BIOINFORMATICS

Vol. 20 no. 16 2004, pages 2751-2758 doi:10.1093/bioinformatics/bth322



A neural network method for prediction of β -turn types in proteins using evolutionary information

Harpreet Kaur and G. P. S. Raghava*

Institute of Microbial Technology, Sector-39A, Chandigarh, India

Received on March 9, 2004; revised and accepted on April 29, 2004 Advance Access publication May 14, 2004

ABSTRACT

Motivation: The prediction of β -turns is an important element of protein secondary structure prediction. Recently, a highly accurate neural network based method Betatpred2 has been developed for predicting β -turns in proteins using positionspecific scoring matrices (PSSM) generated by PSI-BLAST and secondary structure information predicted by PSIPRED. However, the major limitation of Betatpred2 is that it predicts only β -turn and non- β -turn residues and does not provide any information of different β -turn types. Thus, there is a need to predict β -turn types using an approach based on multiple sequence alignment, which will be useful in overall tertiary structure prediction.

Results: In the present work, a method has been developed for the prediction of β -turn types I, II, IV and VIII. For each turn type, two consecutive feed-forward back-propagation networks with a single hidden layer have been used where the first sequence-to-structure network has been trained on single sequences as well as on PSI-BLAST PSSM. The output from the first network along with PSIPRED predicted secondary structure has been used as input for the secondlevel structure-to-structure network. The networks have been trained and tested on a non-homologous dataset of 426 proteins chains by 7-fold cross-validation. It has been observed that the prediction performance for each turn type is improved significantly by using multiple sequence alignment. The performance has been further improved by using a second level structure-to-structure network and PSIPRED predicted secondary structure information. It has been observed that Type I and II β -turns have better prediction performance than Type IV and VIII β -turns. The final network yields an overall accuracy of 74.5, 93.5, 67.9 and 96.5% with MCC values of 0.29, 0.29, 0.23 and 0.02 for Type I, II, IV and VIII β -turns, respectively, and is better than random prediction.

Availability: A web server for prediction of β -turn types I, II, IV and VIII based on above approach is available

at http://www.imtech.res.in/raghava/betaturns/ and http:// bioinformatics.uams.edu/mirror/betaturns/ (mirror site).

Contact: raghava@imtech.res.in

INTRODUCTION

Protein secondary structure prediction is an intermediate step in overall tertiary structure prediction. The secondary structure of a protein consists of helices, β -strands and coil. The coil region in a protein includes tight turns, bulges and random coil structures (Chou, 2000). Tight turns are believed to be important structural elements involved in molecular recognition processes between proteins, in interactions between peptide substrates and receptors, and in protein folding (Rose et al., 1985; Takano et al., 2000). Among tight turns, β turn is predominant one that plays a vital role in protein folding, stability and recognition and is an important component of β -hairpin structure. On an average, β -turns constitute about 25% of the residues in globular proteins (Kabsch and Sander, 1983). They can be classified into nine different types depending on the ϕ , ψ angles of the two central residues.

The consideration of prediction of tight turns can enhance the usefulness of secondary structure prediction methods. Interestingly, in past, methods have been developed for prediction of tight turns based on statistical approaches, Sequencecoupled model, neural networks and support vector machine (Chou, 1997a; Kaur and Raghava, 2002a, 2003a,b,c; Chou and Blinn, 1997; Shepherd et al., 1999). Among these, a number of methods are available for prediction of β -turns (Kaur and Raghava, 2002a, 2003a; Chou and Blinn, 1997; Shepherd et al., 1999; Chou, 1997b; Zhang and Chou, 1997; Cai et al., 2002; Lin et al., 2002). To provide an adequate ranking, an extensive evaluation of all the existing β -turn prediction methods has also been carried out on a uniform dataset of 426 non-homologous protein chains (Kaur and Raghava, 2002b). Recently, we have described a method, Betatpred2 (http://www.imtech.res.in/raghava/betatpred2/) for β -turn prediction, which uses the multiple sequence alignment for prediction (Kaur and Raghava, 2003a). The method has the highest prediction accuracy among all the existing methods.

^{*}To whom correspondence should be addressed.

However, the major limitation of Betatpred2 method is that it predicts whether a residue is in β -turn or not and does not differentiate between different β -turn types. Based on β -turn/non-turn knowledge, we cannot assign the ϕ , Ψ values to residues if we try to build up a complete three-dimensional structure for a given primary sequence. This means that we need to predict different β -turn types so that it will be useful in assigning the ϕ , Ψ angles to a residue corresponding to the predicted β -turn type. Thus, it is important to develop a method, which can predict different β -turn types with high accuracy.

