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A survey on helminthic infection in mice (*Mus musculus*) and rats (*Rattus norvegicus* and *Rattus rattus*) in Kermanshah, Iran

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Article Info	Abstract
<p>Article history:</p> <p>Received: 09 June 2012 Accepted: 15 September 2012 Available online: 15 June 2013</p> <p>Key words:</p> <p>Helminth Iran Kermanshah Mice Rats</p>	<p>Parasitic infections of rodents can compromise scientific research as well as the health of the animals and humans. Based on previous studies, infection rate of parasitic helminths is different in various regions of Iran. The current survey was aimed to determine endoparasitic helminths infection in 138 trapped rodents of Kermanshah county, Iran. Mice and rats were trapped using metal snares from January to October 2011 and euthanized. Rodents included 110 <i>Mus musculus</i> (79.00%), 23 <i>Rattus norvegicus</i> (17.00%), and five <i>Rattus rattus</i> (4.00%). The gastrointestinal and respiratory tracts were removed and examined to identify parasitic helminths. The results indicated that 42.02% of examined rodents were infected with eight helminths species, i.e. <i>Trichuris muris</i> (14.49%), <i>Syphacia obvelata</i> (13.76%), <i>Syphacia muris</i> (2.89%), <i>Aspicularis tetrapetra</i> (5.07%), <i>Heterakis spumosa</i> (5.07%), <i>Capillaria hepatica</i> eggs (3.62%), <i>Hymenolepis diminuta</i> (12.30%), and <i>Cystisercus fasciolaris</i>, the larva of <i>Taenia taeniformis</i> (4.34%). Given the results of this study, we concluded that examined rodents were more infected with nematodes than other helminths. As rodents are usually infected with a number of zoonotic parasites, hence control of these animals has an important role in safeguarding public health.</p> <p>© 2013 Urmia University. All rights reserved.</p>

مطالعه بر روی آلودگی‌های کرمی در موش خانگی و موش صحرائی قهوه‌ای و سیاه در شهرستان کرمانشاه، ایران

چکیده

آلودگی‌های انگلی در جوندگان می‌تواند مطالعات علمی و همچنین سلامت حیوانات و انسان را تحت تأثیر قرار دهد. بر پایه بررسی‌های قبلی میزان آلودگی‌های کرمی جوندگان در نواحی مختلف ایران متفاوت می‌باشد. مطالعه حاضر به تعیین کرم‌های انگلی داخلی در ۱۳۸ جونده به دام افتاده در شهرستان کرمانشاه در ایران پرداخته است. موشها و رتوها توسط تله‌های فلزی از ژانویه تا اکتبر ۲۰۱۱ به دام انداخته و سپس آسان گشتی شدند. این جوندگان شامل ۱۱۰ قطعه موس موسکولوس یا موش خانگی (۷۹/۰۰ درصد)، ۲۳ قطعه رتوس نروژیکوس یا موش صحرائی قهوه‌ای (۱۷/۰۰ درصد) و پنج قطعه رتوس رتوس یا موش صحرائی سیاه (۴/۰۰ درصد) بودند. مجاری گوارشی و تنفسی جوندگان جهت آلودگی‌های کرمی مورد بررسی قرار گرفت. نتایج نشان داد که ۴۲/۰۲ درصد از جوندگان مورد مطالعه با هشت گونه کرم شامل؛ تراکیوریس موریس (۱۴/۴۹ درصد)، سیفاسیا ابولاتا (۱۳/۷۶ درصد)، سیفاسیا موریس (۲/۸۹ درصد)، آسیکیولاریس تتراپترا (۵/۰۷ درصد)، هتراکیس اسپوموزا (۵/۰۷ درصد)، تخم کاپیلاریا هپاتیکا (۳/۶۲ درصد)، هیمنولیس دیمینوتا (۱۲/۳۰ درصد) و سیستی سرکوس فاسیولاریس یا لارو تنیا تنیه‌فورمیس (۴/۳۴ درصد) آلوده بودند. از نتایج حاصله می‌توان دریافت که در جوندگان مورد مطالعه آلودگی با نماتودها بیش از سایر کرم‌ها بود. از آنجائیکه جوندگان با شماری از انگل‌های زئونوز آلوده بودند، لذا کنترل این حیوانات نقش مهمی در حفظ سلامت عمومی خواهد داشت.

