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2 **Are plants with anti-cancer activity resistant to crown gall? : A**  
3 **test of hypothesis**

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18 **Abstract**

19 The Crown gall tumour assay (CGTA) is one of several bench top bioassays recommended  
20 for the rapid screening of plants with anti-cancer activity. The rationale for the use of the  
21 bioassay is that the tumorigenic mechanism initiated in plant tissues by *Agrobacterium*  
22 *tumefaciens* is in many ways similar to that of animals. Several plant species with anti-cancer  
23 activity have already been discovered using this bioassay. However till date no explicit test  
24 of an association between anti-cancer activity of plants and their resistance to crown gall  
25 formation has been demonstrated. Demonstration of an association could have exploratory  
26 potential when searching for plants with anti-cancer activity. In this paper, we determined  
27 whether or not a statistically significant association between crown gall resistance and anti-  
28 cancer activity exists in plants found in existing published data sets. Our results indicate  
29 that plants with anti-cancer activity have a higher proportion of their species resistant to  
30 crown gall formation compared to a random selection of plants. We discuss the implications  
31 of our results especially when prospecting for newer sources of anti-cancer activity in plants.

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34 **Key words:** anti-cancer activity, crown gall resistance, NCI

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## 42 **Introduction**

43 Bioprospecting for plants with anticancer activity has been a major focus in the search for  
44 plant-based cures [1]. The National Cancer Institute (NCI) alone has reportedly screened  
45 over 35,000 plant species for anti-cancer activity [2], [3], [4], [5], [6]. Two of the three most  
46 important anti-cancer compounds available today, namely taxol and camptothecin, were the  
47 result of this endeavor [7], [8],[9], [10].The screening of a large number of plants, involving  
48 over half-a-dozen solvent extraction systems, and testing them on dozens of cancer cell  
49 lines, has often been time-consuming [5].

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51 In an effort to minimize the screening process and hasten the pace of drug discovery, the  
52 NCI developed a number of rapid bench top assays to short-list potential plants, which then  
53 could be targeted for more advanced screening [11]. One of those bench top bioassays was  
54 the crown gall tumour assay (CGTA) [12]. Crown gall is a neoplastic plant disease caused by  
55 *Agrobacterium tumefaciens* [13]. Infected plants, exhibit tumorigenic growth symptoms in stem  
56 collars and other parts of the plant. Crown gall is a common disease of dicot plants including  
57 many woody shrubs and various herbaceous plants. In this bioassay, the ability of plant  
58 extracts to inhibit tumours induced by *A. tumefaciens* in model systems such as potato tuber  
59 discs is evaluated [14]. The rationale for employing this bioassay rests on the fact that the  
60 tumorigenic mechanism induced by *A. tumefaciens* in plants is in many ways similar to that of  
61 animals [15], [16]. The use of this bioassay has resulted in many short lists of plants with  
62 anti-cancer activity, and has helped with the discovery of novel compounds from plants [17],  
63 [18], [14], [11], [19]. McLaughlin et al. (1991) indeed were able to show an association  
64 between the inhibition of crown gall formation on potato discs and the *in vivo* 3PS anti-  
65 tumour activity by the plant extracts.

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67 From the above results, it follows that plants intrinsically resistant to crown gall infection  
68 could, in principle at least also be associated with anti-cancer activity. To the best of our  
69 knowledge, there have been no attempts to evaluate this hypothesis. A test of the prediction  
70 and demonstration of an association between crown gall resistance and anti-cancer activity  
71 could have immense exploratory potential in the search for newer plants as sources of anti-  
72 cancer activity. We have examined the association between crown gall resistance and anti-  
73 cancer activity in plants and now report on the results of that study.

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## 75 **Materials and Methods**

76 We compiled a database of 1193 species (comprised of 588 genera and 138 families) of dicot  
77 plants based on their resistance or susceptibility to crown gall infection as reported in Cleene  
78 et al., (1976). Species were assigned a qualitative score of either crown gall resistance (+) or  
79 susceptible (-). For the purpose of this analysis, we used data on only those 1110 species for  
80 which the information was complete.

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82 We then compiled a list of 38 plant species that were reported to possess anti-cancer activity  
83 from a variety of published sources (Plants for future Database, [www.pfaf.org](http://www.pfaf.org) and other  
84 references mentioned in Table 1). All studies sourced here were based on either an *in vitro* or  
85 an *in vivo* assay for anti-cancer activity.