The method, 'Betaturns' proposed in this paper is aimed at prediction of different types of β -turns. The method is based on artificial neural network (ANN) trained on position-specific scoring matrices (PSSMs) obtained from PSI-BLAST. The ANN has been trained separately for each β -turn type on a non-homologous dataset of 426 protein chains. We have focused mainly on prediction of β -turn types I, II, IV and VIII. The remaining turn types I', II', VIa1, VIa2 and VIb are very few and are not enough for a reliable prediction. For instance, the number of turn types I', II', VIa1, VIa2 and VIb present in the dataset are only 304, 165, 44, 17 and 70 respectively out of total 7153 β -turns. Thus, these turn types have been combined into one set, called NS (non-specific) turn type. For each β -turn type, two networks have been used consecutively—the first 'sequence-to-structure' network trained on PSSM and the second 'structure-to-structure' network trained on output obtained from first network and PSIPRED predicted secondary structure states. The method Betaturns based on this approach is available as web server at http://www.imtech.res.in/raghava/betaturns/

MATERIALS AND METHODS

The dataset

In the present study, the dataset is comprising of 426 non-homologous protein chains as described by Guruprasad and Rajkumar (2000). In this dataset, no two protein chains have >25% sequence identity. The structure of these proteins is determined by X-ray crystallography at \leq 2.0 Å resolution. Each chain contains at minimum one β -turn.

Assignment of β -turns

The PROMOTIF (Hutchinson and Thronton, 1996) program has been used to assign β -turn types in proteins. It uses the β -turn types classification scheme proposed by Hutchinson and Thornton (1994) which categorizes β -turns into nine types: I, II, I', II', IV, VIa1, VIa2, VIb and VIII based on ϕ and Ψ angles of two central residues (Table 1).

7-fold cross validation

A prediction method is often developed by cross-validation or jack-knife method (Rost and Sander, 1993). The present method involves PSI-BLAST to generate position specific

Table 1. Dihedral angles of central residues (i + 1, i + 2) for β -turn types

Turn type	Dihedral angles (°)								
	ϕ_{i+1}	ψ_{i+1}	ϕ_{i+2}	ψ_{i+2}					
I	-60	-30	-90	0					
I'	60	30	90	0					
II	-60	120	80	0					
II'	60	-120	-80	0					
IV	-61	10	-53	17					
VIa1	-60	120	-90	0					
VIa2	-120	120	-60	0					
VIb	-135	135	-75	160					
VIII	-60	-30	-120	120					

scoring matrices and the whole process is time consuming due to which the jack-knife method (individual testing of each protein in the dataset) was not feasible, so a more limited cross-validation technique has been used. For each β -turn type, the prediction method has been trained and tested using 7-fold cross-validation technique, whereby the whole set is divided into seven sets, each containing equal number of proteins. The method has been trained on five sets, validated on sixth set to prevent over training and finally the performance is measured on the remaining seventh set.

Neural network architecture

In the present study, networks have been trained separately for different types of β -turns. For each β -turn type, two feedforward back-propagation networks (i) sequence-to-structure and (ii) structure-to-structure network, both with a single hidden layer containing 10 units have been used. The number of hidden units has been optimized. The target output has one unit, representing the β -turn type for the input pattern. The input to the first network is either single sequence encoded as binaries (0 or 1) or PSI-BLAST PSSM (real numbers) with window of nine residues where the prediction is made for the central residue. Output from the sequence-to-structure network is fed to a second structure-to-structure network with an input layer consisting of four units for each window position where one unit codes for output obtained from first net and the remaining three units are the probabilities of three secondary structure states (helix, strand and coil) obtained from PSIPRED output.

For the neural network implementation and to generate the neural network architecture and the learning process, the publicly available free simulation package SNNS (Zell and Mamier, 1997), version 4.2, from Stuttgart University is used. It allows incorporation of the resulting networks into an ANSI C function for use in stand-alone code. A linear activation function has been used. Weights have been modified using the back-propagation learning algorithm with a sum of square error function (SSE) (Rumelhart *et al.*, 1986). The magnitude

of the error sum in the training and validation set is monitored in each cycle of the training. The ultimate number of cycles is determined where the network converges.

Multiple sequence alignment and secondary structure

PSIPRED (Jones, 1999) uses PSI-BLAST (Altschul *et al.*, 1997) to detect distant homolog of a query sequence and generate position-specific scoring matrix as part of the prediction process, and here we have used these intermediate PSI-BLAST generated position specific scoring matrices as a direct input to the first level network. The matrix has $21 \times M$ elements, where M is the length of the target sequence, and each element presents the frequency of occurrence of each of the amino acids at one position in the alignment.