واژه‌های کلیدی: ایران، کرم، کرمانشاه، موش خانگی، موش صحرائی

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Introduction

Rodents are one of the most successful, abundant, and destructive group of animals causing direct and indirect damage to agricultural products both at pre and post-harvest stages. In addition, they are responsible for transmitting various agents, including a number of helminths parasites, to human and domestic animals.¹⁻³ Infection in human generally occurs directly through contact with rodent excrement, ingesting food contaminated with their fur, feet, urine or fecal dropping, rodent's bites and indirectly through bites from ectoparasitic vectors such as flea and ticks.² Wild rodents serve as reservoir host and have greater ability to harbor a number of endoparasitic agents that play important role in human and livestock health.^{2,4-6} Helminths such as *Trichinella*, *Angiostrongylus*, *Capillaria*, *Hymenolepis*, *Railleitina*, *Echinococcus*, *Schistosoma*, *Paragonimus*, and *Echinostoma* that reported from rodents are importance in public health.⁶⁻⁸ In addition, some of rodents' endoparasites such as *Capillaria hepatica* and *Angiostrongylus cantonensis* cause severe syndromes in humans and other animals.^{9,10} Thus, investigation on rodents' parasites in different geographical areas has medical and veterinary importance to prevent transmission of diseases to humans and animals.¹¹ Several studies have been conducted on parasites of wild rodents from different part of the world that reveal the occurrence of a rich parasite diversity including the endoparasitic helminths fauna^{5,12-14} and ectoparasitic arthropods fauna, as well.¹⁵ In Iran, there are some reports on the occurrence of parasitic infection in different species of rodents in some areas.¹⁶⁻²⁰ In addition, it is demonstrated that some rodents' species are reservoir of cutaneous leishmaniasis²¹⁻²³ and visceral leishmaniasis²⁴ However, little is known about helminths' infection in some areas of Iran such as Kermanshah. The present study reports, for the first, the prevalence of mice and rats parasitic helminths in this region.

Materials and Methods

Study area. Rodents for this survey obtained from both urban and rural area of Kermanshah county, southwest Iran (34°18'N, 47°3'E and 1420 m above sea level), and then examined for parasitic helminthic infection. Kermanshah is situated between two cold and warm regions and enjoys a moderate and mountainous climate. It rains most in winter and is moderately warm in summer. The annual rainfall is 500 mm. The average temperature in the hottest months is above 22 °C.²⁵

Animals. A total number of 138 rodents belonging to three species were collected from 56 locations of Kermanshah county from January to October 2011. These animals were trapped using metal snares, and different baits such as fresh cucumber, cheese and walnut. Traps were set at outdoors in agricultural, horticultural and animal

farms, dry riverbeds, parks and other suitable places in both urban and rural areas. Trapped rodents were transferred to Parasitology Laboratory, School of Veterinary Medicine, Razi University, Kermanshah, and then euthanized. Each rodent sex was recorded, and identification of the species was confirmed on the basis of morphological characteristics with reference to keys²⁶ in Zoology Department of Razi University. After dissection, internal organs (esophagus, stomach, small and large intestines, liver, lungs, peritoneum, urinary bladder, pectoral and abdominal cavity) of each rodent's carcass were removed and examined for adult or larval stages of helminths under stereomicroscope. Parasites were removed carefully from infected organs, cleared, stained, and identified by using appropriate systematic keys.²⁷ In addition, some tissue smears prepared for screening of *Capillaria hepatica* eggs from infected livers.

Results

During the course of the study, out of 138 captured animals, 110 were *Mus musculus* (43 female; 67 male), 23 were *Rattus norvegicus* (10 female; 13 male), and 5 were *Rattus rattus* (1 female; 4 male) (Table 1). Forty two percent (58/138) of examined animals were infected with different helminths whose 21.10% (30/73) trapped in urban areas and others, 20.90%, (28/65) captured in rural areas.

