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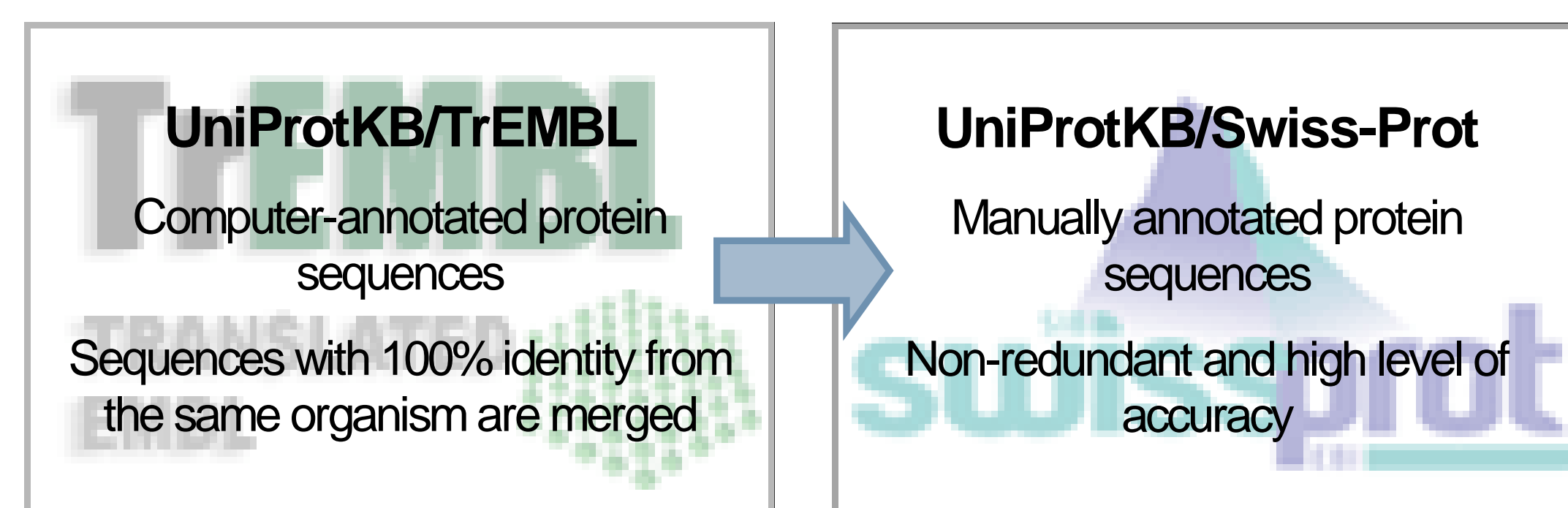
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Introduction to UniProt

• UniProt (Universal Protein Resource: <http://www.uniprot.org>) provides a central resource of protein sequences with functional annotation. The UniProt Consortium is a collaboration between the Swiss Institute of Bioinformatics (SIB), the European Bioinformatics Institute (EBI) and the Protein Information Resource (PIR).

• UniProt Knowledgebase (UniProtKB) contains the manually annotated UniProtKB/Swiss-Prot section and the automatically annotated UniProtKB/TrEMBL section.



Why frogs and fish?

• *Xenopus laevis* (African clawed frog) and *Danio rerio* (zebrafish) are both model organisms in the laboratory, as they are easy to accommodate and can produce large numbers of externally-developing embryos that can be easily viewed and manipulated.

• With a generation time of 2 to 3 months, *D. rerio* is amenable to genetic analysis, and the Sanger Institute began sequencing the zebrafish genome in 2001. The extensive similarity between the zebrafish and human genomes means many human developmental and disease genes have counterparts in the zebrafish.

• *X. laevis* has a much longer generation time of 1-2 years, and its tetraploid genome creates difficulties for genetic analysis. The related species *Xenopus tropicalis* (Western clawed frog) is diploid with a relatively small genome

	<i>X. laevis</i>	<i>X. tropicalis</i>
Ploidy	Allotetraploid	Diploid
Genome Size	3.1 x 10 ⁹ bp	1.7 x 10 ⁹ bp
Adult size	10 cm	4.5 cm
Generation time	1-2 years	4 months
Egg size	1-1.3 mm	0.7 - 0.8 mm

and a shorter generation time, so is much better suited for genetic studies. Sequencing of the *X. tropicalis* genome has begun and is supported by cDNA and EST sequencing projects, making it an ideal time to focus on Xenopus curation.

The Challenge of Duplicated Genomes

• Both *Danio rerio* and *Xenopus laevis* have undergone whole genome duplications, resulting in duplicated genes for multiple loci.

• Protein sequences encoded by duplicated *X. laevis* and *D. rerio* genes are annotated as separate UniProtKB/Swiss-Prot entries.

