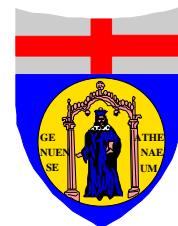


*INCON / 10° ICMAA, Guarujà, Brazil, September 26-29, 2010*

# **MECHANISTIC APPROACHES TO THE PREVENTION OF MUTATION AND CANCER**

**Silvio De Flora**

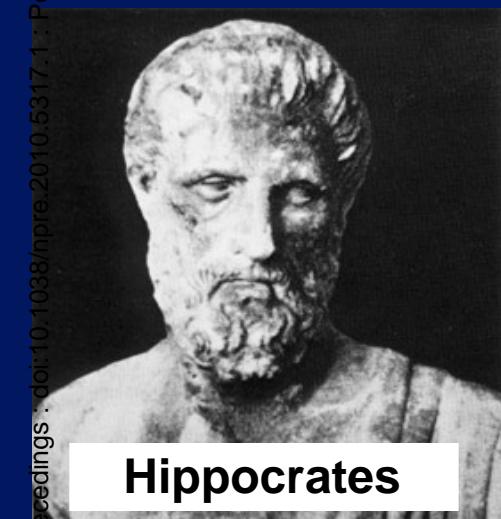


**University of Genoa, Italy**

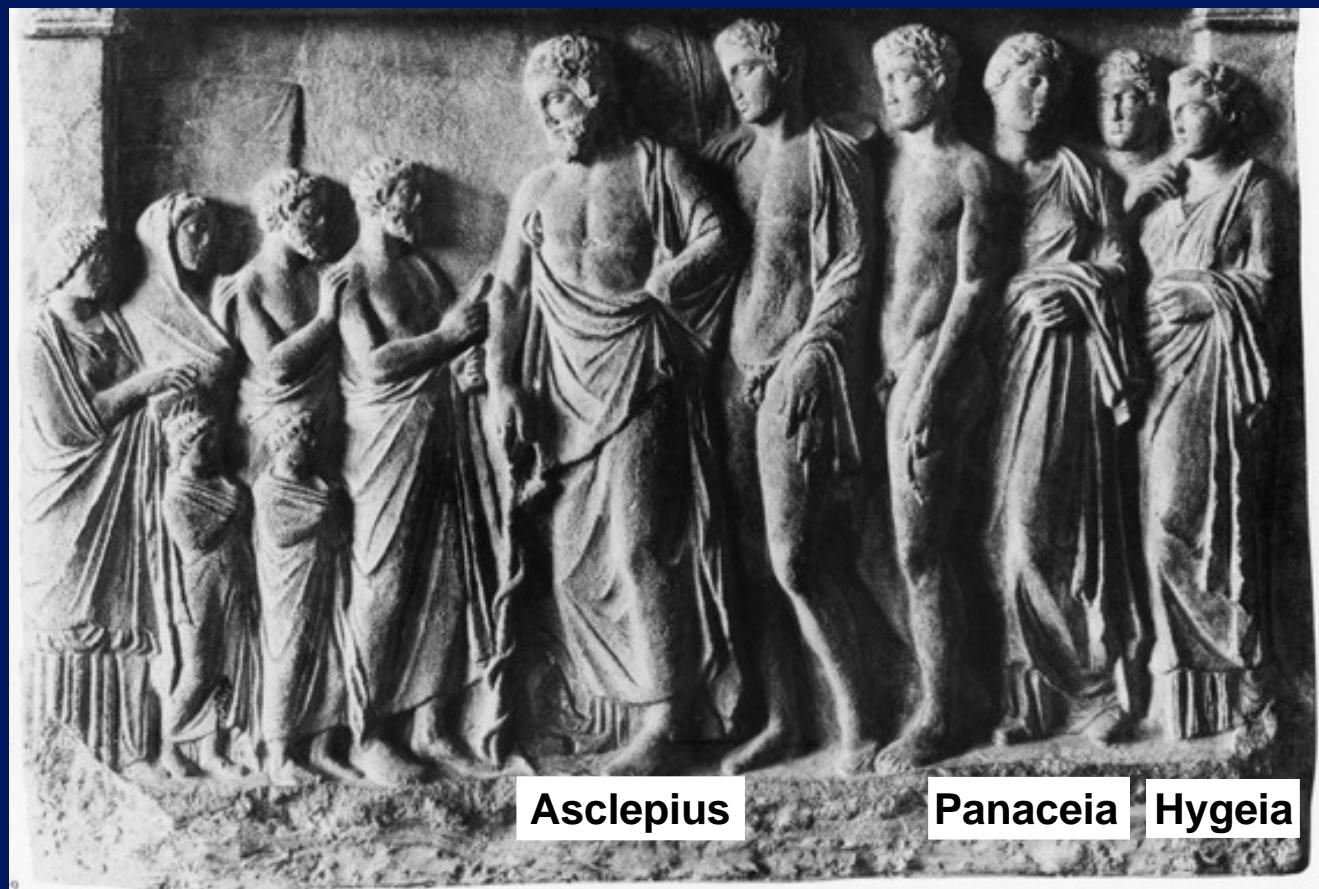
**Department of Health Sciences  
Section of Hygiene and Preventive Medicine**

# THE HIPPOCRATIC OATH

I swear by Apollo the Physician and Asclepius and Hygeia and Panaceia and all the gods and goddesses, making them my witnesses, that I will fulfill according to my ability and judgment this oath.....