Figure 1 shows infection rate based on mice species and sex and Fig. 2 demonstrates the distribution of infected (I) and none infected (N) rodents based on geographical distribution. Five species of nematodes (*Syphacia obvelata*, *Syphacia muris*, *Aspicularis tetrapetra*, *Heterakis spumosa* and *Trichuris muris*), one species of adult cestodes (*Hymenolepis diminuta*) and one species of larvae form of cestodes (*Cysticercus fasciolaris*) were identified. In addition, the eggs of *Capillaria hepatica* were identified from 3.62% of examined rodents. Of 73 trapped rodents in urban areas, 23 (32.00%) were infected with nematodes, 14 (19.00%) with cestodes, and 7 (9.50%) showed mixed infection. These results in rural areas were; 23 rodents (35.00%), 9 rodents (14.00%), and 4 rodents (6.00%), respectively. Also, a total number of 58 infected rodents, 36 (62.06%) were female, and 22 (37.94%) were male. The results indicated that examined rodents were more infected with nematodes than cestodes ($p \leq 0.05$, $\chi^2 = 23.725$, $df = 2$). *Trichuris muris* had the highest prevalence and *Syphacia muris* the least abundant. Infection rates in those mice involved with *Aspicularis tetrapetra* (5.07%), *Heterakis spumosa*, (5.07%) and *Cysticercus fasciolaris* (4.34%), were nearly similar.

Table 1. Distribution of trapped rodents based on sex and location (F: Female, M: Male).

Location of sampling	<i>Mus musculus</i>		<i>Rattus norvegicus</i>		<i>Rattus rattus</i>		Total
	M	F	M	F	M	F	
Urban area	22	31	7	10	1	2	73
Rural area	21	36	3	3	0	2	65
Total	43	67	10	13	1	4	138

In Table 2, infection rates of examined rodents with different helminths based on animals' species are shown. From 84 identified *S. obvelata*, 12 helminths (14.00%) were male and 72 helminths (86.00%) were female. Also, 70.00% (7 helminths) of *S. Muris* was female, and 30.00% (3 helminths) was male. The ratios for *A. tetrapetra*, *H. spimusa* and *T. muris* were 27.00% (12 worms) male and 73.00% (33 helminths) female, 54.00% (30 worms) male and 46.00% (26 helminths) female, and 33.00% (34 helminths) male and 67.00% (70 helminths) female, respectively. Figure 3 shows some of parasitic helminths removed from examined mice. No infection was observed in the esophagus, stomach, and lung of examined animals.

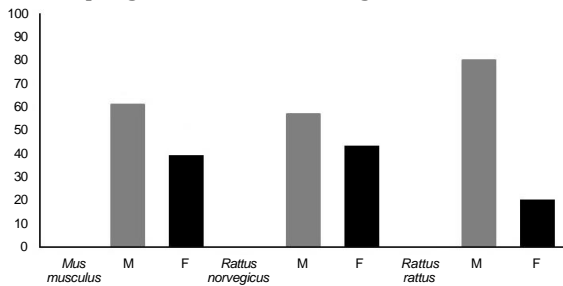


Fig. 1. Infection rate (%) based on species and sex ($\chi^2 = 137.087$, $df = 2$), (F: Female, M: Male).

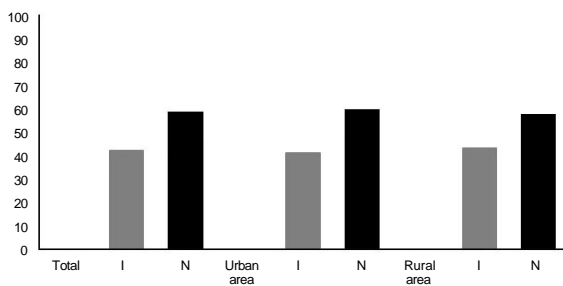


Fig. 2. Distribution rate (%) of infected (I) and non-infected (N) mice based on locations of sampling ($\chi^2 = 0.069$, $df = 1$).

Discussion

The present study gives the first overview on the endoparasitic helminths infection of trapped rodents in Kermanshah, Iran of which reporting eight species of helminths from three species of rodents. Data from previous studies on helminths parasites in rodents of other regions of Iran were partially comparable with the helminths fauna in the present study. In this study, the number of trapped

Mus musculus was significantly higher than other species of rodents ($\chi^2 = 137.087$, $df = 2$, $p \leq 0.05$), but no significant differences between infection rate in urban and rural areas was observed ($\chi^2 = 0.069$, $df = 1$). Also, infection with nematodes was significantly higher than cestodes ($\chi^2 = 23.725$, $df = 2$, $p \leq 0.05$), while the trematodes were absent in examined mice, resembling the results of another report by Malsawmtluangi and Tandon in India.⁶

