

# A norbornyl route to aminocyclohexitols: syntheses of diverse aminocarbasugars and ‘confused’ aminocarbasugars

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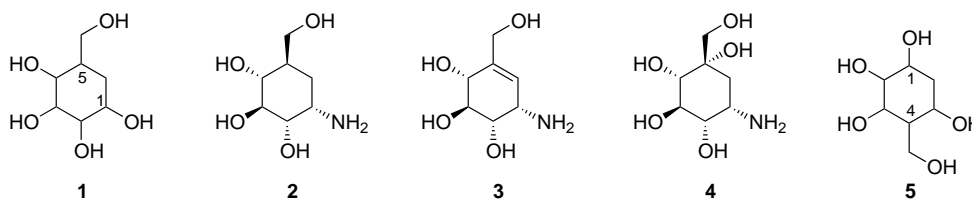
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**Abstract**—New syntheses of a range of aminocyclitols, of the aminocarbasugar and ‘confused’ aminocarbasugar type, from simple, readily available norbornyl derived building-blocks is reported. Our synthetic approaches incorporate features that provide for large product diversity in terms of the placement of the amino group on the cyclohexitol matrix.

Structural entities having the cyclohexitol (polyhydroxylated cyclohexanoids) or amino cyclohexitol core have aroused widespread synthetic interest because of the diverse biological activity exhibited by them ranging from glycosidase inhibition to mediation of cellular communication.<sup>1</sup> Carbasugars (also known as pseudosugars) like **1**, which has been known since 1966,<sup>2a</sup> are an important subclass among cyclohexitols which were projected and then proved to be potent glycomimics. While synthetic interest in carbasugars has remained undiminished during the past four decades,<sup>1,2</sup> a search for their more potent siblings has led to aminocarbasugars from Nature as well as through syntheses.<sup>3,4</sup> Among the better-known examples of naturally occurring aminocarbasugars are vali-

aminocarbasugars have been synthesized in recent years and subjected to biological evaluation and interest in these entities remains unabated.<sup>4</sup>

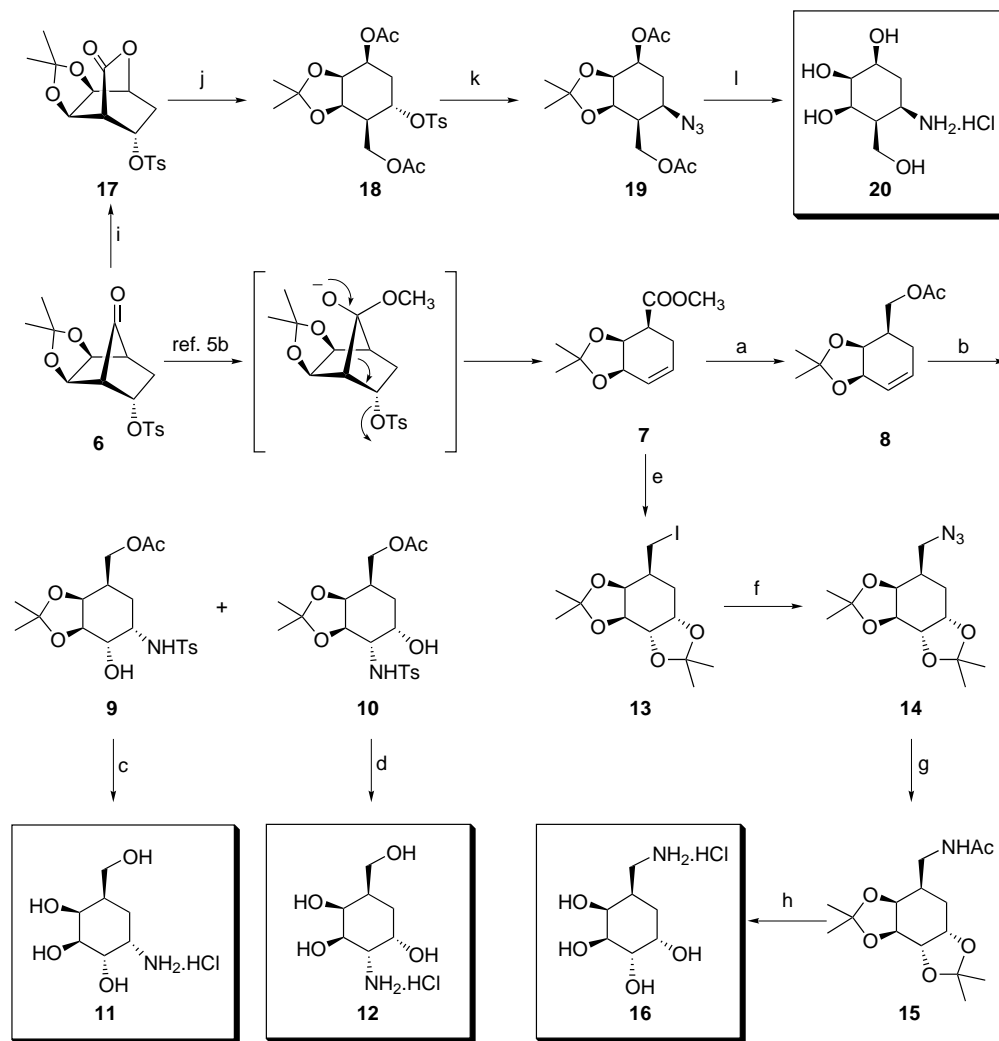
We have recently delineated<sup>5a-d</sup> general and versatile synthetic approaches based on the bicyclo[2.2.1]heptane (norbornyl) system towards carbasugars **1** and their close siblings, the hitherto unknown ‘confused’ carbasugars **5** (swapping of the *para* hydroxymethyl and hydroxy groups of **1**).<sup>5d</sup> This ready access to **1** and **5** through flexible routes that provide considerable latitude in terms of functional group maneuverability opened opportunity to access a range of new aminocarbasugars and ‘confused’ aminocarbasugars and these endeavors form the theme of this letter.