Hippocrates



Asclepius

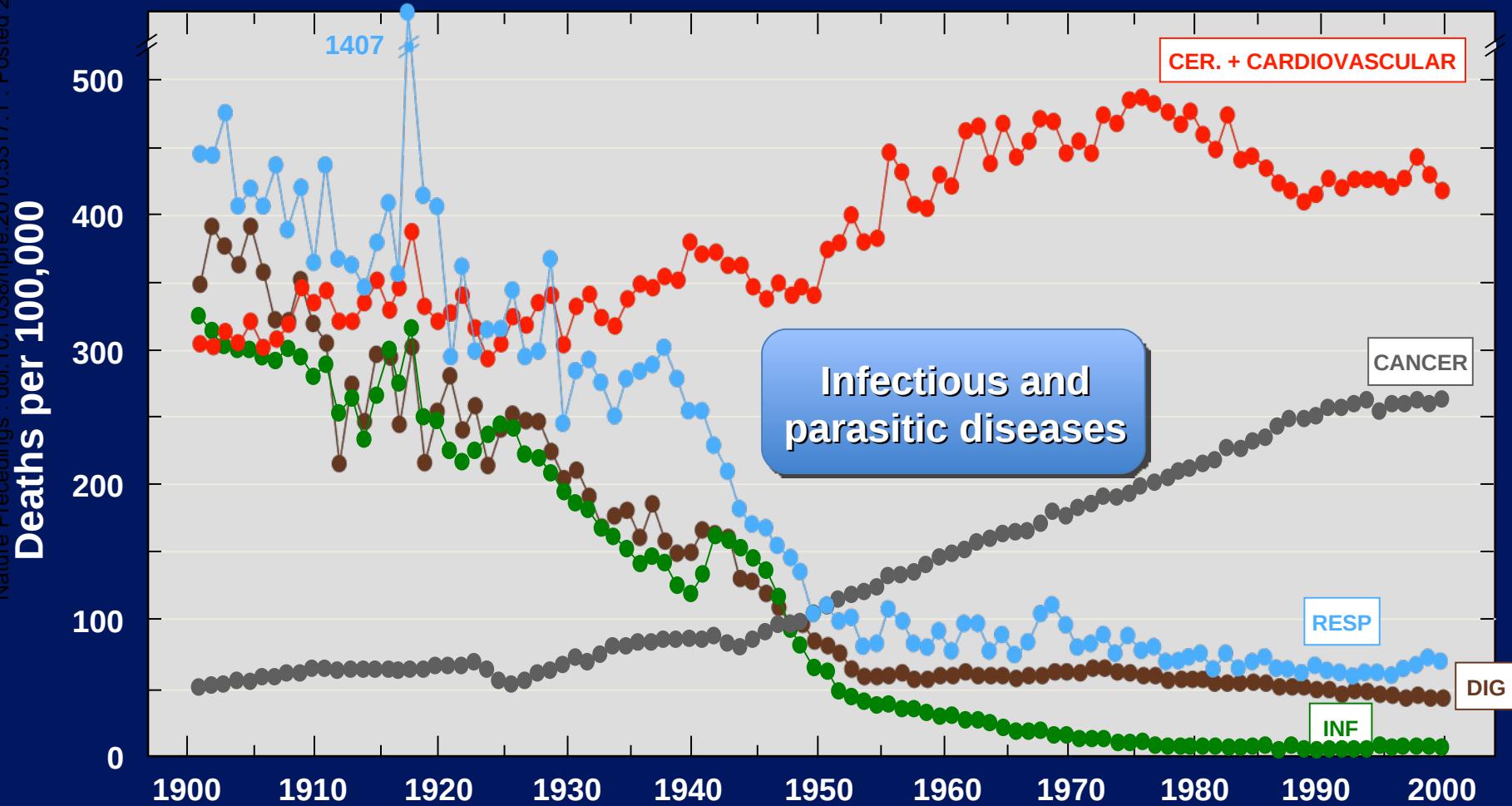
Panaceia Hygeia

# THE EPIDEMIOLOGICAL REVOLUTION OF THE 20th CENTURY

S. De Flora, A. Quaglia, C. Bennicelli & M. Vercelli, FASEB J. 19, 892–897, 2005

Nature Precedings : doi:10.1038/npre.2010.5317.1 : Posted 23 Nov 2010

## ITALY, 1901–2000 (RAW MORTALITY DATA)

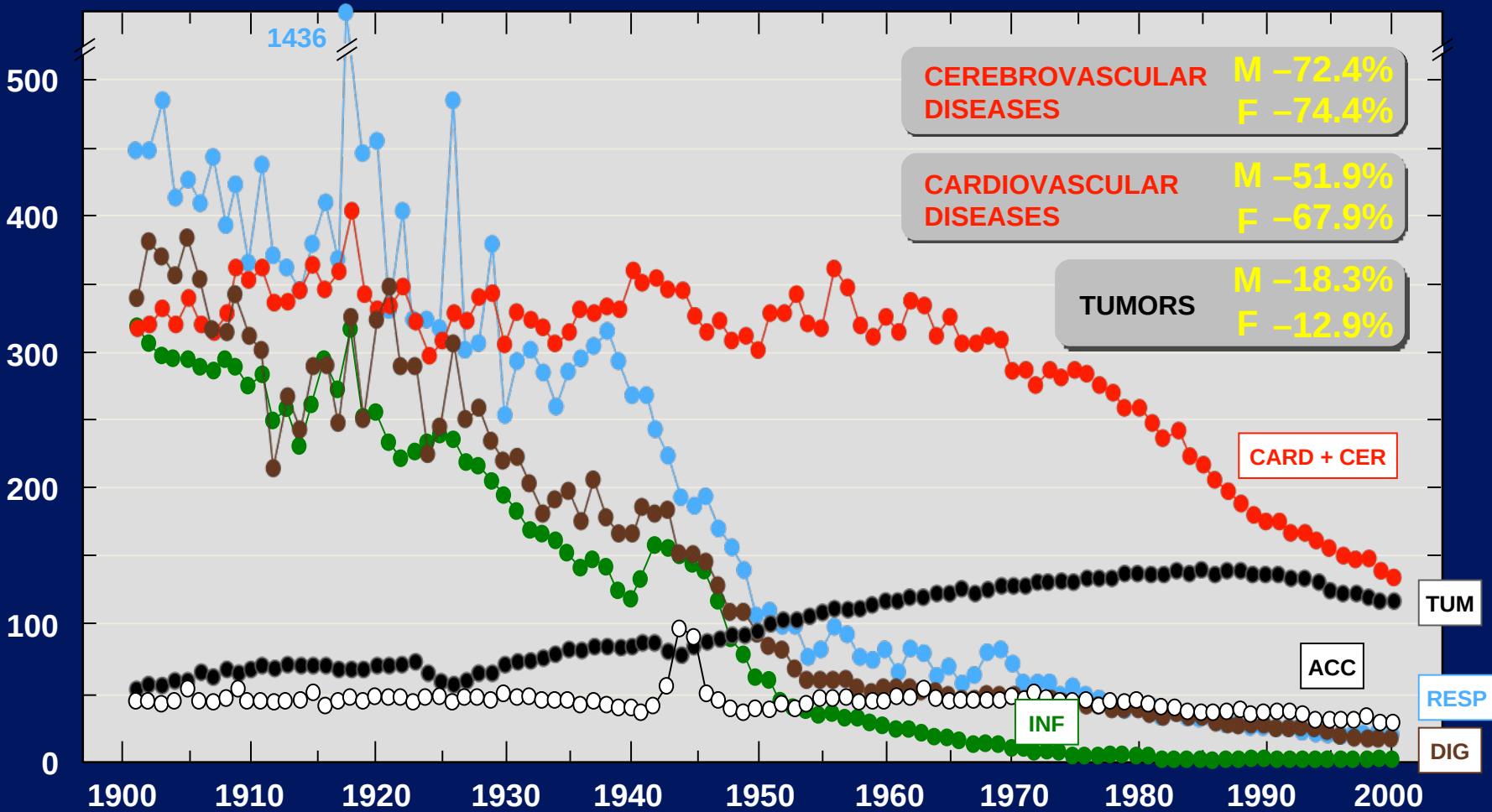


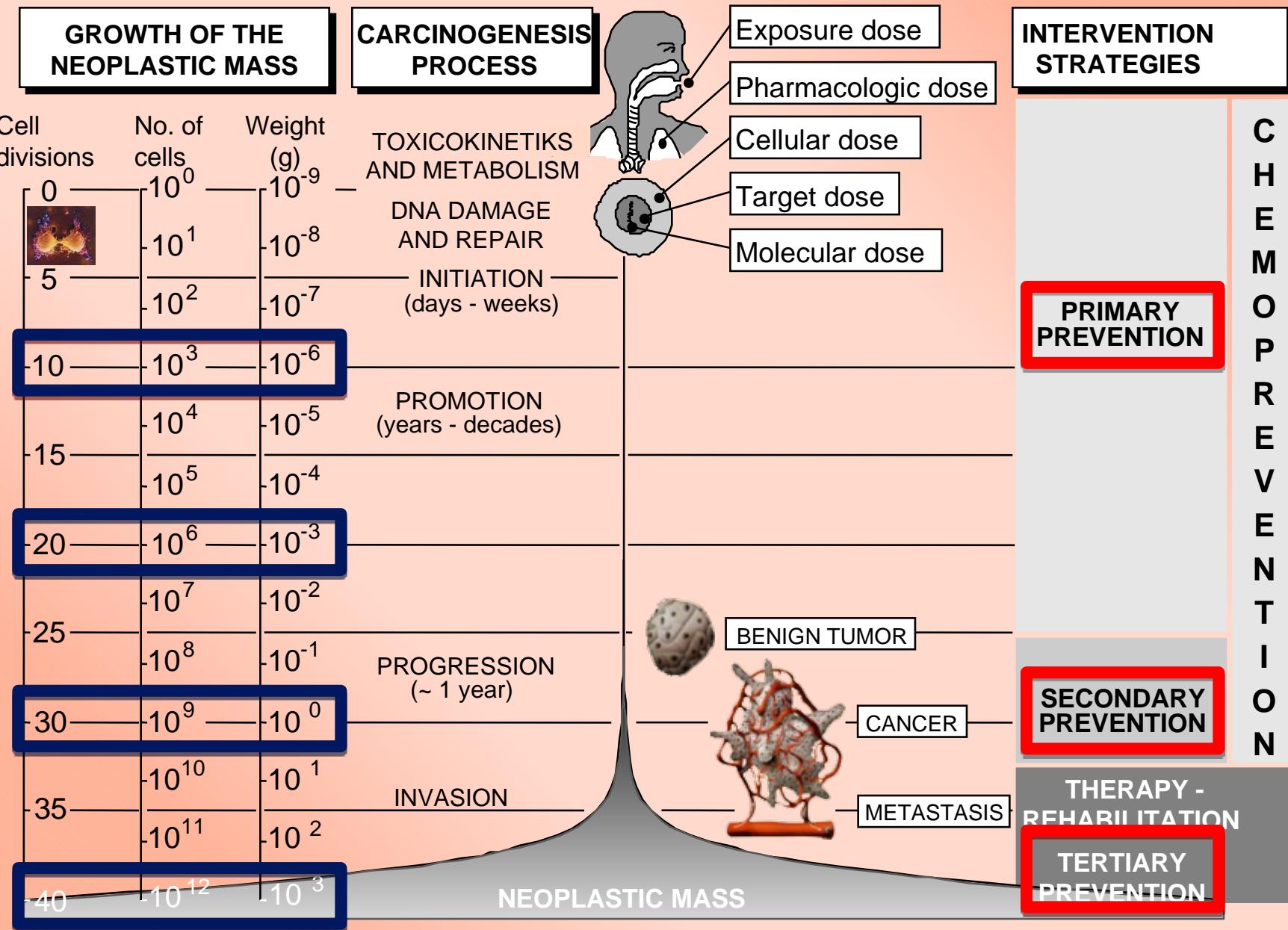
# THE EPIDEMIOLOGICAL REVOLUTION OF THE 20th CENTURY

S. De Flora, A. Quaglia, C. Bennicelli & M. Vercelli, FASEB J. 19, 892–897, 2005

## ITALY, AGE-STANDARDIZED MORTALITY DATA

Nature Precedings : doi:10.1038/npre.2010.5317.1 : Posted 23 Nov 2010





# MECHANISMS OF CANCER CHEMOPREVENTIVE AGENTS

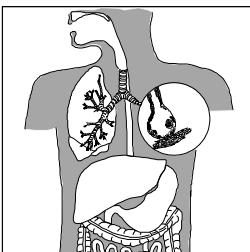
S. De Flora and C. Ramel, Mutat. Res., 202, 285–306, 1988

## PRIMARY PREVENTION

2010

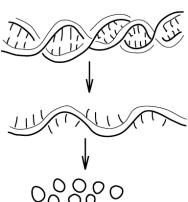
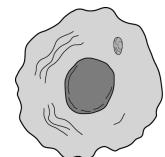
### Inhibition of mutation and cancer initiation in the extracellular environment or in nontarget cells

- 1.1. Inhibition of uptake of mutagens/carcinogens
  - 1.1.1. Inhibition of penetration
  - 1.1.2. Removal from the organism
- 1.2. Inhibition of the endogenous formation of mutagens and carcinogens
  - 1.2.1. Inhibition of the nitrosation reaction
  - 1.2.2. Modification of the intestinal microbial flora
- 1.3. Complexation, dilution and/or deactivation of mutagens/carcinogens outside cells
  - 1.3.1. By physical or mechanical means
  - 1.3.2. By chemical reaction
  - 1.3.3. By enzyme-catalyzed reaction
- 1.4. Favoring absorption of protective agents
- 1.5. Stimulation of trapping and detoxification in nontarget cells



### Inhibition of mutation and cancer initiation in target cells

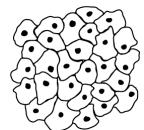
- 2.1. Modification of transmembrane transport
  - 2.1.1. Inhibition of cellular uptake
  - 2.1.2. Stimulation of extrusion outside cells
- 2.2. Modulation of metabolism
  - 2.2.1. Inhibition of activation of pro-mutagens/procarcinogens by Phase I enzymes
  - 2.2.2. Induction of Phase I detoxification and Phase II conjugation pathways, or acceleration of decomposition of reactive metabolites
  - 2.2.3. Stimulation of activation, coordinated with detoxification and blocking of reactive metabolites
- 2.3. Blocking or competition
  - 2.3.1. Trapping of electrophiles by either chemical reaction or enzyme-catalyzed conjugation
  - 2.3.2. Antioxidant activity and scavenging of reactive species
  - 2.3.3. Protection of DNA nucleophilic sites
- 2.4. Inhibition of cell replication
- 2.5. Maintenance of DNA structure and modulation of DNA metabolism and repair
  - 2.5.1. Increase of fidelity of DNA replication and repair
  - 2.5.2. Stimulation of repair and/or reversion of DNA damage
  - 2.5.3. Inhibition of error-prone repair pathways
  - 2.5.4. Correction of hypomethylation
  - 2.5.5. Inhibition of histone deacetylation
  - 2.5.6. Blocking of telomerases or inhibition of their activity
- 2.6. Control of gene expression
  - 2.6.1. Targeted inactivation of oncogenes
  - 2.6.2. Inhibition of oncogene expression
  - 2.6.3. Inhibition of oncogene sequences or activity
    - 2.6.3.1. Inhibition of translation targeted to oncogene mRNA
    - 2.6.3.2. Inhibition of transcription of specific DNA sequences
    - 2.6.3.3. Blocking of target genes
    - 2.6.3.4. Farnesyltransferase inhibition
  - 2.6.4. Neutralization or post-translational modification of oncogene products
  - 2.6.5. Replacement of deleted tumor suppressor genes
  - 2.6.6. Mimicking the DNA binding of tumor suppressor genes by antiidiotypic antibodies
  - 2.6.7. Killing of cells lacking tumor suppressor genes



S. De Flora and L.R. Ferguson, Mutat. Res., 591, 8–15, 2005

### 3. Inhibition of tumor promotion

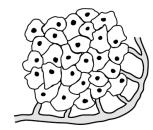
- 3.1. Inhibition of genotoxic effects (see 1 and 2)
- 3.2. Antioxidant activity and scavenging of free radicals
- 3.3. Anti-inflammatory activity
  - 3.3.1. Cyclooxygenase inhibition
  - 3.3.2. Lipoxygenase inhibition
  - 3.3.3. Inhibition of inducible nitric oxide synthase
  - 3.3.4. Leukotriene receptor antagonism
- 3.4. Inhibition of proteases
- 3.5. Inhibition of cell proliferation
  - 3.5.1. Inhibition of ornithine decarboxylase
  - 3.5.2. Promoting proteasomal degradation of cyclins
  - 3.5.3. Interference with multiple signaling pathways
- 3.6. Induction of cell differentiation
- 3.7. Modulation of cell apoptosis
- 3.8. Signal transduction modulation
- 3.9. Protection of intercellular communications



## SECONDARY PREVENTION

### 4. Inhibition of tumor progression

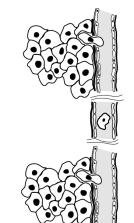
- 4.1. Inhibition of genotoxic effects (see 1 and 2)
- 4.2. Antioxidant activity and scavenging of free radicals
- 4.3. Inhibition of proteases
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- 4.5. Effects on the hormonal status
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  - 4.5.4. Decrease in ovarian hormones by dietary isoflavones
  - 4.5.5. Inhibiting the pituitary secretion of luteinizing hormone
  - 4.5.6. Preventing conversion of testosterone into dehydrotestosterone by 5α-reductase
  - 4.5.7. Selective androgen receptor antagonism
- 4.6. Effects on the immune system
- 4.7. Inhibition of angiogenesis
- 4.8. Antineoplastic activity by either mechanical, physical, chemical, or biological means



## TERTIARY PREVENTION

### 5. Inhibition of invasion and metastasis

- 5.1. Antioxidant activity and scavenging of free radicals
- 5.2. Signal transduction modulation
- 5.3. Inhibition of cell proliferation (see 3.4)
- 5.4. Modulation of cell apoptosis
- 5.5. Induction of cell differentiation
- 5.6. Inhibition of angiogenesis
- 5.7. Effect on cell-adhesion molecules
- 5.8. Inhibition of proteases involved in basement membrane degradation and modulation of the interaction with the extracellular matrix
- 5.9. Activation of antimetastasis genes