PSIPRED method has been used for predicting secondary structure, which gives the reliability indices for all the three secondary structure states (helix, strand and coil) for each residue.

Performance measures

Both threshold dependent and independent measures have been used to assess the performance of the method.

Threshold dependent measures

Five different parameters have been used to measure the performance of prediction method. These five parameters can be derived from the four scalar quantities: TP_i (true positives: number of correctly classified β -turn type i), TN_i (true negatives: number of correctly classified non- β -turns), FP_i (false positives: number of non- β -turns incorrectly classified as β -turn type i) and FN_i (false negatives: number of β -turn type i incorrectly classified as non- β -turns or some other turn type), where i = I, II, IV, VIII and IV. Following five parameters have been calculated at different threshold or cutoff values.

- (1) Prediction Accuracy (Accⁱ) = $[(TP_i + TN_i)/t] \times 100$, where $t = TP_i + TN_i + FP_i + FN_i$ is the total number of examples including β -turn types and non- β -turns.
- (2) Sensitivity $(S_n^i) = [TP_i/(TP_i + FN_i)] \times 100$ is the percentage of observed β -turn types that are predicted correctly.
- (3) Specificity $(S_p^i) = [TN_i/(TN_i + FP_i)] \times 100$ is the percentage of observed non- β -turns that are predicted correctly.
- (4) Probability of correct prediction (P_C^i): The probability of correct prediction is the percentage of predicted β -turn types that are predicted correctly.

$$P_C^i = \left(\frac{\mathrm{TP}_i}{\mathrm{TP}_i + \mathrm{FP}_i}\right) \times 100$$

(5) Matthews correlation coefficient (MCC_i): The commonly used parameter, prediction accuracy may be misleading due to disparity in the number of β -turns

types and non- β -turns; hence, it is possible to achieve high accuracy by predicting all non- β -turn residues as non- β -turns. Thus, there is a need to use more robust measures to evaluate a method. One of the best performance measures that accounts for unbalancing (both over- and under-prediction) is the Matthews correlation coefficient (Matthews, 1975). The correlation coefficient is defined as

$$MCC_i = \frac{(TP_i)(TN_i) - (FP_i)(FN_i)}{\sqrt{(TP_i + FP_i)(TP_i + FN_i)(TN_i + FP_i)(TN_i + FP_i)}}$$

The MCC is a number between -1 and 1. If there is no relationship between the predicted values and the actual values, the correlation coefficient is 0 or very low (the predicted values are no better than random numbers). As the strength of the relationship between the predicted values and actual values increases so does the correlation coefficient. A perfect fit gives a coefficient of 1.0. Thus, the higher the correlation coefficient the better is the prediction performance.

Performance with respect to random prediction

Another useful approach is to compare the accuracy of predictions with respect to predictions generated by random. Here, we have calculated the total number of residues that are expected to be predicted correctly by randomly generated predictions. The requisite formula is

$$R_{\text{total}}^i = \frac{(\text{TP}_i + \text{FP}_i)(\text{TP}_i + \text{FN}_i) + (\text{TN}_i + \text{FP}_i)(\text{TN}_i + \text{FN}_i)}{t}$$

To measure how well a method is performing compared with random (R_{total}^{i}), the normalized percentage better than random (S_{i}) has been calculated for each β -turn type.

$$S_i = \frac{(TP_i + TN_i) - R_{total}^i}{t - R_{total}^i} \times 100$$

Perfect predictions score $S_i = 100\%$, predictions that are no better than random score $S_i = 0\%$ (Shepherd *et al.*, 1999).

Threshold independent measures

The performance measures described so far are threshold dependent. One problem with the threshold dependent measures is that they measure the performance at a given threshold. They fail to use all the information provided by a method. For instance, the false positive rate varies with the threshold value. Thus, the receiver operating characteristic (ROC), a trade off between sensitivity and specificity and which is a threshold independent measure has been used to assess the performance. For a prediction method, ROC plot is obtained by plotting all sensitivity values (true positive rate) on the y-axis against their equivalent (1 — specificity) values (false positive rate) for all available thresholds on the x-axis. The curve always goes through two points (0,0 and 1,1). 0,0 is where the classifier finds no positives. In this case, it always gets the negative cases

right but it gets all positive cases wrong. The second point is 1,1 where everything is classified as positive. So the classifier gets all positive cases right but it gets all negative cases wrong. A classifier that randomly guesses has ROC which lies somewhere along the diagonal line connecting 0,0 and 1,1. An important index of ROC curve is its area. A random classifier has an area of 0.5, while an ideal one has an area of 1 (Deleo, 1993).

