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Modulation of the Bovine Delayed-Type Hypersensitivity Responses to Defined Mycobacterial Antigens by a Synthetic Bacterial Lipopeptide

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Bovine tuberculosis (TB) is a disease of economic and zoonotic importance caused by the intracellular bacterium Mycobacterium bovis. In the United Kingdom, there has been a steady rise in the incidence of bovine TB during the last decade, and the cost of control measures currently exceeds £25 million per annum. Bovine TB is a zoonotic disease, and prior to introduction of milk pasteurization and disease control measures in England, up to 6% of human deaths (2,500 people per annum) resulting from TB were attributed to M. bovis infection (19). Furthermore, the zoonotic impact of bovine TB remains an important consideration in developing countries in which limited or no control measures are in place (26, 27). The early and specific diagnosis of this disease in cattle is therefore of vital importance.

The control of bovine TB in the United Kingdom relies on detection of infected animals by the single comparative intradermal tuberculin test (SCITT) (32). This test detects the development of a delayed-type hypersensitivity (DTH) response to intradermal injection of a purified protein derivative (PPD) prepared from an M. bovis culture. The reaction to bovine PPD (PPD-B) is compared to the skin reaction induced by a Mycobacterium avium-derived PPD to provide a measure of sensitization to environmental mycobacteria. Cattle presenting with a positive skin test are slaughtered (test and slaughter control policy).

Recent advances in the detection of bovine and human TB have demonstrated that the use of defined protein and peptide antigens can overcome specificity limitations of purified protein derivatives in the detection of bovine tuberculosis when the antigens are used in blood-based tests. Since the use of these specific antigens as skin test reagents could have practical advantages, we investigated the potential of Mycobacterium bovis-specific antigens to stimulate delayed-type hypersensitivity (DTH) responses in cattle experimentally infected with M. bovis. A cocktail of the recombinant antigens ESAT-6, MPB83, and MPB64 failed to stimulate in vivo DTH in cattle that had been experimentally infected with M. bovis despite the fact that the antigens were recognized in vitro by the same animals. However, it was possible to stimulate antigen-specific bovine DTH responses by using ESAT-6 in combination with a synthetic bacterial lipopeptide. This lipopeptide stimulated the release of the proinflammatory cytokine tumor necrosis factor alpha from monocyte-derived bovine dendritic cells in vitro, thereby providing a possible mechanism for its DTH-enhancing properties.

The purpose of the present study was to investigate the diagnostic potential of defined M. bovis antigens as skin tests reagents in cattle. We report here that a protein cocktail comprising recombinant forms of the ESAT-6, MPB64, and MPB83 antigens was found to be a potent DTH-inducing antigen combination in a guinea pig model, yet it failed to induce DTH responses in M. bovis-infected cattle despite the fact that it was recognized in vitro in the same animals. However, the failure of the defined mycobacterial antigens to induce bovine DTH responses was overcome by coadministration of a synthetic bacterial lipopeptide with proinflammatory properties.

**MATERIALS AND METHODS**

**Bacterial strains.** M. bovis strain AN5 was supplied by the Tuberculin Production Unit, Veterinary Laboratories Agency (VLA), Addlestone, United Kingdom. M. bovis strain AF2122/97 was supplied by the TB Diagnostic Unit (VLA). Both strains were stored frozen in Middlebrook 7H9 culture media (Becton Dickinson, Oxford, United Kingdom). The number of viable CFU was determined by plated an aliquot of a strain on Middlebrook 7H10 agar (Becton Dickinson) supplemented with 10% (vol/vol) oleic acid-albumin-dextrose-catalase (Becton Dickinson). Agar plates were also supplemented with 0.2% (vol/vol) oleic acid-albumin-dextrose-catalase (Becton Dickinson). Agar plates were also supplemented with 0.2% (vol/vol) oleic acid-albumin-dextrose-catalase (Becton Dickinson).
glycerol (Sigma, Poole, United Kingdom) for growth of AN5 and with 4.16 g of pyruvic acid (Sigma) per liter for growth of AF2122/97.

