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Goverdhan Mehta,* Kotapalli Pallavi and Jayant D. Umarye

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A total synthesis of the structure corresponding to the novel tricyclic diterpene guanacastepene C has been realized in which a Knoevenagel cyclization serves as a key step to annulate the six-membered C-ring on a stereochemically secured bicyclic hydroazulene precursor.

Guanacastepenes are a structurally diverse and biologically potent group of diterpenoids based on a linearly fused 5,7,6- tricarbocyclic ring system. The first member of this family, guanacastepene A 1 was isolated from an unidentified endophytic fungus growing on the tree Daphnopsis americana in Costa Rica by Clardy et al.^{1a} in 2000 and its structure was elucidated by X-ray crystallography. Subsequently, several additional guanacastepenes, B-O, have been isolated from the same fungus and structurally characterized, among which guanacastepene C $\mathbf{2}$ is a close sibling of $\mathbf{1}$ with minor stereo- and functional group variations.^{1b,2} Several guanacastepenes, more notably 1, have been shown to exhibit impressive activity against methicillin-resistant Staphylococcus aureus (MRSA) and vancomycin-resistant Enterococcus faecalis (VREF). Additionally, guanacastepene A 1 exhibited moderate activity against Gram-positive bacteria, poor activity against Gram-negative bacteria and hemolytic activity against human red blood cells (RBC), thus undermining its therapeutic potential.^{1c} However, the basic bioactivity attributes of guanacastepenes hold considerable promise and devising means to access their structural variants could be a useful and productive synthetic enterprise. Indeed, guanacastepenes have drawn very widespread attention³ from synthetic chemists in the past few years, culminating in the total synthesis⁴ of guanacastepene A 1. Our group has also been in the fray and earlier we reported^{3f,j} model studies directed towards the synthesis of 1 as well as access to several structural analogs³⁰ of guanacastepenes. Following these early leads and successes, we decided to persist with efforts in the arena and have now accomplished the first total synthesis of guanacastepene C 2.



Department of Organic Chemistry, Indian Institute of Science, Bangalore, India. E-mail: gm@orgchem.iisc.ernet.in; Fax: +91-80-23600283; Tel: +91-80-22932850

[†] Electronic supplementary information (ESI) available: ¹H NMR and ¹³C NMR spectral data of selected compounds. See http://dx.doi.org/ 10.1039/b506931a

Guanacastepenes A 1 and C 2 share an identical hydrophobic southern zone (lighter shade) but vary in small but subtle ways in the hydrophilic northern zone (darker shade). More specifically, the C13 hydroxyl group in 2 is free (cf. C13 β -acetate in 1) and trans disposed with respect to the C12 isopropyl group, and the C15 substituent in guanacastepene C is in the reduced hydroxymethyl form (cf. C15 aldehyde in 1). In our previous synthetic studies^{3f} towards guanacastepenes, we had outlined the construction of stereo-defined and adequately functionalized AB-ring segment 5, present in these natural products, from the readily available tricyclic enone 3 via 4, Scheme 1. The stereochemical and functionalization pattern present in 5 appeared to be more amenable towards the synthesis of guanacastepene C 2 rather than guanacastepene A 1 and therefore we embarked on a venture targeted towards the former, which through many trials and tribulations has eventuated in the total synthesis of the natural product.

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The key issue in the evolution of the AB-ring synthon 5 towards the natural product 2 was the annulation of six-membered ring C and our previous experience had shown the inadequacy of the RCM and aldol based approaches devised by us³⁰ in delivering the requisite functional and stereochemical array on the annulated ring. Consequently, we opted to explore a Knoevenagel cyclization based protocol for the C-ring annulation on 5. LAH reduction of previously described³⁰ keto ester 5 furnished the diol 6^5 in modest yield. The two hydroxyl groups in 6 needed to be differentiated with the selective protection of the secondary hydroxyl group but, as direct protection manoeuvres were unsuccessful, a more circuitous protocol was devised to deliver 7,5 Scheme 2. This required the initial protection of the primary hydroxyl group as the PMB derivative, followed by the protection of the secondary hydroxyl group as the TBS derivative and selective removal of the PMB protection with DDQ.^{5,6} IBX oxidation⁷ of 7 was smooth and led to the aldehyde 8.⁵ To set up the Knoevenagel cyclization, bicyclic aldehyde 8 was condensed (Wittig-Horner-Emmons reaction) with the dianion derived from ethyl 4-diphenylphosphinoyl-3-oxobutanoate⁸ to append a 5C fragment and furnish 9 as a mixture of keto-enol tautomers, Scheme 2. Regioselective hydrogenation of 9 led to 10 quantitatively.⁵ TBS deprotection of







Scheme 2 *Reagents and conditions:* i, LiAlH₄, THF, 0 °C to rt, 2 h, 55%; ii, a. PMBCl, THF, NaH, TBAI, 0 °C to rt, 5 h, 67%; b. TBSOTf, 2,6-lutidine, DCM, 0 °C, 15 min, 68%; c. DDQ, DCM–H₂O (25:1), rt, 2 h, 95%; iii, IBX, toluene–DMSO (4:1), rt, 3 h, 92%; iv, Ph₂POCH₂COCH₂COOEt, NaH, *n*BuLi, THF, 0 °C to rt, 6 h, 86%; v, H₂, EtOAc, 5% Pd/C (10% w/w), 30 min, rt, quant.; vi, a. 6 N H₂SO₄, THF–H₂O (4:1), rt, 18 h, 80%; b. 2,2-DMP, PPTS, rt, 45 min, 91%; vii, PCC, NaOAc, molecular sieve powder (4 Å), DCM, rt, 12 h, 80%; viii, DBU, benzene, 65 °C, 12 h, 82%.