A measure of statistical significance

When comparing different prediction approaches, we need to know whether the differences in performance measures (prediction accuracies or MCC values) among them are statistically significant or not. Statistics theory gives us a method to compute the 'significance interval' for the difference between two population proportions (Daniel, 1987).

In this case, the 'proportion' is the percentage of cases in test dataset, which have been predicted correctly. If we assume that the prediction accuracies of two algorithms are p_1 and p_2 for two test datasets of r_1 and r_2 number of examples, respectively, and the test data are randomly selected, then we can say that we are 100% confident that the two accuracies are really different if

$$|p_1 - p_2| > I$$
,

where

$$I = z \left(1 + \frac{a}{2} \right) \cdot \sqrt{p_1 (1 - p_1)/r_1 + p_2 (1 - p_2)/r_2},$$

z is the inverse cumulative normal distribution. The larger the difference between two prediction accuracies, the more significant it is. The above equation (Zhang $et\ al.$, 1992) has been used to determine whether the difference in the accuracies or other measures is statistically significant or not.

Segment overlap measure (SOV)

The method predicts the β -turn type at residue level. The single-residue predictions do not completely reflect the quality of a prediction. One should take into account the average length of predicted β -turn type. The SOV is a measure for evaluation of prediction method by secondary structure segment rather than individual residues. To address the overlapping between the observed and predicted β -turn type residues, SOV (Zemla *et al.*, 1999) has been calculated for each β -turn type as:

$$SOV_i = \frac{1}{N} \sum_{S} \frac{\min ov(S1; S2) + \delta}{\max ov(S1; S2)} \times \text{len}(S1),$$

where S1 and S2 are the observed and predicted β -turn type i; len(S1) is the number of residues in the segment S1 of β -turn type i; min ov(S1; S2) is the length of actual overlap of S1 and S2 or the extent for which both segments have residues in β -turn type i; max ov(S1; S2) is the length of the total

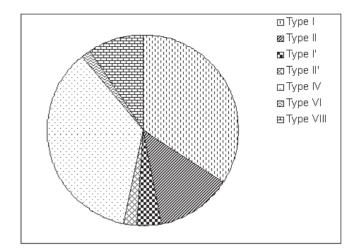


Fig. 1. Distribution of different types of β -turns in the dataset.

extent for which either of the segments S1 or S2 has a residue in β -turn type i; δ is the integer value defined as equal to the min{[max ov(S1; S2) min ov(S1; S2)]; min ov(S1; S2); int[len(S1)/2]; int[len(S2)/2]}; N is the number of residues in particular β -turn type i and sum is taken over all the pairs of segments $\{S1; S2\}$, where S1 and S2 have at least one residue in β -turn in common.

RESULTS

Distribution

From the dataset of 426 protein chains, 7153 number of β -turns have been located and categorized as shown in Figure 1. Of all the β -turn types, Type IV is the most frequently occurring turn type (\sim 35.4%) followed by Type I turns (34.1%), which are two to three times more common than Type II (12.7%). The mirror image types I' and II' are rare, comprising only 4.2 and 2.3%, respectively. Other β -turn types are very few.

Selection of neural network architecture

A number of architectures and parameters have been tried to search the best architecture and parameters for prediction. It has been observed that ANN with nine input units and a single hidden layer with 10 units perform best for all β -turn types, so in this study we have used this architecture. All the networks have been trained and tested separately for each β -turn type using 7-fold cross-validation procedure. The prediction performance measures have been averaged over seven sets.

Prediction using single sequence

The ANN has been trained and tested with single sequences encoded as binary bits (0 and 1) and the results are shown in Table 2. Type I and II β -turns have been predicted with an averaged accuracy of 70.3 and 89.6% respectively and overall their performance is better than other β -turn types. The corresponding MCC values are 0.22 and 0.24. The MCC of

Table 2. The results of β -turn types predictions by using single sequence with and without secondary structure information

