

ARHI (DIRAS 3), an Imprinted Tumor Suppressor Gene, Binds to Importins, and Blocks Nuclear Translocation of Stat3

Shaoyi Huang¹, In Soon Chang², Wenbo Lin³, Robert Z. Luo¹, Zhen Lu¹, Yiling Lu⁴, Ke Zhang², Warren S-L. Liao¹, Tao Tao³, Robert C. Bast Jr.¹, Xiaomin Chen², Yinhua Yu^{1,*}, #

Departments of ¹Experimental Therapeutics, ²Biochemistry and Molecular Biology and ⁴Systems Biology, The University of Texas, M.D. Anderson Cancer Center, Houston, Texas 77030, U.S.A.; ³School of Life Sciences, Xiamen University, Xiamen, Fujian 361005, People's Republic of China

Running title: Function of ARHI in nuclear import of Stat3

Key words: ARHI, importin, nuclear import, Ran, Stat3

* To whom proofs are to be addressed: Yinhua Yu, Department of Experimental Therapeutics, The University of Texas. M.D. Anderson Cancer Center, 1515 Holcombe Blvd. Box 354, Houston, TX 77030. Telephone: 713-792-3790, FAX: 713-745-2107, E-mail: yyu@mdanderson.org

#Y. Yu's current address is Obstetrics and Gynecology Hospital of Fudan University, Shanghai, China 200011.

ABSTRACT

ARHI (DIRAS3) is an imprinted tumor suppressor gene whose expression is lost in the majority of breast and ovarian cancers. Unlike its homologs Ras and Rap, ARHI functions as a tumor suppressor. Our previous study showed that ARHI can interact with transcription activator Stat3 and inhibit its nuclear translocation in human breast and ovarian cancer cells. To identify proteins that interact with ARHI in nuclear translocation, we have performed proteomic analysis and identified several importins that can associate with ARHI. To further explore this novel finding, we have purified 10 GST-importin fusion proteins (importin 7, 8, 13, β 1, α 1, α 3, α 5, α 6, α 7 as well as mutant α 1). Using a GST-pull down assay, we found that ARHI can bind strongly to most importins; however, its binding is significantly reduced with an importin α 1 mutant which contains an altered nuclear localization signal (NLS) domain. In addition, an ARHI N-terminal deletion mutant (NTD) exhibits much less binding to all importins than does wild type ARHI. ARHI and NTD proteins were purified and tested for their ability to inhibit nuclear importation of proteins in HeLa cells. ARHI protein inhibits interaction of Ran-importin complexes with GFP fusion proteins that contain an NLS domain and a beta-like import receptor binding domain, blocking their nuclear localization. Addition of ARHI also blocked nuclear localization of phosphorylated Stat3 β . By GST-pull down assays, we found that ARHI could compete for Ran-importins binding. Thus, ARHI-induced disruption of importin binding to cargo proteins including Stat3 could serve as an important regulatory mechanism that contributes to the tumor suppressor function of ARHI.

INTRODUCTION

Transport of macromolecules between the nucleus and cytoplasm is critical for the normal function of eukaryotic cells. Two groups of karyopherins - importins and exportins - mediate RanGTPase-dependent transport through the nuclear pore (1). During malignant transformation, aberrant nucleocytoplasmic transport of transcription factors (such as Stat3 and E2F1) (2, 3) and their regulatory kinases (such as Sgk and Erk/MAPK) (4) occurs through impaired nuclear import, enhanced export, suppression of degradation, and sequestration in protein aggregates. Conversely, secreted factors such as CCN proteins, EGF, FGFs and their receptors are often detected in the nucleus of cancer cells. Nuclear localization of these molecules has been correlated with tumor progression and poor prognosis for patient survival (5, 6).

The classical nuclear import pathway consists of importin α and β . Whereas importin α interacts with a nuclear localization signal (NLS) in the cargo, importin β binds to the autoinhibitory domain on importin α and mediates the transport of the trimeric complex from the cytoplasm to the nucleus through the nuclear pores. Once inside the nucleus, the small GTP binding protein Ran (RanGTP) dissociates the complex by interacting with importin β . Importin α and β are shuttled separately back to the cytoplasm (7). There are more than 20 members in the importin family, including importin $\alpha 1, \alpha 3, \alpha 4, \alpha 5, \alpha 6, \alpha 7$, importin $\beta 1, 7, 8, 9$, and 13 (8, 9, 10, 11, 12). Importin α s are composed of a flexible N-terminal importin- β -binding (IBB) domain. The flexible IBB domain interacts either in trans with importin β or in cis with the cNLS-binding groove (8). Importin β s have in common an N-terminal Ran binding domain. Importins direct the import of various cargoes and may have different functions. For example, importin β /importin 7 heterodimer is a functional nuclear import receptor for histone H1 (10);

importin β , transportin, importin 7, and importin 9 promoted efficient import of c-Jun into the nucleus; importin alpha, by contrast, inhibited nuclear import of c-Jun in vitro (11). Importin 13, a recently identified importin β family member, regulates nuclear import of the glucocorticoid receptor in airway epithelial cells (12).

Ran is a small Ras-like GTP-binding protein that switches between a GTP- and a GDP-bound form by GTP hydrolysis and nucleotide exchange (13). The GTPase Ran plays a crucial role in nucleo-cytoplasmic transport of tumor suppressors, proto-oncogenes, signaling molecules and transcription factors. The RanGTPase cycle provides directionality to nucleocytoplasmic transport, regulating interactions between cargoes and nuclear transport receptors of the importin- β family. The common principle underlying these diverse functions throughout the cell cycle is thought to be anisotropy of the distribution of RanGTP (the RanGTP gradient), driven by the chromatin-associated guanine nucleotide exchange factor RCC1 (14).

ARHI is a maternally imprinted tumor suppressor gene that encodes a 26 kD protein with 55-62% homology to Ras and Rap (15). In contrast to Ras, ARHI contains a 34 amino acid N-terminal extension and inhibits growth, motility and invasion of cancer cells (15, 16). Our previous study showed that ARHI can interact with transcription activator Stat3 and inhibit its nuclear translocation and transcription activity in human breast and ovarian cancer cells (17). Deletion of the N-terminus of ARHI (NTD) significantly compromised its inhibitory activity, suggesting that this unique extension contributes to ARHI's inhibitory effects on Stat3-mediated transcriptional activities (17). To identify additional ARHI-interacting proteins, we have performed proteomic analysis and found that ARHI is complexed with several importin proteins. We have explored the possibility that ARHI might displace cargo proteins, including Stat3, and inhibit their nuclear localization.

RESULTS

ARHI interacts with importins. Our previous studies have shown that, when ARHI and Stat3 were both expressed in SKOV3 cells, ARHI formed a complex with Stat3 in the cytoplasm and prevented interleukin-6–induced Stat3 accumulation in the nucleus (17). To identify the protein(s) interacting with ARHI in the nuclear import process, we performed proteomic analysis. Wild-type ARHI and its NTD mutant were expressed in SKBr3 breast cancer cells with a dual adenovirus system (18). Cells infected with the LacZ adenovirus served as a control. ARHI- or NTD-protein complexes were immunoprecipitated using a specific ARHI antibody and co-immunoprecipitated proteins were analyzed by mass spectrometry. Several novel interacting proteins were identified in complexes with wild type ARHI that bound less intensely to the NTD ARHI mutant. Three of these proteins (Fig. 1A) belong to the family of importins (importin 7, importin 9, importin α re-exporter). These results were further confirmed by co-immunoprecipitation assays. As shown in Figure 1B and 1C, in SKBr3 and HeLa cells, ARHI antibody could immunoprecipitate importin 7, 9 and β 1, and importin 7 antibody could also immunoprecipitate ARHI (Fig. 1D). NTD:importin binding was much reduced compared to wild-type ARHI:importin interaction.