10 led to concomitant acetonide deprotection and intramolecular hemiketalization and the 1,2-diol moiety in the resultant product was reprotected to give 11, Scheme 2. PCC oxidation of 11 furnished the β -keto ester precursor 12⁵ for the Knoevenagel cyclization. As envisaged, 12, on exposure to DBU, underwent smooth cyclization to generate the C-ring and delivered 13⁵ embodying the complete tricarbocyclic framework of guanacastepenes, Scheme 2.

In the dienone ester 13, we had in place the three rings and all the functional elements for further elaboration to the natural product 2 and the main task ahead was functional group adjustments. LAH reduction of 13 led to 14 as a predominant C5 α -OH isomer (*vide infra*), albeit in moderate yield, Scheme 3. The diol moiety in 14 was protected as the diacetate 15 and acid hydrolysis of the acetonide functionality gave a 4:1 mixture of the diol 17 and the triol 16. Fortunately, triol 16 provided good crystals for X-ray crystal structure determination⁹ and this among



Scheme 3 Reagents and conditions: i, LiAlH₄, THF, -78 °C to rt, 1 h, 65%; ii, Ac₂O, NEt₃, DMAP, DCM, 5 h, 92%; iii, 4 N H₂SO₄, THF–H₂O (4:1), rt, 8 h, 75% of 17 and 20% of 16; iv, DDQ, THF, 65 °C, 6 h, 80%.

other things revealed the undesired nature of the C5 hydroxyl stereochemistry. DDQ mediated¹⁰ chemoselective allylic oxidation of **17** furnished **18**,⁵ formally a 5-*epi*-guanacastepene C diacetate, Scheme 3. The elaboration of **14** to **18** paved the way for a final assault on the natural product **2** and mandated the inversion of C5 α -OH stereochemistry.

Consequently, diol 14 was subjected to the standard Mitsunobu protocol to furnish the dibenzoate 19 in fair yield, Scheme 4. The acetonide protecting group was now disengaged to give 20 and further chemoselective oxidation¹⁰ of the allylic hydroxyl group delivered the guanacastepene C dibenzoate 21, Scheme 4. At this stage hydrolysis of the benzoate ester groups was expected to deliver the natural product 2. However, for reasons not quite obvious to us, this seemingly straightforward transformation frustrated us to no end and forced us to explore an alternate circuitous route. Thus, the dibenzoate 19 was hydrolyzed to the diol 22 and the hydroxyl groups were reprotected as the diacetate 23, Scheme 4. Deprotection of the acetonide group, even under the most carefully monitored reaction regime, led to some concurrent acetate hydrolysis and a 62:38 mixture of the desired diol diacetate 24 and the triol 25 was obtained. Chemoselective allylic oxidation of 24 led to the guanacastepene C diacetate 26 whose spectral data² was distinctly different from its C5-epimer 18.⁵ Further careful hydrolysis of 26 delivered a compound corresponding to the reported structure of guanacastepene C 2, Scheme 4. However, as a direct spectral comparison with the natural product was not possible, our arrival at 2 has been inferred from the internal consistency of our synthetic sequence and the limited spectral data.11

In summary, we have crafted an access to the 5,7,6-fused tricyclic framework of guanacastepene diterpenes from a readily available hydroazulenic precursor employing a Knoevenagel cyclization as the key step and further elaborated this advanced intermediate towards the diterpenoid natural product guanacastepene C.



Scheme 4 *Reagents and conditions:* i, PPh₃, C₆H₅COOH, DIAD, THF, -78 °C to rt, 2 h, 78%; ii, 2 N HCl, THF, rt, 6 h, 61%; iii, DDQ, THF, 65 °C, 6 h, 76%; iv, LiAlH₄, THF, 0 °C to rt, 30 min, 84%; v, Ac₂O, NEt₃, DMAP, DCM, rt, 5 h, 90%; vi, 4 N H₂SO₄, THF–H₂O (4:1), rt, 6 h, 44% of **24** and 28% of **25**; vii, DDQ, THF, 65 °C, 6 h, 85%; viii, K₂CO₃, MeOH, 0 °C to 15 °C, 2 h, 70%.

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- 11 The original spectra of the natural product were not available from the authors¹ for direct comparison and full spectral data are not recorded in the communication.^{1b} We and others have already commented^{2,4c,d} on the inconsistent nature of the ¹H and ¹³C NMR data in this series of compounds due to dynamic conformational changes. Synthetic **2** is no exception in this regard and this makes the independent characterization of **2** difficult. However, the comparative ¹H NMR data on the C5- α -OH and the C5- β -OH series of compounds presented in Schemes 3 and 4, respectively, and the X-ray crystal structure of **16** clearly point to our acquisition of **2**.