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87 Based on these two datasets, we analyzed the average proportion of species resistant to  
88 crown gall: Using a bootstrap analysis involving repeated sampling with replacement  
89 (PopTools version 2.6.2); [20], we randomly selected 100 species from the database and

90 determined the proportion of species resistant to crown gall. The process was repeated 100  
91 times. For each of the repeats, we computed the proportion of species resistant to crown  
92 gall. A frequency distribution of the proportion of species resistant to crown gall was then  
93 plotted and the overall mean proportion of species resistant along with the standard  
94 deviation was computed. The proportion of species resistant to crown gall from among the  
95 38 species reported to possess anti-cancer activity was calculated. For each of the 38 plants,  
96 we inferred their resistance or otherwise to crown gall from the database assembled from  
97 Cleene et al., (1976) and computed the proportion of species that were resistant.

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99 Finally, we performed a test of significance between the two proportions (for species drawn  
100 randomly *vs* species possessing anti-cancer activity) using a one-tailed student t-test.

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## 102 **Results and discussion**

103 The frequency distribution of proportion of species resistant to crown gall for the randomly  
104 drawn species (N=100 from 1110 species, repeated 100 times) was nearly normally  
105 distributed, with an overall mean proportion of  $0.41 \pm 0.051$  (Figure 1). The proportion of  
106 species resistant to crown gall among plants exhibiting anti-cancer activity (N=38) was 0.81,  
107 which was significantly higher than that of randomly selected plants (one tailed t-test,  
108  $p < 0.001$ ).

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111 Plants with anti-cancer activity therefore appear to have a higher proportion of species  
112 resistant to crown gall than randomly selected species. While the result suggests that an  
113 association between crown gall resistance and anti-cancer activity in plants exists, a more

114 robust demonstration would have been to set up a 2 X 2 matrix of crown gall  
115 (resistance/susceptibility) and anti-cancer activity (present/absent) and then statistically  
116 evaluating the association. Demonstration of such an association may have more accurately  
117 shown if plants with anti-cancer activities are more likely to be found in plants that are  
118 resistant to crown gall compared with a randomly chosen set of species. Unfortunately,  
119 because of a well-recognized positive bias in publications, papers often only report studies  
120 where anti-cancer activity was observed, seldom publishing studies with no activity.  
121 Consequently, a 2 X 2 matrix with data cells corresponding to crown gall  
122 resistance/susceptibility and anti-cancer activity (absence) is deficient thus limiting the  
123 association analysis.

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125 Though not as directly demonstrative as would have been desired, our results nevertheless  
126 provide a useful first step in working towards a more robust test of the association. Results  
127 of further analysis could pave the way for the development of algorithms that make the  
128 search for anti-cancer activity in plants in a more directed manner.

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278 **Figure and table legends**

279 Figure 1: Frequency distribution of proportion of species resistant to crown gall formation in  
280 randomly selected plants (see text for details). The mean proportion of species resistant to  
281 crown gall for a random collection of plants and that for plants with anti-cancer activity is  
282 also indicated.

283 Table 1: List of plant species with anti-cancer activity along with information on crown gall  
284 resistance or susceptibility.