	Results of present study β -turn types						BTPRED results ^a (Shepherd <i>et al.</i> , 1999) β -turn types			
	ρ-turn typ I	II	IV	VIII	NS	ρ-turn typ I	II	IV	VIII	
Accuracy	70.3	89.6	64.3	96.4	97.8	91.2	95.5	95.7	96.8	
	$(72.7)^{b}$	(92.0)	(65.4)	(96.8)	(98.2)					
Sensitivity	64.7	46.6	60.8	2.1	4.8	46.6	58.4	18.0	2.2	
	(65.3)	(48.5)	(62.4)	(1.3)	(4.4)					
Specificity	70.8	91.3	64.7	99.1	99.2					
	(73.6)	(94.1)	(65.7)	(99.5)	(99.7)					
Probability of correct prediction	18.7	17.8	15.6	7.0	8.3	13.9	12.2	3.3	9.3	
	(20.1)	(21.0)	(16.2)	(8.9)	(9.7)					
MCC	0.22	0.24	0.16	0.02	0.05	0.219	0.253	0.062	0.033	
	(0.24)	(0.25)	(0.17)	(0.02)	(0.09)					
Better-than-random	16.9	21.3	11.1	1.8	4.9	18.1	18.9	4.5	2.6	
	(19.1)	(23.2)	(12.4)	(1.4)	(6.1)					

^aPrediction results of BTPRED with predicted secondary structure.

Type II β -turn is higher than Type I β -turn, however, the sensitivity of former is 20% lower than that of latter. The prediction of both Types I and II β -turns has been found to be 17 and 21% better than random prediction. The prediction performance of Type IV β -turn is less than Type I and II β -turns, however, is better than VIII and NS β -turns. Approximately 61% of Type IV β -turn has been predicted correctly with MCC of 0.16. The prediction of Type IV β -turn has been found to be 11% better than random. Among all β -turn types, Type VIII and NS β -turns show the least performance. Type VIII β -turn has MCC of 0.02 and is only 2% better than random. Its sensitivity and probability of correct prediction is <10%. The prediction of non-specific β -turns is 5% better than random with MCC of 0.05.

Prediction using single sequence and secondary structure

The outputs obtained from the first network along with the PSIPRED predicted secondary structure information has been used as input for structure-to-structure network. The results averaged over seven sets are presented in parentheses in Table 2. Improvement with secondary structure information can be seen for all β -turn types. There is a gain of 1–2% in accuracy and sensitivity of all β -turn types. For Type I and II β -turns, MCC is raised from 0.22 to 0.24 and 0.24 to 0.25 respectively. Their prediction is found to be 20 and 23% better than random. There is a marginal improvement for Type VIII β -turn after using secondary structure information. We have obtained final MCC values of 0.24, 0.26, 0.17, 0.03 and 0.08 for β -turn types I, II, IV, VIII and NS respectively. Moreover, the results are also better than BTPRED (Table 2). The MCC values and better-than-random scores are higher than that obtained with BTPRED except for Type II and VIII

 β -turns for which both BTPRED and present work shows comparable performance.

Prediction using multiple alignment

To further enhance the prediction performance, the multiple sequence alignment (MSA) in the form of PSI-BLAST PSSM has been used as input to ANN. The results are shown in Table 3. The prediction accuracy of all β -turn types increases by 2–3% except for Type VIII and NS β -turns. The maximum improvement in sensitivity has been obtained for NS β -turn type followed by Type II β -turn. For Type I and II β -turns, nearly 65 and 50% of turns have been predicted correctly. There is no change in sensitivity of Type IV and VIII β -turns even after using MSA. The final probability of correct prediction of Type I and II β -turns is 20.8 and 21.8% which are respectively 2 and 4% higher than that obtained with sequence alone. Moreover, the improvement has found to be statistically significant at 95% confidence level. The final MCC values achieved are 0.25, 0.26, 0.18, 0.02 and 0.13 for β -turn types I, II, IV, VIII and NS respectively and are 1–8% higher than with single sequence. The performance of all β turn types has been found to be better than random prediction with Type I and II β -turns better-than-random score being 20 and 27% respectively.

Prediction using multiple alignment and secondary structure

An output obtained from first network (trained on PSI-BLAST PSSM) and secondary structure predicted by PSIPRED is applied to the second network. The results are shown in Table 3. Using secondary structure along with MSA has resulted in a significant increase of 9 and 11% in sensitivities of Type I and IV β -turns with MCC 0.29 and 0.23 respectively.

^bValues in parentheses are prediction results obtained using PSIPRED predicted secondary structure.