Infection rate in male animals (62.06%) was higher than female (37.94%), this is irreconcilable with finding of Milazzo *et al.* who found no significant differences between male and female rodents.⁸ However Rogriguez *et al.* found higher prevalence of helminths in male rodents.²⁸ Of identified nematodes from infected rodents, the number of female helminths (208 helminths) was higher than male helminths (91 helminths) ($\chi^2 = 45.873$, $df = 1$, $p \leq 0.05$). *Trichuris muris* was the most common nematode removed from infected animals, but no significant differences were found among this parasite and *S. obvelata* and *H. diminuta*, which were nearly similar to those findings of other reports in Iran^{11,29,32} and other locations of the world.^{8,33-35} *Hymenolepis nana*,³⁶ *Syphacia hodarae n. sp.*,³⁷ *Trichuris navonae n. sp.*,³⁸ *Gongylonema monigi*,¹⁹ *Physaloptera spp.*,^{32,39} *Nippostrongylus brasiliense*,⁴⁰ *Angiostrongylus cantonensis*,^{41,42} *Strongylus ratti*,²⁷ and larvae of *Taenia endorasicus*⁴³ were reported in some previous investigations, but were absent in current study. In addition, Asmar *et al.* and Kia *et al.* reported *Strongyloides ratti* and *Physaloptera sexulatus* whom were not observed in this study.^{19,44} Singla *et al.* indicated that *C. fasciolaris* were the common helminths in rodents of Panjab, India, and its prevalence was much higher (35.20%) than our finding (4.34%).² In other investigations, infestation of rodents with ectoparasites^{35,45} and protozoa (e.g. *Cryptosporidium spp.* and *Sarcocystis spp.*)^{30,39} have also been reported.

Some of the recovered parasites from rodents in this study were of zoonotic importance helminths, including *C. hepatica*, *H. diminuta*, *S. obvelata*, and *Taenia taeniaformis* larva. *Hymenolepis nana* is the zoonotic helminth commonly reported in Iran,⁴⁶ but it was not found in this study. Also *H. diminuta* has already been reported in human.^{47,48} This parasites are common in children and sometimes produce disorders in the hosts.⁴⁹ *Capillaria hepatica*, which was found in this survey, is very important in human causing a lethal syndrome which has already been reported from different countries.¹⁰ These zoonotic parasites have been

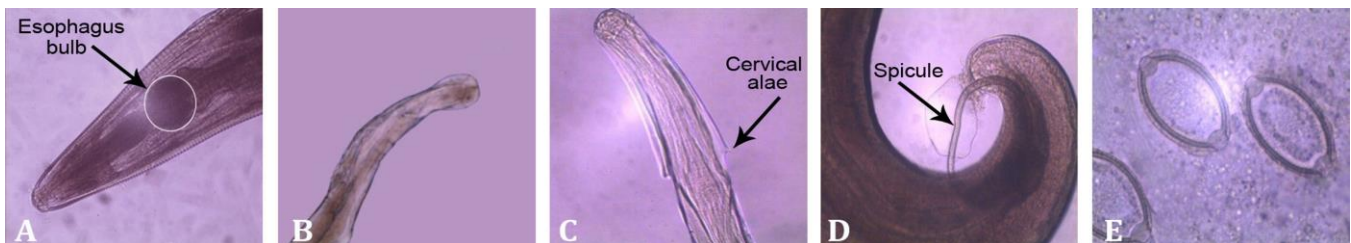


Fig 3. A) Anterior end and esophagus bulb of *Syphacia obvelata*, 400 ×; B) Anterior end and cervical alae of *Syphacia muris*, 100×; C) Anterior end of *Aspicularis tetrapetra*, 400 ×; D) Posterior end of *Trichuris muris* (male), 400 ×; E) *Capillaria hepatica* eggs in the liver of infected rodents, 400 ×.

Table 2. Infection rates (%) of identified helminths in examined mice.