ARHI can bind to multiple importins through the NLS domain. To investigate the ARHI:importin interaction, 10 GST- importins fusion proteins (importin α 1, α 1 mutant, α 3, α 5, α 6, α 7, importin β 1, and importin 7, 8, and 13) were purified. GST protein was used as a negative control. ARHI bound strongly to most of importins in GST pull-down assays. Binding of ARHI to importin α 1 was stronger than binding of ARHI to an importin α 1 mutant that contained an altered nuclear localization signal (NLS) domain (Fig. 2A).

N-terminal of ARHI may mediate the binding with importins. Our previous studies demonstrated that deletion of the unique 34 amino acid N-terminal extension of ARHI nearly abolished its inhibitory effect on cancer cell growth (16) and its ability to block the DNA binding activity of Stat3 (17). Thus, the ARHI N-terminal extension could be important for its biological function. In the proteomic analysis and immunoprecipitation assays (Fig. 1), NTD protein exhibited much less binding to importins. To test further the possible role of the N-terminus of ARHI in binding importins, equal amounts of ARHI complexes and NTD complexes were tested in GST-pull down assays. As presented in Figure 2B, wild type ARHI bound more strongly to most importins than did the truncated NTD mutant, consistent with the possibility that the 34 amino acid N-terminal extension of ARHI mediates ARHI-importin binding.

ARHI can block protein nuclear localization signal. Recent studies have shown that interaction of importins with the Ran protein is required to facilitate transport of cargo proteins into the nucleus (13). Since ARHI and Ran belong to the same family of small G-proteins, ARHI therefore may antagonize importins' interaction with Ran. To test this hypothesis, we have purified ARHI and NTD proteins and assessed their effects on nuclear transport of cargo proteins in HeLa cells. In nuclear import assays, an NLS-GFP fusion protein could be imported into the nucleus in association with importin α 7, importin β 1 and RanGDP (Fig. 3), whereas the BIB (beta-like import receptor binding)-GFP fusion protein only needed help from importin 7 and RanGDP for nuclear localization (Fig. 4). ARHI protein blocked the interaction of NLS with Ran-importin β and α complexes, and also the interaction of BIB with Ran-importin 7

complexes, preventing their nuclear translocation (Figs. 3, 4). In comparison, ARHI-NTD protein did not block these interactions or nuclear transport (Figs. 3, 4).

ARHI can block Stat3 protein nuclear localization signal. Using a similar GST pull-down assay, Stat3 protein was shown to bind importins $\beta 1$, $\alpha 1$, $\alpha 3$, $\alpha 6$, and $\alpha 7$, but not the other importins (Fig. 5A). After IL-6 stimulation, the level of pStat3 greatly increased. Importins bound both to phosphorylated Stat3 (after IL-6 stimulation) and non-phosphorylated Stat3 (no IL-6 stimulation), but importin binding to pStat3 was much stronger. To investigate if ARHI could block Stat3 protein nuclear translocation, we have produced a phosphorylated Stat3 β protein (residues 127-722), which contained the binding site to importins. This protein was labeled with FITC and evaluated in the nuclear import assay. As shown in Figure 5B, pStat3 β protein alone was concentrated at nuclear pores; when importins and Ran were added to the system, pSTAT3 β was distributed in the nucleus. Addition of purified ARHI protein blocked Ran-dependent pSTAT3 β nuclear translocation.

ARHI can compete for importin-Ran binding. ARHI (DIRAS3) and Ran share a 22% sequence identity and are expected to have a similar overall structure. Although the crystal structure of ARHI is not available, those of the closest neighbors in the Ras family, DIRAS1 and DIRAS2, are. We obtained a homology model for ARHI via the SWISS-MODEL server (<http://swissmodel.expasy.org/>). The model was generated using the DIRAS2 structure (PDB code: 2ERX) as the template and it covers residues 37 to 206 of the protein's 229 residues. As shown in Figure 6A, when the ARHI model is superimposed onto the crystal structure of Ran

(PDB code: 1IRB, chain A) (26), the structural conservancy is clear. A DaliLite (<http://www.ebi.ac.uk/DaliLite/>) pairwise comparison resulted in a Z-score of 23.4 and C α RMSD of 2.4 Å, over 165 aligned residues. In the crystal structure of the importin β :Ran complex (26), Ran utilizes an extensive surface in binding to importin β (Fig. 6B). To determine whether ARHI binds to the same region of importin β as Ran and could compete for importin-Ran binding, we have performed GST-pull down assays and found Ran strongly bound to both importin β 1 and 13, ARHI protein could reduce at least 50% of these binding ability (Fig. 6C), but NTD protein could not.

DISCUSSION

Stat3, a latent transcription factor, transduces signals from the cell surface to the nucleus and activates gene transcription, triggering proliferation, resistance to apoptosis, motility, invasion and angiogenesis (21, 22). Recent studies clearly demonstrate that Stat3 is required for both tumor initiation and promotion (21). Stat3 is frequently phosphorylated, activated in the majority of breast and ovarian cancers (23), where cytokines and growth factors such as IL-6 bind to specific receptors and activate JAK2 which, in turn, phosphorylates Stat3 and prompts translocation of pStat3 to the nucleus. The classic NLS-importin pathway has been reported to mediate the nuclear translocation of Stat3. Liu L et al. reported that the Stat3 nuclear import is mediated by importin- α 3 (24), whereas other studies have shown that the regulation of Stat3 nuclear import is through importins α 5 and α 7 (25). Few studies have considered factors which regulate translocation of pStat3.

From our previous studies, re-expression of the putative tumor suppressor gene ARHI in cancer cells markedly inhibited binding of pStat3 to Stat response elements in target gene promoters and downregulated Stat3-dependent promoter activity without significantly affecting Stat3 phosphorylation (17). When ARHI and Stat3 were co-expressed in SKOV3 ovarian cancer cells, ARHI formed a complex in the cytoplasm with Stat3 and prevented IL-6-induced Stat3 translocation to the nucleus (17). Our present study has elucidated the mechanism by which ARHI prevents nuclear translocation of pStat3. Nuclear import assays have shown that pStat3 can be translocated to the nucleus in the presence of Ran and importins (Fig 5B), whereas non-phosphorylated Stat3 could not (data not shown). ARHI protein bound to importins (Figs. 1-2) and blocked nuclear translocation of pStat3 (Fig. 5B). ARHI also blocked nuclear translocation of NLS-GFP and BIB-GFP in the presence of Ran and appropriate importins. Thus, ARHI might affect nuclear localization of Stat3, but might also affect transport of other proteins that are required for oncogenesis.