Table 3. Prediction results of network using PSI-BLAST PSSM with and without secondary structure information

	Multiple alignment (Sequence-to-structure network) β -turn types					Multiple alignment and secondary structure (Structure-to-structure network) β -turn types				
	I	II	IV	VIII	NS	I	II	IV	VIII	NS
Accuracy	72.8	91.9	67.7	96.2	97.8	74.5 (74.1)	93.5 (92.6)	67.9 (67.2)	96.5 (96.4)	98.1 (98.0)
Sensitivity	65.0	50.2	60.6	2.4	13.2	74.1	52.8	72.0	2.8	13.3
Specificity	73.7	93.4	68.5	98.9	99.1	(73.8) 75.5 (74.6)	(51.9) 94.8 (94.2)	(71.8) 66.0 (66.0)	(2.6) 98.7 (98.4)	(13.3) 99.4 (99.4)
Probability of correct prediction	20.8	21.8	16.9	7.2	15.0	22.1	25.5	18.6	7.2	23.7
MCC	0.25	0.26	0.18	0.0	0.13	(21.7) 0.29 (0.29)	(25.0) 0.29 (0.29)	(18.3) 0.23 (0.23)	(7.0) 0.02 (0.02)	(23.5) 0.17 (0.17)
Better-than-random	19.8	26.7	13.5	1.9	11.9	22.7 (22.2)	31.5 (31.2)	17.0 (16.8)	1.9 (1.8)	16.0 (16.0)

Values in parentheses correspond to the prediction results obtained by excluding the proteins that were used to develop PSIPRED.

For Type II β -turn, there is 2–3% improvement in sensitivity and probability of correct prediction. The final MCC is 0.29, which is 31.5% better than random prediction. There is no improvement for Type VIII β -turn. For Type NS β -turn, MCC improves from 0.13 to 0.17 and this can be contributed to a significant increase of 9% in its probability of correct prediction. All these values indicate that use of secondary structure along with MSA considerably increases the number of true positives and true negatives and decreases over- and under-predictions. Such improvements are also found to be statistically significant at 95% confidence level.

To check whether the prediction performance with secondary structure information is due to PSIPRED or not, the results have been cross validated by removing those proteins from the dataset that were used to develop PSIPRED. The results given in parentheses in Table 3 show negligible difference in performance measures.

SOV

Since, the prediction is done at residue level, the predicted turn may be displaced by one or two residues and useful predictive information, may be contained in these displaced predictions. To account for such predictions, SOV, which is a segment-based measure of prediction assessment, has been calculated for turn types I, II, IV and VIII. This score measures the extent of segment overlap with a deviation of residues at both ends. Overlap accuracies SOV through 7-fold cross-validation with multiple sequence alignments and secondary structure are 49.6, 41.9, 51.6 and 28.2% for β -turn types I, II, IV and VIII, respectively (Table 4).

ROC

Performance of networks trained on PSSM and secondary structures for each β -turn types has also been evaluated by

Table 4. SOV values of β -turn types with network trained using PSI-BLAST PSSM and secondary structure information

β -turn type	SOV (in		
I	49.6		
II	41.9		
IV	51.6		
VIII	28.2		

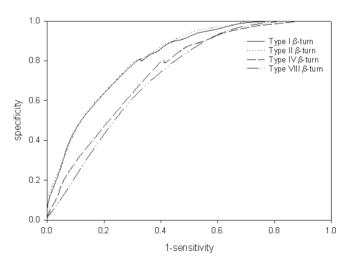


Fig. 2. ROC curves for turn types I, II, IV and VIII of network trained on multiple alignment with secondary structure information.

calculating the area under the ROC curve. Figure 2 shows the ROC curves for turn types I, II, IV and VIII. The corresponding areas under the curves are as follows: Type I β -turn 0.746; Type II β -turn 0.759; Type IV β -turn 0.713; and Type VIII

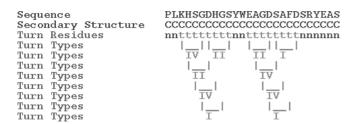


Fig. 3. Sample β -turn type predictions. Row 1 is the amino acid sequence; row 2 is the secondary structure states predicted by PSIPRED (H, helix; E, strand; and C, coil) and Row 3 is the predicted β -turn residues (t, β -turn residues; n, non- β -turn residues). The predicted β -turn residues are further classified as Types I, II, IV, VIII and non-specific marked by roman numerals I, II, IV VIII and NS, respectively.

 β -turn 0.662. These values reflect the better prediction of type I and II β -turns in comparison to type IV and VIII β -turns.

Comparison with BTPRED

A comparison of present method with BTPRED shows that the results obtained in this work are better than that of BTPRED. In BTPRED study, Shepherd *et al.* has reported MCC 0.22 and better-than-random score 18% for Type I β -turn, which are 7 and 5% less than that achieved in this study. Similarly, the prediction of Type II β -turn is 31.5% better than random prediction and is 19% higher than that of BTPRED. Even for Type IV β -turn, the results obtained in present study are better than BTPRED. However, the performance of Type VIII β -turn is inferior to BTPRED.