# MECHANISMS OF CANCER CHEMOPREVENTIVE AGENTS

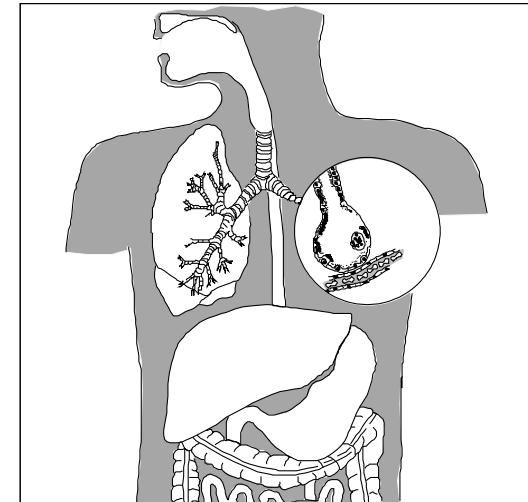
S. De Flora and C. Ramel, Mutat. Res., 202, 285–306, 1988

S. De Flora and L.R. Ferguson, Mutat. Res., 591, 8–15, 2005

## PRIMARY PREVENTION

### ***1. Inhibition of mutation and cancer initiation in the extracellular environment or in nontarget cells***

- 1.1. Inhibition of uptake of mutagens/carcinogens
  - 1.1.1. Inhibition of penetration
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  - 1.2.2. Modification of the intestinal microbial flora
- 1.3. Complexation, dilution and/or deactivation of mutagens/carcinogens outside cells
  - 1.3.1. By physical or mechanical means
  - 1.3.2. By chemical reaction
  - 1.3.3. By enzyme-catalyzed reaction
- 1.4. Favoring absorption of protective agents
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# MECHANISMS OF CANCER CHEMOPREVENTIVE AGENTS

S. De Flora and C. Ramel, Mutat. Res., 202, 285–306, 1988

S. De Flora and L.R. Ferguson, Mutat. Res., 591, 8–15, 2005

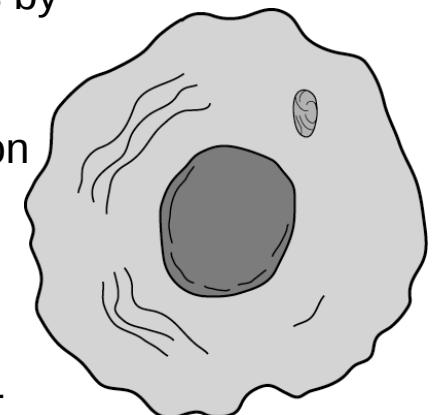
## PRIMARY PREVENTION (cont.)

### 2. *Inhibition of mutation and cancer initiation in target cells*

- 2.1. Modification of transmembrane transport
  - 2.1.1. Inhibition of cellular uptake
  - 2.1.2. Stimulation of extrusion outside cells

- 2.2. Modulation of metabolism
  - 2.2.1. Inhibition of activation of promutagens/ procarcinogens by Phase I enzymes
  - 2.2.2. Induction of Phase I detoxification and Phase II conjugation pathways, or acceleration of decomposition of reactive metabolites
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- 2.3. Blocking or competition
  - 2.3.1. Trapping of electrophiles by either chemical reaction or enzyme-catalyzed conjugation
  - 2.3.2. Antioxidant activity and scavenging of reactive species
  - 2.3.3. Protection of DNA nucleophilic sites
- 2.4. Inhibition of cell replication



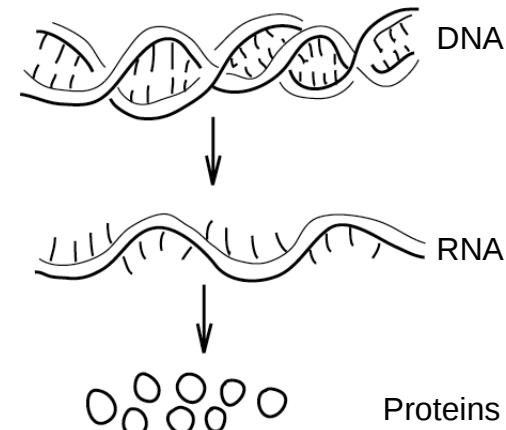
# MECHANISMS OF CANCER CHEMOPREVENTIVE AGENTS

S. De Flora and C. Ramel, Mutat. Res., 202, 285–306, 1988

S. De Flora and L.R. Ferguson, Mutat. Res., 591, 8–15, 2005

## PRIMARY PREVENTION (cont.)

- 2.5. Maintenance of DNA structure and modulation of DNA metabolism and repair
  - 2.5.1. Increase of fidelity of DNA replication and repair
  - 2.5.2. Stimulation of repair and/or reversion of DNA damage
  - 2.5.3. Inhibition of error-prone repair pathways
  - 2.5.4. Correction of hypomethylation
  - 2.5.5. Inhibition of histone deacetylation
  - 2.5.6. Blocking of telomerases or inhibition of their activity
- 2.6. Control of gene expression
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    - 2.6.3.1. Inhibition of translation targeted to oncogene mRNA
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  - 2.6.5. Replacement of deleted tumor suppressor genes
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  - 2.6.7. Killing of cells lacking tumor suppressor genes



# MECHANISMS OF CANCER CHEMOPREVENTIVE AGENTS

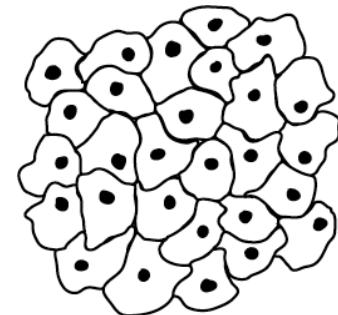
S. De Flora and C. Ramel, Mutat. Res., 202, 285–306, 1988

S. De Flora and L.R. Ferguson, Mutat. Res., 591, 8–15, 2005

## PRIMARY PREVENTION (cont.)

### 3. *Inhibition of tumor promotion*

- 3.1. Inhibition of genotoxic effects (see 1 and 2)
- 3.2. Antioxidant activity and scavenging of free radicals
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  - 3.3.1. Cyclooxygenase inhibition
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# MECHANISMS OF CANCER CHEMOPREVENTIVE AGENTS

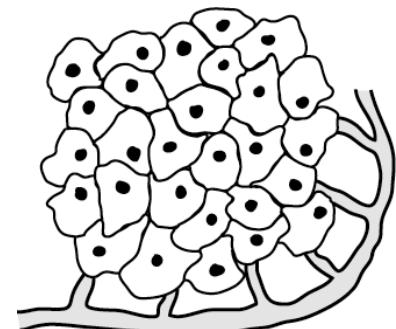
S. De Flora and C. Ramel, Mutat. Res., 202, 285–306, 1988

S. De Flora and L.R. Ferguson, Mutat. Res., 591, 8–15, 2005

## SECONDARY PREVENTION

### 4. *Inhibition of tumor progression*

- 4.1. Inhibition of genotoxic effects (see 1 and 2)
- 4.2. Antioxidant activity and scavenging of free radicals
- 4.3. Inhibition of proteases
- 4.4. Signal transduction modulation
- 4.5. Effects on the hormonal status
  - 4.5.1. Selective estrogen receptor modulation
  - 4.5.2. Aromatase inhibition
  - 4.5.3. Selective blocking of prostaglandin E<sub>2</sub> receptors
  - 4.5.4. Decrease in ovarian hormones by dietary isoflavones
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# MECHANISMS OF CANCER CHEMOPREVENTIVE AGENTS

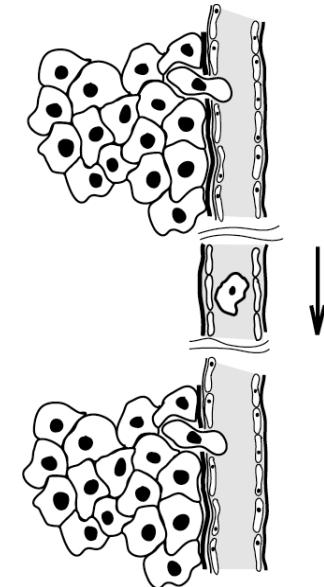
S. De Flora and C. Ramel, Mutat. Res., 202, 285–306, 1988

S. De Flora and L.R. Ferguson, Mutat. Res., 591, 8–15, 2005

## TERTIARY PREVENTION

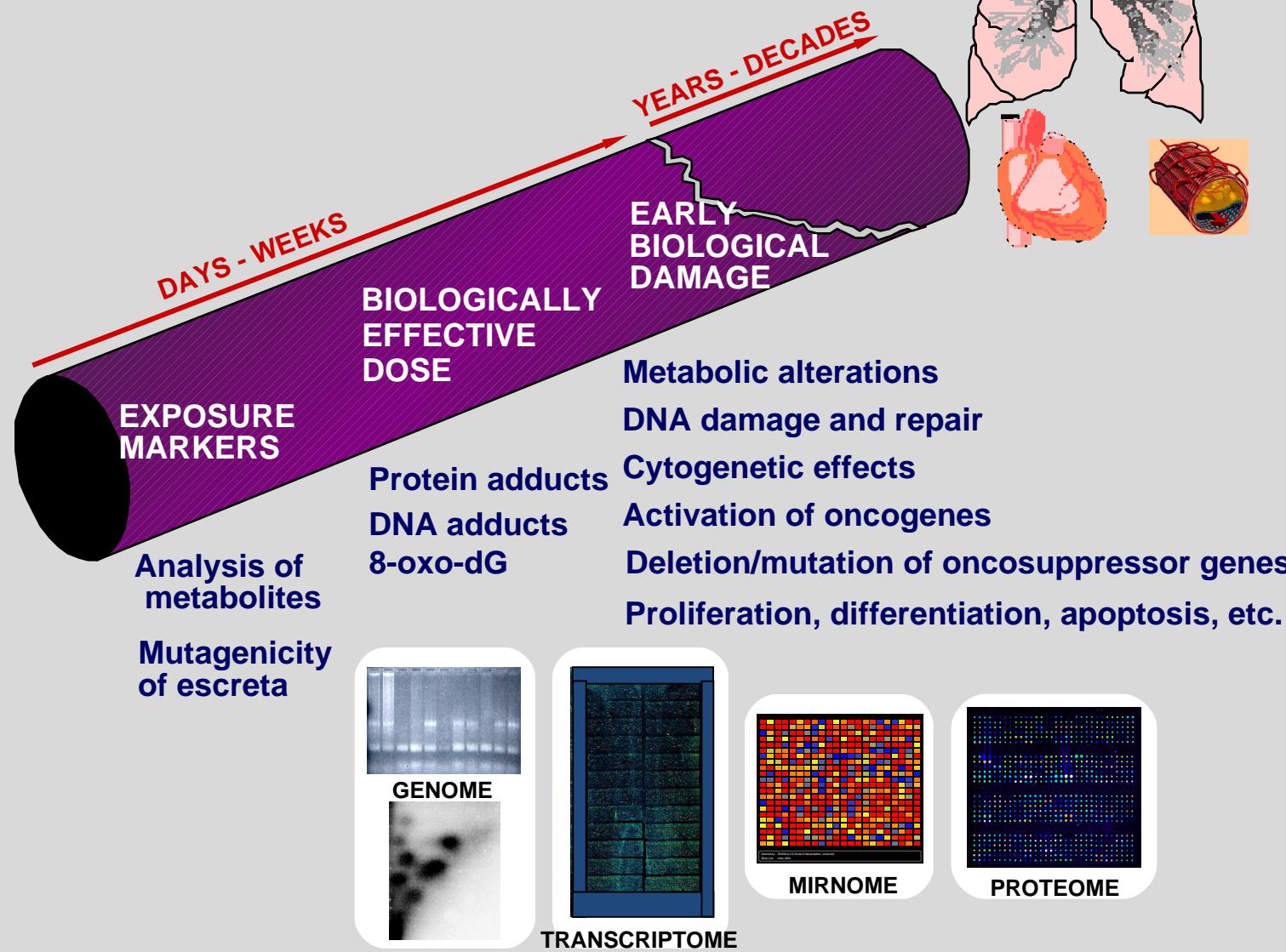
### 5. *Inhibition of invasion and metastasis*

