Mean (φ,ψ) R.m.s.(φ,ψ)	(-141,138) (8,2)	(-65,151) (11,3)	(-62,147) (6,14)	(89,–2) (3,32)	(-95,144) (21,29)	(-70,85) (23,18)	(-78,-40) (26,9)	(-59,-52) (11,4)		

The corresponding values do not have any meaning for set VII, which has structurally dissimilar conformations. The number of examples in each set is given in parentheses along with the set number in the first column.

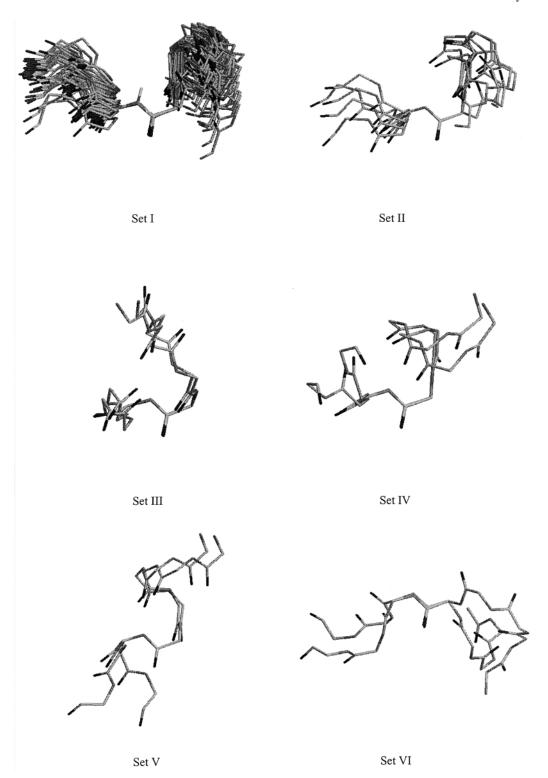
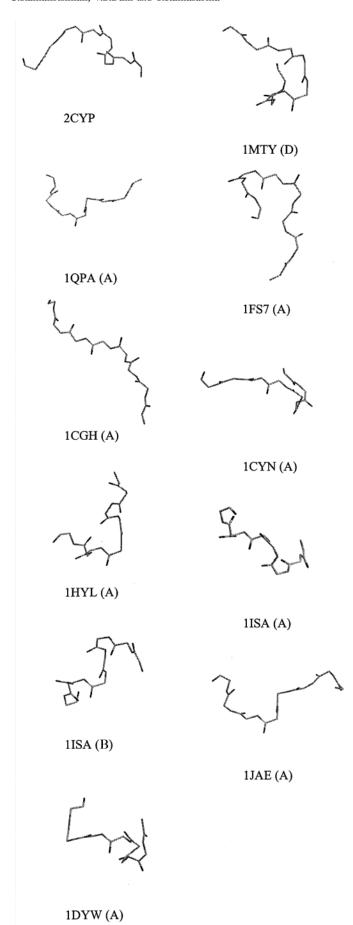


Fig. 1. Wire-frame diagrams of the backbone atoms in the segment GXXXXGKT (S) in the proteins given in sets I-VI of Table II.

groups in Table I which had only one entry, the choice was unambiguous; (2) for those groups having more than one entry, one with the best resolution shown in bold face in Table I had been picked up as the representative of the group/sub-group. These collectively gave 107 examples which were regrouped solely on the basis of similarity of the Ramachandran angles (ϕ, ψ) . Of the new sets thus

obtained, 53 (out of a total of 107) entries constituted the major set. Another set had five entries, while seven others had two entries each. The last set comprised 35 entries, without any structural similarity among them. In any particular set, proteins with high overall sequence homology could be found, although the sequences of the variable region were different. These are as follows: (1AYL, 1OEN);



(1A4R, 1MH1); (1DPF, 1TX4); (1CIP, 1AS0), (1AGP, 1CTQ, 1RVD, 421P, 821P); pairs [(2CYP, 1CCG); (1MHY (D), 1MTY (D)], as well as [1DT0 (A), 1ISA (B)]. Since the structures in such cases are expected to be similar, the entries that had the best resolution were retained. These were 1AYL, 1MH1, 1TX4, 1CIP, 1CTQ, 2CYP, 1MTY (D) and 1ISA (B). The final grouping thus obtained is given in Table II, which has 45 proteins in set I, five in set II, two each in sets III, IV, V and VI and the remaining 38 in set VII. The mean and r.m.s. (ϕ, ψ) values of sets I–VI are given in Table III and these are small enough to warrant structural similarity among the members. The r.m.s. has no relevance for the last set (set VII).