Web server Betaturns

A web server 'Betaturns' (http://www.imtech.res.in/raghava/betaturns/) has been developed for the prediction of β -turn types I, II, IV, VIII and NS based on neural network and multiple sequence alignment approach. The SNNS generated network for each turn type is converted into C program and is used as an interface.

The output consists of target sequence, PSIPRED predicted secondary structure and predicted β -turn types such as I, II, IV, VIII and NS. Turn residues are predicted as four residues block with turn types indicated by roman numerals I, II, IV, VIII for turn types I, II, IV and VIII respectively or 'NS' for non-specific beta-turn category which does not belong to any of the four turn types. A sample of the prediction output is shown in Figure 3.

DISCUSSION

It is known that using information from sequence alignment significantly improves protein secondary structure prediction rather than single sequence. Typically, more divergent profiles yield better prediction (Przybylski and Rost, 2002). A neural network based method Betatpred2 (Kaur and Raghava, 2003a) for predicting β -turns from the amino acid

sequence based on multiple sequence alignment has recently been developed. The better performance of Betatpred2 over BTPRED (Shepherd *et al.*, 1999) has resulted from neural network training on PSI-BLAST generated position specific scoring matrices. For Betatpred2, MCC value increases from 0.31 with single sequence to 0.37 with multiple sequence alignment. Further, by incorporating secondary structure information, the final MCC achieved is 0.43 and is the best achieved so far. However, the method Betatpred2 predicts only β -turn or non-turn residues and does not provide any information of β -turn types. In the present work, the approach of multiple sequence alignment and secondary structure has been extended for prediction of different types of β -turns and a method betaturns based on such approach has been developed for predicting β -turn types I, II, IV and VIII.

Two different inputs coding to the network have been used. One is based on single sequence with amino acids as binary bits and the other is multiple sequence alignment in the form of PSI-BLAST generated PSSM. In both the cases, a second 'structure-to-structure' network has been trained on secondary structure obtained from PSIPRED. It has been found that for all β -turn types, the performance of network with MSA is superior to that of sequence alone. With MSA, Type I and II β -turns have MCC 0.25 and 0.26, which is 19.8 and 26.7% better than random prediction. The probability of correct prediction is low for all β -turn types. This is due to the fact that in the dataset the number of examples having a particular β -turn type is far less than the number of negative examples, which results in a large number of false positive predictions and thus a lower probability of correct prediction. Using secondary structure along with MSA improves the performance of all β turn types with final MCC of 0.29, 0.29, 0.23, 0.02 and 0.17 for I, II, IV, VIII and NS β -turns. There is 2–4% improvement in probability of correct prediction except for Type NS, which shows a significant increase of 9%. The performance is 22.7, 31.5, 17, 1.9 and 16% better than random prediction. These results clearly shows the ability of the method to predict Type I and II β -turns (MCC \sim 0.29) and Type VIII (MCC \sim 0.02) and NS (MCC \sim 0.17). The results are according to our expectations; the more numerous and well-defined β -turn Types I and II are predicted more accurately than the less numerous VIII and NS β -turns. The performance for Type IV β -turn is intermediate that of Types I, II and VIII, NS β -turns. Overall, the performance is better than BTPRED.

To conclude, the β -turn type prediction method described in this paper yields predictions that are significantly more accurate than previous methods and this improvement can be contributed to the use of multiple sequence alignment and secondary structure information in neural network training.

ACKNOWLEDGEMENTS

The authors are thankful to Council of Scientific and Industrial Research (CSIR) and Department of Biotechnology (DBT),

Government of India for financial assistance. We are also thankful to the developers of SNNS and PSIPRED. This report has IMTECH communication No. 012/2003.