Accession	Entry name	Status	Protein names	Gene names	Organism
Q9W707	FXF1B_XENLA	★	Forkhead box protein F1-B (FoxF1-B) (FoxF1b) (Fork head domain-related protein 13) (xFD-13)	foxf1-B	<i>Xenopus laevis</i> (African clawed frog)
Q9W706	FXF1A_XENLA	★	Forkhead box protein F1-A (FoxF1-A) (FoxF1a) (Fork head domain-related protein 13) (xFD-13)	foxf1-A	<i>Xenopus laevis</i> (African clawed frog)

A and B nomenclature is used to distinguish the duplicated sequences

Accession	Entry name	Status	Protein names	Gene names	Organism
Q6PCS4	SN25B_DANRE	★	Synaptosomal-associated protein 25-B (SNAP-25) (Synaptosome-associated protein 25.2) (SNAP-25.2)	snap25b (Snap) (snap25.2)	<i>Danio rerio</i> (Zebrafish) (Brachydanio rerio)
Q5T266	SN25A_DANRE	★	Synaptosomal-associated protein 25-A (SNAP-25A) (Synaptosome-associated protein 25.1) (SNAP-25.1)	snap25a (Snap) (snap25.1) (si:dkeyp-814.6)	<i>Danio rerio</i> (Zebrafish) (Brachydanio rerio)

• For *D. rerio*, the UniProtKB A/B nomenclature matches ZFIN. For *X. laevis*, the A/B naming is taken from the literature where available, or assigned by a curator when the literature doesn't specify.

Curation of a *X. tropicalis* TrEMBL Entry

For both *Xenopus* and zebrafish, protein and gene nomenclature is generally propagated from the human ortholog, with species-specific names added as synonyms

Identifiers from large-scale projects are recorded

Functional annotation is attributed to individual papers

Evidence tags are used throughout the entry to show the source of the annotation

Isoforms are annotated based on all available sequences and literature

GO annotation is added manually using the Protein2GO editor. IEA annotations are generated from automatic methods, including KW:GOterm mapping

Computer-generated keywords (KW) are verified by a curator, and further keywords are added manually

Evidence tags show the source of annotation

Internal comments are used to explain annotation and point to related annotated entries

- We use a protein-by-protein approach to curation.
- PubMed searches, requests from users, cross-reference updates and sequence revisions are all used to identify which proteins are of priority to curate.
- Entries are curated using our flat-file editor, CRISP.

```
CRISP [rFXR1X1.new* - E:curation]
File Edit View Find Options Project Language Tools Window Help | Analyze Get Seq Lines Entry Info Projects Setup
-----
rFXR1X1.new*
-----
ID FXR1_XENTR Unreviewed; 674 AA.
AC Q5B356; Q28C19;
DT 12-APR-2005; integrated into UniProtKB/TrEMBL.
DT 12-APR-2005; sequence version 1.
OT 02-MAY-2006; entry version 9.
DE Fragile X mental retardation syndrome-related protein 1 homolog[EC5];
DE AltName: Full=FXR1p[EC6];
GN Name=FXR1[EC5]; ORFNames=Teg078n09.1[EC1];
OC Xenopus tropicalis (Western clawed frog) (Silurana tropicalis);
OC Eukaryota; Metazoa; Chordata; Craniata; Vertebrata; Euteleostomi;
OC Amphibia; Batrachia; Anura; Mesobatrachia; Pipoidae; Pipidae;
OC Xenopodinae; Xenopus; Silurana.
OX NCBI_TaxID=8364;
RN [1][E1];
RP NUCLEOTIDE SEQUENCE [LARGE SCALE MRNA].
RC TISSUE=Egg[EC10];
RG Sanger Xenopus tropicalis EST/cDNA project;
RL Submitted (MAR-2005) to the EMBL/GenBank/DBJ databases.
RN [2][E12,EC4];
RP NUCLEOTIDE SEQUENCE [LARGE SCALE MRNA] OF 4-674 (ISOFORM 2).
RC TISSUE=Embryo[EC12];
RG NIH - Xenopus Gene Collection (XGC) project;
RL Submitted (MAR-2005) to the EMBL/GenBank/DBJ databases.
RN [3][EC4];
RP IDENTIFICATION (ISOFORM 1), SUBCELLULAR LOCATION, AND TISSUE SPECIFICITY.
RX PubMed:15968590; DOI=10.1387/jdb.0159741b;
RA Blonden L., van 't Padje S., Severijnen L.-A., Destree O.,
RA Oostra B.A., Willemsen R.;
RT "Two members of the Fxr gene family, Fmr1 and Fxr1, are differentially
RT expressed in Xenopus tropicalis.";
RL Int. J. Dev. Biol. 49:437-441(2005).
```

References include both large scale projects (including XGC/ZGC and Sanger cDNA projects), and sequences and papers from individual labs. A PubMed search is performed to find additional characterization papers. All available papers are manually curated and added to a TrEMBL entry

A wide range of functional data is taken from papers. Data is also propagated from similar UniProtKB/Swiss-Prot entries, and predicted by sequence analysis tools

Most cross-references are added automatically. Some (E.g. PROSITE, EMBL/DBJ/GenBank, and MOD X-ref lines) can be added or modified by a curator

Reciprocal links to Xenbase gene pages were introduced at the end of 2006

Key residues, domains and motifs are annotated based on sequence analysis tools (such as Anabelle), similarity to existing UniProtKB/Swiss-Prot entries, and information in the literature

The longest isoform is usually displayed. Where conflicts exist, for frogs and fish the consensus sequence is normally displayed

Acknowledgements

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