The precise mechanism by which ARHI disrupts the interaction of pStat3, Ran and importins remains to be elucidated. ARHI is a member of Ras superfamily, but contains a unique 34 residue extension at the N-terminus. Like Ras, ARHI can bind to GTP with high affinity, but has low intrinsic GTPase activity (16). ARHI associates with the cell membrane after prenylation at the C-terminal cysteine residue. Mutation of the conserved CAAX box at the C-terminus led to a loss of its membrane association ability and a modest decrease in its ability to inhibit cell growth (16). Most strikingly, deletion of ARHI's unique N-terminal extension nearly abolishes its inhibitory effect on cell growth (16, 17). In the present study, the NTD mutant lost much of the ability to bind importins and did not block the nuclear translocation of

NLS-GFP or BIB-GFP. This suggests that interaction of the N-terminal extension of ARHI to importin may be required to prevent effective interaction of Ran-importin complexes with NLS and BIB. Interestingly, C-terminal deletion mutants of ARHI localize in the nucleus (data not shown), raising the possibility that accumulation of pStat3 in the cytoplasm in the presence of wild type ARHI protein could result from direct binding of ARHI to Stat3, which we have demonstrated (17), or to lack of an effective nuclear transport mechanism. As wild type ARHI exhibits a nuclear localization signal on its N-terminal extension, the ARHI might compete with cargo for binding to importins in the presence of Ran. Once bound, ARHI could trap importins in the cytoplasm as the small G protein is tethered to cell membranes through its prenylated C-terminus. Our present study showed that the structural conservancy between ARHI and Ran is clear (Fig. 6A), ARHI protein could compete for at least 50% of importin-Ran binding ability (Fig. 6C), suggesting that ARHI binds to the same region of importin β as Ran does and ARHI-induced disruption of Ran-importin binding could serve as an important regulatory mechanism that contributes to the tumor suppressor function of ARHI.

MATERIALS AND METHODS

Cell lines and reagents. SKBr3 human breast cancer cell line was maintained as described (16). HeLa cells were grown at 37°C to near confluence in Dulbecco's Modified Eagle's Medium containing 10% fetal calf serum. Constructs of Ran and importin α 1 mutant, as well as anti-importin β 1 antibody were provided by Dr. Karsten Weis (University of California Berkeley). Constructs of GST, GST-importin 7, GST-importin 8, GST-GFP, GST-GFP-BIB, GST-GFP-NLS were provided by Dr. Keith Yamamoto (University of California San Francisco). Anti-importin 7 and anti-importin 9 antibodies were obtained from Dr. Dirk Görlich (University of Heidelberg,

Germany). Anti-Ran and anti-GST antibodies were purchased from Upstate Biotechnology Inc. (Billerica, MA). Anti-ARHI antibody was produced by our group as previously described (16). Adenovirus constructs expressing LacZ, ARHI and NTD were prepared (18). The reagents for a nuclear import assay (digitonin, ATP, GTP, creatine phosphate, creatine kinase) were purchased from Sigma (St. Louis, MO).

Proteomics analysis. Wild-type ARHI and its NTD mutant were re-expressed in the breast cancer cell line SKBr3 with a dual adenovirus system (18). LacZ adenovirus-infected cells were used as a control. ARHI interacting proteins were immunoprecipitated using an ARHI monoclonal antibody and further purified using Protein G beads (Pierce, Rockford, IL). Proteins were separated on a 15% SDS-PAGE gel and analyzed by mass spectrometry at the University of Texas M.D. Anderson Cancer Center Proteomics Facility.

Western blot analysis. Confluent SKBr3 cells were lysed and the cell lysates were clarified by centrifugation for 5 min at 14,000 rpm at room temperature. Equal amounts of total cellular proteins were electrophoresed in 15 % SDS/PAGE and transferred to polyvinylidene fluoride membrane (Millipore, Billerica, MA). The membrane was immunoblotted with antibodies and signals developed by using an ECL system (Amersham, Piscataway, NJ).

Fusion protein expression and purification. GST, GST-importin α 1, α 3, α 5, α 6, α 7, GST-importin α 1 mutant, GST-importin β 1, 7, 8, and 13 were all expressed in BL21 (DE3) *Escherichia coli* (Stratagene). Overnight bacterial cultures in LB/Ampicillin were diluted 1/100

and grown to an OD_{595} of 0.6 to 0.8 at 37°C. Cultures were cooled, induced with 0.25 mM IPTG overnight at 16°C and harvested by centrifugation. Cell pellets were re-suspended in lysis buffer (50 mM Tris, pH 8.0, 150 mM NaCl, 1 mM EDTA, 1 mM DTT, 1 µg/ml leupeptin, 1 µg/ml pepstatin, 1 µg/ml aprotinin, and 1 mM PMSF), sonicated, and clarified at 40,000 x *g* for 30 min at 4°C. After centrifugation, the supernatant fraction was further purified on an AKTA FPLC system using a glutathione sepharose column and a Superdex 200 gel filtration column. Tyrosine phosphorylated Stat3 β (residues 127-722) (pStat3 β) was purified as published (19), briefly, the core fragment of STAT3 β was co-expressed with the Elk kinase in *E. coli*. The identity of the phosphoprotein was confirmed by using mass spectrometry.

The ARHI and NTD cDNA sequences were PCR amplified from pcDNA3-ARHI plasmid DNA and cloned into the pQE30 vector. The importin 7 coding sequence was retrieved from the GST-importin 7 plasmid by restriction digestion and inserted into the pQE30 vector. For the nuclear import substrates, GST-GFP-BIB and GST-GFP-NLS proteins were purified by glutathione sepharose beads (Amersham) according the manufacturer's instructions. pQE-Ran, pQE-importin 7, pQE-ARHI, and pQE-NTD were expressed in SG13009 competent cells and purified according to Nachury and Weis (20). Importin β 1 was cleaved from GST- β 1 fusion by PreScission protease (GE Healthcare, Piscataway, NJ) and importin α 7 was cleaved from GST- α 7 fusion using TEV protease (Invitrogen, Carlsbad, CA), and the mixture was loaded on a HiTrap Q cartridge (Amersham) to separate GST from the importin proteins. All fusion proteins were concentrated using Centricon 30 concentrators, aliquoted, flash frozen in liquid nitrogen, and stored at -80 °C.

Labelling of pStat3 β . pStat3 β proteins were FITC-labeled using a FITC Protein Labeling kit (Pierce) according manufacturer's instructions.

GST pull-down assay. GST-importin fusion proteins (2-4 μ g) were mixed with 10-30 μ g of cell lysates from SKBr3 cells in binding buffer (20 mM Hepes, pH 7.9, 150 mM NaCl, 0.5 mM EDTA, 10 % Glycerol, 0.1% Triton X-100, 1 mM DTT). The mixture was incubated with shaking at 4°C for 1 h. Fifteen microliters of glutathione-sepharose beads, pre-equilibrated in TEE buffer (50 mM Tris, pH 7.9, 1 mM EDTA, 1 mM EGTA) were added to the mixture and incubated with shaking at 4°C for another 1 h. The beads were collected and washed 4-5 times with Washing buffer (20 mM Hepes, pH 7.5, 100 mM KCl, 2.5 mM MgCl₂, 1% glycerol, 1% Triton X-100) and once with ice-cold PBS. The beads were resuspended in 20 μ l of 2X SDS loading buffer and then boiled for 5 min. The soluble proteins were separated on 15% SDS-PAGE gels for Coomassie blue staining or western-blotting analysis.

