

1 **Title:**

2 **Characteristics of *Staphylococcus aureus* infections to consider in designing an**
3 **effective vaccine.**

4

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14

15 **Running title:**

16 **Designing a *S. aureus* vaccine**

17

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28 **ABSTRACT**

29

30 *Staphylococcus aureus* is a very versatile and adaptable microorganism. It can potentially
31 infect virtually any host tissue. Given the appropriate conditions it can become a life-
32 threatening pathogen, or a commensal colonizer of the nose. Extensive antibiotic use for
33 infection control facilitated the rise of antibiotic resistance, stressing the need for alternate
34 forms of control. Vaccine efforts in other pathogens have proved successful, but so far *S.*
35 *aureus* candidate vaccines have not been as effective. Here we review *S. aureus* factors
36 involved in pathogenesis that could help develop a successful vaccine, like host nasal
37 colonization and immune evasion factors. An effective multicomponent vaccine could
38 incorporate antigenic fragments from several *S. aureus* proteins, preferably involved in
39 colonization, immune evasion and/or toxicity.

40

41 **KEYWORDS:** *S. aureus*, Vaccine, Nasal colonization, Immune evasion.

42

43 **INTRODUCTION**

44

45 *Staphylococcus aureus* is a natural inhabitant of mammalian skin and certain mucous
46 epithelia. It is an opportunistic pathogen and has the ability to infect virtually every tissue
47 in the body of animals and humans, especially those at risk of infection like wounds, or
48 with diminished immunological protection like secretory glandular tissue (28). It is a very
49 versatile and adaptable organism; it can become pathogenic causing bacteremia or establish
50 a commensal relationship in humans without causing overt disease, as is the case in nasal

51 colonization. However, given the appropriate conditions each and every strain of *S. aureus*
52 can become a life-threatening pathogen (29).

53 Infection control usually requires antibiotics; however, their extensive use has facilitated
54 the emergence of strains with antibiotic resistance (21). Resistance to methicillin and
55 vancomycin has been observed in recent years in hospital and community acquired *S.*
56 *aureus* infections (5, 44). Methicillin resistance in *S. aureus* is mediated by the acquisition
57 of an exogenous gene, *mecA*, that encodes a β -lactam-resistant penicillin-binding protein
58 (PBP), termed PBP 2a (or PBP2') (22). There are two known types of vancomycin
59 resistance, complete (vancomycin-resistant *S. aureus*, VRSA) and intermediate resistance
60 (vancomycin intermediate resistant *S. aureus*, VISA). The VRSA resistance is mediated via
61 the apparent acquisition of the *vanA* gene that allows synthesis of modified peptidoglycan
62 precursors with decreased affinity for vancomycin. In VISA, genetic mutations that result
63 in production of a much thicker cell wall makes it very difficult for vancomycin to enter the
64 cell (10).

65 In this review we will present a summary of *S. aureus* nasal colonization and immune
66 evasion mechanisms used to overcome host responses, as well as strategies used in vaccine
67 design. Based in this information, we then suggest desirable characteristics that future
68 vaccine candidates may incorporate in their design.

69

70 **Nasal colonization**

71

72 The association between *S. aureus* nasal carriage and staphylococcal disease was first
73 reported by Danbolt in 1931 (45), which numerous studies confirmed afterwards (48, 53,
74 54). Several studies including historical medical controls have reported great reductions of
75 surgical site infections in patients pre-treated to remove *S. aureus* from their noses -nasal
76 decolonization- (7, 25). Although nasal carriage is one of the most important risk factors for
77 nosocomial and surgical site infections, randomized controlled trials have failed to confirm
78 a significant reduction in infection rates after nasal decolonization (24). Therefore,
79 clearance of nasal *S. aureus* is not a completely effective method for infection control. It is
80 possible that after nasal clearance, *S. aureus* that reside in other parts of the body are the
81 source of infection. *S. aureus* cells can survive for months on many types of surface (26),
82 and propagate from there through the hands of the patient (or relatives or caregivers) to the
83 site of infection, or even back to the nasal niche by nose picking (52).

84 Longitudinal studies distinguish at least three nasal carriage patterns in healthy individuals:
85 persistent carriage (about 20%), intermittent carriage (30%), and non-carriage (50%) (14,
86 47, 54). Persistent carriers have higher single-strain *S. aureus* loads (their "persistent
87 strain") and higher risk of developing staphylococcal infections (34), while intermittent
88 carriers may carry different strains over time (14, 47). Furthermore, after inoculation with a
89 mix of *S. aureus* strains, non-carriers quickly eliminate all strains, whereas persistent
90 carriers eliminate all strains except for their "persistent strain" when it was present in the
91 inoculation mix (33). It is important to notice that non-carriers who become infected from
92 exogenous *S. aureus* strains have a four-fold increased mortality rate compared with *S.*

93 *aureus* nasal carriers (53). The host immune response that kept non-carriers noses free from
94 *S. aureus* is not effective enough to prevent other *S. aureus* infections.