Mycobacterial antigens. The recombinant M. bovis antigens ESAT-6, MPB70, and MPB83 were produced as previously described (43). Recombinant MPB64 was provided by D. Bakker, Animal Health Science, Boxtel, The Netherlands. PPD-B and avian PPD (PPD-A) were produced and supplied by the Tuberculins Production Unit (VLA) according to procedures prescribed by the European Pharmacopoeia (12). For assessment of the purity of antigens, samples were separated on 8 to 18% precast gel sodium dodecyl sulfate (SDS)-polyacrylamide gel electrophoresis (PAGE) gels (Amersham-Pharimacia Biotech, Little Chalfont, United Kingdom), and electrophoresis was performed by using a Multiphor II system (Amersham-Pharimacia Biotech). Sample purity was assessed by performing total-protein staining of gels with a commercial silver staining kit (Amersham-Pharimacia Biotech) and by using a standard Western blot procedure to probe for the presence of contaminating Escherichia coli proteins with rabbit anti-E. coli polyclonal antibody sera (Dako Ltd., Ely, United Kingdom).

Protease treatment of PPD-B was performed with a lypophilized aliquot of international standard bovine tuberculin PPD (NIBSC, Potters Bar, United Kingdom). The PPD-B standard was reconstituted at a concentration of 1 mg/ml and then incubated (1-ml aliquot) in the presence of 5 μl of a 10-mg/ml solution of proteinase K (Sigma) for 5 min at 37°C. The sample was then incubated at 65°C for 5 min to heat inactivate the enzyme. The proteinase K treatment was repeated two more times. The sample was then incubated in the presence of 10 μl of a 10 mg/ml solution of pronase (Roche Diagnostics Ltd., Lewes, United Kingdom) for 90 min at 25°C. Finally, the pronase was heat inactivated at 100°C for 10 min.

Protein concentrations of noncommercially derived materials were determined with a bicinchoninic acid protein assay kit (Perbio Science UK Ltd., Tattenhall, United Kingdom).

Cattle and guinea pig experimental infections. Guinea pigs that weighed 350 to 400 g and were free of intercurrent infection were obtained from Charles Tattenhall, United Kingdom. Guinea pigs were infected with 100 CFU of M. bovis AN5 in 0.25 ml of phosphate-buffered saline (PBS) by intramuscular injection into the iliacus muscles of the croup region of the right hind leg. The route of infection and the strain of M. bovis and the route of infection and the strain of M. bovis were derived from CD14+ monocytes that were isolated from PBMC and cultured with granulocyte-macrophage colony-stimulating factor and interleukin-4 (IL-4) as previously described by Hope et al. (23). After 3 days of culture, the synthetic bacterial lipopeptide Pam3CSK4 (EMC Microcollections, Tubingen, Germany) (42, 45), where PamC is a palmitoyl-Cys[i(11)]-3,23,4-dipalmitoyl)-propyl residue, was added to the DC. Supernatants were harvested 48 h later and assayed for the presence of bovine tumor necrosis factor alpha (TNF-α) by using a luminometric enzyme-linked immunosorbent assay (ELISA) protocol.

TNF-α luminoimetric ELISA. Antibodies to TNF-α were generated by immunization of mice with recombinant bovine TNF-α and hybridomas generated in a manner similar to that described previously for IL-12 (22). A pair of monoclonal antibodies, designated CC327 and biotinylated CC328, were shown to recognize recombinant and native bovine TNF-α in ELISA (J. C. Hope, L. S. Kwong, and C. J. Howard, unpublished data) when a protocol similar to that developed for assaying IL-12 (22) was used. U-bottom black 96-well plates (Porvair, Shepperton, United Kingdom) were coated with 2 μg of CC327 per ml overnight at room temperature. The plates were washed with PBS containing 0.01% Tween 20 (wash buffer) and then blocked by using sodium casein (1 mg/ml in PBS) for 1 h at room temperature. The plates were washed, and DC culture supernatant or a COS cell-derived TNF-α standard (Institute for Animal Health) was added to the appropriate wells. Following a 1-h incubation at room temperature, the plates were washed and then incubated in the presence of biotinylated CC328 (1 μg/ml) for an additional 1 h. The plates were washed and then incubated in the presence of horseradish peroxidase-labeled streptavidin (Amersham-Pharimacia Biotech) for 45 min at room temperature. Following the final wash, the Super Signal ELISA femto maximum-sensitivity substrate (Perbio) was added to each well, and the number of relative light units was determined 1 min later with an Anthos LUCY 1.0 lumimeter (AnthosLabtec, Salzbur, Austria). The number of biological units of TNF-α per milliliter was calculated by comparison with the bovine standard.

