

## Antiviral Effects, of a synthetic Aluminium-Magnesium Silicate, on Avian Influenza Virus .

<sup>1</sup>Ezeibe, M. C. O.,<sup>2</sup> Egbuji, A. N.,<sup>1</sup>Okoroafor, O. N., <sup>1</sup>Eze,J.I., <sup>1</sup>Ijabo, O., <sup>1</sup>Ngene,A.A, <sup>3</sup>Eze, I. C. <sup>3</sup>Ugonabo, J. A. C.,<sup>1</sup>Sanda, M. E. and <sup>1</sup>Mbuko, I. J.

1 Department of veterinary medicine, University of Nigeria, Nsukka.

2 Viral research laboratory, National Veterinary Research Institute, Vom, Nigeria.

3 Department of Microbiology, University of Nigeria, Nsukka.

### Summary.

Effects of a synthetic Aluminium-Magnesium Silicate [AMS] on *Avian Influenza Virus (AIV)* were tested. Equal amounts of H<sub>5</sub>N<sub>1</sub> AIV samples and of AMS were mixed, left one hour, at room temperature before centrifuging. The supernatants were remeasured and tested for viral titre, for Mean Death Time (MDT) and Embryo Mortality Rate (EMR) of chicken eggs.

Volumes of the viral samples reduced at rate of  $23.4 \pm 5.48$  %. Viral titres reduced significantly ( $P = 0.001$ ) from HA,  $73 \pm 32.72$  to  $1.4 \pm 0.43$  . Also, mortality of infected embryos reduced from 100 % to 65% while MDT of those that died, increased significantly ( $P = 0.001$ ) from  $76 \pm 4.38$  to  $130 \pm 17.27$  hours .When incubation with AMS was repeated on portions of same sample ,MDT increased from 64 to 104 hours with the portion incubated once .Two AIV portions on which incubation with AMS was repeated could not kill chick embryos.

### Background.

*Avian Influenza Viruses* are enveloped, single stranded RNA viruses. Two different protein projections cover their envelopes. These are the Haemagglutinin (H) and the Neuraminidase (N) antigens. Combinations of the H and the N antigens is an important property of *Influenza viruses* because, it allows for reassortment when two *Influenza viruses* replicate in same cell at the same time.

Influenza viruses are classified into types, A, B and C, based on antigenic differences of their nucleocapsid and of their matrix proteins<sup>1</sup>. *Avian Influenza Viruses* are *Influenza A viruses*<sup>2</sup>.

*Influenza A viruses* are subdivided serologically, into sixteen H and nine N subtypes. All the subtypes have been identified in avian species and they coexist with their natural hosts (water fowls and sea birds) in perfect harmony<sup>2</sup>. Disease outbreaks occur in domestic birds when they come in contact with these carriers or with infected pigs<sup>2</sup>. Infected birds shed *Influenza viruses* in their secretions and excretions. So, domestic birds can also get infected when they contact secretions or excretions of infected wild birds or fomites<sup>3</sup>.

*Avian Influenza* viruses cause two main forms of disease, distinguished by low or high virulence in domestic poultry. The low pathogenic form causes only ruffled feathers and drop in egg production

while the Highly Pathogenic *Avian Influenza Viruses* (HPAIV) spread rapidly between flocks, affect many organs of infected birds and lead to high mortality within 48 hours<sup>3</sup>.

Genetic characterization of *Avian Influenza Virus* samples isolated from outbreaks in Nigeria, in 2007 revealed a new reassortant virus (H<sub>5</sub>N<sub>1</sub>)<sup>4</sup>. Reassortment occurs when an *Influenza virus* infects a human being or a pig that is already infected with another *Influenza virus*. Such reassortment of antigens (viral shift), can produce *Influenza virus strains*, different from available vaccine strains. Such viral shift led to human pandemics of 1918 and of 1957 - 1958. So, isolation of a new *Influenza Virus* subtype in Nigeria is of great concern to the country, both for her poultry industry and for public health.

AMS is safe when ingested by man or animals. For this reason, it is used in many pharmaceutical formulations<sup>5</sup>. It is used in tableting drugs because, its molecules have one of their ends positively charged and the other, negatively charged<sup>6</sup>. Viruses have electrical charges too. Some have net positive electrical charges while others have net negative charges<sup>7</sup>.