95 Recently, a study of anti-staphylococcal antibodies profile showed that levels of IgG and
96 IgA against 17 different *S. aureus* antigens were equal in intermittent carriers and non-
97 carriers but not in persistent carriers. This suggests there are only 2 types of nasal carriers:
98 persistent and non-persistent carriers (46).

99 Nasal carriage patterns are most likely determined by host and bacterial factors. No relation
100 has been observed between carriage rate and seasonality, temperature, or relative humidity
101 (30, 31, 54). Genetic studies have shown that a simple Mendelian trait probably does not
102 explain host carrier states (1, 3, 36). However, there are observed differences in bacterial
103 attachment to the nasal epithelia of carriers and non-carriers that suggest host factors
104 (genetic and/or environmental) can determine carrier state (2). Personal environmental
105 factors probably have a larger influence: carrier states are usually shared among household
106 members and most mothers carry the same strain as their children (36), suggesting that
107 close contact helps adaptation of the pathogen to its host. Even the anatomy of the nose
108 may influence carrier state (9, 38).

109 In summary, the existence of non-carriers suggest that there is an immune host response,
110 and probably some genetic host factors as well, that is effective in preventing *S. aureus*
111 colonization. Even in carriers, there is a balance between host and pathogen that allows
112 only a specific strain to colonize and prevents colonization from other strains. However,
113 this balance is lost when *S. aureus* manages to thwart host defenses and invades the host. It

114 is likely that genetic changes in the strain are partly responsible for the newly developed
115 abilities of the strain to overcome the host immune response (20), probably aided by host
116 changes, like wounds or diminished immune defenses, that facilitate infective processes.
117 It may be possible to elicit an immune response through vaccination that allows the host to
118 defend against invading bacteria, one that mimics the immune response of non-carriers.
119 Studies in host immune responses have identified *S. aureus* molecules that react strongly to
120 sera of non-carriers. Identified molecules are usually involved in immune evasion and
121 colonization mechanisms by *S. aureus* (13, 49).

122

123 **Immune evasion**

124

125 Usually after host internalization, a microorganism and its products are taken up by
126 macrophages and other antigen-presenting cells and transported to lymph nodes, where B
127 cells are stimulated to differentiate and secrete antibodies that neutralize toxins and
128 promote more efficient phagocytosis of bacterial cells. Antibodies to *S. aureus* antigens can
129 be detected in all humans, and titers usually rise after infection (13, 16, 40). However, these
130 antibodies and immunological memory seem to be inadequate to prevent subsequent
131 infections, which reflect the great capacity of *S. aureus* to compromise immune responses.
132 *S. aureus* has an impressive number of immune evasion factors to overcome host defense
133 mechanisms (11, 17). It is important to notice that many of these factors have multiple,
134 often redundant roles: if one of them is rendered inactive through mutation or antibody
135 targeting, its function can still be carried away by another redundant factor (Figure 1).

136 After the physical barrier of the skin is breached and the bacterium starts to grow inside the
137 host, the innate immune response is activated (50). *S. aureus* is particularly adept in
138 evading innate host defense, as evidenced by the abundance of mechanisms that the
139 bacterium uses to evade killing by phagocytes (17).

140 Complement activation is part of the innate immune response, and *S. aureus* has several
141 bacterial products that interfere with its function by (i) the recruitment or mimicking of
142 complement regulators, (ii) the modulation or inhibition of complement proteins by direct
143 interactions, and (iii) the inactivation by enzymatic degradation (27). Phagocyte function is
144 also altered by *S. aureus*, expressed *S. aureus* molecules can block phagocyte receptor
145 function. Bacteria may hide from recognition by producing protective coats, such as
146 capsular polysaccharide or biofilm. After ingestion by professional or non-professional
147 phagocytic cells, the bacteria use mechanisms to decrease the efficiency of antimicrobial
148 mechanisms and to survive killing mechanisms. Intracellular persistence provides a
149 protective niche from professional phagocytes and extracellular antibiotics, and can
150 promote recrudescence infection (19). *S. aureus* often produce toxins that lyse phagocytes
151 and superantigen toxins that overstimulate the immune system (11, 17). The tight control of
152 expression is also essential for pathogenesis. The expression of toxins, colonization and
153 immune evasion factors is controlled by complex regulatory networks that include the
154 quorum-sensing *agr* system, transcriptional regulators of the *sar* family, the two-
155 component regulatory systems *ArlRS* and *SaeRS*, and the alternative sigma factor SigB

156 (35). A more detailed review of *S. aureus* molecules that contribute to immune evasion or
157 alter host immune function is presented elsewhere (11, 17).