RESULTS

Screening of candidate antigens in a guinea pig DTH model. The homogeneity of recombinant antigens ESAT-6, MPB64, MPB70, and MPB83 was confirmed prior to investigation in animals by SDS-PAGE analysis (Fig. 1). While ESAT-6,
MPB64, and MPB70 produced single bands at the expected molecular masses, an additional band at a molecular mass of 50 kDa was obtained for MPB83 along with the expected product at a molecular mass of 25 kDa (Fig. 1A, lane 4). However, this band, as well as the 25-kDa product, reacted with an MPB83-specific monoclonal antibody (data not shown) but not with anti-\textit{E. coli} antibodies (Fig. 1B, lane 4). We therefore concluded that this 50-kDa product was a dimeric form of recombinant MPB83.

The DTH-inducing potential of these antigens was first tested in a guinea pig skin test model to help define an optimized antigen combination that could be used for further investigations in cattle. Each of the antigens stimulated DTH in a proportion of the guinea pigs tested, and ESAT-6 induced DTH responses most frequently (Fig. 2) (for MPB64 there were responses in three of eight animals; for MPB70 and MPB83 there were responses in four of eight animals; and for ESAT-6 there were responses in six of eight animals). The combination of ESAT-6 and MPB83 stimulated DTH responses in all of the guinea pigs (12.7 ± 0.5 mm) (Fig. 2). However, the optimal protein cocktail contained ESAT-6, MPB64, and MPB83 since this cocktail induced significantly stronger DTH responses ($P > 0.05$) than the ESAT-6–MPB83 combination (14.5 ± 1.5 mm). Addition of MPB70 to this cocktail resulted in no significant increase in the strength of the responses observed (14.4 ± 1.7 mm). In contrast to PPD-B, which gave an increase in skin thickness of 15.6 ± 1.3 mm, none of these proteins induced skin test responses in BCG-vaccinated guinea pigs when the proteins were tested individually or in combination (data not shown), thereby confirming their high specificity.

**DTH response to mycobacterial antigens in cattle.** The most promising protein combination identified in guinea pigs (ESAT-6, MPB64, and MPB83) was then tested as a skin test reagent in five calves experimentally infected with \textit{M. bovis} approximately 16 weeks postinfection. PBMC were cultured in the presence of protein cocktail (5 \mu{g} of each component protein per ml), PPD-B, or PPD-A (10 \mu{g}/ml) for 6 days, and incorporation of $[^3\text{H}]$thymidine was determined during the final 18 h of culture. Responses to the protein cocktail were considered positive if the SI (antigen-stimulated proliferative response/no-antigen response) was ≥2 and proliferation was ≥1,000 cpm. Skin test responses in the same animals were determined by intradermal administration of the protein cocktail (20 \mu{g} of each component antigen) or PPD-B plus PPD-A (100 \mu{g}). The increase in skin induration was measured 72 h later.

![FIG. 2. DTH responses to mycobacterial antigens in guinea pigs.](image)

![FIG. 3. Lymphocyte transformation assay and DTH responses to mycobacterial antigens in cattle.](image)

4 mm greater than the response induced by PPD-A, thereby confirming the tuberculin skin test-positive status of the animals (Fig. 3B). In contrast, in comparison to the PBS control in the same animals, the protein cocktail induced no DTH response in three of the calves and a poor response (2 mm) in the remaining calves (Fig. 3B).

**Bacterial lipopeptide can promote antigen-specific DTH responses in cattle.** We postulated that since we had determined that antigen-specific T cells were present in the peripheral blood of our \textit{M. bovis}-infected cattle (Fig. 3A), the inability of the protein cocktail to induce bovine DTH was likely to be due in part to a failure to recruit effector T cells to the site of
antigen administration due to an inability to induce proinflammatory cytokines. Therefore, we investigated whether potential proinflammatory stimuli were able to promote the DTH responses induced by recombinant mycobacterial protein antigens. As PPD itself has been shown to induce proinflammatory cytokines during DTH responses (9), we first assessed whether nonprotein components of PPD were able to enhance specific DTH responses to ESAT-6, MPB64, and MPB83. We therefore formulated these proteins together with protease-treated PPD-B, which by itself does not stimulate DTH in M. bovis-infected cattle or guinea pigs (data not shown), and we tested the DTH-inducing properties of this antigen formulation in three M. bovis-infected calves. However, protease-treated PPD-B failed to promote DTH responses to the three recombinant antigens in these calves (data not shown).