These electrical charges could make *Influenza Virus* particles adsorb onto molecules of AMS. If this happens, first stage of viral infection, which is adsorption to hosts' cells may be inhibited.

Natural AMS has impurities<sup>6</sup>. These could cause adverse reactions on treated animals if the natural ore is ingested at the high doses that may be required to treat viral diseases. So, two medicinal minerals which occur in Nigeria, Aluminium Silicate {Al<sub>4</sub>(SiO<sub>4</sub>)<sub>3</sub>} and Magnesium Silicate {Mg<sub>2</sub>SiO<sub>4</sub>} were reacted to get a synthetic AMS:  $Al_4(SiO_4)_3 + 3Mg_2SiO_4 \rightarrow 2Al_2Mg_3(SiO_4)_3$ .<sup>8</sup>

### Summary of Methods:

Samples of the H<sub>5</sub>N<sub>1</sub> AIV, isolated from outbreaks of *Bird Flu* in Nigeria were incubated with the AMS. To a measured volume of each viral sample, equivalent amount of the synthetic AMS (on volume to weight basis) was added. They were thoroughly mixed and allowed to stand at room temperature for one hour before they were centrifuged at 3000 revolutions per minute for ten minutes. Volumes of their supernatants were remeasured and tested for viral titre by the haemagglutination method<sup>9</sup>. Also, 0.1ml of each supernatant was inoculated into yolk sac of two embryonated chicken eggs to determine Mean Death Time (MDT) and Embryo Mortality Rates (EMR) of infected chicken embryos. The eggs were candled every 12 hours to check for embryo mortality. Embryos that remained alive 208 hours post infection (PI), were recorded to have survived the infection with the AIV.

As controls, intact portions of each of the AIV samples were used for HA test along with supernatants of their portions incubated with the synthetic AMS, on same plates. Also, portions of each of the intact viruses were used for egg inoculation along with supernatants of their portions incubated with the AMS. Means of the viral titres and of the MDT of the two groups were used to test the null hypothesis that AMS has no effect on titre and pathogenicity of AIV, by the Student T – test (n =10).

Portions of a sample of the, H<sub>5</sub>N<sub>1</sub> AIV which gave high HA titre were used to test effect of repeating incubation of AIV with AMS, on titre of the virus and on MDT and EMR of chicken embryos inoculated with it. One portion was incubated with the AMS once. To a second portion, after the centrifugation,

equal amount of the AMS was mixed with the supernatant and the incubation process was repeated once. In the third portion, incubation with the AMS was repeated twice. The three portions were then tested for viral titre and for MDT and EMR of chicken embryos inoculated with AIV.

### **Findings.**

Volumes of the viral samples reduced at a mean rate of  $23.4 \pm 5.48$  % following incubation with the AMS. Also, incubation with the AMS significantly ( $P = 0.001$ ) reduced titre of the H<sub>5</sub>N<sub>1</sub> AIV from a mean HA,  $73 \pm 32.72$  to  $1.4 \pm 0.43$  and the EMR ( $P = 0.001$ ) from 100 to 65 % .Incubating AIV with the AMS increased MDT of chicken embryos inoculated with the virus significantly ( $P = 0.002$ ) from  $76 \pm 4.38$  to  $128.00 \pm 18.36$  hours.

Repeating incubation of AIV with the AMS reduced titres of the viral samples from HA, 128 to HA, 2 in both the portion incubated with AMS once and in the two portions incubated twice and thrice respectively. EMR of chicken embryos was 100% in the control and in the group inoculated with AIV portion incubated with AMS once, but their MDT increased from 64 in the control to 104 hours in the group inoculated with AIV portion incubated with the AMS once . In the group of eggs inoculated with portions of the AIV incubated with AMS twice and thrice respectively, there was no embryonic death, 208 hours PI.