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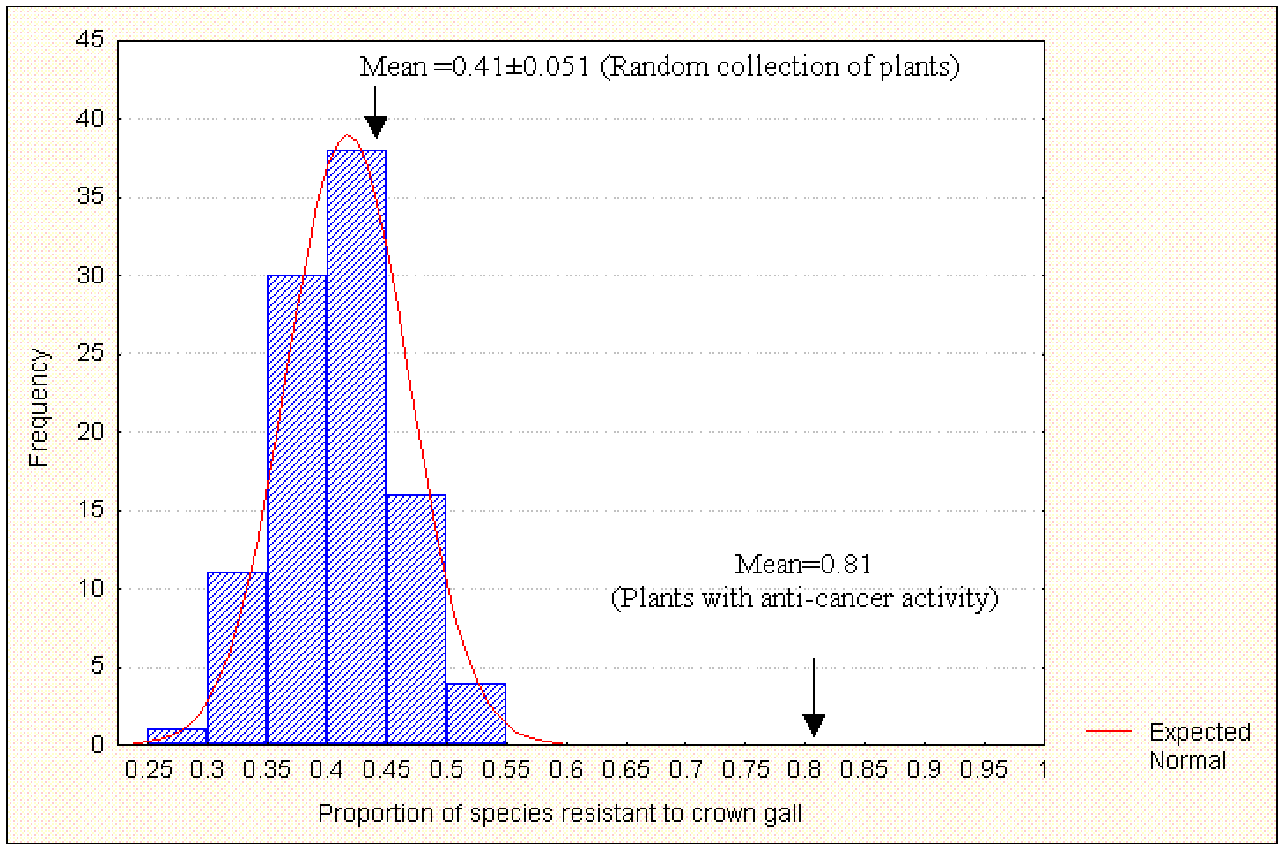


Figure 1: Frequency distribution of proportion of species resistant to crown gall formation in randomly selected plants (see text for details). The mean proportion of species resistant to crown gall for a random collection of plants and that for plants with anti-cancer activity is also indicated.

341 Table 1: List of plant species with anti-cancer activity along with information on crown gall resistance or susceptibility.  
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| Sl no | Common name      | Scientific name                | Family        | Anti-cancer activity |  | Crown gall resistance or susceptibility (inferred from Cleene et al., 1976) |
|-------|------------------|--------------------------------|---------------|----------------------|--|---|
|       |                  |                                |               | Reference            | In vivo or in vitro anticancer assay   |   |
| 1     | Aster            | <i>Aster sp</i>                | Compositae    | [21]                 | Epifriedelinol shows anti-cancer activity  | Resistant   |
| 2     | Birch            | <i>Betula alleghaniensis</i>   | Betulaceae    | [22]                 | Induces apoptosis in human melanoma and neuroblastoma cells                            | Resistant   |
| 3     | Blueberry        | <i>Vaccinium sps</i>           | Ericaceae     | [23]                 | Induces Phase-II Xenobiotic detoxification enzymes                                     | Resistant   |
| 4     | Cactus           | <i>Opuntia microdasys</i>      | Cactaceae     | [24]                 | Induces apoptosis and cell cycle arrest of cancer cells.                               | Resistant   |
| 5     | Dahlia           | <i>Dahlia rosea</i>            | Compositae    | [25]                 |  | Resistant   |
| 6     | Gossypium        | <i>Gossypium</i>               | Malvaceae     | [26]                 | Cytotoxic to murine B16 melanoma and L1210 lymphoma cells                              | Resistant   |
| 7     | Hydrangea        | <i>Hydrangea serrata</i>       | Hydrangeaceae | [25]                 |  | Resistant   |
| 8     | Maple            | <i>Acer sp</i>                 | Sapindaceae   | [27]                 | Possesses activity against Walker 256 and Sarcoma 180 cell lines                       | Resistant   |
| 9     | Rhododendron     | <i>Rhododendron indicum</i>    | Ericaceae     | [28]                 | Cytotoxic against Spodoptera frugiperda cell line Sf-9                                 | Resistant   |
| 10    | Sequoia          | <i>Sequoia sempervirens</i>    | Taxodiaceae   | [29]                 | Shows Brine Shrimp Lethality   | Resistant   |
| 11    | Spruce           | <i>Picea sps</i>               | Pinaceae      | [30]                 | Inhibits growth of LNCaP tumors in Mice.   | Resistant   |
| 12    | Maidenhair tree  | <i>Ginkgo biloba</i>           | Ginkgoaceae   | [31]                 | Inhibits DNA damage  | Resistant   |
| 13    | Golden-rain tree | <i>Koelreuteria paniculata</i> | Sapindaceae   | [32]                 | Tyrosine kinase inhibition   | Resistant   |
| 14    | Holly            | <i>Ilex aquifolium</i>         | Aquifoliaceae | [33]                 | Ursolic Acid Inhibits Cyclooxygenase-2 Transcription in Human Mammary Epithelial Cells | Resistant   |
| 15    | Hornbeam         | <i>Carpinus betulus</i>        | Betulaceae    | [34]                 | Active against human melanoma cells  | Resistant   |
| 16    | Serviceberry     | <i>Amelanchier spp.</i>        | Rosaceae      | [35]                 | Berry extract induces cell-cycle arrest.   | Resistant   |
| 17    | Barberry         | <i>Berberis vulgaris</i>       | Berberidaceae | [36]                 | Berberine affects the structure of filamentous actin cytoskeleton of the B16 cells.    | Resistant   |