REFERENCES

- Altschul, S.F., Madden, T.L., Alejandro, A.S., Zhang, J., Zhang, Z., Miller, W. and Lipman, D.J. (1997) Gapped BLAST and PSI-BLAST: a new generation of protein databases and search programs. *Nucleic Acids Res.*, 25, 3389–3402.
- Cai, Y.D., Liu, X.J., Xu, X.B. and Chou, K.C. (2002) Support vector machines for the classification and prediction of beta-turn types. *J. Pept. Sci.*, **8**, 297–301.
- Chou, K.C. and Blinn, J.R. (1997) Classification and prediction of *β*-turn types. *J. Protein Chem.*, **16**, 575–595.
- Chou, K.C. (1997a) Prediction and classification of α -turn types. *Biopolymers*, **42**, 837–853.
- Chou, K.C. (1997b) Prediction of β-turns. J. Pept. Res., 49, 120–144.
 Chou, K.C. (2000) Prediction of tight turns and their types in proteins.
 Anal. Biochem., 286, 1–16.
- Daniel, W.W. (1987) *Biostatistics: A Foundation for Analysis in the Health Science*. John Wiley & Sons, New York.
- Deleo, J.M. (1993) Receiver operating characteristic laboratory (ROCLAB): Software for developing decision strategies that account for uncertainty. In: *Proceedings of the Second International Symposium on Uncertainty Modelling and Analysis*. IEEE, Computer Society Press, College Park, MD, pp. 318–325.
- Guruprasad,K. and Rajkumar,S. (2000) β and γ -turns in proteins revisited: a new set of amino acid dependent positional preferences and potential. *J. Biosci.*, **25**, 143–156.
- Hutchinson, E.G. and Thornton, J.M. (1994) A revised set of potentials for β -turn formation in proteins. *Protein Sci.*, **3**, 2207–2216.
- Hutchinson, E.G. and Thornton, J.M. (1996) PROMOTIF—a program to identify and analyze structural motifs in proteins. *Protein Sci.*, **5**, 212–220.
- Jones, D.T. (1999) Protein secondary structure prediction based on position-specific scoring matrices. J. Mol. Biol., 292, 195–202.
- Kabsch, W. and Sander, C. (1983) Dictionary of protein secondary structure: pattern recognition of hydrogen-bonded and geometrical features. *Biopolymers*, **22**, 2577–2637.
- Kaur,H. and Raghava,G.P.S. (2002a) BetaTPred: prediction of β-turns in a protein using statistical algorithms. *Bioinformatics*, 18, 498–499.

- Kaur, H. and Raghava, G.P.S. (2002b) An evaluation of β -turn prediction methods. *Bioinformatics*, **18**, 1508–1514.
- Kaur, H. and Raghava, G.P.S. (2003a) Prediction of β -turns in proteins from multiple alignment using neural network. *Protein Sci.*, **12**, 627–634.
- Kaur, H. and Raghava, G.P.S. (2003b) A neural-network-based method for prediction of gamma-turns in proteins from multiple sequence alignment. *Protein Sci.*, **12**, 923–929.
- Kaur, H. and Raghava, G.P.S. (2003c) Prediction of α-turns in proteins using PSI-BLAST profiles and secondary structure information. *Proteins*, **55**, 83–90.
- Lin, T.H., Wang, G.M. and Wang, Y.T. (2002) Prediction of beta-turns in proteins using the first-order Markov models. *J. Chem. Inf. Comput. Sci.*, **42**, 123–133.
- Matthews, B.W. (1975) Comparison of the predicted and observed secondary structure of T4 phage lysozyme. *Biochim. Biophys. Acta*, **405**, 442–451.
- Przybylski, D. and Rost, B. (2002) Alignments grow, secondary structure prediction improves. *Proteins*, **46**, 197–205.
- Rose, G.D., Gierasch, L.M. and Smith, J.A. (1985) Turns in peptides and proteins. *Adv. Protein Chem.*, **37**, 100–109.
- Rost,B. and Sander,C. (1993) Prediction of protein secondary structure at better than 70% accuracy. *J. Mol. Biol.*, **232**, 584–599.
- Rumelhart, D.E., Hinton, G.E. and Williams, R.J. (1986) Learning representations by back-propagation errors. *Nature*, **323**, 533.
- Shepherd, A-J., Gorse, D. and Thornton, J.M. (1999) Prediction of the location and type of β -turn types in proteins using neural networks. *Protein Sci.*, **8**, 1045–1055.
- Takano, K., Yamagata, Y. and Yutani, K. (2000) Role of amino acid residues at turns in the conformational stability and folding of human lysozyme. *Biochemistry*, 39, 8655–8665.
- Zell, A. and Mamier, G. (1997) *Stuttgart Neural Network Simulator*. Version 4.2. University of Stuttgart, Stuttgart, Germany.
- Zemla, A., Venclovas, C., Fidelis, K. and Rost, B. (1999) A modified definition of Sov, a segment based measure for protein secondary structure prediction assessment. *Proteins*, 34, 220–223.
- Zhang, C.T. and Chou, K.C. (1997) Prediction of β -turns in proteins by 1-4 & 2-3 Correlation Model. *Biopolymers*, **41**, 673–702.
- Zhang, X., Mesirov, J.P. and Waltz, D.L. (1992) Hybrid system for protein secondary structure prediction. J. Mol. Biol., 225, 1049–1063.