158

159 **Vaccine designs.**

160

161 Is a *S. aureus* vaccine feasible? This is not a question with an easy answer. Recovery from
162 a *S. aureus* infection does not appear to confer immunity against subsequent infections,
163 which cast doubts in the feasibility of generating a better protective immune response than
164 the one induced after natural infection. However, work in the prevention of bovine mastitis
165 (32) showed a 50-70% protection level when using killed bacteria combined with α - and β -
166 toxin toxoids, indicating that it may be possible to generate an immune response with an
167 improved level of protection. Since it is not appropriate to use whole killed *S. aureus*
168 vaccine preparations in humans, alternatives have to be found.

169 Several reviews covering vaccine development have been published (12, 39, 42).

170 Concisely, the few vaccine candidates that have advanced to clinical trials have failed to
171 show positive results, even after having shown excellent results in animal models of
172 infection. Lessons learned in other pathogens might not be transferable to *S. aureus*. For
173 example, vaccines based in capsular polysaccharides of other bacterial pathogens have
174 proven successful (15, 18), whereas in *S. aureus* failed to show protection (43). Important
175 efforts are directed towards creating staphylococcal subunit vaccines (39), although one
176 could argue that a vaccine carrying only one bacterial factor as immunogen might not work
177 due to the ample role redundancy of *S. aureus* molecules (Figure 1).

178 Another approach consists of letting the human immune response choose the best target for
179 vaccination (8, 13, 49, 51). Sera of carriers vs. non-carriers, or healthy vs. infected human
180 subjects is probed against a library of expressed *S. aureus* proteins, with the idea of identify
181 potential targets already recognized by a "good" human immune response to be used as
182 candidate vaccines. This approach has its strength in that bacterial proteins recognized by
183 human sera with a high antibody titer and opsonic activity against *S. aureus in vitro*, are
184 more likely to perform better as vaccine components than bacterial proteins not recognized.
185 Its weakness resides in that selected targets are only as effective as the antibodies used to
186 select them. The fact that non-carriers that successfully fend off *S. aureus* from colonizing
187 their noses can still become severely ill from a *S. aureus* infection means their antibodies
188 are not completely capable of excluding *S. aureus* under all conditions.
189 There is also the possibility that an identified *S. aureus* candidate antigen in its fully
190 functional state is poorly antigenic, resulting in few or no induced antibodies in the host.
191 Mice immunized with fibronectin-binding protein (FnBP) were able to resist an *S. aureus*
192 infection challenge (6, 41), but were unable to block binding of FnBP to host fibronectin. It
193 was found that native FnBP is poorly immunogenic, but after binding to fibronectin, the
194 FnBP-fibronectin complex is actually immunogenic (37). Therefore, induced antibodies
195 were actually recognizing the FnBP-fibronectin complex instead of preventing its binding.
196 It was later found that biologically inactive FnBP fragments were actually capable of
197 inducing antibodies that could recognize native FnBP and prevent its binding to fibronectin,
198 therefore being better immunogens than native FnBP (4, 23) (Figure 2).

199 Therefore, it is possible that an effective vaccine could incorporate antigenic fragments
200 from several *S. aureus* proteins, a multicomponent vaccine, preferably involved in
201 colonization, immune evasion and/or toxicity (Table 1). These antigenic fragments would
202 be biologically inactive proteins that could still induce protective antibodies. The absence
203 of biologically active antigens can increase safety and the targeting of multiple *S. aureus*
204 proteins at once decreases the chances of *S. aureus* immune evasion.

205

206 **CONCLUSION**

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208 A successful vaccination protocol could greatly contribute to *S. aureus* infection control,
209 providing a much needed relief in reliance of antibiotic therapy alone. Also, advances in the
210 field will help develop effective passive immunization protocols, which are useful in cases
211 where prophylactic vaccination is not possible. Much work is still needed, but the field has
212 greatly progressed lately, with a growing number of research groups working in *S. aureus*
213 vaccine development, as observed after a Pubmed search in *S. aureus* vaccines
214 (<http://www.ncbi.nlm.nih.gov/sites/entrez/> access date: July 2009). It is likely that *S.*
215 *aureus* infection control can be achieved soon.