For easy comprehension of the structural grouping, the line diagrams of the backbone of GXXXXGKT (S) segments of the proteins in sets I–VI, drawn with the peptide unit spanning residues 5–(X) and 6–(G) as the common internal frame of reference, are shown in Figure 1. The sickle-like folding with overlap of the atoms of the backbone is seen with members of the set I (Figure 1). Nearly the same structure of segment 5–8 is found in set II, but that of segment 1–4 is different (Table III). Set VII is comprised of structures of differing conformations indicating the flexibility of Walker sequences to acquire random folding. Out of the 38 examples in this set, only those which have resolution of 1.8 Å or better are shown in Figure 2.

Differing structures with same Walker sequence

Structural differences between segments having the same Walker sequence are also perceivable from the foregoing data. There are nine such examples in the present data set. The PDB codes along with the Ramachandran angles at the eight positions of Walker sequence of these nine pairs are given in Table IV. Large differences in Ramachandran angles are observed at four different locations within the segment (shown in bold face in the table). These are as follows: (i) 1VOM–2MYS (A), (ii) 1F5N (A)–1DG3 (A), (iii) 1FP6 (A)–1G20 (E) and (iv) 1EFT–1EFU (A) all at locations 3/4; (v) 1MMO (D)–1MMO (E) at locations 4/5; (vi) 1ISA (A)–1ISA (B), (vii) 1D9X (A)–1D9Z (A) and (viii) 1BMF (D)–1BMF (E) at locations 5/6; and (ix) 1G3I (A)–1G3I (S) at locations 6/7. These large changes arise owing to a flip of the peptide unit spanning the two residues.

There are more than two examples with differing conformations for two of the sequences. The first example consists of 1EFT, 1EFU (A) and 1ETU, which have the same sequence, GHVDHGKT (iv in Table IV), but the peptide unit between residues 3 and 4 in 1EFT and 1EFU (A) is flipped. The conformation of the third member of this group, 1ETU, does not match in entirety with the other two examples. A close examination reveals that the conformations of 1EFT and 1ETU differ only in the segment 18–20. The second example is of 1BMF (D, E), 1MAB (B) and 1E1R (E) having the same sequence GGAGVGKT. The peptide unit between locations 5 and 6 in 1BMF (E) and 1BMF (D) is flipped, as also is the one between locations 6 and 7 in 1BMF (D) and 1MAB (B) (viii in

Fig. 2. Wire-frame diagrams of the backbone atoms in the segment GXXXXGKT (S) in the proteins belonging to set VII of Table II. Only those examples occurring in protein structures which have a resolution of 1.8 Å or better are shown.

Table IV Ramachandran angles (ϕ, ψ) (°) in the segment GXXXXGKT for those examples with same sequence for XXXX but with different conformations