Specie of rodent	<i>Syphacia obvelata</i>	<i>Syphacia muris</i>	<i>Aspicularis tetrapetra</i>	<i>Heterakis spumosa</i>	<i>Trichuris muris</i>	<i>Capillaria hepatica</i> eggs	<i>Hymenolepis diminuta</i>	<i>Cystisercus fasciolaris</i>	Mixed infections
<i>Mus musculus</i>	14.54	4.54	6.36	3.64	16.36	0.90	9.09	4.54	10.90
<i>Rattus norvegicus</i>	4.34	0	0	13.04	8.69	13.04	30.43	3.34	8.70
<i>Rattus rattus</i>	20.00	0	0	0	0	20.0	0	0	0

reported from rodents in variable prevalence of different areas worldwide such as Siberia,⁹ Switzerland⁵⁰ and Iran.¹⁸ Because of important role of rodents in spreading different parasitic agents and destroying the food crops, control programs are needed for reducing their adverse impact.

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References

- Singh YP, Gangwar S, Kumar D, et al. Rodent pests and their management in the northeastern hill region. Research bulletin. No 37, ICAR research complex for NEH region, Barapani, Meghalaya 1995; 35.
- Singla LD, Singla N, Parshad VR, et al. Rodents as reservoirs of parasites in India. *Integrative Zool* 2008; 3: 21-26.
- Oldham JN. The helminth parasites of common rats. *J Helminthol* 1931; 9:49-60.
- Durden LA, Hu R, Oliver JH, et al. Rodents' ectoparasites from two locations in northwestern Florida. *Vec Ecol* 2000; 25: 222-228.
- Stojcevic D, Mihaljevic Z, Marinculic A. Parasitological survey of rats in rural regions of Croatia. *Veterinarni Medicina* 2004; 49(3): 70-74.
- Malsawmtuangi C, Tandon V. Helminth parasite spectrum in rodent hosts from bamboo growing areas of Mizoram, north-east India. *J Parasitol Dis* 2009; 33(1-2): 28-35.
- Khalil LF. The helminth parasites of rodents and their importance. In *Proceedings: The second symposium on recent advances in rodent control*. Sheraton, Kuwait. 1986; 141-149.
- Millazzo C, Ribasa A, Casanova JC, et al. Helminths of the brown rat (*Rattus norvegicus*) (Berkenhout, 1769) in the city of Palermo, Italy. *Helminthologia* 2010; 47(4): 238-240.
- Chechulin AI, Karpenko SV, Panov VV. Ecology of *Hepaticola hepatica* infection in rodents in southern west Siberia. *Contem Prob Ecol* 2011; 4(4): 423-427.
- Fuehrer HP, Petra Igel P, Auer H. *Capillaria hepatica* in man: An overview of hepatic capillariosis and spurious infections. *Parasitol Res* 2011; 109: 969-979.
- Kia EB, Shahryary-Rad E, Mohebbali M, et al. Endoparasites of rodents and their zoonotic importance in Germi, Dashte Mogan, Ardabil province, Iran. *Iranian J Parasitol* 2010; 5 (5): 15-20.
- Claveria FG, Causapin J, Guzman MA, et al. Parasite diversity in *Rattus spp.* caught in wet markets. *South As J Trop Med Publ Health* 2005; 36: 1-4.
- Hasegava H, Koboyoshi J, Ozturu M. Helminth parasites collected from *Rattus rattus* on Lanyu, Taiwan. *J Helminthol Soc Washington* 1994; 61 (1): 95.
- Gomez Villafane IE, Robles MR, Busch M. Helminth communities and host-parasite relationships in argentine brown rat (*Rattus norvegicus*). *Helminthologia* 2008; 45(3): 126-129.
- Nava S, Lareschi M, Voglino D. Interrelationship between ectoparasites and wild rodents from north-eastern Buenos Aires province, Argentina. *Memorias do Instituto Oswaldo Cruz* 2003; 98(1): 45-49.
- Sadjjadi SM, Massoud J. Helminth parasites of wild rodents in Khuzestan province, southwest of Iran. *J Vet Parasitol* 1999; 13(1): 55-56.
- Mowlavi GH. Study on the parasitic infections of rats in Tehran. MSPH Thesis. School of public health and institute of public health research, Tehran University of Medical Sciences, Tehran, Iran. 1991; 1-2.
- Mohebbali M, Rezaei H, Faranak A, et al. A survey on parasitic fauna (helminths and ectoparasites) of rodents in Meshkin Shahr district, northwest Iran. *J Fac Vet Med Univ Tehran* 1997; 52(3): 23-25.
- Kia EB, Hoamyouni MM, Faranak A, et al. Study on endoparasites of rodents and their zoonotic importance in Ahvaz, southwest Iran. *Iran J Public health* 2001; 30 (1-2): 49-52.
- Fasihi-Harandi M. Study on the fauna of parasites of wild rodents in northern Isfahan. MSPH Thesis. School of public health and institute of public health research, Tehran University of Medical Sciences, Tehran, Iran. 1992; 132.
- Edrissian GH, Ghorbani M, Tahvildar-Bidruni GH. *Meriones persicus*, another probable reservoir zoonotic cutaneous leishmaniasis in Iran. *Trans R Soc Trop Med Hyg* 1975; 69(5-6): 517-519.
- Yaghoobi- Ershadi MR, Akhavan AA, Mohebbali M. *Rhombomys opimus* and *Meriones libycus* (Rodentia: Gerbillidae) are the main reservoir hosts in a new focus of zoonotic cutaneous leishmaniasis in Iran. *Trans R Soc Trop Med Hyg* 1996; 90: 503-504.
- Javadian E, Dehestani M, Nadim A, et al. Confirmation of *Tatera indica* (Rodentia: Gerbillidae) as the main reservoir host of zoonotic cutaneous leishmaniasis in the west of Iran. *Iran J Public Health* 1998; 27(1-2): 55-60.
- Mohebbali M, Poormohammadi B, Kanani P, et al. Rodents (*Gerbillidea-Cricitidae*), another animal host of