Nuclear import assay. HeLa cells were grown on glass cover slips for 40-42 h. Cells were then washed twice with ice-cold Transport buffer (20 mM HEPES, pH 7.3, 110 mM KOAc, 5 mM NaOAc, 2 mM MgOAc, 2 mM DTT, 1 mM EGTA), incubated with 40 μ g/ml digitonin in Transport buffer for 6 min on ice, washed twice in cold Transport buffer, and kept on ice for 20 min. Cover slips containing permeabilized cells were incubated for 30 min at 30°C with a 30 μ l nuclear import reaction mixture [2 μ M GFP-tagged import substrate or 4 μ M FITC labeled p-Stat3 β and an ATP-regenerating system (1 mM ATP, 5 mM creatine phosphate, 10 U/ml creatine kinase, 0.5 mM GTP)]. Some experiments were complemented with 3 μ M RanGDP, 3

μM importin $\alpha 7$, 2 μM importin $\beta 1$, 3 μM ARHI or 3 μM ARHI-NTD. After the import reactions were completed, the cells were rinsed once in PBS, fixed in 4% paraformaldehyde in PBS, and analyzed by fluorescence microscopy.

ACKNOWLEDGEMENTS

This work has been supported by grants from the National Cancer Institute, CA 80957 (Y.Yu) and CA 64602 (R.C.Bast), and from the National Foundation for Cancer Research LF 2004-00009224HM (R.C.Bast).

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TITLES AND LEGENDS TO FIGURES:

Figure 1. ARHI-importin form complexes. (A) Proteomic analysis. Wild-type ARHI and its NTD mutant were expressed in the SKBr3 breast cancer cells using a dual adenovirus system. LacZ adenovirus infected cells served as the control. A: ARHI complexes; B: NTD complexes; C: LacZ control. Arrows point to protein bands corresponding to the indicated novel binding proteins. (B-D) Immunoprecipitation of ARHI-importin complexes. Protein lysates from (B) SKBr3 or (C) HeLa cells infected with ARHI, NTD, or LacZ adenoviruses were incubated with anti-ARHI antibody. The immunoprecipitates were analyzed by Western blotting with anti-importin (Imp) 7, 9, β 1 antibodies. Mouse IgG was used as control. (D) Proteins immunoprecipitated with anti-importin 7 antibodies were analyzed by Western analysis with anti-ARHI antibody.

Figure 2. ARHI-importin binding analyzed by GST pull-down assays. (A) SKBr3 cells were infected with ARHI adenovirus and lysed. Ten GST-importin (Imp) fusion proteins and ARHI antibody were used to detect ARHI-importin binding. GST protein served as a control. Loading was measured with anti-GST antibody. (B) Wild type ARHI binds more strongly to most importins than does the NTD mutant. Equal amounts of ARHI or NTD protein lysates (induced by adenovirus in SKBr3 cells) were used in GST-pull down assays. Nine GST fusion proteins and ARHI antibody were used to detect the binding. GST protein did not bind to both ARHI and NTD.

Figure 3. Nuclear import assay for NLS-GFP. NLS-GFP protein was mixed with RanGDP,

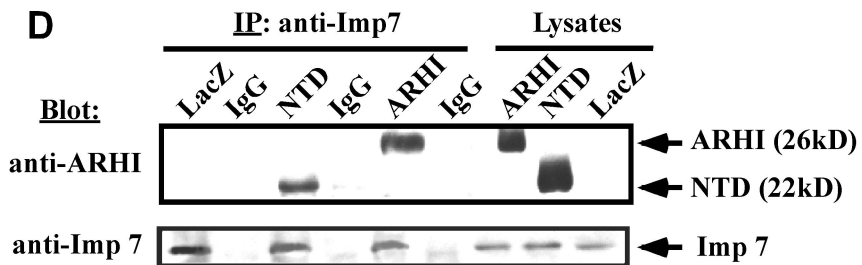
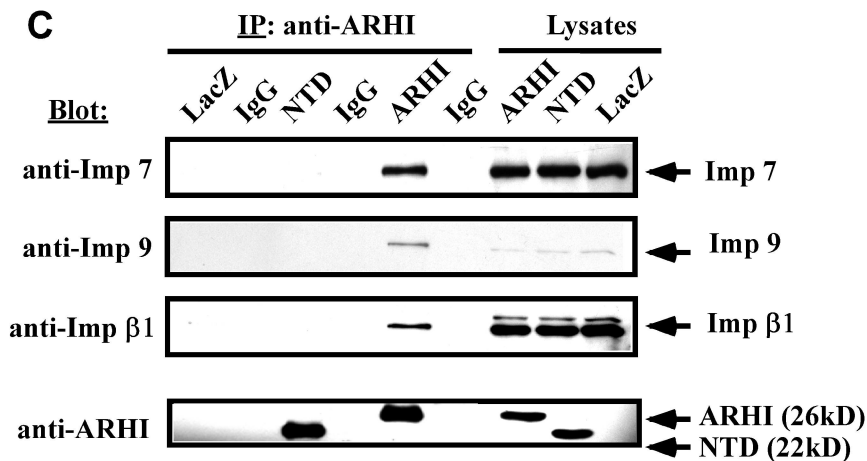
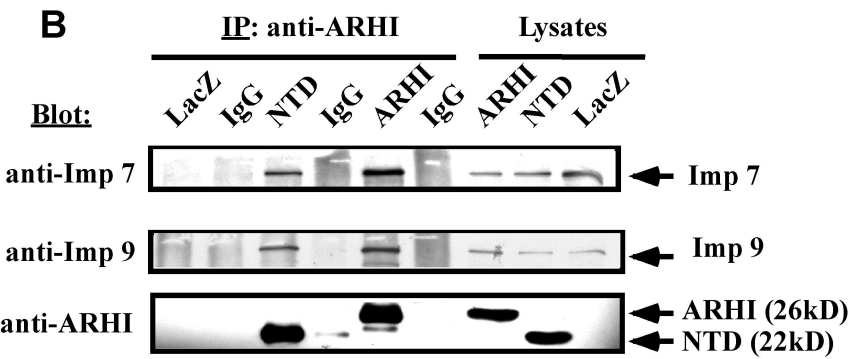
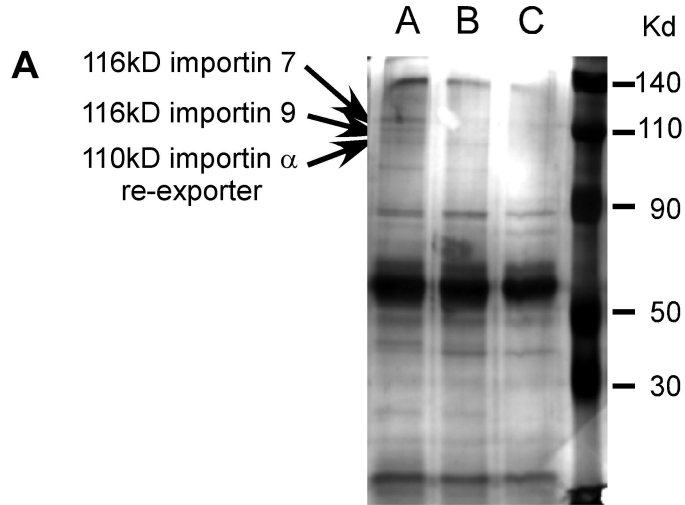
importin (Imp) $\alpha 7$, importin $\beta 1$, and incubated with permeabilized HeLa cells. Import assays were also performed with ARHI and NTD proteins.