damine **2**,<sup>3a</sup> valienamine **3** and valioline **4** and some of these and their derivatives have found commercial use. In aminocarbasugars **2–4**, the amino group is located on C-1. However, replacement of any of the other hydroxy groups by an amino group on the carbasugar platform **1** offers vast opportunities for structural and stereochemical variation, a point not missed by the enterprising synthetic community. As a result, many new types of

Our first foray towards aminocarbasugars and ‘confused’ aminocarbasugars emanated from the readily available 7-norbornanone derivative **6**, which undergoes smooth fragmentation to functionalized cyclohexanoid **7**, as recently reported by us.<sup>5b</sup> Elaboration of **7** to **8** was straightforward and further catalytic aminohydroxylation<sup>6</sup> proceeded stereoselectively and with good regioselectivity to furnish a mixture (4:1) of **9** and **10** (Scheme 1).<sup>7</sup> Reductive *N*-detosylation and removal of the protective groups in **9** and **10** led to **11** and **12**, respectively (Scheme 1).<sup>7</sup> While **11** turned out to be carbagalactovalidamine, whose synthesis has been described by two

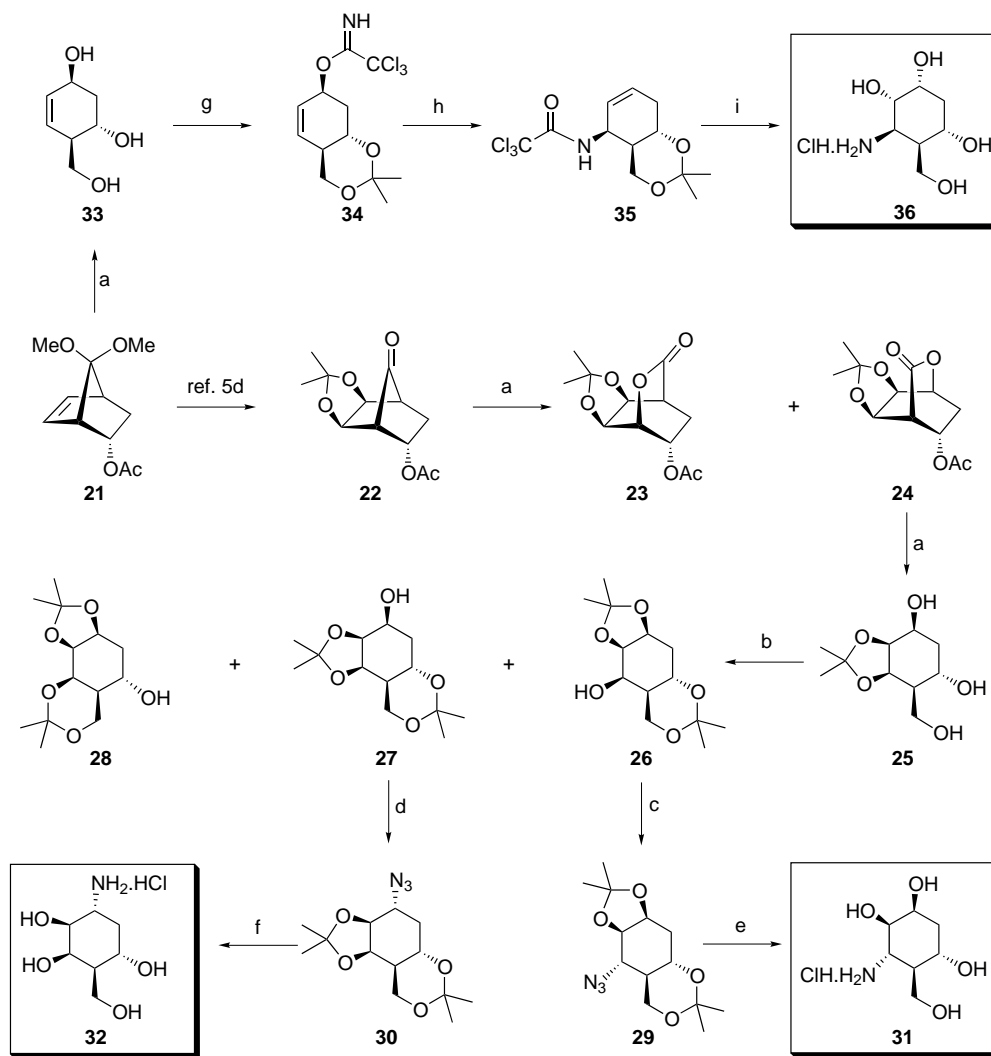
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**Scheme 1.** Reagents and conditions: (a) Ref. 5b; (b) Chloramine-T, OsO<sub>4</sub>, <sup>t</sup>BuOH–H<sub>2</sub>O (4:1), rt, 12 h, 70% (**9:10**; 4:1); (c) i. Ac<sub>2</sub>O, DMAP, DCM, rt, 2 h, 95%, ii. Na-Naphthalenide, DME, –60°C, 4 h, 65%, iii. 1N HCl, Δ, 20 h, 90%; (d) i. Ac<sub>2</sub>O, DMAP, DCM, rt, 2 h, 93%, ii. Na-Naphthalenide, DME, –60°C, 4 h, 45%, iii. 1N HCl, Δ, 20 h, quant.; (e) Ref. 5c; (f) NaN<sub>3</sub>, DMF, 100°C, 5 h, 92%; (g) i. H<sub>2</sub>, Pd/CaCO<sub>3</sub>, EtOH, rt, 3 h, ii. Ac<sub>2</sub>O, DMAP, DCM, rt, 60% (two steps); (h) 1N HCl, Δ, 20 h, 92%; (i) MCPBA, Na<sub>2</sub>CO<sub>3</sub>, DCM, rt, 4 h, 85% (87:13 regioisomeric mixture of lactones); (j) i. LAH, THF, 0°C–rt, 2 h, 85%, ii. Ac<sub>2</sub>O, DMAP, DCM, rt, 92%; (k) NaN<sub>3</sub>, DMF, 90°C, 5 h, 82%; (l) i. H<sub>2</sub>, Pd/CaCO<sub>3</sub>, EtOH, rt, 3 h, 80%, ii. 1N HCl, Δ, 25 h, 90%.