|    |                 |                                |                |            |   |            |
|----|-----------------|--------------------------------|----------------|------------|---|------------|
| 18 | Mahonia         | <i>Mabonia fremontii</i>       | Berberidaceae  | [37]       | Protoberberine shows antimutagenic activity by inhibiting Topoisomerase I   | Resistant  |
| 19 | Linden          | <i>Viburnum dilatatum</i>      | Caprifoliaceae | [38]       | Iridoids glucosides exhibits moderate inhibitory activity against HeLa S3 cancer cells                                    | Resistant  |
| 20 | Larch           | <i>Larix decidua</i>           | Pinaceae       | [39]       | LaPSvS1 showed good antiangiogenic activity in CAM-assay.   | Resistant  |
| 21 | Magnolia        |                                | Magnoliaceae   | [28]       | Induces apoptosis in leukemia cells   | Resistant  |
| 22 | Pine            | <i>Pinus sp</i>                | Pinaceae       | [7]        | Cell cycle arrest: inhibits tubulin diassembly  | Resistant  |
| 23 | Douglas fir     | <i>Pseudotsuga menziesii</i>   | Pinaceae       | [40]       | Induces growth inhibition in human lung carcinoma cells   | Resistant  |
| 24 | Bald Cypress    | <i>Taxodium distichum.</i>     | Cupressaceae   | [41]       | Cell cycle arrest: inhibits tubulin diassembly  | Resistant  |
| 25 | Hemlock         | <i>Conium maculatum</i>        | Umbelliferae   | [42]       | Inhibits malignant tumours especially breast cancer.  | Resistant  |
| 26 | Redbud          | <i>Cercis canadensis</i>       | Redbud         | [42], [25] | Antileukemia  | Resistant  |
| 27 | Smoke tree      | <i>Cotinus cogygria</i>        | Anacardiaceae  | [43]       | Gallic acid has been shown to display selective cytotoxicity against tumor cells, and to induce apoptosis in tumor cells. | Resistant  |
| 28 | Yew             | <i>Taxus baccata</i>           | Taxaceae       | [7]        | Cell cycle arrest: inhibits tubulin diassembly  | Resistant  |
| 29 | Andromeda       | Andromeda sp                   | Ericaceae      | [25]       |   | Resistant  |
| 30 | Mimosa          | <i>Albizia julibrissin</i>     | Leguminosae    | [24]       | Shows marked inhibitory action against Bel-7402 Cancer cell line.   | Resistant  |
| 31 | Mountain laurel | <i>Kalmia latifolia</i>        | Ericaceae      | [44]       | Shows cytotoxicity against 9KB cell lines   | Resistant  |
| 32 | Euonymus        | <i>Euonymus alatus apterus</i> | Celastraceae   | [21]       | Dulcitol inhibits growth of cancerous cells   | Suceptible |
| 33 | Rose            | <i>Rosa roxburghii</i>         | Rosaceae       | [45]       |   | Suceptible |
| 34 | Russian- olive  | <i>Elaeagnus angustifolia</i>  | Elaeagnaceae   | [46]       | Inhibits several stages in colon carcinogenesis.  | Suceptible |
| 35 | Almond          | <i>Prunus dulcis</i>           | Rosaceae       | [47]       | Betulinic acid showed antiproliferative activity toward MCF-7 cells ( $GI_{50} = 0.27\mu M$ )                             | Suceptible |
| 36 | Walnut          | <i>Juglans sps</i>             | Juglandaceae   |            | Plumbagin is a potent inhibitor of the NF-B activation.   | Suceptible |
| 37 | Ficus           | <i>Ficus citrifolia</i>        | Moraceae       | [48]       |   | Suceptible |
| 38 | Wisteria        | <i>Wisteria sinensis</i>       | Fabaceae       | [25]       |   | Suceptible |



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