

1 ***Chromobacterium violaceum* ATCC12472: Multi-drug**  
2 ***and ethidium bromide resistant***

3

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9 **Abstract**

10

11 *Chromobacterium violaceum* is an opportunistic human pathogen causing a range of gastric  
12 infections and occasionally septicemia. This Gram-negative bacillus is a common inhabitant of  
13 soil and water in tropical and subtropical areas of the world. Infection occurs after contamination  
14 of damaged skin exposed to soil or environmental water. Alternatively, systemic infection can  
15 follow the aspiration or ingestion of contaminated water. The major features of infections by *C.*  
16 *violaceum* are, in generally, rapid clinical course, multiple visceral abscesses, and high  
17 mortality. Genomic data on the type strain ATCC 12472 has provided a comprehensive basis for  
18 detailed studies of pathogenicity, virulence and drug resistance genes. In this study, the  
19 susceptibility of this organism was tested on a variety of drugs at different concentrations, in  
20 solid and liquid media. *C. violaceum* shown to be resistance to ampicillin, penicillin, rifampicin,  
21 erythromycin, vancomycin and also to ethidium bromide. The bacteria was susceptible to  
22 gentamicin, tetracycline, chloramphenicol, cotrimoxazole, kanamycin, streptomycin and nalidixic  
23 acid, at the tested concentrations.

24 **Key words:** *Chromobacterium violaceum*; multi-drug resistance; ethidium bromide.

25 **Introduction**

26

27 *Chromobacterium violaceum* is a common inhabitant of soil and water in tropical and sub-  
28 tropical regions. This Gram negative bacillus is considered as a saprophyte, but occasionally it  
29 can act as an opportunistic pathogen for animals and man (Crosse *et al.* 2006; Fombuena *et al.*  
30 1998; Hassan *et al.* 1993; Ti *et al.* 1993). In immunocompromised individual the localized lesion  
31 might lead to septicaemic infections with abscesses in multiple internal organs, urinary tract  
32 infection and diarrhoea. The most common feature of this infection is sepsis, followed by  
33 cutaneous involvement and liver abscesses (Midani & Rathore, 1998). Lung abscesses have  
34 also been reported (Sirinavin *et al.* 2005; Tee *et al.* 2006; Teoh *et al.* 2006). Mortality rate is  
35 very high for patients with disseminated infections (Ang, 2004). The first case in humans was  
36 reported in Malaysia in 1927, and the medical literature contains reports of approximately 150  
37 human cases since then (Midani & Rathore, 1998; Ray *et al.* 2004; Teoh *et al.* 2006). This  
38 bacteria is able to compete with other bacteria in the environment and possess a highly  
39 effective survival mechanism (Brito *et al.* 2004; Duran & Menck, 2001). Infection treatment is  
40 very difficult and currently there are no vaccines. A successful treatment with ciprofloxacin and  
41 amikacin has been reported (Ray *et al.* 2004).

42 The resistance of prokaryotic organisms to many unrelated hydrophobic  
43 chemotherapeutic drugs is termed multi-drug resistance (MDR). A variety of MDR efflux  
44 systems are widely distributed among prokaryotic microorganisms, including pathogenic  
45 bacteria (Aeschlimann, 2003; Lewis, 1994; Lewis *et al.* 1994; Marquez, 2005). They belong to  
46 three different families of transport proteins: the major facilitator superfamily (MFS) (Saidijam *et al.*  
47 2006; Vardy *et al.* 2004), the resistance nodulation division (RND) family (Gotoh *et al.* 1999;  
48 Piddock, 2006; Saier *et al.* 1994), and the small multidrug resistance (SMR) family of small  
49 translocases (Jack *et al.* 2000; Paulsen *et al.* 1996; Tate, 2006).

50 Genomic data of *C. violaceum* ATCC 12472 (<http://www.brgene.lncc.br/cviolaceum/>) has  
51 provided a comprehensive basis for detailed studies of pathogenicity, virulence and drug  
52 resistance genes (Brito *et al.* 2004; Fantinatti-Garboggini *et al.* 2004). The genome contains  
53 extensive but incomplete arrays of ORFs coding for proteins associated with mammalian  
54 pathogenicity, possibly involved in the occasional but often fatal cases of human *C. violaceum*

55 infection (Consortium, 2003). In this study, we provided experimental data of drug resistance of  
56 *C. violaceum* ATCC12472 on a variety of antibiotics, in different concentrations, in solid and  
57 liquid media. Ethidium bromide was also tested, in order to confirm the resistance to this toxic  
58 compound, as indicated by the genome data.

59

## 60 **Material and Methods**

61

62 *Chromobacterium violaceum* ATCC12472 was grown for 12 hours on shaking in Luria  
63 Bertani (LB) broth medium and in LB agar plates at 30°C. For the susceptibility assay, fourteen  
64 different drugs, including ethidium bromide, were tested at different concentrations when *C.*  
65 *violaceum* were cultivated in liquid and in solid LB medium. Antimicrobial susceptibility was also  
66 tested using plate disc diffusion method. Experiments were repeated on triplicates and shown  
67 agreement.