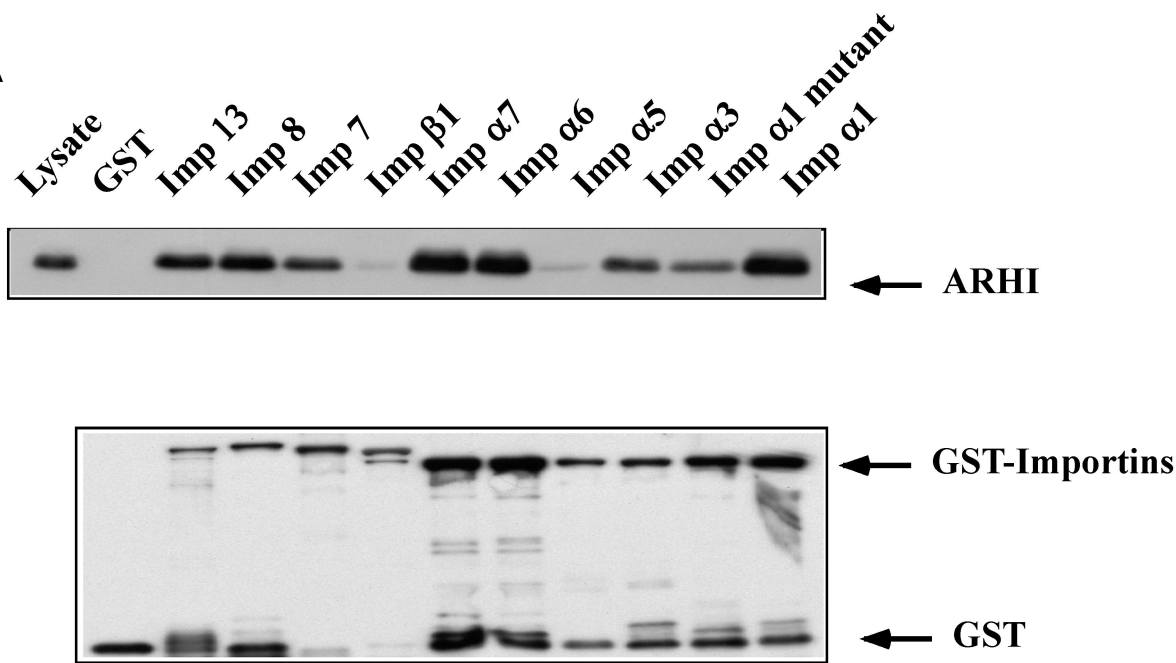
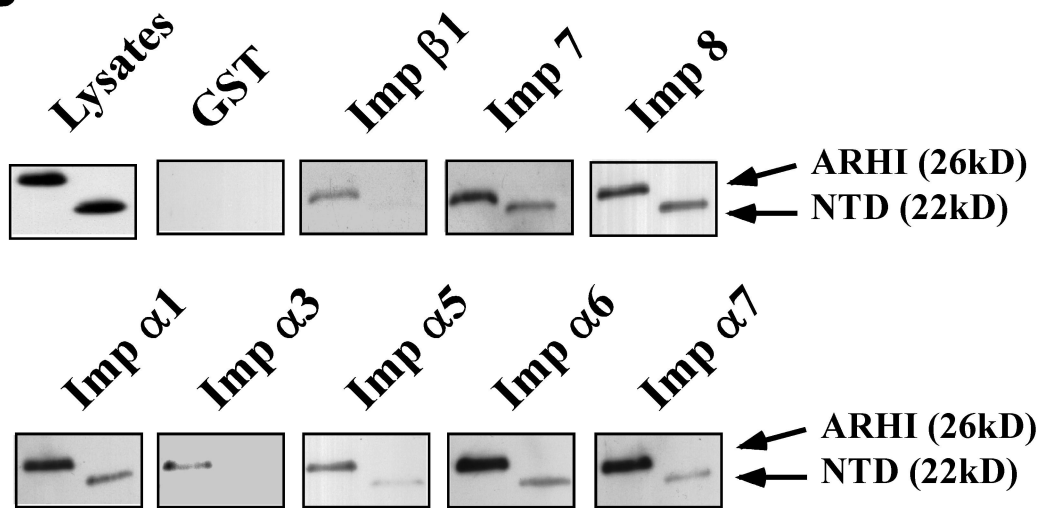
Figure 4. Nuclear import assay for BIB-GFP. BIB NLS-GFP protein was mixed with RanGDP, importin (Imp) $\alpha 7$ and incubated with permeabilized HeLa cells. Import assays were also performed with ARHI and NTD proteins.