216

217 Table 1. Possible targets for vaccine development

218	<i>aaa</i>	Autolysin / adhesin from <i>S. aureus</i>
219	<i>atl</i>	<i>S. aureus</i> autolysin
220	<i>aur</i>	Zinc metalloproteinase aureolysin, Aur
221	<i>bbp</i>	Bone sialo-protein binding protein
222	<i>cap5 / cap8</i>	Capsular polysaccharide
223	<i>chp</i>	Chemotaxis inhibitory protein of <i>S. aureus</i>
224	<i>clfA, ClfB</i>	Clumping factor A and B
225	<i>can</i>	Collagen binding protein
226	<i>coa</i>	Staphylococcal coagulase
227	<i>crtM, crtN</i>	Carotenoid pigment, staphyloxanthin
228	<i>dltc</i>	DltC, from Dlt operon, DltABCD
229	<i>eap</i>	Extracellular adherence protein
230	<i>ebh</i>	extracellular matrix (ECM) binding protein homologue
231	<i>ebps</i>	Elastin binding protein
232	<i>ecb</i>	Extracellular complementbinding protein
233	<i>efb</i>	Extracellular fibrinogenbinding protein
234	<i>emp</i>	Extracellular matrix protein-binding protein
235	<i>fbpa</i>	Fibrinogen binding protein
236	<i>fnbA, fnbB</i>	Fibronectin-binding proteins A and B
237	<i>hla, hly</i>	Alpha-hemolysin (α -hemolysin)
238	<i>hld</i>	Delta-hemolysin
239	<i>hlgA, hlgB, hlgC</i>	Gamma-hemolysin subunits A, B, and C
240	<i>icaD, icaB, icaC</i>	Polysaccharide intercellular adhesin, PIA
241	<i>isdA, isdB</i>	Iron-regulated surface determinants of <i>S aureus</i> ,
242	<i>lukS-PV, lukF-PV</i>	Leukocidin S-PV and F-PV, Panton Valentine leukocidin
243	<i>lukD, lukE</i>	Leukocidin D and E
244	<i>mprF</i>	Multiple peptide resistance factor
245	<i>pls</i>	plasmin sensitive protein PLS
246	<i>psm</i>	Phenol-soluble modulinkinlike peptides
247	<i>rap</i>	RANIII activating protein
248	<i>sak</i>	Staphylokinase
249	<i>sasg</i>	<i>S. aureus</i> surface protein G
250	<i>sbi</i>	IgG-binding protein
251	<i>scn</i>	Staphylococcal inhibitor of complement
252	<i>sea, seb, secn, sed, see, seg,</i>	Staphylococcal enterotoxins
253	<i>seh, sei, sej, sek, sel, sep</i>	
254	<i>spa</i>	Protein A
255	<i>ssl5</i>	Staphylococcal superantigen-like 5, SSL5
256	<i>ssl7</i>	Staphylococcal superantigen-like 7, SSL7
257	<i>tst</i>	Toxic shock syndrome toxin-1, TSST1
258	<i>vbnp</i>	Vibronectin binding protein
259	<i>vwbp</i>	Von Willebrand factor binding protein
260	<i>eno</i>	α -enolase, Laminin binding protein

261 Figures:

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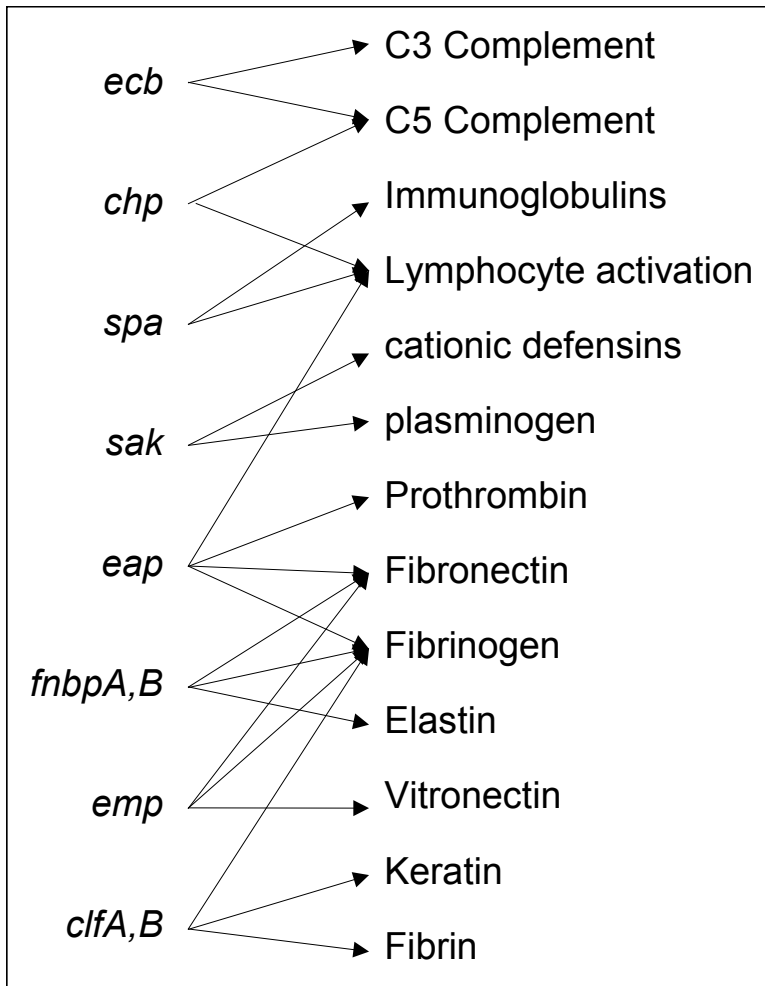
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276 **Figure 1.** Redundancy in *S. aureus*. Expressed products of *S. aureus* genes involved in