No.	Sequence, PDB code (chain) and	(φ,ψ) at posit	ion						
	segment locations	1	2	3	4	5	6	7	8
(i)	GESGAGKT, 1VOM, 179–186	(150,155)	(-73,171)	(-59,131)	(86,4)	(-67,-26)	(117,18)	(-61,-47)	(-67,-37)
	GESGAGKT, 2MYS(A), 179–186	(178,174)	(-72,166)	(-35,-42)	(-101,2)	(-61,-25)	(82,57)	(-82,-47)	(-44,-48)
(ii)	GLYRTGKS, 1F5N(A), 45–52	(155,173)	(-53,147)	(-63,157)	(60,34)	(-111,13)	(79,28)	(-66,-52)	(-60,-41)
	GLYRTGKS, 1DG3(A), 45–52	(149,178)	(-75,161)	(-60,-42)	(-54,-22)	(-71,-14)	(65,31)	(-67,-17)	(-60,-32)
(iii)	GKGGIGKS, 1FP6(A), 9–16	(155,176)	(-64,163)	(-73,131)	(74,13)	(-79,-24)	(130,20)	(-75,-25)	(-76,-35)
	GKGGIGKS, 1G20(E), 9–16	(137,156)	(-53,138)	(-80,-35)	(-62,10)	(-97,-49)	(134,39)	(-61,-46)	(-70,-32)
(iv)	GHVDHGKT, 1EFT, 18–25	(147,165)	(-65,164)	(-66,133)	(75,18)	(-91,-20)	(125,31)	(-67,-46)	(-73,-34)
	GHVDHGKT, 1EFU(A), 18–25	(175,176)	(-63,141)	(-45,-41)	(-102,24)	(-101,4)	(84,40)	(-68,-44)	(-67,-47)
	GHVDHGKT, 1ETU, 18–25	(-126, -166)	(-120,-41)	(143,82)	(133,-27)	(-47,-37)	(119,10)	(-49,-47)	(-57,-46)
(v)	GLRSDGKT, 1MMO(D), 487–494	(-80,42)	(-125,158)	(-79,161)	(-62,137)	(125,-29)	(74,23)	(-135,-59)	(-69,136)
	GLRSDGKT, 1MMO(E) ^a , 487–494	(-84,50)	(-129,148)	(-70,170)	(-70,-1)	(-86,-2)	(65,27)	(-145,-53)	(-80,138)
(vi)	GTAFEGKT, 1ISA(A), 44–51	(81,8)	(-112,173)	(-70,-13)	(-74,-23)	(-69,134)	(92,-12)	(-87,162)	(-79,166)
	GTAFEGKT, 1ISA(B), 44–51	(83,9)	(-107,174)	(-67,-19)	(-68,-21)	(-64,-25)	(-100,31)	(-126,159)	(-76,161)
(vii)	GATGTGKT, 1D9X(A), 39–46	(-122,133)	(-75,149)	(-68,161)	(92,–34)	(-73,173)	(-92,68)	(-52,-48)	(-70,-48)
	GATGTGKT, 1D9Z(A), 39–46	(-110,129)	(-68, -169)	(-68,156)	(48,26)	(-99,2)	(76,84)	(-97,-53)	(-41,-41)
(viii)	GGAGVGKT, 1BMF(E), 156–163	(93,153)	(-125,-107)	(-86,159)	(78,–37)	(-86,133)	(-48,30)	(-41,-58)	(-69,22)
	GGAGVGKT, 1BMF(D), 156–163	(-174,145)	(-75,176)	(-65,127)	(68,41)	(-112,-8)	(128,5)	(- 52,-60)	(-64,-43)
	GGAGVGKT, 1MAB(B), 156–163	(-153,160)	(-76,116)	(-100,105)	(69,89)	(-160,-25)	(179, -158)	(88, -106)	(-45,-52)
	GGAGVGKT, 1E1R(E), 156–163	(-170,-87)	(111, -176)	(-74,-71)	(66,17)	(-131,23)	(79,17)	(-51,-54)	(-63,-35)
(ix)	GPTGVGKT, 1G3I(A), 57–64	(-161,179)	(-82, -172)	(-80,136)	(43,65)	(-124,8)	(126,–23)	(-34,-51)	(-80,-40)
	GPTGVGKT, 1G3I(S), 57–64	(-174,158)	(-47, -165)	(-118,92)	(141,–19)	(73,-4)	(82,98)	(-120,-23)	(-89,-33)

^aThe entry 1MMO (E) has been taken in the place of 1MTY (D) of Table II since both of these are structurally and sequentially highly homologous.

Table V. Distribution of the different types secondary structures flanking Walker sequence GXXXXGKT (S) in the examples given in Tables I and II

Secondary str Walker sequen	ucture flanking nce	Examples in Table I	Examples in Table II
Preceding	Following		
α	α	62	9
α	β	19	4
α	X	91	9
β	α	418	60
β	β	21	8
β	X	4	3
X	α	9	0
X	β	23	3
X	X	2	1

 $\alpha = \alpha$ -Helix; $\beta = \beta$ -strand; $X = \text{neither } \alpha \text{ nor } \beta$.

Table IV, shown as overlapping boxes). The fourth entry, 1EIR (E), has an altogether different conformation.

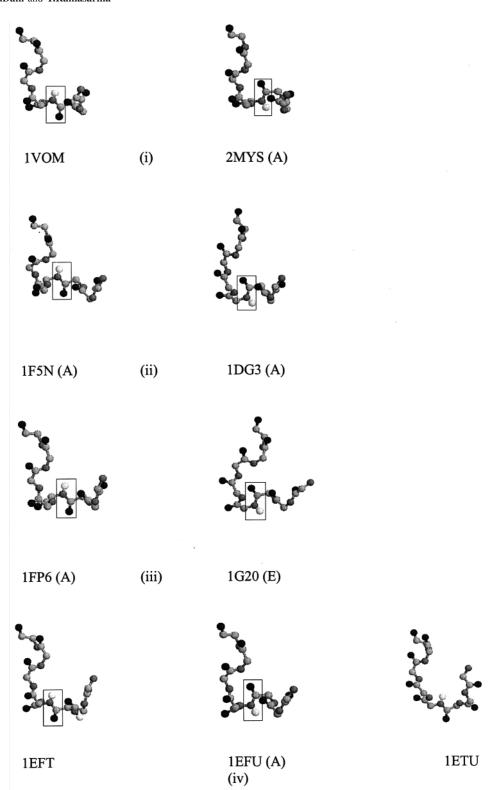
The last entry in Table IV corresponds to the sequence GPTGVGKT occurring in the two chains A and S of the protein 1G3I and the conformations are different. In this case the peptide unit between locations 6 and 7 show a rotation of $\approx 90^{\circ}$ about the virtual C^{α} – C^{α} bond, instead of a flip, as is found in the other examples.