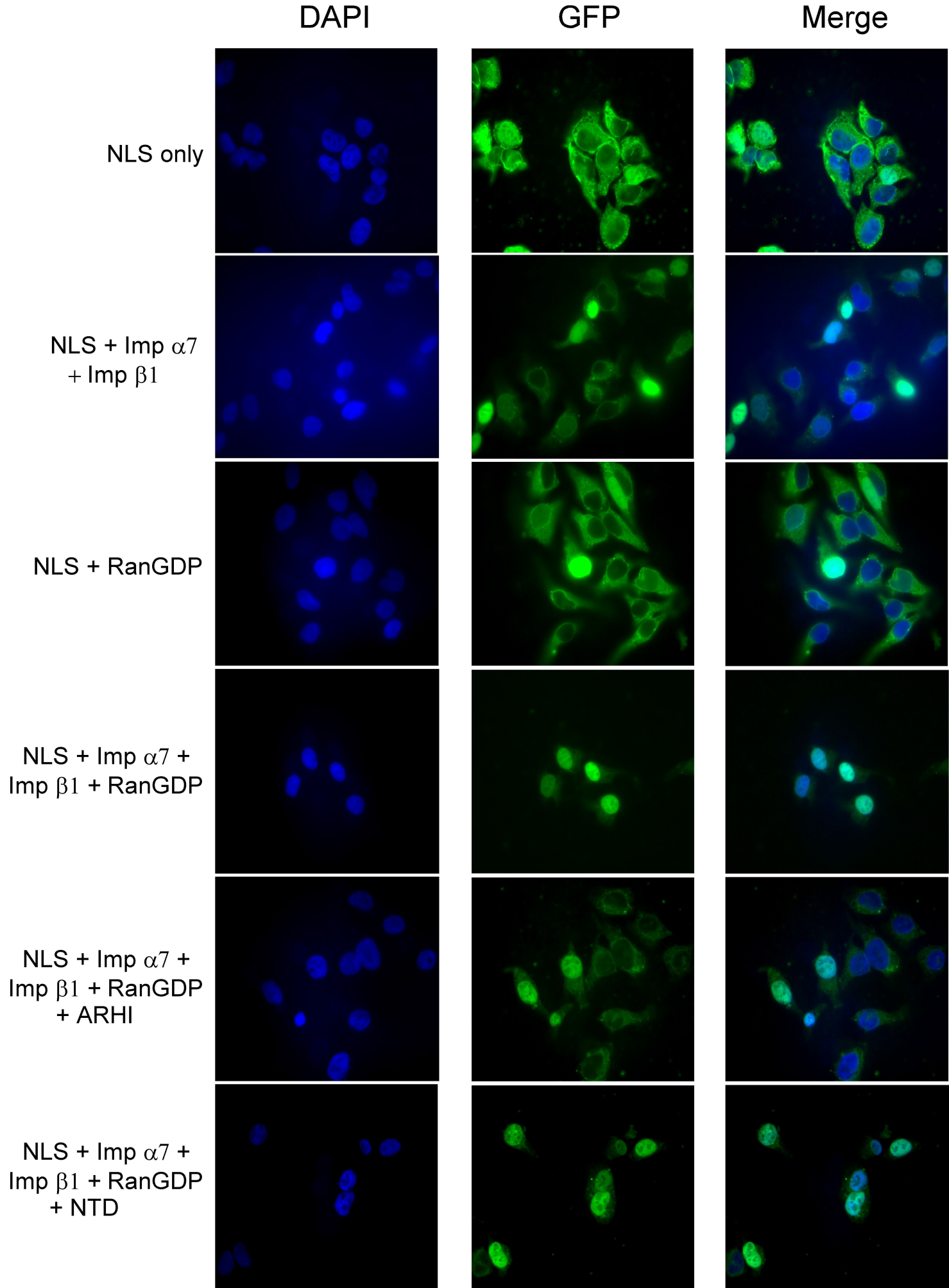
Figure 5. ARHI can block pStat3 β nuclear translocation. (A) Stat3-importin binding analyzed by GST-pull down assays. SKBr3 cells were treated with or without IL-6 (10 ng/mL) for 30 minutes before cells were harvested for cell lysate preparation. Ten GST-importin (Imp) fusion proteins and Stat3/pStat3 antibodies were used to detect binding between Stat3 and importins. GST protein was used as a control. (B) Nuclear import assay for pStat3 β -GFP. Purified pStat3 β and RanGDP, importin $\alpha 7$, and importin $\beta 1$ were incubated with permeabilized HeLa cells. Import assays were also performed with ARHI protein.

Figure 6. (A) Superimposition of the ARHI structural model (homology modeling, in red) and the Ran crystal structure (PDB code: 1IBR, chain A, in green). (B) A semi-transparent surface diagram of Ran (in the same orientation as in A), showing the side-chains of the residues (in red) involved in binding to importin β (26). Both diagrams were made using PyMOL (<http://pymol.sourceforge.net/>). (C) ARHI competes for importin-Ran binding. GST or GST-RanQ69L fusion proteins were bound to glutathione-Sepharose 4B, then incubated with Importin 13 (Imp 13)-His/Importin $\beta 1$ (Imp $\beta 1$)-His at 4°C for 4 h, treated with ARHI-His or

NTD-His at 4°C for 4h, the samples were separated with SDS-PAGE, then blotted with monoclonal antibodies to His or polyclonal antibody to importin 13.



A**B**

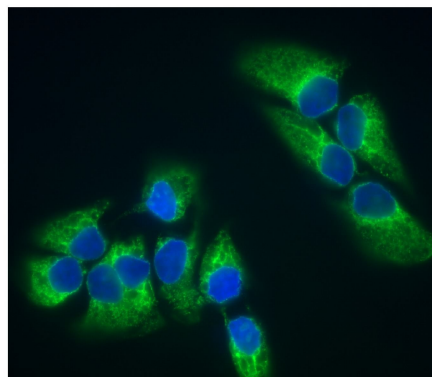
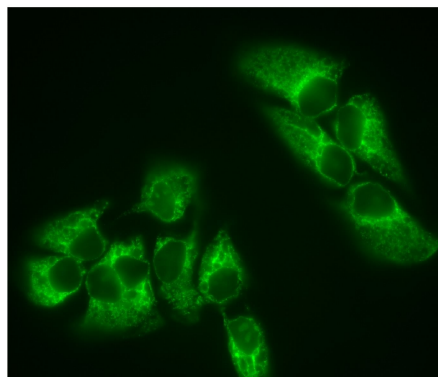