groups recently,<sup>4h,k</sup> the minor aminocarbasugar **12** is a new entity. A new carbasugar with an amino group in the side arm could also be accessed from the versatile precursor **7** through conversion to the iodomethyl derivative **13** and azide ion-mediated displacement to **14**. Catalytic hydrogenation to the amine functionality, characterized as amide **15** and deprotection furnished the new carbagalactosamine **16** (Scheme 1).<sup>7</sup> *endo*-Keto-tosylate **6** also proved to be an effective precursor for ‘confused’ aminocarbasugars. Baeyer–Villiger oxidation of **6** was regioselective (87:13) and lactone **17** was the dominant product (Scheme 1). LiAlH<sub>4</sub> reduction and acetylation gave protected cyclohexitol **18**. Displacement of the tosylate with azide ion in **18** was smooth and delivered the azido compound **19** (Scheme 1). Finally, the reduction of the azide functionality in **19** and a deprotection sequence delivered the first ‘confused’ aminocarbasugar **20** (Scheme 1).<sup>7</sup>

The successful acquisition of **20** and the inherent flexibility of our simple access to the ‘confused’ carbasugars presented an opportunity to generate diversity through placement of the amino group at various positions on the cyclohexitol framework. For this purpose, *endo*-keto-acetate **22**, obtained<sup>5d</sup> from abundantly available **21**, was found to be most serviceable. Baeyer–Villiger oxidation of **22** led to the regioisomeric lactones **23** and **24** in which the later predominated (13:87) (Scheme 2). The ‘confused’ carbasugar monoacetone derivative **25** obtained from **24** was subjected to bis-acetonide formation and led to a mixture (43:47:10) of bis-acetonides **26–28**,<sup>7</sup> leaving one hydroxyl group free in each one of them for further manipulation (Scheme 2). The hydroxyl group in the two major bis-acetonides **26** and **27** was transformed to the mesylate leaving group and then displaced with azide ion to furnish azido compounds **29** and **30**, respectively (Scheme 2). Reduction of the azido