The ball and stick diagrams of these nine examples with a flipped peptide unit shown within a box are given in Figure 3. The overlap of the polypeptide backbones appears good. The examples of pairs i-vi correspond to the flip occurring at the middle peptide unit of the well-known $4\rightarrow1$ hydrogenbonded β -turns of types I and II (Venkatachalam, 1968; Gunasekaran *et al.*, 1998). However, the flip of the peptide unit observable in pairs vii–ix does not correspond to the β -

turn flip as the values of (ϕ, ψ) are far different from those characteristic of β -turn ranges. Further, the $4 \rightarrow 1$ hydrogen bond is also absent. Notwithstanding the flip, the same overall backbone structure is retained.

The examples of nucleotide-binding proteins are arranged in Figure 3 with the nucleotide bound forms on the left and the free forms on the right. These are as follows: myosin ATPase [1VOM-2MYS (A)], guanylate binding protein [1F5N (A)-1DG3(A)], nitrogenase, [1FP6(A)-1G20(E)] elongation factor Tu [1EFT-1EFU(A), uvrB protein (A)-DNA helicase $[1D9Z(A)-1D9X(A)], F_1$ -ATPase [1BMF(D)-1BMF(E)] and 1MAB(B)] and the HSLUV protease chaperone complex [1G3I(A)-1G3I(S)]. Wherever the nucleotide is bound the N-H of the flipped peptide unit projects inwards of the loop. In the case of F₁-ATPase (Abrahams et al., 1994), this N–H forms a hydrogen bond with P=O of the β -phosphate. It appears that the presence or absence of the nucleotide makes the difference between the two structural forms. The residues of Walker sequence in such proteins not only bind to the nucleotide phosphates but also show consequent localized structural changes. This feature has important implications in the biochemical events that occur at this site.

In the case of proteins with oxygen-related reactions, the difference appears to be present in the polypeptides as isolated. The two proteins of methane monooxygenase, showing a flip at position 4/5, are derived from two organisms. The Walker sequence is present only in the Fe-form of superoxide dismutase and the two identical subunits of this enzyme protein exhibit this flip of a peptide unit. It is possible that the O=O group may act as the P=O in nucleotide phosphate in protein–substrate interactions. No relationship has so far been found between the peptide flips in Walker sequences and the activities of these proteins.



The secondary structures flanking the Walker sequence

The foregoing analysis indicated that the variable region is unlikely to determine the conformation of the Walker sequence A found in many nucleotide-binding proteins. The characteristic loop structure of the Walker sequence in these proteins is known to be preceded by a β -strand and followed by an α -helix (see, for an example, Abrahams *et al.*, 1994). It was therefore of interest to examine the occurrence of the flanking secondary structure of Walker sequences in proteins listed in

Tables I and II. For this purpose, segments of eight residues on either side of Walker sequences were examined for the presence of secondary structures ($\alpha = \alpha$ -helix; $\beta = \beta$ -strand; X = neither α nor β ; W = Walker sequence A). All nine possible combinations do occur and their distribution is given in Table V. The majority of the examples fall into the category of β -W- α . This structural motif is present in all cases in the sets 1, 2 and 6 and some in set 7 of Table II. Interestingly, each of these proteins can bind to nucleotides leading to