Effects the AMS had on volume of the viral samples, on the viral titres and on MDT and EMR of chicken embryos inoculated with the *Avian Influenza Virus* are as on tables 1, 2 and 3

**Table 1: Effect of incubating H<sub>5</sub>N<sub>1</sub> Avian Influenza Virus with a Synthetic Aluminium – Magnesium Silicate on Volume of the viral samples.**

Viral samples	Volume before incubation	Volume after incubation	% Reduction
	With AMS (ml)	with AMS (ml)	
1	13	10	21.1
2	9	6.5	21.8
3	6	3.5	30
4	3	2.5	16.7
Mean	7.78± 4. 27	5.6± 3.38	23.4±5. 48

Incubating AIV with the AMS reduced volume of the viral samples significantly [P = 0. 001].

**Table 2:Effect of Aluminium – Magnesium Silicate on Titre of H<sub>5</sub>N<sub>1</sub> Avian Influenza Virus**

Virus samples	HA Titre		
	Control	Incubated with AMS	
1	256	4	
2	256	0	
3	128	0	
4	32	2	
5	32	2	
6	8	0	
7	8	2	
8	4	2	
9	4	2	
10	2	0	
Mean		73±32.72	1.4 ± 0.43

Incubating AIV with AMS reduced the viral [HA] titre (P = 0.001).

Table 3: Effect of Aluminium – Magnesium Silicate on Mean Death Time and on Embryo Mortality Rates of Chicken Eggs inoculated with H<sub>5</sub>N<sub>1</sub> Avian Influenza Virus.

Virus samples	MDT(Hours)		EMR(%)	
	Incubated with AMS	Control	Incubated with AMS	Control
1	112	88	100	100
2	52	88	100	100
3	76	40	100	100
4	64	88	100	100
5	148	76	50	100
6	148	76	100	100
7	136	76	100	100
8	208	76	0	100
9	208	76	0	100
10	118	76	0	100
Mean	128.00 ±18.36	76±4.38	65 ± 15.00	100 ± 0.00

Incubating AIV with AMS increased MDT (P = 0.002) and reduced EMR (P = 0.001) of Embryonated chicken eggs inoculated with the virus.

### Discussion.

Brooks<sup>10</sup> defined antiviral agents as substances which kill viruses or substances which inhibit replication or pathogenicity of viruses. Stern *et al*<sup>11</sup> had reported that pathogenicity of myxoviruses reside in their haemagglutinin antigen. Since the synthetic Aluminium – Magnesium Silicate inhibited HA titre which is activity of the haemagglutinin antigen of H<sub>5</sub>N<sub>1</sub>, Avian Influenza virus, reduced the rate at which the virus killed chick embryos and delayed death even in the embryos that were killed, it suggests the AMS had antiviral effect against the virus. Reduction in volume of the viral samples suggests that the synthetic AMS adsorbed onto water molecules in the samples. This action of the synthetic AMS agrees with reported effect of natural AMS on water<sup>5</sup>. Reduction in volume of viral samples should lead to increase in the viral titre if the AMS had no effect on the virus. However, the viral titres reduced in spite of the reduction in volume. This suggests that the electrostatic attraction

between the AMS molecules and the viral particles was more than that between them and water molecules. Reduction in titre of the HPAI virus and inhibition of the viral activities by AMS as seen in these experiments agree with earlier results of effect of the synthetic AMS on *Peste des Petits Ruminants Virus*<sup>12</sup>, on *Infectious Bursal Disease Virus*<sup>13</sup>, on *Egg drop syndrome 76 virus*<sup>14</sup>, on *Canine Parvovirus*<sup>15</sup> and on *Newcastle Disease Virus*<sup>16</sup>.

AMS is safe when ingested by man and by animals<sup>5,17</sup>. It is used to bind drugs to make tablets used in treating both humans and animals<sup>6</sup>. Foster and Smith<sup>18</sup> reported that AMS has been in use for treatment of ulcer for many years. Also, the synthetic AMS tested in this study, was got by reaction of Aluminium Silicate and Magnesium Silicate<sup>8</sup> which are medicines being used to treat both animals and human beings. So, the synthetic AMS should be safe for treating viral diseases of man and of animals. It can be used for systemic treatment of infectious diseases, because, once there is inflammation of mucous membranes of the stomach or of the intestines even unabsorbable substances can pass into the blood stream<sup>19</sup>. Also, simple sugars are reported to carry drugs across intact mucous membranes by active transport<sup>20</sup>. Dextrose monohydrate incorporated in the AMS<sup>8</sup> would carry the AMS across even uninflamed mucous membranes. Ability of the AMS to inhibit replication or pathogenicity of the HPAI virus as suggested by the significant increase in MDT and decrease in EMR recorded in these experiments shows that it could be a good candidate for development of a drug for control of Bird Flu in poultry and for management of human cases of Avian Influenza.