DAPI

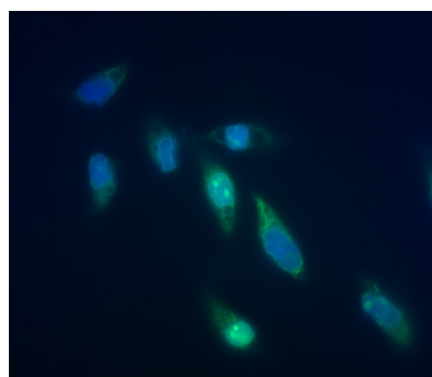
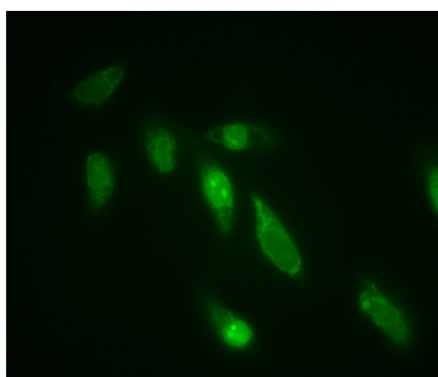
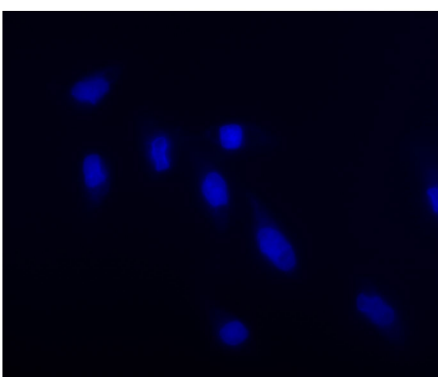
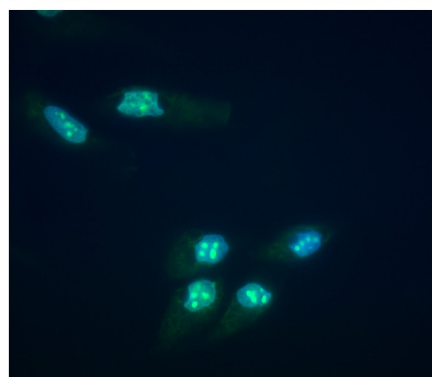
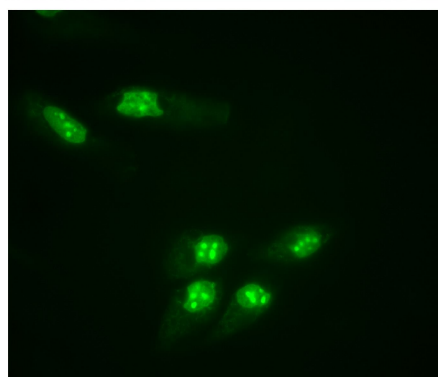
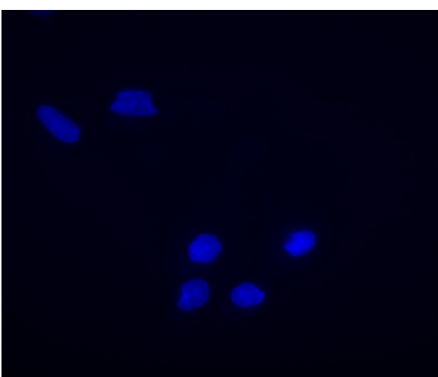
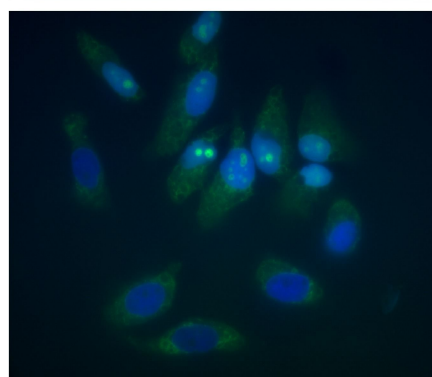
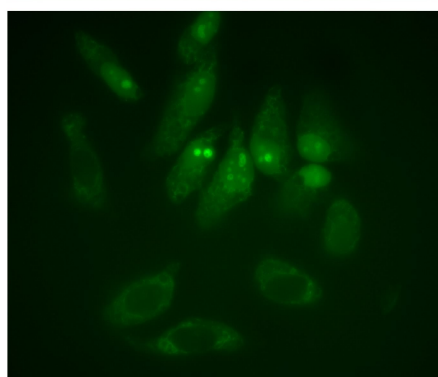
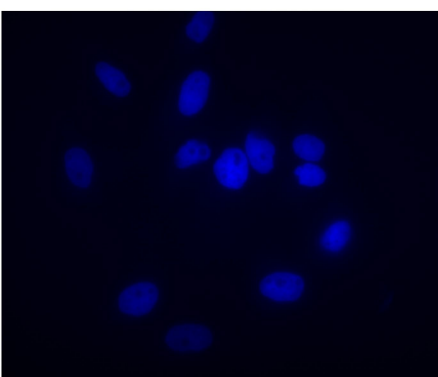
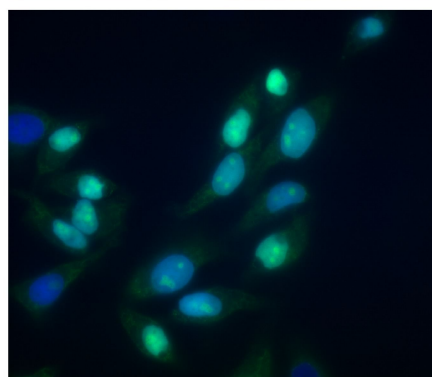