277 pathogenicity, colonization and/or immune evasion usually interact with several targets.

278 Additionally, host factors are usually targeted by different bacterial factors. Redundancy

279 assures infection processes can continue even when some bacterial factors are neutralized

280 by host responses or vaccination

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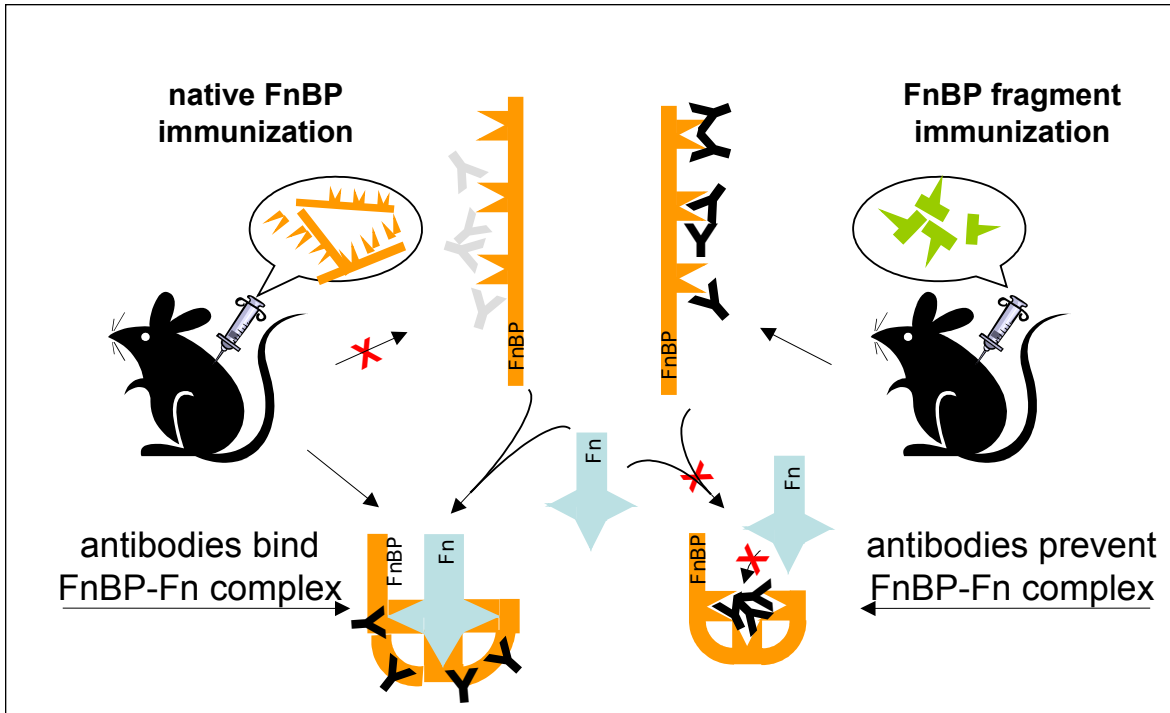
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295 **Figure 2.** Immunization with FnBP. Immunization with native FnBP induces antibodies

296 that fail to prevent binding of FnBP to Fn, by failing to recognize native FnBP; but

297 recognizes the FnBP-Fn complex. Immunization with biologically inactive FnBP fragments

298 induces antibodies effective in preventing binding of FnBP to Fn, by recognizing native

299 FnBP and failing to bind the FnBP-Fn complex.

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