68

## 69 **Results**

70 *Chromobacterium violaceum* ATCC12472 strain is resistant to a variety of antibiotics and  
71 is also resistant to ethidium bromide, as shown in Tables 1-3. This organism is more sensitive to  
72 gentamycin and tetracycline at concentrations higher than 10µg/ml. It is very resistant to  
73 rifampicin, vancomycin, and ampicillin, even at concentrations higher than 130µg/ml. This strain  
74 was found sensitive to erythromycin at concentration higher than 50µg/ml when it was cultivated  
75 in LB broth. However, in LB agar plates it was resistant to erythromycin at concentrations higher  
76 than 130µg/ml.

77

## 78 **Discussion**

79 *Chromobacterium violaceum* ATCC12472, a wild type strain, showed to be resistant to  
80 ethidium bromide even at concentrations higher than 50µg/ml. It has been reported that  
81 mutants of *Bacillus subtilis* was found to be resistant to 10 µg of ethidium bromide per ml  
82 (Bishop & Brown, 1973). *E-coli* mutants have been reported to be resistant to ethidium bromide  
83 at concentration higher than 75µg/ml, when *mdfA* was over-expressed. MdfA is a putative  
84 membrane transport protein and belongs to the MFS superfamily (Edgar & Bibi, 1997). There

85 are a large number of open reading frames associated with various mechanisms of drug  
86 resistance in the *C. violaceum* genome. Genes associated with bacitracin, bicyclomycin,  
87 chloramphenicol, kasugamycin, and methylenomycin were also found (Fantinatti-Garboggini *et al.*  
88 *et al.* 2004), comprising a remarkable feature of this organism. Families of drug extrusion  
89 translocases have been identified in the *C. violaceum* genome, based on sequence homology.  
90 These are the MFS (major facilitator super family), SMR (small multidrug resistance), RND  
91 (resistance nodulation cell division), ABC (ATP-binding cassette), and MATE (multidrug and  
92 toxic compound extrusion) family (Fantinatti-Garboggini *et al.* 2004). EmrE, the product of the  
93 *emrE* gene homolog identified in the *C. violaceum* genome is an SMR-related protein. The most  
94 striking feature of EmrE is that it is capable of transporting substrates of widely varying  
95 structure, provided that overall the compound is highly hydrophobic and carries a positive  
96 charge. EmrE couples the extrusion of toxins to the influx of protons down their electrochemical  
97 gradient, with probably two or more protons transported per toxin molecule (Arkin *et al.* 1996;  
98 Lebediker & Schuldiner, 1996; Schuldiner *et al.* 1996; Venkatraman *et al.* 2002; Yerushalmi *et al.*  
99 *et al.* 1996). Presence of a large number of open reading frames associated with various  
100 mechanisms in *C. violaceum* genome may help to explain the resistance of this organism to  
101 ethidium bromide and to a variety of antibiotics tested.

102

### 103 **Conclusion**

104 In this study, we have tested drug resistance of *C. violaceum* ATCC12472 strain at  
105 different concentrations of different antibiotics. *C. violaceum* is resistance to a wide broad range  
106 of antibiotics, and was found to be resistant to ethidium bromide at concentrations higher than  
107 50µg/ml. The presence of multi-drug resistance genes found in *C. violaceum* genome may  
108 explain such ability to survive, and may represents a challenging for infection treatment.

109

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230

Table 1. Test of Antibiotics resistance to *Chromobacterium*

231

*violaceum* (ATCC12472) in disc plate.

<b>Antibiotic</b>	<b>Concentration (<math>\mu\text{g/ml}</math>)</b>	<b>*Susceptibility of <i>C. violaceum</i> ATCC12472C.</b>
Ampicilin	10	+
Penicilin	10	+
Gentamicin	10	-
Tetracycline	30	-
Chloramphenicol	30	-
Cotrimoxazole	25	-

232

\* (+) Resistant; Sensitive(-).

233

Table 2. Test of antibiotics resistance to *Chromobacterium violaceum* (ATCC12472) in solid

234

media (LB agar)

<b>Antibiotic</b>	<b>Concentration (<math>\mu\text{g/ml}</math>)</b>	<b>*Susceptibility of <i>C. violaceum</i> ATCC12472</b>
Rifampicin	10	+
	20	+
	30	+
	50	+
	100	+
	150	+
	200	+
Erythromycin	50	+
	100	+
	150	+
Ethidium bromide	10	+
	15	+
	20	+
	25	+
	40	+

235

\* (+) Resistant; Sensitive(-).

236

Table 3. Test of antibiotics resistance to *Chromobacterium*

237

*violaceum* (ATCC12472) in liquid media (LB broth).

Antibiotic	Concentration ( $\mu\text{g/ml}$ )	*Susceptibility of <i>C. violaceum</i> ATCC12472
Ampicilin	50	+
	100	+
	150	+
Tetracycline	10	-
Kanamycin	50	-
Chloramphenicol	50	-
Streptomycin	50	-
Nalidixic Acid	15	-
Vancomycin	50	+
	75	+
	100	+
	150	+
Erythromycin	50	+
	75	-
	100	-
	150	-
Rifampicin	50	+
	100	-
	150	-
	200	-
Carbamycin	20	+
	50	+
	100	-
	150	-
Ethidium bromide	1	+
	10	+
	30	+
	60	+

238

\* (+) Resistant; Sensitive(-).