- 5.1. Antioxidant activity and scavenging of free radicals
- 5.2. Signal transduction modulation
- 5.3. Inhibition of cell proliferation (see 3.4)
- 5.4. Modulation of cell apoptosis
- 5.5. Induction of cell differentiation
- 5.6. Inhibition of angiogenesis
- 5.7. Effect on cell-adhesion molecules
- 5.8. Inhibition of proteases involved in basement membrane degradation and modulation of the interaction with the extracellular matrix
- 5.9. Activation of antimetastasis genes



## DISEASES

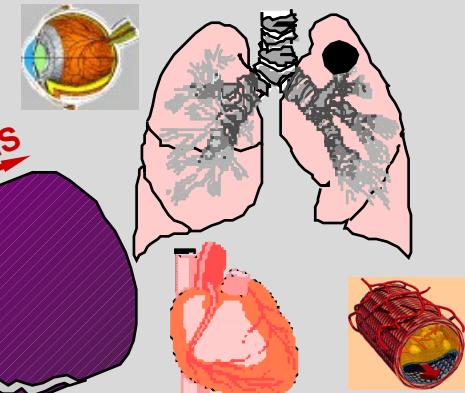
# DECODING THE BLACK BOX



# PROTECTIVE FACTORS



# DISEASES



DAYS - WEEKS

YEARS - DECADES

## BIOLOGICALLY EFFECTIVE DOSE

## EARLY BIOLOGICAL DAMAGE

Metabolic alterations

DNA damage and repair

Cytogenetic effects

Activation of oncogenes

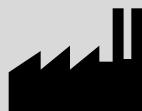
Deletion/mutation of oncosuppressor genes

Proliferation, differentiation, apoptosis, etc.

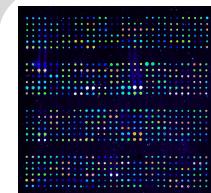
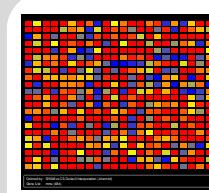
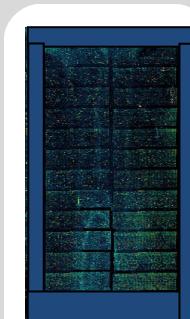


Analysis of metabolites

Mutagenicity of escreta



RISK FACTORS



## Pathological conditions

# Cancer / Physical agents



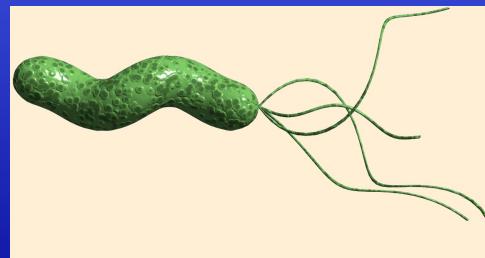
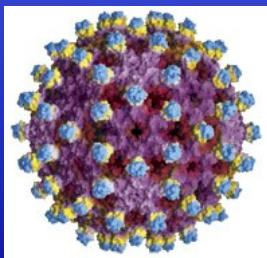
## Pathological conditions

# Cancer / Chemicals and complex mixtures



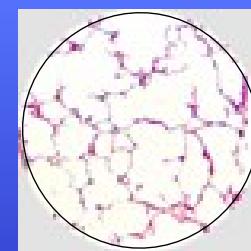
## Pathological conditions

# Cancer / Microbial diseases



## Pathological conditions

### COPD

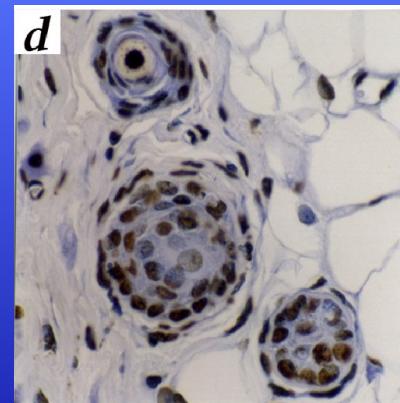


**(A. Izzotti et al., FASEB J. 17, 1127-29, 2003)**

**(R. Balansky et al., Carcinogenesis 30, 1398-401, 2009)**

## Pathological conditions

# Alopecia



(R. Balansky et al., PNAS USA 103, 7823-28, 2006)

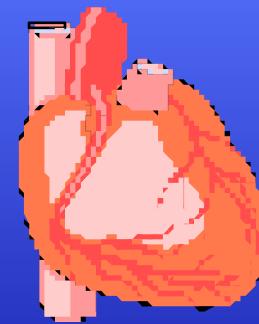
## Pathological conditions

# Atherosclerosis



## Pathological conditions

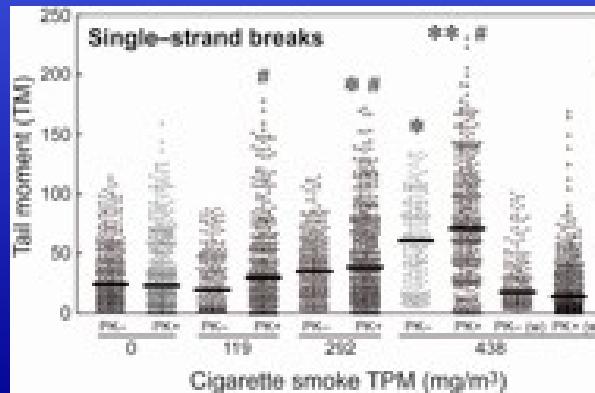
# Heart diseases



# Pathological conditions

## Neurodegenerative diseases

(S. La Maestra et al., 2010)



## Pathological conditions

# Eye diseases



**(A. Izzotti et al., Am. J. Med. 114, 638-646, 2003)**

## Pathological conditions

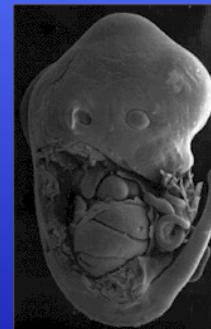
# Rare genetic diseases

**(A. Izzotti et al., Neurology 71, 610-2, 2008)**



## Physiological situations

### Pregnancy



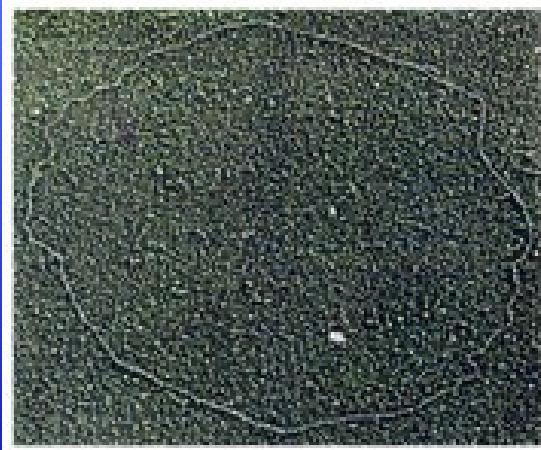
**(A. Izzotti et al., FASEB J. 17, 1127-9, 2003)**

## Physiological situations

### Perinatal period



# Physiological situations



## Aging

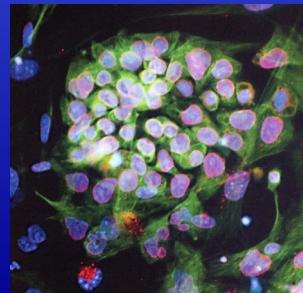


**(R. Balansky et al., Cancer Res. 56, 1642-7, 1996)**

## Physiological situations

# Stem cells

**(S. De Flora et al., Int. J. Oncol. 29, 521-529, 2006)**



# GENOMIC CHANGES IN MOUSE LUNG AT BIRTH

A. Izzotti et al., Mutat. Res. (Rev. Genetic Toxicol.), 544, 441-449, 2003

UNTREATED  
PREGNANT  
MICE

8-oxo-dGuo

1.9  
 $P < 0.05$

0.9  
NS

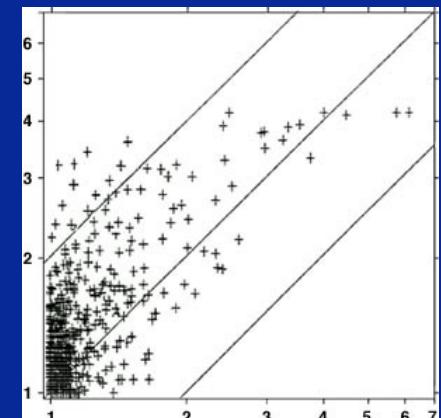
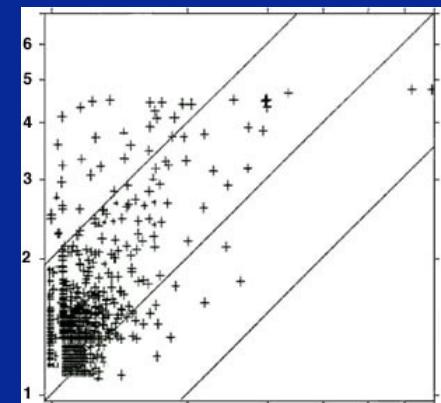
DNA adducts

5.0  
 $P < 0.001$

2.0  
NS

Newborn mice / fetuses

Expression of 746 genes

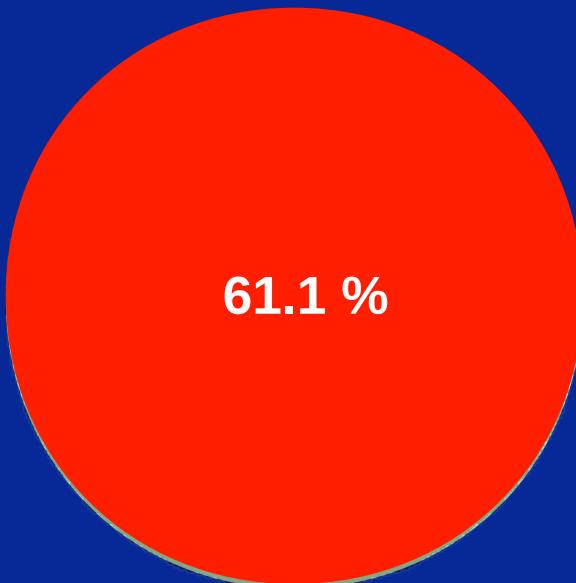


# MICE EXPOSED TO SMOKE AFTER BIRTH

Nature Precedings : doi:10.1101/0.53171; Posted 23 Nov 2010

UNTREATED  
PREGNANT  
MICE

LUNG TUMORS



NAC-TREATED  
PREGNANT  
MICE

17.0 %

LUNG  
EMPHYSEMA

HYPERPLASIA  
OF BLADDER  
EPITHELIUM

16.7 %



6.4 %

20.4 %



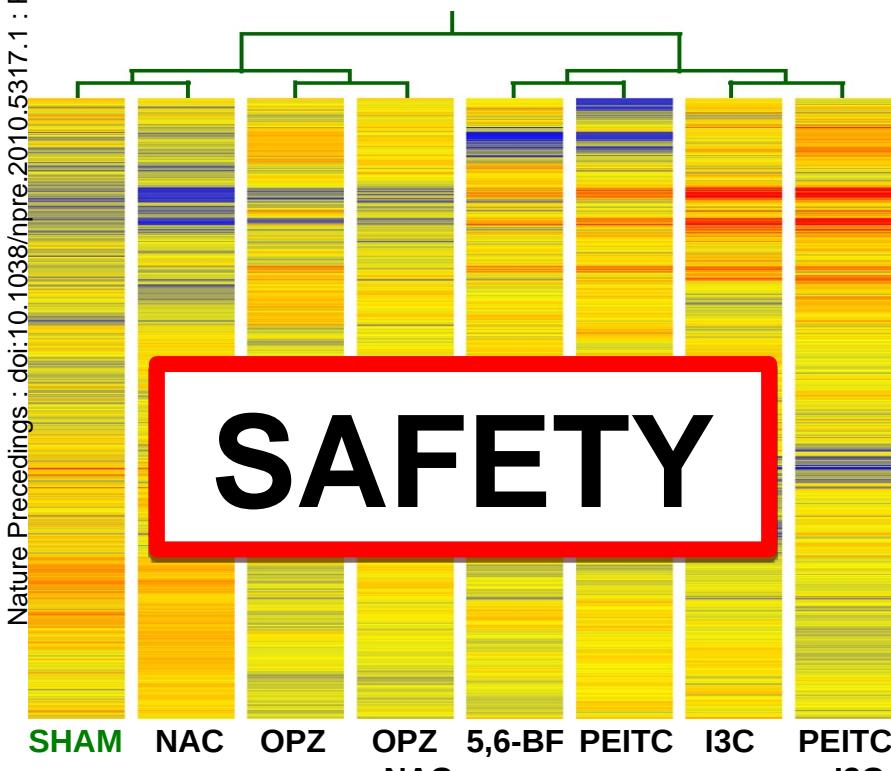
R. Balansky et al., Carcinogenesis 30, 1398-1401, 2009