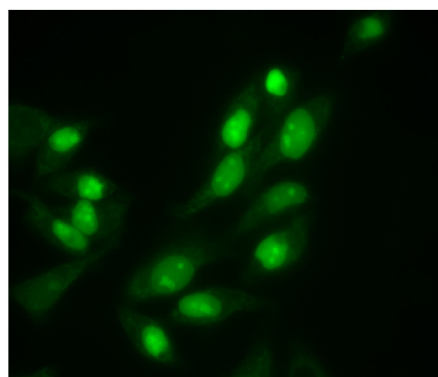
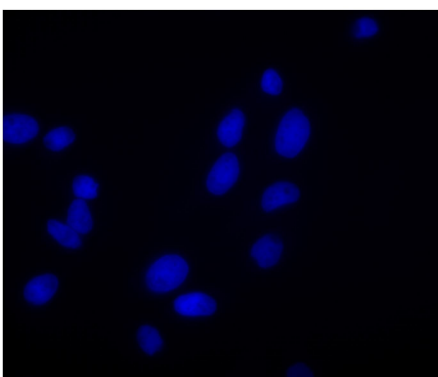
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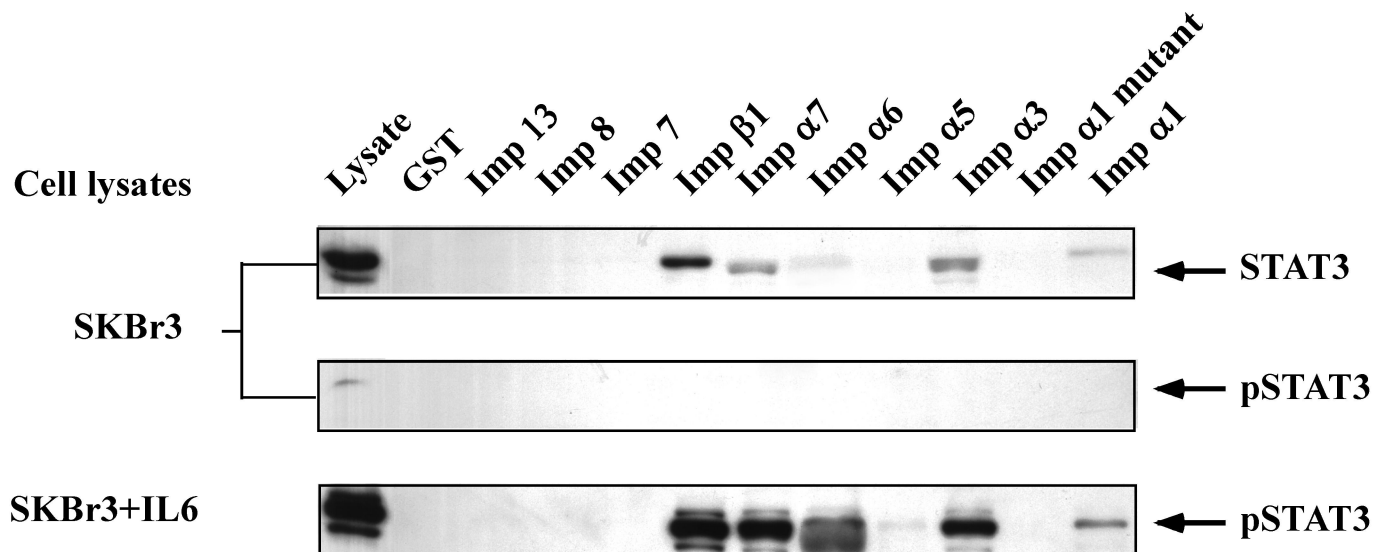
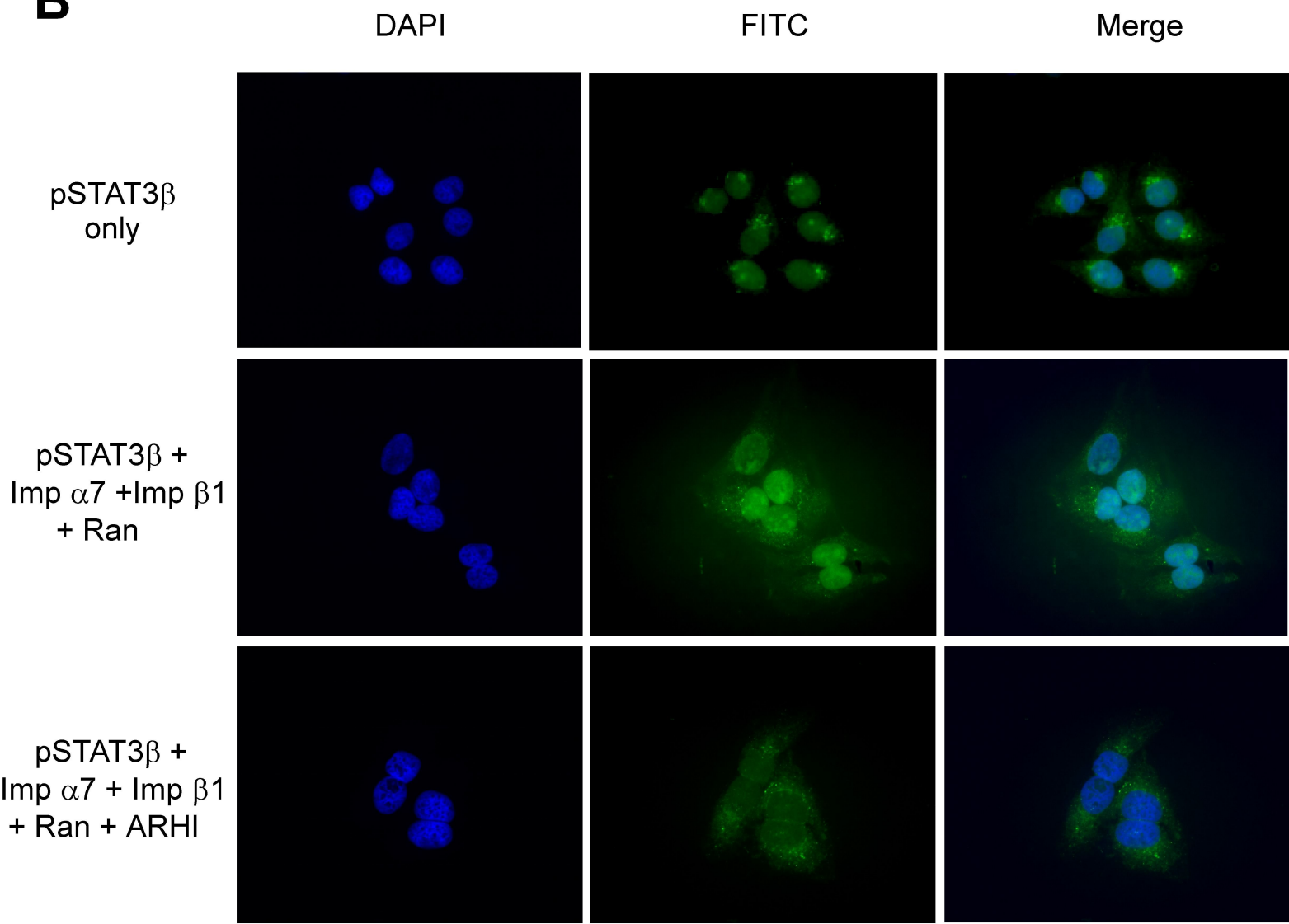
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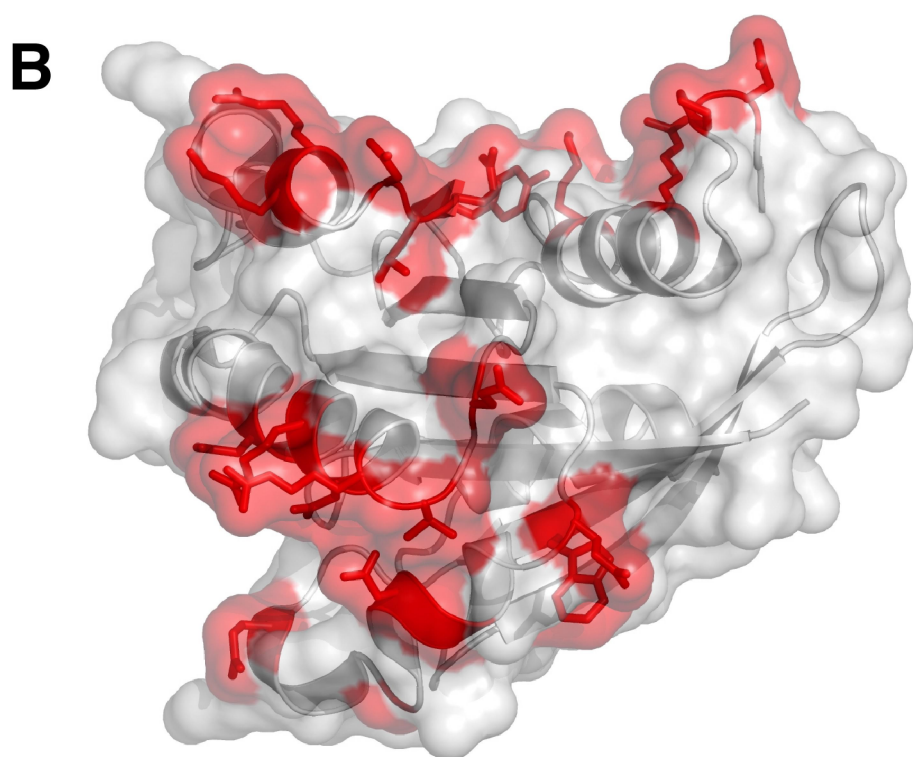
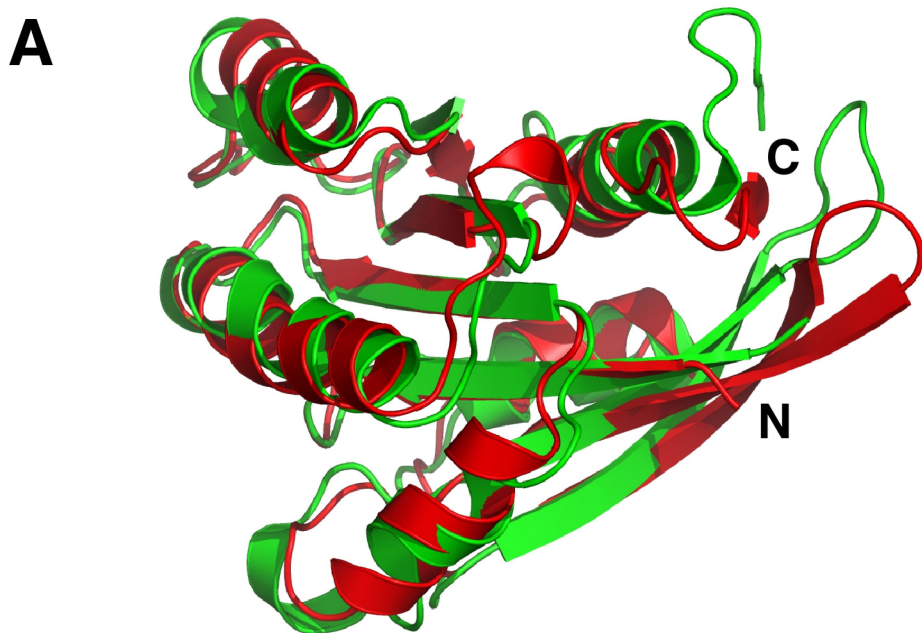
BIB only



BIB + Ran

BIB + Ran
+ Imp 7BIB + Ran
+ Imp 7
+ ARHIBIB + Ran
+ Imp 7
+ NTD

A**B**



C

