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Abstract

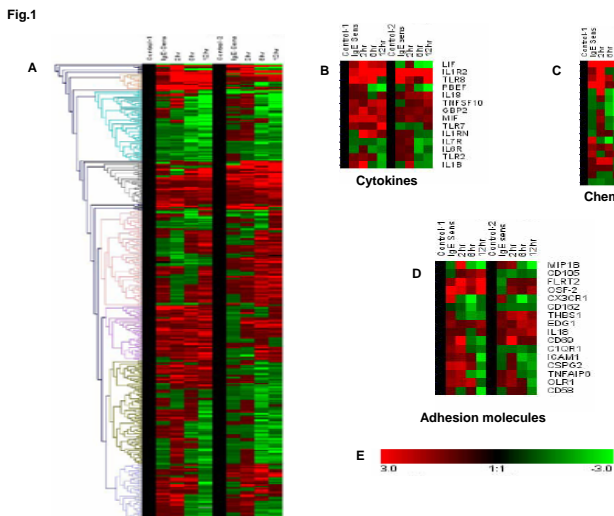
High-density oligonucleotide microarray is a promising approach for high throughput analysis. It has been extensively used in many areas of biomedical research. Immunoglobulin E (IgE) mediated allergic response (type-1 hypersensitivity) is one of the most powerful reactions of the immune system. Tissue Mast Cells (MCs) and circulating basophils are the major effector cells in these reactions. By dissecting the regulatory circuitry of mast cells by analyzing the genome wide effects of antigen stimulation triggered by FcεRI, offers a potential for finding novel genes as 'targets' for therapeutic intervention. In this work, we tried to study the gene expression pattern in IgE sensitized and FcεRI cross linked cord blood derived MCs using one of the latest techniques, high density oligonucleotide expression probe array (HG-Focus array, Gene Chip, Affymetrix, Santa Clara, CA). Microarray hybridization of RNA from cord blood derived MCs revealed coordinated changes in gene expression in response to IgE stimulation and receptor cross linking at different time points. Among the most prominent findings, we observed 2 to 32-fold increased expression of different transcripts. Real-time PCR confirmed reliability of microarray data. This enabled us to classify and cluster genes by functional families as well as to understand known genes in signaling pathways. These results defined a list of primary candidates for finding novel genes as 'targets' for therapeutic intervention.

Experimental set-up and Data Analysis

Total RNA isolated from cord blood derived human mast cells was processed and hybridized to the HG-Focus Gene Array (Affymetrix, Santa Clara, CA). Gene Chip image files were processed using the Microarray Analysis suite 5.0 (Affymetrix). Data of 8400 genes from each time point were used for each time point and the results from duplicate chips work, genes termed 'significantly changed' in response to IgE were those that met the criteria: All genes induced at a change call (P<0.05) of either 'I' (increase) or 'D' (decrease), but not 'NC' (no change). A further constraint was that in at least one experimental condition were included in subsequent analyses. Expressions were clustered by average linkage hierarchical clustering using the GeneSifter clustering prior to hierarchical clustering yielded similar results (data not shown). Genes were then analyzed using the NetAffx analysis centre database (Affymetrix) to confirm the reliability of microarray results.

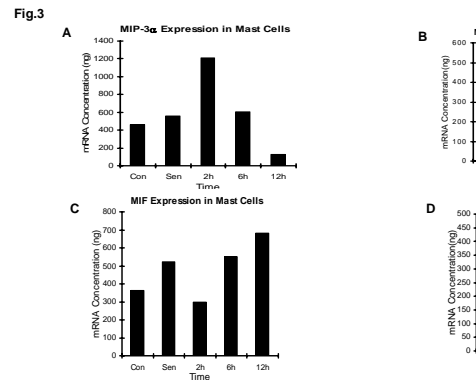
Results

Hierarchical clustering of differentially expressed genes



Changes in gene expression in human mast cells stimulated via FcεRI. Fig.1 (A) Clustering of 395 genes that exhibited a 2-32 fold change in expression over control in duplicates of human cord blood derived mast cells that were activated by IgE sensitization and FcεRI cross linking for different time points (2hr, 6hr and 12hr). Hierarchical clustering was applied using Genesis. Genes were selected for this analysis if their expression level deviated from that in the unstimulated mast cells by 2 fold change in at least 1 time point. The values from different time points and its duplicates were analyzed. Changes in gene expression were depicted according to the color scale shown at the bottom (E). The results are displayed in a table format, in which each row represents a series of measurements of mRNA levels for a single gene, and each column represents the measured mRNA levels for all of the genes in a single sample of cells. Each cell is colored to reflect expression of the corresponding gene in a specific cell sample, relative to its expression level prior to sensitization. Green color represents decreased expression; red color represents increased expression. As indicated, the scale extends from ratios of -3 to 3 in fold change units. Genes for (B) Cytokines, (C) Chemokines and (D) Adhesion molecules whose expression changed significantly.

Real-time PCR: Validation of microarray results



Real-Time PCR for some genes (selected from microarray's result) expressed in mast cells. Mast cells were sensitized by human IgE, and then cross-linked with anti-human IgE for 2hr. Total RNA was extracted. Light-Cycler Real-Time PCR was performed following the protocol for each gene. mRNA were calculated using respective standard curves. Fig.3. (A) MIP-3α expression and (D) COX-2 expression.

Conclusion

In our study, we compared the levels of expression of thousands of genes; expressed in mast cells. Sensitization with IgE triggers the upregulation of several chemokines and cytokines, which has traditionally been linked to their role in immune dysfunctions widely. Thus, in view of the differential gene expression pattern of human mast cells involved in innate immune responses, but may also play a key role in initiating allergic reactions. Future studies should be focused on models that can validate the potential roles of mast cell molecules as potential targets for therapeutic intervention in allergic and inflammatory diseases.

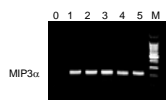
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Electrophoresis of PCR products

Validity of real-time PCR amplicons were checked by agarose gel electrophoresis. Figure.2 shows 210bp amplicon of MIP3α gene.

Fig.2. Lane 0 :Negative control, Lane1-5: RNA from Control, Sensitized, Cross-linked 2hr, Cross-linked 6hr and Cross-linked 12hr respectively. Lane M :Molecular size marker 100bp DNA ladder.



Acknowledgements

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Microarray of the HG-Focus Gene Array (Affymetrix, Santa Clara, CA) analysis was performed with scaled to an average intensity of 1 with R value = 0.91. In this work, we used the following criteria: All genes induced at a change call (P<0.05) of either 'I' (increase) or 'D' (decrease), but not 'NC' (no change). A further constraint was that in at least one experimental condition were included in subsequent analyses. Expressions were clustered by average linkage hierarchical clustering using the GeneSifter clustering prior to hierarchical clustering yielded similar results (data not shown). Genes were then analyzed using the NetAffx analysis centre database (Affymetrix) to confirm the reliability of microarray results.

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