We next investigated the potential DTH-enhancing properties of a synthetic bacterial lipopeptide, Pam3CSK4. This model lipopeptide contains acyl moieties that mimic those identified in bacterial lipoproteins (47), and it has been shown to be capable of inducing TNF-α from murine and human monocytes (4, 21), a proinflammatory cytokine important in the in vivo recruitment of T cells (25). This lipopeptide was tested in combination with ESAT-6, the immunodominant antigen of the protein cocktail, in seven calves experimentally infected with M. bovis. Each of these calves exhibited positive recognition of ESAT-6 in vitro, as determined by the T-cell proliferation assay (median SI, 5.9; SI range, 2.0 to 50.6). As expected, ESAT-6 alone failed to induce a DTH response in any of these calves. However, when ESAT-6 was coadministered with the lipopeptide, DTH responses were induced in four of the animals (Fig. 4A). The modulation of ESAT-6-induced DTH responses by the lipopeptide in these infected calves was demonstrated to be antigen specific since the lipopeptide control in these calves induced no measurable response (Fig. 4A) and, furthermore, when ESAT-6 and the lipopeptide were administered alone or in combination, they stimulated no measurable DTH response in four SCITT-negative uninfected calves (reaction sizes, 0 mm).

To investigate a molecular mechanism by which the synthetic lipopeptide could stimulate enhanced inflammatory responses, we quantified the presence of the proinflammatory cytokine TNF-α in culture supernatants of monocyte-derived bovine DC stimulated with Pam3CSK4. As shown in Fig. 4B, Pam3CSK4 was able to induce bovine DC to produce TNF-α in a dose-dependent manner.

**DISCUSSION**

Defined antigens of M. bovis have demonstrated much promise in overcoming specificity limitations of PPD for diagnosing bovine TB when they are used in blood-based in vitro assay formats (8, 14, 35, 36, 43). However, investigation of such antigens as bovine skin test reagents has not been reported previously and therefore was the subject of the present study. ESAT-6, MPB64, and MPB70 have previously been shown to be capable of stimulating DTH responses in guinea pigs that were infected or sensitized with either live Mycobacterium tuberculosis, M. bovis BCG, or heat-killed M. bovis strains (1, 11, 15, 18, 31, 34). The demonstration that recombinant forms of these antigens also induced DTH in M. bovis-infected guinea pigs in the present study confirmed that DTH-inducing antigens may be useful as skin test reagents in cattle. In addition, we also identified MPB83 as another promising DTH-inducing antigen whose potential for diagnosing bovine TB through detection of cellular immune responses has been demonstrated previously only in vitro (14, 43).

Having defined an antigen cocktail that induced DTH responses in guinea pigs that were comparable to those induced
by PPD-B, we tested the ability of this cocktail to detect *M. bovis* infection in the target host. However, this cocktail failed to stimulate measurable DTH in infected calves. These calves were not deficient in the ability to mount DTH responses since PPD-B induced strong skin responses in each of them. This observation is analogous to the results of a clinical study in which the workers investigated MPT64 as a skin test reagent in TB patients. Like the proteins tested in the present study, MPT64 was selected for a clinical investigation due to its ability to induce specific and potent DTH responses in *M. tuberculosis*-infected guinea pigs (18, 20). However, it also failed to induce DTH in the majority of the patients tested (46). We considered that the explanation for the exquisite sensitivity of guinea pigs to induce DTH compared with the sensitivities of other hosts may be related to differences in the cellular compositions of the dermis and epidermis. The cells present in the skin that have an important role in mediating DTH include mast cells, which increase vascular permeability through the release of vasoactive amines (2, 16), and Langerhans cells, which are potent antigen-presenting cells of the DC lineage (39). However, a review of literature indicated that the densities of Langerhans cells in the epidermis of guinea pigs and in the epidermis of cattle are equivalent (6, 38), while the densities of mast cells in cattle dermis and human dermis are actually greater than the density observed in guinea pigs (17, 28). Therefore, while the differences in sensitivity between guinea pig and cattle (and human) DTH remain unclear, these data suggest that caution should be used when attempting to extrapolate quantitative DTH data between these species.