# EXPRESSION OF 4858 GENES IN MOUSE LUNG

A. Izzotti et al., Mutat. Res. 591, 212–223, 2005

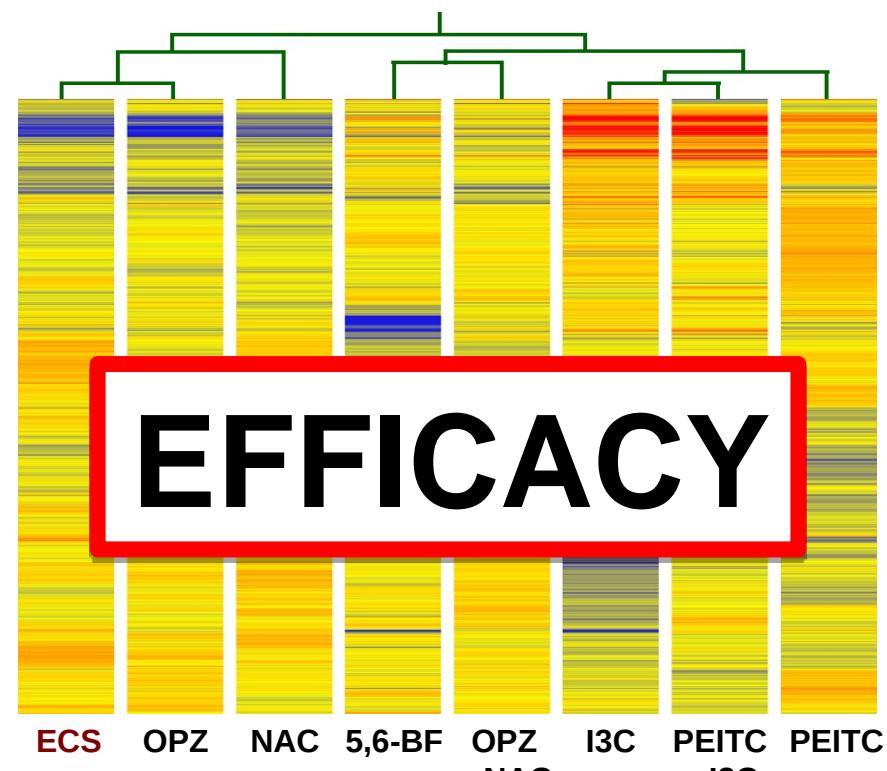
Nature Precedings : doi:10.1038/npre.2010.5317.1 : Posted 23 Nov 2010

SMOKE-FREE MICE



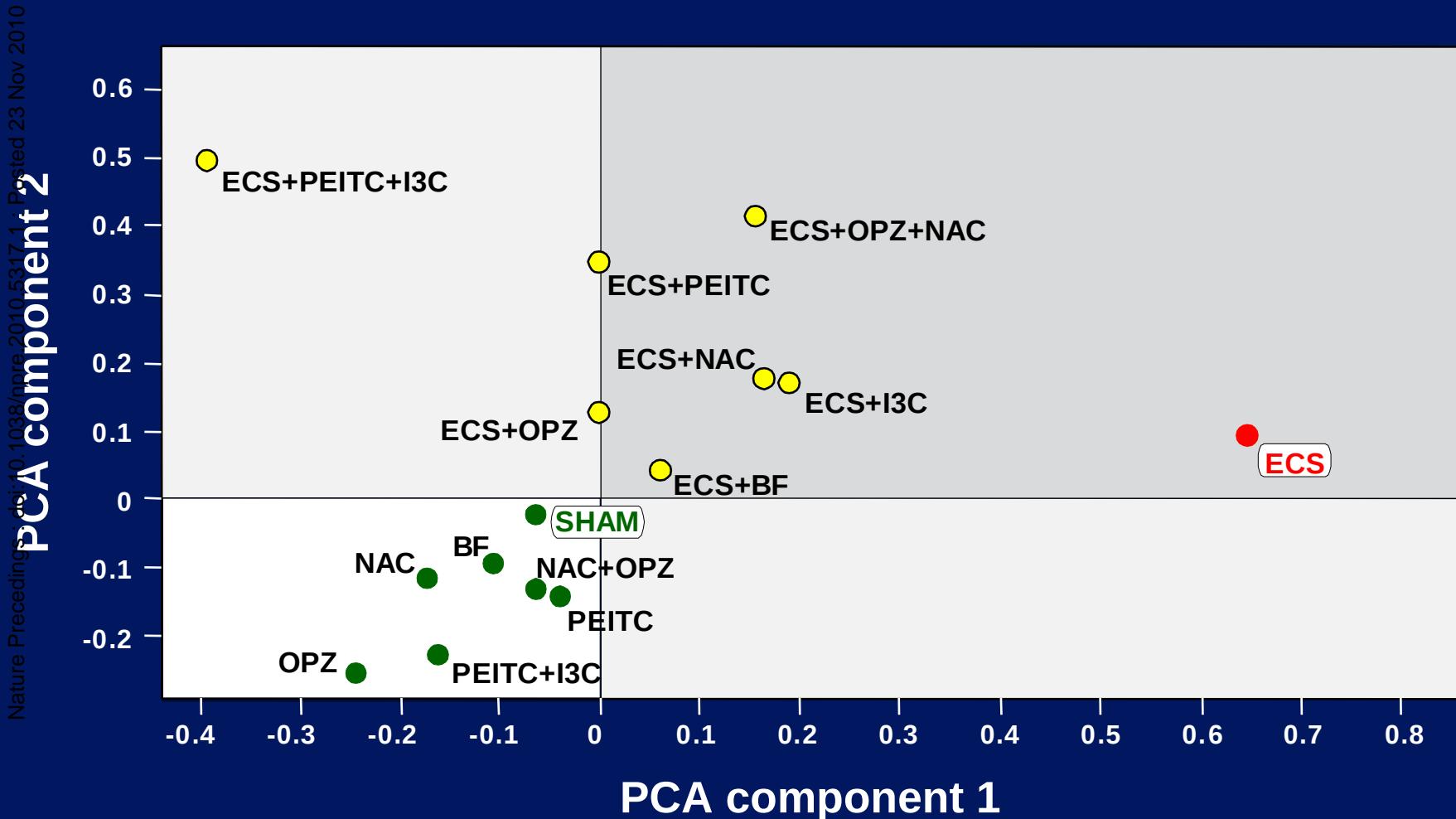
SAFETY

SMOKE-EXPOSED MICE



EFFICACY

# EFFECT OF CIGARETTE SMOKE (ECS) AND CHEMOPREVENTIVE AGENTS ON miRNA EXPRESSION IN RAT LUNG



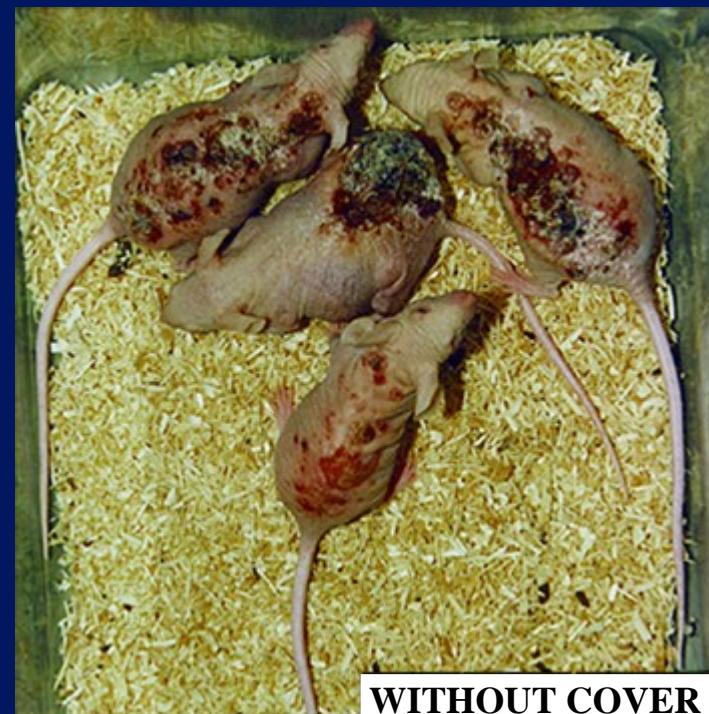
# EXPOSURE OF HAIRLESS MICE TO HALOGEN LAMPS