**Scheme 2.** Reagents and conditions: (a) Ref. 5d; (b) Me<sub>2</sub>CO, Amberlyst-15, rt, 2 h, 78% (**26:27:28**; 43:47:10); (c) i. MsCl, Py, rt, 10 h, ii. NaN<sub>3</sub>, DMF, 100°C, 5 h, 65% (two steps); (d) i. MsCl, Et<sub>3</sub>N, DMAP, DCM, rt, 1 h, ii. NaN<sub>3</sub>, DMF, 100°C, 4 h, 70% (two steps); (e) i. H<sub>2</sub>, Pd/CaCO<sub>3</sub>, EtOH, rt, 3 h, ii. 1N HCl, Δ, 20 h, 80% (two steps); (f) same as (e), 85% (two steps); (g) i. Me<sub>2</sub>CO, Amberlyst-15, rt, 2 h, 81.5%, ii. CCl<sub>3</sub>CN, DBU, 0°C, 1 h, 93%; (h) i. K<sub>2</sub>CO<sub>3</sub>, *p*-xylene, reflux, 12 h, 70%; (i) i. OsO<sub>4</sub>, NMMO, Me<sub>2</sub>CO–H<sub>2</sub>O–*t*BuOH (5:5:2), rt, 2 days, 92%, ii. 1N HCl, Δ, 36 h, quant.

group in **29** and **30** through catalytic hydrogenation and deprotection delivered the ‘confused’ aminocarbascugars **31**<sup>7</sup> and **32**,<sup>7</sup> respectively (Scheme 2).

An interesting stereochemical variant could also be added to the design of aminocyclitols from **21**. We have recently shown that **21** could be readily transformed to the trihydroxy-cyclohexene **33** via regioselective Baeyer–Villiger oxidation and hydride reduction.<sup>5d</sup> Acetonide protection in **33** and reaction with trichloroacetonitrile led to the allyl trichloroacetimidate **34** and set the stage for an Overman rearrangement<sup>8</sup> to allyl trichloroacetamide **35** under thermal activation in the presence of a base (Scheme 2). Catalytic OsO<sub>4</sub>-mediated dihydroxylation on **35** was stereoselective and further deprotection led to a stereochemically diverse ‘confused’ aminocarbascugar **36**.<sup>7</sup>

The availability of so many new and diverse range of aminocyclitols **16**, **20**, **31**, **32** and **36** led us to explore in

a preliminary way their glycosidase inhibitory activity against a set of six commonly used enzymes ( $\alpha$ - and  $\beta$ -glucosidase,  $\alpha$ - and  $\beta$ -galactosidase, and  $\alpha$ - and  $\beta$ -mannosidase) following the standard protocol followed by us.<sup>9</sup> However, we have not observed any notable inhibition to report at this time and clearly more screening is warranted.

In short, we have amplified the utility of the norbornyl building blocks **6** and **22** by obtaining a range of aminocarbascugars and ‘confused’ aminocarbascugars, the latter as new entities for the first time, through simple, regio- and stereocontrolled operations.