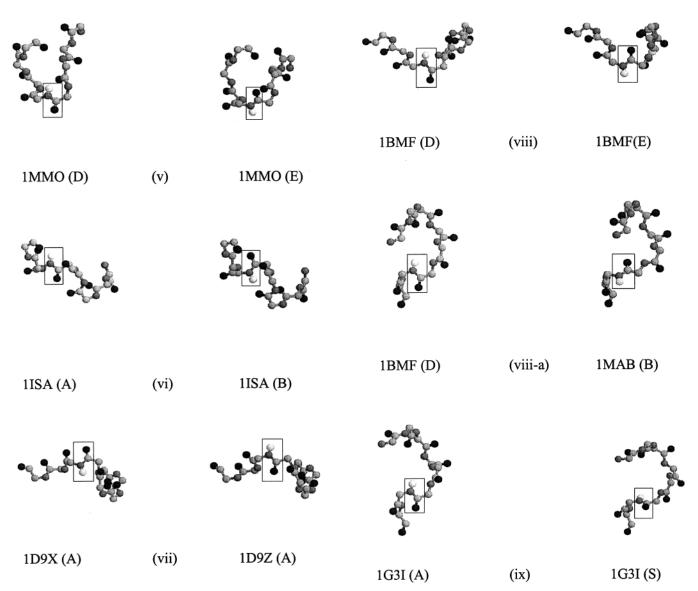


Fig. 3. Ball and stick diagrams of the backbone atoms in the segment GXXXXGKT (S) in the examples given in Table II. The flip of the peptide unit can be seen in the box, shown at the stated positions; 3 and 4 for i–iv; 4 and 5 for v; 5 and 6 for vi–viii; 6 and 7 for viii-a and ix. Shown as a white ball in this peptide unit, the hydrogen atom has been geometrically fixed.

hydrolysis of the terminal phosphate to provide energy for accompanying reactions (e.g. ATPases) in a large number of cases and in some cases transfer the phosphate to acceptors (kinases). This is true of the examples of proteins in the miscellaneous set 7. Hence it appears that the structural motif β –W– α , but not W alone, is the determining factor for nucleotide binding. The examples in sets 3, 4 and 5 of Table II, although small in number (only two each), show distinctive motifs of X–W– β , α –W– β and α –W– α , respectively.

Discussion

The noteworthy observation in this study is that the Walker sequence is present in many proteins and is not limited to those that bind and/or use nucleotides in their actions. Because of the belief that it provides the loop for phosphate binding, the so-called P-loop, this sequence was looked for only in such proteins and was invariably found. A search in the PDB files for its general occurrence, undertaken in this study, revealed its broad distribution (Tables I and II). The diversity

of these proteins is truly amazing. These include peroxidases (of cytochrome, lignin), proteases (cathepsin, collagenase, serine protease), enzymes (methane monooxygenase, superoxide dismutase, α-amylase, glutamate dehydrogenase, carbonic anhydrase, Taq polymerase, etc.), binding proteins (lectin, trypsin inhibitor) and miscellaneous proteins (α-toxin, cyclophilin B, enterotoxin). An examination of the structures of cytochrome peroxidase and superoxide dismutase indicated that these sequences are present at some distance from the active metal centers. It is to be ascertained whether Walker sequences in these proteins are utilized in their actions or their presence is incidental. It becomes obvious that the Walker sequence is more widely distributed and presence of the P-loop seems to be restricted to the nucleotide binding proteins.

The second observation in this study is the sharing of a common loop structure in proteins of the major group which use and bind nucleotide phosphates. These include kinases, phosphatases, ATPases, heat shock proteins, transfer/transport

ATPases, permeases, myosin motor domain and elongation factor. The variable quartet (XXXX) has little influence on the bend as seen from the minor variation of overlap in this region (Figure 1). Indeed, the variable quartet is so highly random in sequence that it gives no clue on the looping. Of these, G (13.3%), A (11.9%), S (9.8%), V (8.4%) and T (5.9%) occur more commonly than other amino acids, but no sequence can be identified with a set or a sub-set of proteins. Thus the formation of the β-turn loop seems to depend less on this sequence and more on the polypeptide chains on either side of the P loop, characteristically a β-sheet at the N-terminus and an α-helix at the C-terminus. The absence of the classical $4\rightarrow1$ hydrogen bond in these loop structures appears to provide more room to surround and manipulate the phosphate chain of nucleotides for exchanging terminal phosphate.

Finally, the minor, local differences in the structures with the same Walker sequence, in our opinion, are of importance as they offer possibilities of participation in the functions of these proteins. These relate to the flip of peptide units in four positions (3-4, 4-5, 5-6, 6-7 in Table IV) in these sequences. The large differences in Ramachandran angles indeed brings to light these structural variants. Three examples are noted in the pairs that show these flips: the same enzyme protein from two different organisms (methyl monooxygenase), the two subunits of a homodimer protein (Fe-superoxide dismutase) and the binding of nucleotide to one of the two subunits (F₁-ATPase, β -subunit). The last example is a case with possible interaction of the substrate and the backbone structure of the enzyme active site and offers interesting mechanistic possibilities. Details of this have been reported elsewhere (Ramasarma and Ramakrishnan, 2002).

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