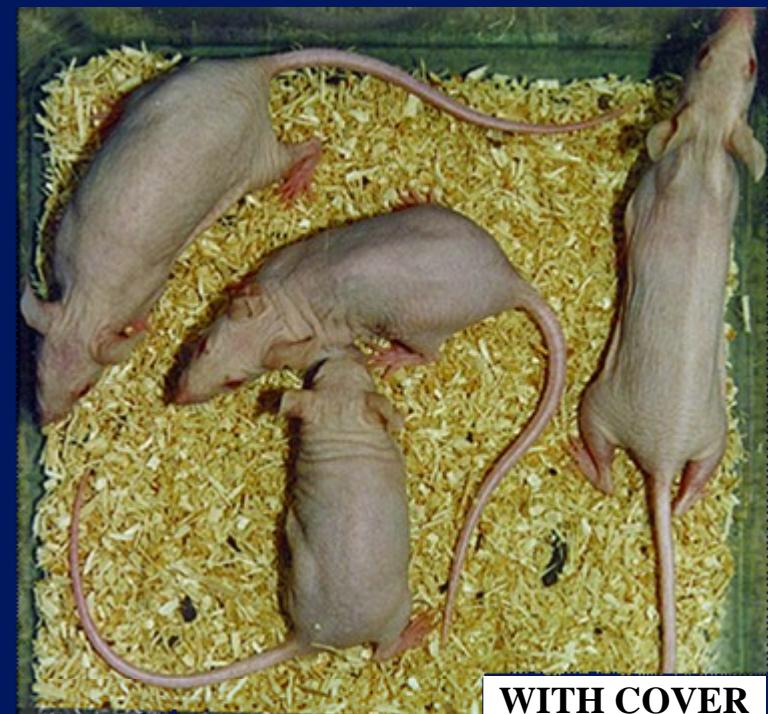
# SKIN CARCINOGENICITY OF HALOGEN LAMPS

S. De Flora & F. D'Agostini, Nature 356, 569, 1992

F. D'Agostini & S. De Flora, Cancer Res. 54, 5081–5, 1994



WITHOUT COVER



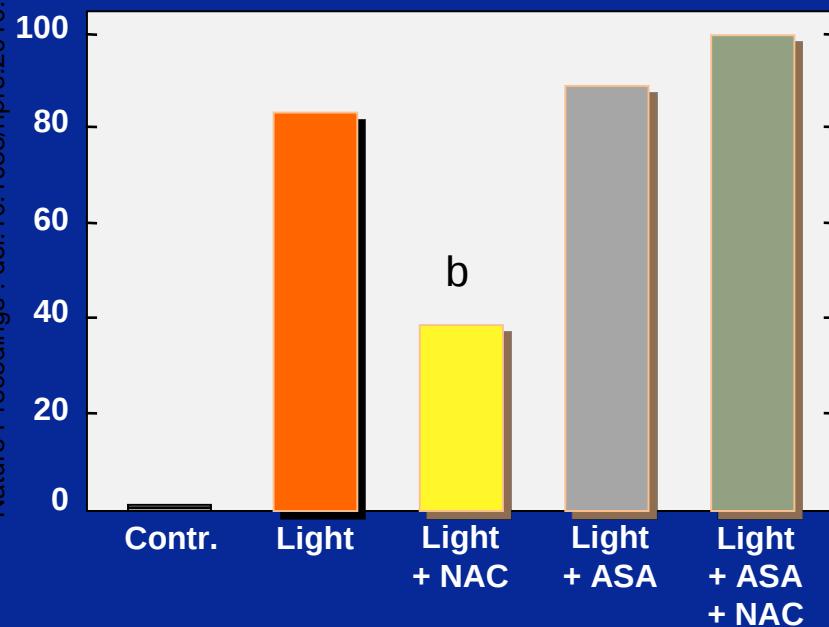
WITH COVER

# MODULATION OF LIGHT-INDUCED SKIN TUMORS BY N-ACETYLCYSTEINE (NAC) AND ASCORBIC ACID (ASA)

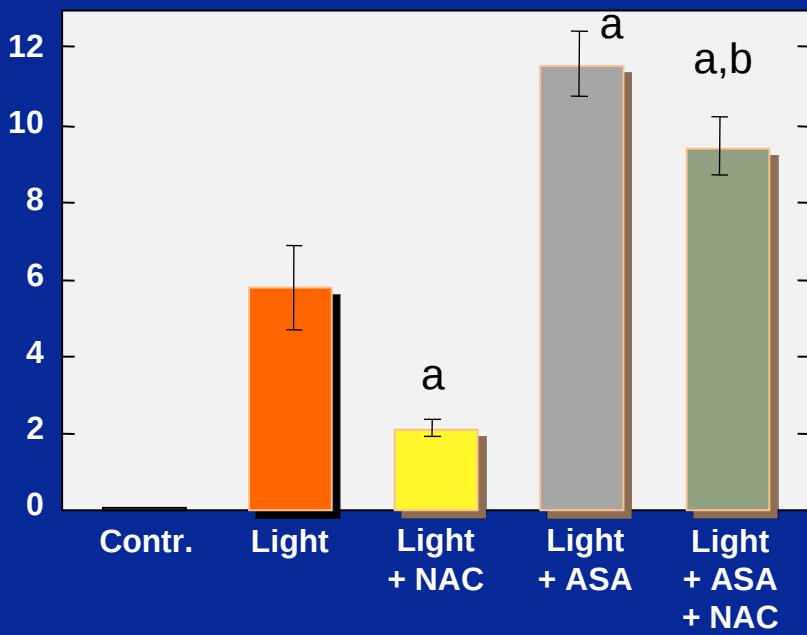
F. D'Agostini et al., Carcinogenesis 26, 657–664, 2005



Incidence (%)



Multiplicity (mean  $\pm$  SE)



<sup>a</sup>P <0.001, as compared with Light; <sup>b</sup>P <0.001, as compared with Light + AsA

# THE PREVENTION OF INFECTION-ASSOCIATED CANCERS

(S. De Flora and P. Bonanni, 2010)

Pathogen	IARC Group	Main associated cancer
<b>Hepatitis viruses</b>		
HBV	1	Hepatocellular carcinoma
HCV	1	Hepatocellular carcinoma
HDV	3	None
<b>Papillomaviruses</b>		
α HPV type 16	1	Cancers at several sites
α HPV types 18, 31, 33, 35, 39, 45, 51, 52, 56, 58, 59	1	Cervical cancer
α HPV type 68	2A	Cervical cancer
α HPV types 26, 30, 34, 53, 66, 67, 69, 70, 73, 82, 85, 97	2B	Cervical cancer
β HPV type 5 and 8	2B	Skin cancer
α HPV type 6 and 11	3	None
Other β and γ HPV types	3	None
<b>Polyomaviruses</b>		
JCV	NA	CNS tumors and colorectal cancer?
MCV	NA	Skin cancer (Merkel cell carcinoma)
SV40	NA	Malignant mesothelioma ?
<b>Herpesviruses</b>		
EBV or HHV4	1	Burkitt's lymphoma, sinonasal angiogenic T-cell lymphoma, immunosuppressor-related non-Hodgkin's lymphoma, Hodgkin's lymphoma, nasopharyngeal carcinoma
KSHV or HHV8	1	Kaposi's sarcoma, primary effusion lymphoma

Pathogen	IARC Group	Main associated cancer
<b>Retroviruses</b>		
HTLV-I	1	Adult T-cell leukemia/lymphoma
HTLV-II	3	None
HIV-I	1	Kaposi's sarcoma, non-Hodgkin's lymphoma, Hodgkin's lymphoma, cervical cancer, anus cancer, conjunctive cancer
HIV-II	2B	Kaposi's sarcoma, non-Hodgkin's lymphoma
HERV-K	NA	Human breast cancer
<b><i>Helicobacter pylori</i></b>	1	Gastric cancer, MALT
<b>Schistosomes</b>		
<i>S. haematobium</i>	1	Urinary bladder cancer
<i>S. japonicum</i>	2B	Colorectal and liver cancers
<i>S. mansoni</i>	3	None
<b>Liver flukes</b>		
<i>Opisthorchis viverrini</i>	1	Cholangiocarcinoma
<i>Opisthorchis felineus</i>	3	None
<i>Clonorchis sinensis</i>	1	Cholangiocarcinoma

Infectious agents cause 17% of all cancers worldwide, 26% in developing world, 8% in developed world

D.M. Parkin, Int.J.Cancer 15, 3030-44, 2005

# THE PREVENTION OF INFECTION-ASSOCIATED CANCERS

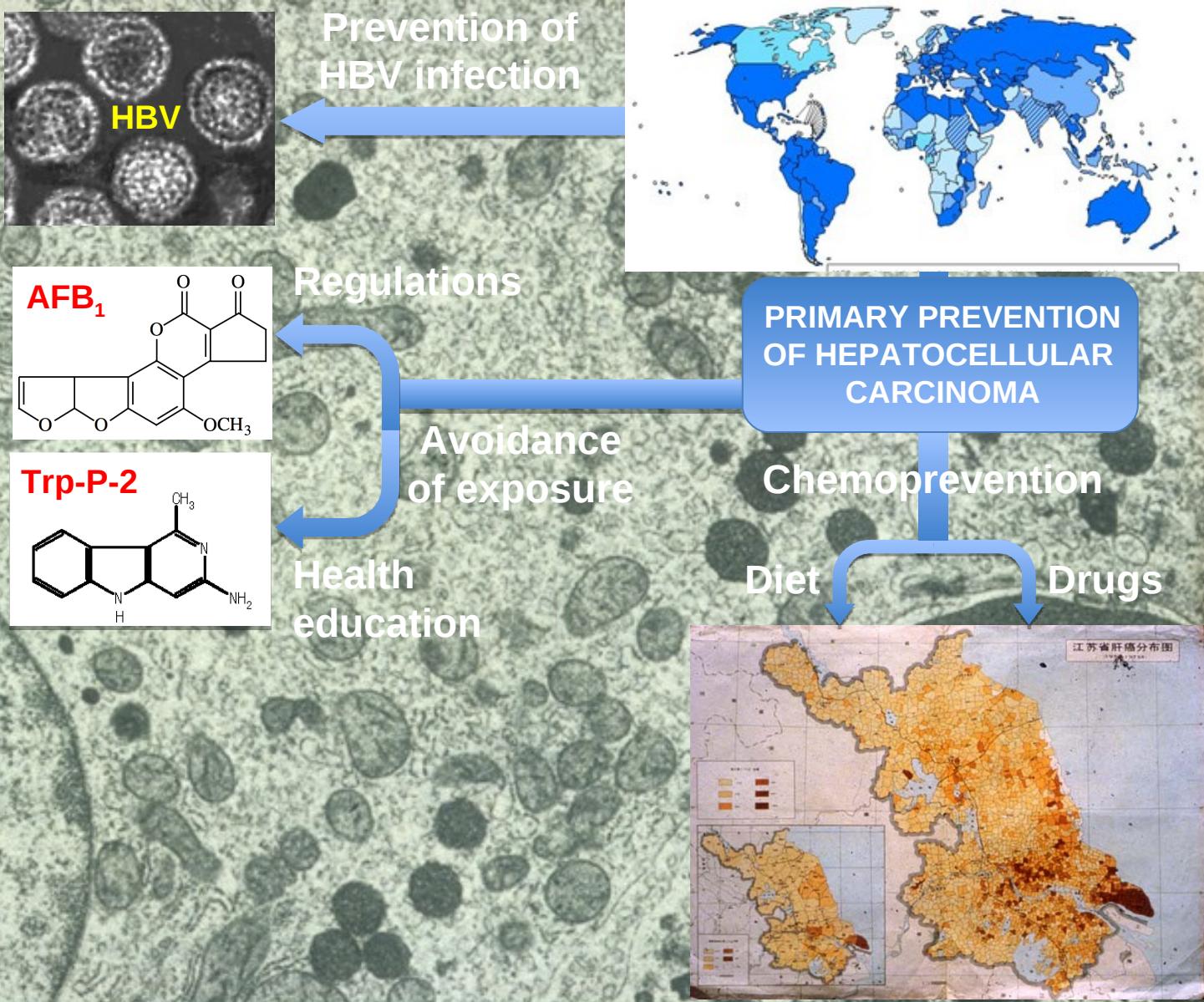
(S. De Flora and P. Bonanni, 2010)