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- All new compounds reported here were racemic and characterized on the basis of spectroscopic data (IR,  $^1\text{H}$ ,  $^{13}\text{C}$  NMR and mass). Selected spectroscopic data ( $J$  in Hz). **11**:  $\delta_{\text{H}}$  (300 MHz,  $\text{D}_2\text{O}$ ): 3.97 (1H, br d,  $J$  2.1), 3.87 (1H, ddd,  $J$  1.8, 4.8, 10), 3.61 (1H, br s), 3.51–3.48 (2H, m), 3.39 (1H, d 1/2 ABq,  $J$  6.3, 11), 1.78 (1H, m, q like), 1.68–1.65 (2H, m);  $\delta_{\text{C}}$  (75 MHz,  $\text{D}_2\text{O}$ ): 71.6 (CH), 69.8 (CH), 68.1 (CH), 62.6 ( $\text{CH}_2$ ), 52.0 (CH), 37.2 (CH), 24.1 ( $\text{CH}_2$ ). **12**:  $\delta_{\text{H}}$  (300 MHz,  $\text{D}_2\text{O}$ ): 4.12 (1H, br d,  $J$  3.3), 3.96 (1H, br s), 3.72 (1H, dd,  $J$  2.4, 10.5), 3.5 (1H, d 1/2 ABq,  $J$  8, 11), 3.37 (1H, d 1/2 ABq,  $J$  6.5, 11), 3.26–3.22 (1H, m), 1.99–1.87 (1H, m), 1.57 (1H, br td,  $J$  3.6, 14.4), 1.47–1.37 (1H, m);  $\delta_{\text{C}}$  (75 MHz,  $\text{D}_2\text{O}$ ): 69.9 (CH), 69.1 (CH), 66.2 (CH), 63.1 ( $\text{CH}_2$ ), 54.4 (CH), 36.8 (CH), 28.7 ( $\text{CH}_2$ ). **16**:  $\delta_{\text{H}}$  (300 MHz,  $\text{D}_2\text{O}$ ): 4.05 (1H, m), 3.98 (1H, br s), 3.65 (2H, m), 3.07 (1H, d 1/2 ABq,  $J$  7, 13), 2.94 (1H, d 1/2 ABq,  $J$  7, 13), 2.21–2.10 (1H, m), 1.65–1.61 (2H, m);  $\delta_{\text{C}}$  (75 MHz,  $\text{D}_2\text{O}$ ): 71.5 (CH), 71.3 (CH), 71.2 (CH), 69.2 (CH), 42.0 ( $\text{CH}_2$ ), 32.9 (CH), 29.0 ( $\text{CH}_2$ ). **20**:  $\delta_{\text{H}}$  (300 MHz,  $\text{D}_2\text{O}$ ): 3.97–3.91 (3H, m), 3.79–3.69 (2H, m, series of d), 3.39–3.33 (1H, m, dd like), 2.11 (1H, td,  $J$  6, 12), 1.33 (1H, m, q like), 1.45 (1H, dt,  $J$  3.3, 7.5);  $\delta_{\text{C}}$  (75 MHz,  $\text{D}_2\text{O}$ ): 84.6 (CH), 73.6 (CH), 68.7 (CH), 66.7 ( $\text{CH}_2$ ), 47.9 (CH), 45.1 (CH), 30.9 ( $\text{CH}_2$ ). **31**:  $\delta_{\text{H}}$  (300 MHz,  $\text{D}_2\text{O}$ ): 4.04 (1H, br s), 3.89 (1H, d 1/2 ABq,  $J$  3.3, 11.7), 3.78–3.63 (3H, m), 3.32 (1H, t,  $J$  11), 2.11 (1H, td,  $J$  4, 13.8), 1.65 (1H, ddt,  $J$  3.3, 7.5, 10.8), 1.56 (1H, dt,  $J$  2.1, 13.8);  $\delta_{\text{C}}$  (75 MHz,  $\text{D}_2\text{O}$ ): 72.2 (CH), 68.9 (CH), 64.5 (CH), 60.7 ( $\text{CH}_2$ ), 53.1 (CH), 47.4 (CH), 38.5 ( $\text{CH}_2$ ). **32**:  $\delta_{\text{H}}$  (300 MHz,  $\text{D}_2\text{O}$ ): 4.04 (1H, t,  $J$  2.7), 3.76 (1H, d 1/2 ABq,  $J$  4.5, 11), 3.66 (1H, dt,  $J$  4.5, 10.8), 3.59 (1H, d 1/2 ABq,  $J$  8.4, 11.1), 3.53 (1H, dd,  $J$  2.7, 10.8), 3.28 (1H, m, dt like), 2.20 (1H, td,  $J$  4.5, 12), 1.59 (1H, dddd,  $J$  2.7, 4.5, 7, 8.4), 1.43 (1H, q,  $J$  12);  $\delta_{\text{C}}$  (75 MHz,  $\text{D}_2\text{O}$ ): 72.9, 69.8, 65.6, 60.4, 49.8, 48.9, 36.8. **36**:  $\delta_{\text{H}}$  (300 MHz,  $\text{D}_2\text{O}$ ): 3.9–3.86 (2H, m), 3.82 (1H, dd,  $J$  3.3, 8.4), 3.69–3.64 (2H, m), 3.57 (1H, d 1/2 ABq,  $J$  5.7, 11), 2.22–2.19 (1H, m), 1.80–1.75 (2H, m);  $\delta_{\text{C}}$  (75 MHz,  $\text{D}_2\text{O}$ ): 68.9 (CH), 68.7 (CH), 66.8 (CH), 59.7 ( $\text{CH}_2$ ), 51.6 (CH), 44.3 (CH), 33.9 ( $\text{CH}_2$ ).
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