## References.

1. Jordan, F. W. T. (1990). *Poultry diseases* 3rd Edit. Bailliere Tindal, London.
2. Webster, R. G. Peiris, M. Chen, H. and Guan, Y. (2006). H<sub>5</sub>N<sub>1</sub> outbreaks and enzootic influenza. *Emerg. Infect. Dis.* **12** : 3 – 8.
3. WHO (2005). Avian influenza A (H<sub>5</sub>N<sub>1</sub>) infection in humans : urgent need to eliminate the animal reservoirs. <http://www.who.int/csr/don/2004.03.02/en/index.html>. Accessed 31 October, 2005. 17. 03 pm
4. Monne, I. et al (2008). Resistant Avian Influenza Virus (H<sub>5</sub>N<sub>1</sub>) in poultry in Nigeria in 2007. *Emerg. Infect. Dis.* **14** (4) : 637 – 640.
5. Verndabilt, R. T. (1992). Inc. Technical literature: Veegum® the versatile ingredient for pharmaceutical formulations pp 1 – 5.
6. Venkatakrisnan, R. (1995). *Geology – An introduction to physical geology*. Worth publishers, New York.
7. Cann, A. J. (1993). *Principles of Molecular virology*. Academic press, USA.
8. Ezeibe, M. C. O. (2006). Admacine®. Federal Republic of Nigeria. Patents and Designs Act. Cap. 344 LDN 1990. No. 16448,

9. Johnson, R. H. (1971). Serological procedures for the study of Feline panleukopaenia. *J. Am. Vet. Med. Assoc.* **158** :876 -884.
10. Brooks, G. F. (1998). Medical microbiology. 21st Edit. *Mc Graw Hill education Inc.* San Francisco.
11. Stern, I. B., Greenberg, M., Gersoni, J. M., and Rozenblatt, S. (1995). The haemagglutination envelope protein of *Canine distemper virus (CDV)* confers cell tropism as illustrated by *CPV – Measles virus* complementation analysis. *J. Virol.* **69** (3) :1661 -1668.
12. Ezeibe, M. C. O. *et al* (2009a). Antiviral effects of Aluminium – Magnesium Silicate on *Peste des Petits Ruminants Virus*. *Anim. Sci. Rep.* **3** (4) : 141 – 147.
13. Ezeibe, M. C. O. *et al* (2009 b ). In vitro and in vivo effects of Aluminium – Magnesium Silicate on *Infectious Bursal Disease Virus* of chickens. *Anim. Sci. Rep.* **3** (4) : 132 – 137.
14. Ezeibe, M. C. O. *et al* (2010 a). Haemagglutination and Haemagglutination – Inhibition titres of *Egg Drop Syndrome 76 Virus* treated with Aluminium – Magnesium Silicate. *Anim. Sci. Rep.* **4** (3) : 87 – 90.
15. Ezeibe, M. C. O. *et al* (2010b). Aluminium – Magnesium Silicate inhibits *Canine Parvovirus* and cures infected dogs. *Health* **2** (10) : 1215 – 1217.
16. Ezeibe, M. C. O. *et al.* (2011). Effects of Aluminium – Magnesium Silicate on *Newcastle Disease Virus* and on recovery of infected chicks. *Int. J. Biol. Chem.* **5** (2): 835 – 839.
17. Windholz M. B. (1978). *The Merck index*. Merck and co. Inc. New Jersey USA.
18. Foster, E. and Smith, K. (2007). Aluminium – Magnesium Silicate used as antacid, antiulcer and to control diarrhea. *Ren. Nutri.* **4** (11) : 50 – 53.
19. Greene, C. E. (1990 ). *Infectious Diseases of dogs and cats*. *W. B. Saunders*, Philadelphia.
20. Murray, K. R. (2000). *Harpers Biochemistry*. *McGraw Hill*, New York.