Pathogen	IARC Group	Main associated cancer
<b>Hepatitis viruses</b>		<b>4.9% of all cancers 85.5% of all HCCs</b>
HBV		
HCV		
HDV		
<b>Papillomaviruses</b>		
α HPV type 16	1	Cancers at several sites
α HPV types 18, 31, 33, 35, 39, 45, 51, 52, 56, 58, 59		
α HPV type 68		
α HPV types 26, 30, 34, 53, 66, 67, 69, 70, 73, 82, 85, 97		
β HPV type 5 and 8	2B	Skin cancer
α HPV type 6 and 11	3	None
Other β and γ HPV types	3	None
<b>Polyomaviruses</b>		
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HIV-II	2B	Kaposi's sarcoma, non-Hodgkin's lymphoma
HERV-K		
<b>Helicobacter pylori</b>		<b>5.5% of all cancers 63.4% of stomach cancers</b>
<b>Schistosomes</b>		
<i>S. haematobium</i>	1	Urinary bladder cancer
<i>S. japonicum</i>	2B	Colorectal and liver cancers
<i>S. mansoni</i>	3	None
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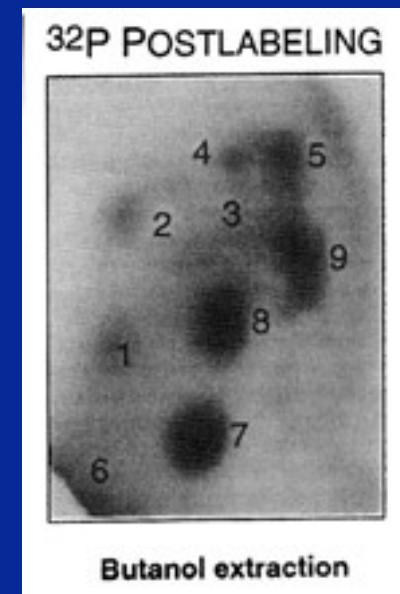
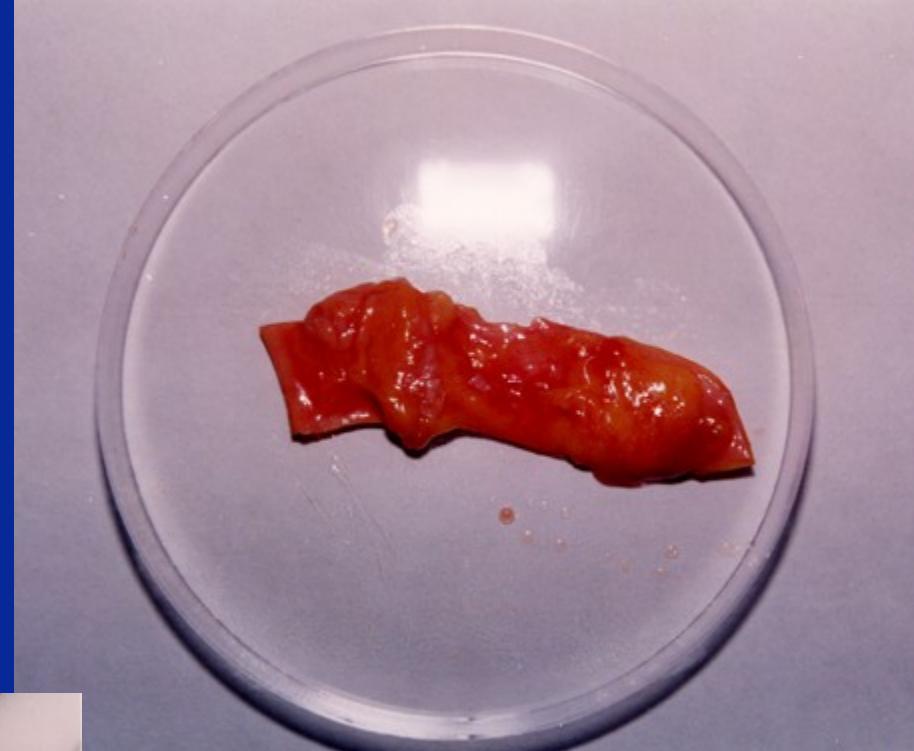
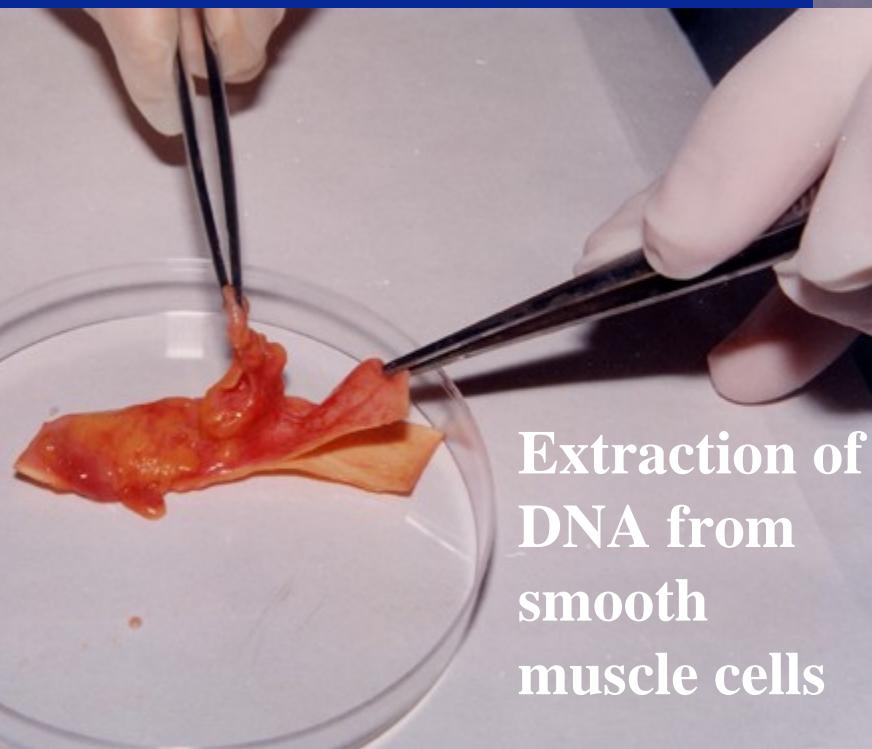
Infectious agents cause 17% of all cancers worldwide, 26% in developing world, 8% in developed world

D.M. Parkin, Int.J.Cancer 15, 3030-44, 2005



- S. De Flora et al, Cancer Res. 47, 4052–8, 1987; Carcinogenesis 10, 1099–1106, 1989  
A.Izzotti et al, Chem.-Biol. Int. 97, 273–285, 1995  
A. Camoirano et al, Cancer Epid. Biol. Biom. 10, 775–783, 2001

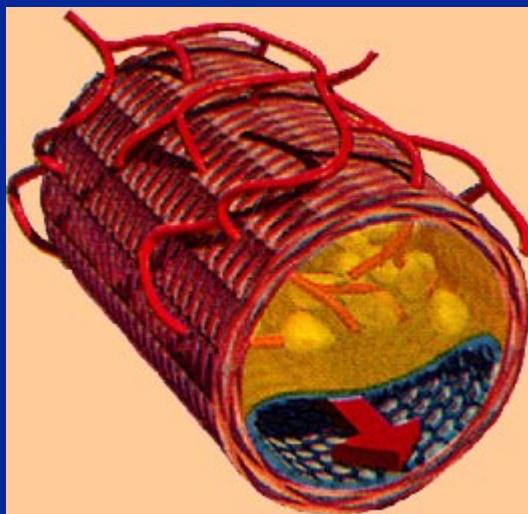
# HUMAN ABDOMINAL AORTA WITH ATHEROSCLEROTIC LESIONS



# MOLECULAR EPIDEMIOLOGY OF ATHEROSCLEROSIS

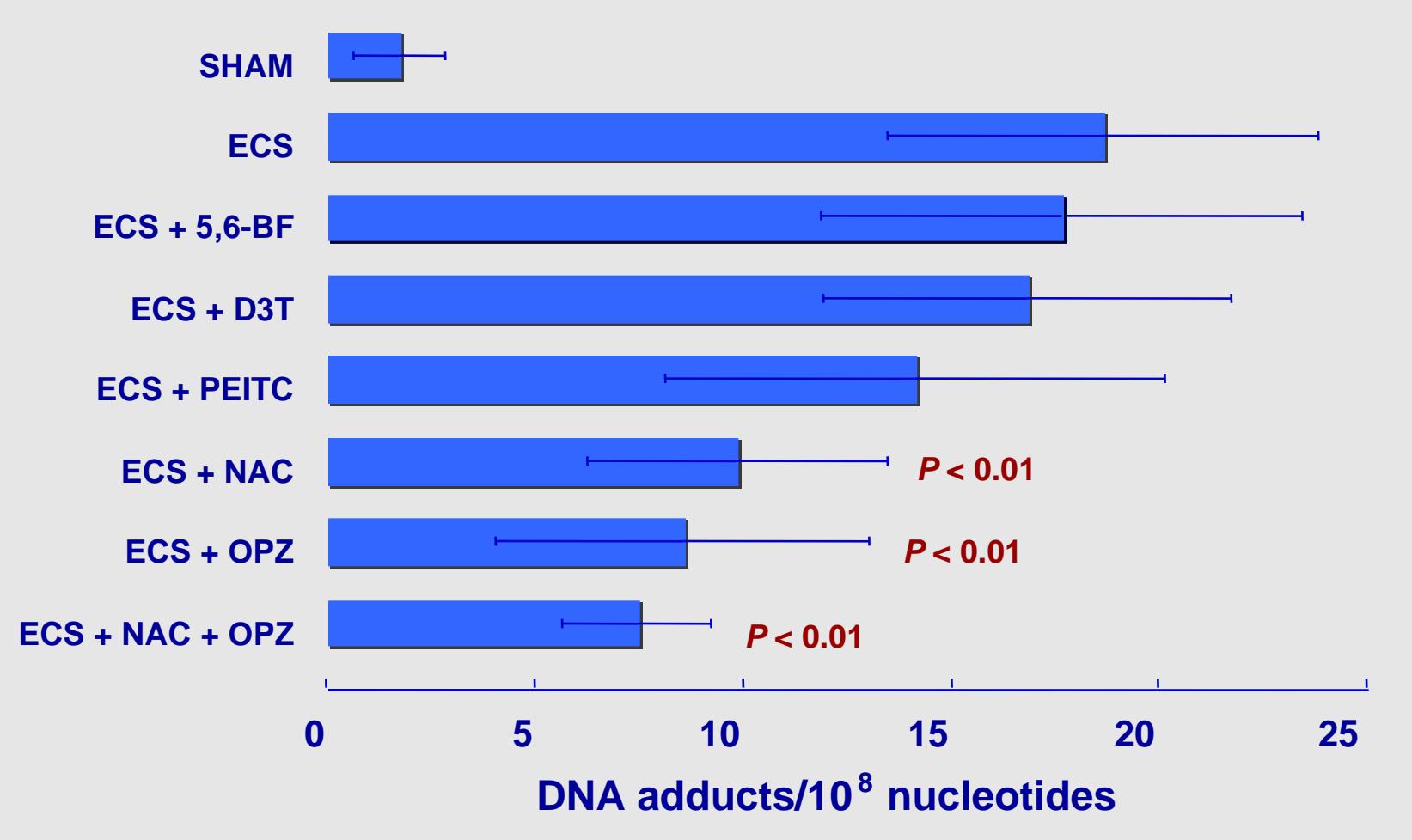
S. De Flora et al., FASEB J. 11: 1021-1031, 1997

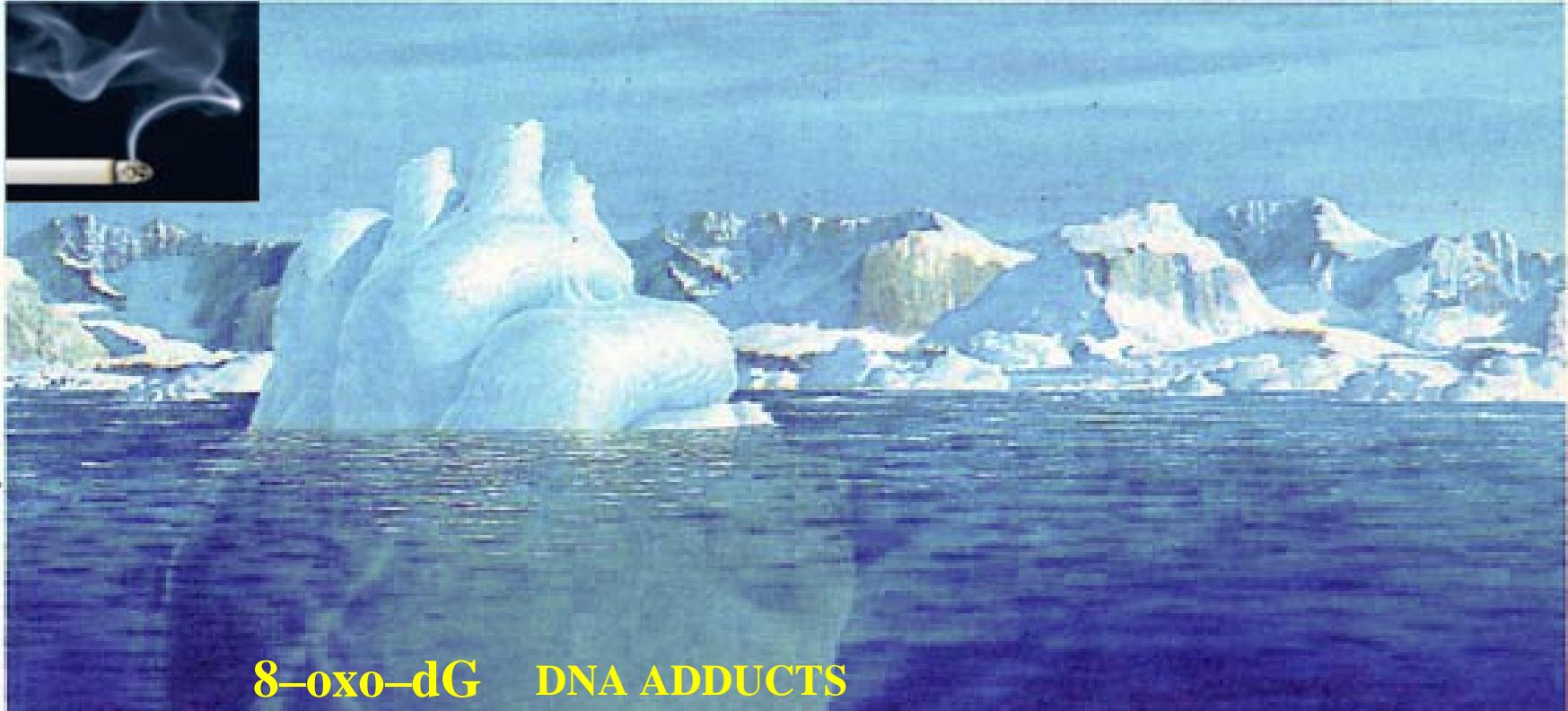
The levels of  $^{32}\text{P}$  postlabelled DNA adducts in the aorta from 85 atherosclerotic patients were significantly correlated with:



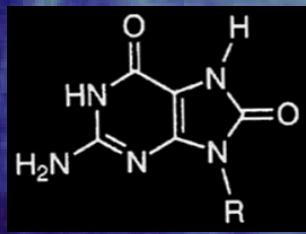
- Age of patients
- Number of cigarettes smoked currently
- High blood pressure
- Blood triglycerides
- Blood cholesterol (total/HDL)
- SFS-positive DNA adducts
- Oxidative DNA damage (8-OH-dG)

# MODULATION OF DNA ADDUCTS BY DIETARY AGENTS IN THE AORTA OF SMOKE-EXPOSED RATS

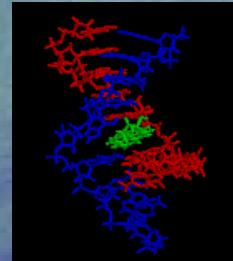




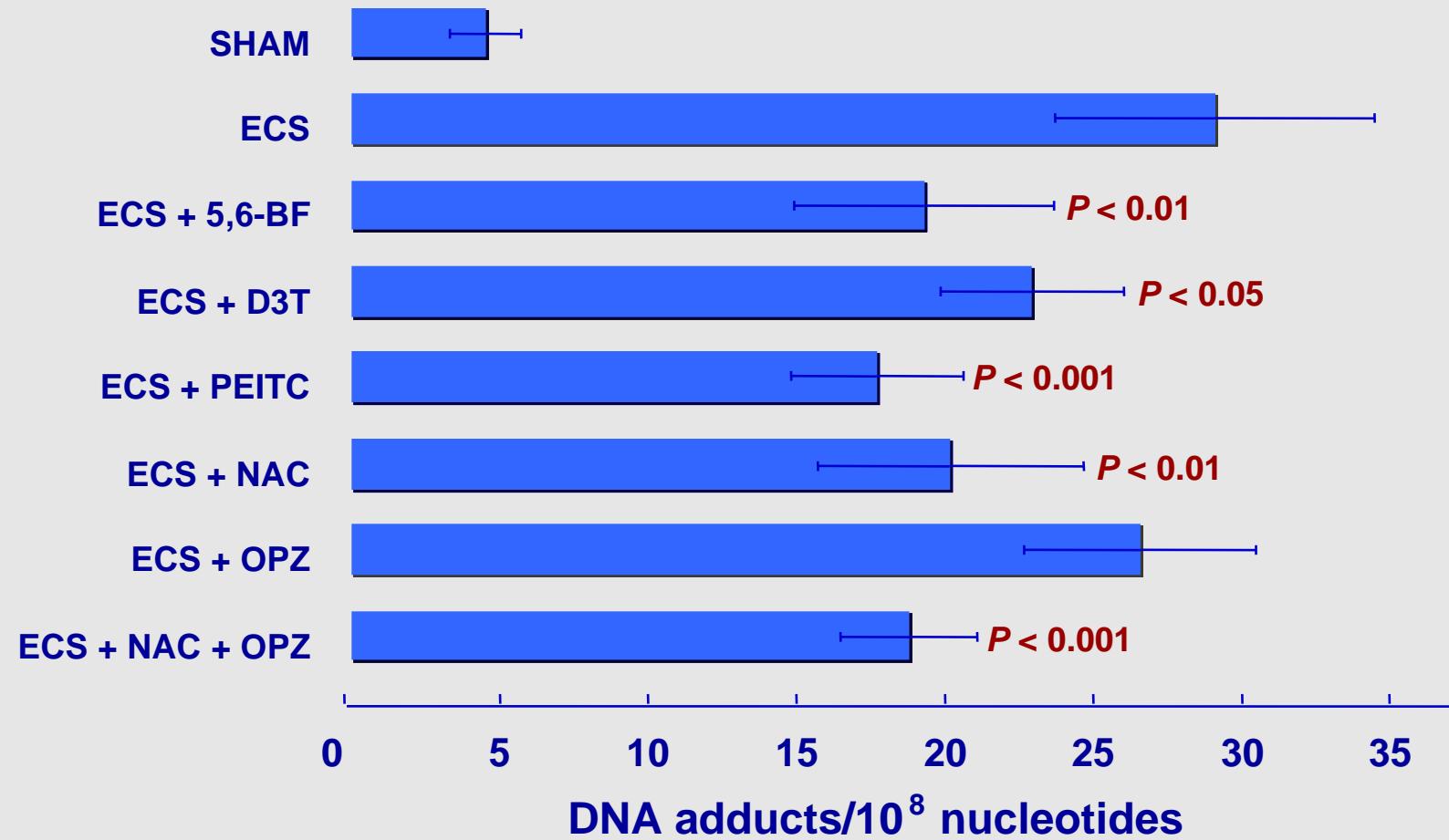
8-oxo-dG



DNA ADDUCTS



# MODULATION OF DNA ADDUCTS BY DIETARY AGENTS IN THE HEART OF SMOKE-EXPOSED RATS



## REQUIREMENTS

EFFICACY

LOW COST  
PRACTICALITY  
TOLERABILITY

## INTERVENTION (Targets)

THERAPY (Cancer patients)

TERTIARY PREVENTION  
(Treated cancer patients)

EARLY INTERVENTION  
(Cancer patients in preclinical or early stage)

PREVENTION OF PROGRESSION  
(Individuals affected by precancerous lesions)

TARGETED CHEMOPREVENTION  
(High risk individuals)

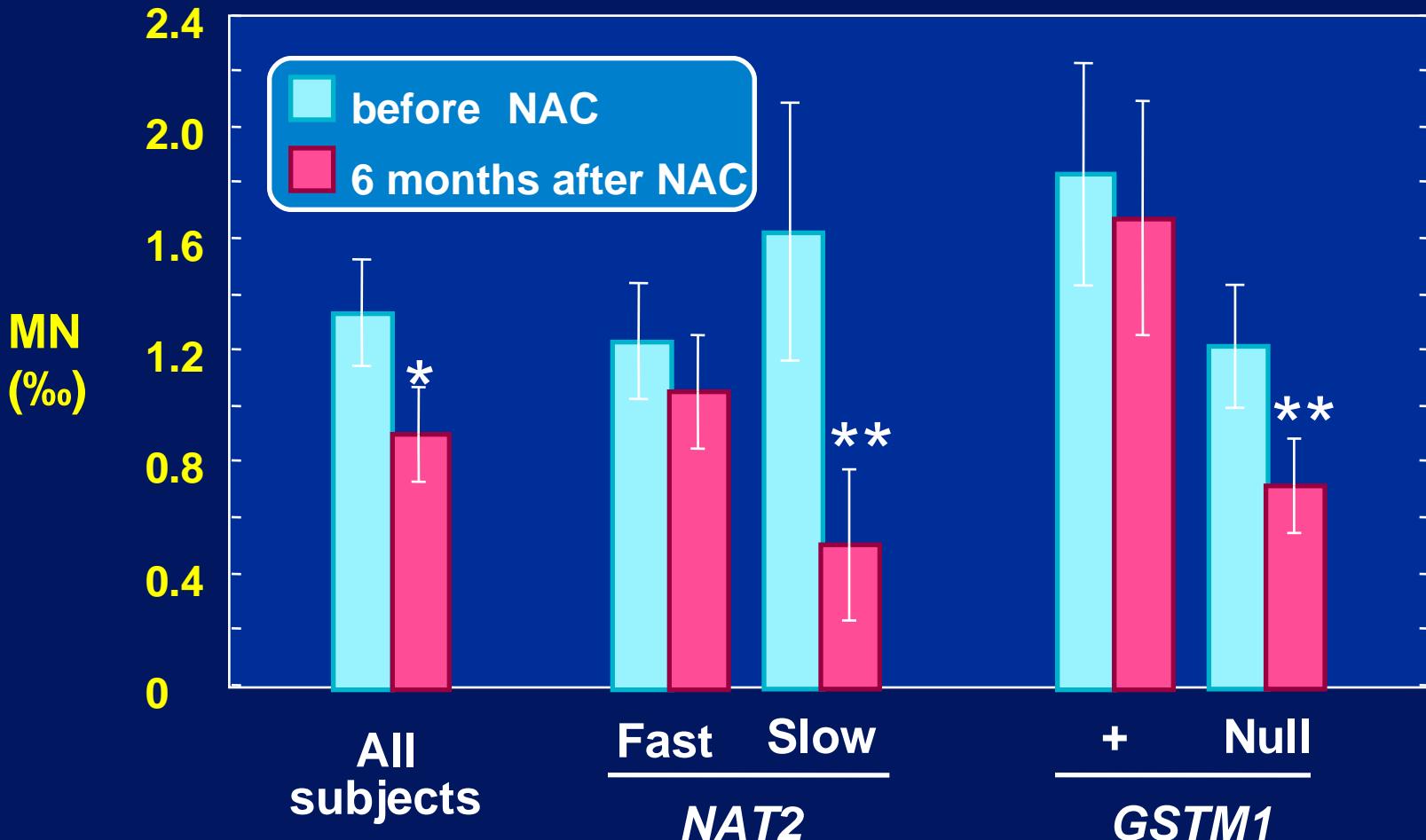
PUBLIC HEALTH INTERVENTION  
(Healthy subjects in the population)

# PHASE II CHEMOPREVENTION TRIAL WITH NAC IN DUTCH SMOKERS

	DNA adducts/ $10^8$ nucleotides in BAL cells	8-oxo-dGuo/ $10^5$ nucleotides in BAL cells	Micronuclei in mouth cells (%)
Placebo	$T_0$ <b><math>6.0 \pm 0.7</math></b>	$4.8 \pm 0.5$	$1.2 \pm 0.3$
	$T_6$ <b><math>5.9 \pm 0.7</math></b>	$3.2 \pm 0.8$	$1.0 \pm 0.2$
NAC	$T_0$ <b><math>6.0 \pm 0.9</math></b>	$4.9 \pm 0.7$	<b><math>1.3 \pm 0.3</math></b>
	$T_6$ <b><math>4.3 \pm 0.8</math></b>	<b><math>1.8 \pm 0.3</math></b>	<b><math>0.9 \pm 0.3</math></b>

Statistically significant as compared to  $T_0$

# PHARMACOGENOMICS / NUTRIGENOMICS OF CHEMOPREVENTIVE AGENTS



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