The calves used in the present study were considered suitable for investigating the DTH-inducing properties of the defined protein cocktail since we were able to demonstrate that in each animal, T cells present in PBMC recognized the cocktail in vitro. However, while in vitro assays demonstrate that antigen-specific T cells have been generated, such assays do not model the inflammatory process that is required to recruit antigen-experienced T cells to sites of local antigen exposure in vivo. It has long been recognized that injection of protein antigens in the absence of a suitable adjuvant results in poor humoral and cell-mediated immune responses. In a recent review of the in vivo recruitment of CD4+ T cells (24), the workers proposed that the role of the adjuvant is to stimulate the release of proinflammatory cytokines, such as TNF-α and IL-1, which promote migration of DC both to the site of antigen administration and subsequently to the draining lymph nodes after digestion of the antigen. The presentation and recognition of the antigen by memory T cells in turn lead to an influx of monocytes, macrophages, and T cells to the site of antigen administration. In the absence of an adjuvant stimulus, presentation of the antigen should be insufficient due to the low level of DC migration that occurs under noninflammatory conditions (24). Therefore, we concluded that the poor performance of the protein cocktail as a skin test reagent in our cattle was probably due in part to the absence of a suitable proinflammatory stimulus.

PPD is itself a potent DTH-inducing antigen that, due to its crude method of manufacture, can contain up to 15% nonprotein components, including DNA and cell wall-derived lipids and carbohydrates (30). Since mycobacterial cell wall components and DNA are both known to be potent adjuvants for cell-mediated responses (3, 40), we speculated that the nonprotein constituents of PPD might provide an important component of the proinflammatory signal required to promote the antigen-specific component of DTH responses. However, PPD-B depleted only of its protein component by treatment with proteases failed to promote bovine DTH responses to the protein cocktail.

Bacterial lipoproteins and lipopeptides from a diverse range of bacteria, including *E. coli*, *Borrelia burgdorferi*, *Treponema pallidum*, *Mycoplasma fermentans*, and *M. tuberculosis*, have been shown to induce proinflammatory cytokine production from monocytes and macrophages through triggering of the innate pattern recognition signal Toll-like receptor 2 (5, 29). Similarly, a synthetic lipopeptide containing an acyl moiety that models that of bacterial lipoproteins (47) has also been shown to stimulate the production of proinflammatory cytokines, including TNF-α (4, 21, 33). Therefore, we investigated this synthetic bacterial lipopeptide, Pam3CSK4, in combination with ESAT-6 and demonstrated that it was capable of promoting antigen-specific DTH responses in more than one-half of the calves tested. The immunomodulatory influence of this lipopeptide was not observed in vitro since Pam3CSK4 did not enhance ESAT-6-stimulated T-cell proliferation when the two reagents were cocultured in the presence of PBMC (data not shown), although this result may reflect the in vitro culture conditions used rather than indicate a complete inability of the lipopeptide to enhance T-cell responses in vitro. Since we were then able to demonstrate that Pam3CSK4 stimulated the production of TNF-α from monocyte-derived DC, we propose that the molecular mechanism for the DTH-enhancing properties of this compound involves an ability to induce TNF-α and other proinflammatory cytokines in vivo, thereby promoting enhanced migration of DC and infiltration of monocytes, macrophages, and T cells. However, to be able to consider formulations based on defined antigens as skin test reagents in cattle for general field applications, further optimization is required. In addition, it is necessary to ensure that the diagnostic sensitivity of such a skin test is at least as good as that of an equivalent blood-based assay by, for example, inclusion of additional antigens, such as CFP-10, which have been shown to be useful in diagnosing bovine TB (10, 41, 44).

In summary, our data indicate that the development of a skin test for bovine TB in which defined antigens are used presents many more challenges than the development of blood-based tests. However, we demonstrated that the limitations of defined mycobacterial antigens to induce DTH responses in cattle may be overcome by using them in combination with a suitable proinflammatory stimulus, although further test optimization, such as optimization of antigen concentrations or the time that the reactions are read, is necessary if practical application of defined antigens as skin test reagents for bovine TB is to be realized.

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Additional references for the topic of lipopeptide modulation of delayed-type hypersensitivity (DTH) and tuberculosis are listed below. These references cover various aspects of the disease, including the role of Mycobacterium tuberculosis, the use of lipopeptides as adjuvants, and the immune responses in vaccinated animals and humans.