- visceral leishmaniasis in Meshkinshahr district, I.R. of Iran. *East Mediterr Health J* 1998; 4(2): 376-378.
25. Chalechaleh A, Karimi I. The prevalence of *Trichomonas vaginalis* infection among patients that presented to hospitals in the Kermanshah district of Iran in 2006 and 2007. *Turk J Med Sci* 2010; 40 (6): 971-975.
 26. Etemad E. Mammals of Iran. Vol I: Rodents and key to their identification. National society of natural source and human environment protection. Publication of Tehran 1978; 288.
 27. Eslami A. *Veterinary Helminthology*. 2nd ed. Vol. II cestoda, Vol. III Nematoda and Acanthocephala. Tehran University Publication. 1997; 825-845.
 28. Rodriguez-Vivas RI, Panti-May JA, Parada-Lopez J et al. The occurrence of the larval cestode *Cysticercus fasciolaris* in rodent populations from the costal ecological reserve, Yucatan, Mexico. *J Helminthol* 2011; 6: 1-4.
 29. Gholami S, Mobedi I, Motavali-Haghighoo F, et al. Study on intestinal helminth parasites of rodents in urban and central area of Mazandaran province. *J Sci Research Mazandaran Med Uni* 2002; 12 (35): 67-73.
 30. Akhtardanesh B, Radfar MH, Bagheri F. A parasitological study on blood, skin and alimentary tract of conventionally maintained laboratory mice and rat. *Tehran Uni Med J* 2010; 68(8): 339-443.
 31. Hamediy Y, Heidari M, Soleimani-Ahmadi M. Intestinal and blood parasites of brown rats in Bandar-Abbas. *Hormozgan Med J* 2003; 7(3): 123-127.
 32. Rasti S, Mobedi I, Dehghani R, et al. Endoparasites fauna of wild and house mice in Kashan county. The second congress of parasitic diseases of Iran. Tehran 1997; 213.
 33. Abdel-Wahed MM, Salem GH, El - Assaly TM. The role of wild rats as a reservoir of some internal parasites in Qalyobia governorate. *J Egypt soc parasitol* 1999; 29(2): 495-503.
 34. Mikail MW, Metwally AM, Allam KA, et al. Rodentes as reservoir host of intestinal helminths in different Egyptian agroeco systems. *J Egypt Soc parasitol* 2009; 39 (2): 633-640.
 35. Fagir DM, El- Rayah A. Parasites of the Nile rat in rural and urban regions of Sudan. *Integrative zool* 2009; 4(2): 179-187.
 36. Macnisha MG, Ryana UM, Behnke JM, et al. Detection of the rodent tapeworm *Hymenolepis microstoma* in humans. A new zoonosis? *Intern J Parasitol* 2003; 33: 1079-1085.
 37. Herrera EJR, Mino MH, Notarnicola J, et al. A new species of *Syphacia* (Nematoda: *Oxyuridae*) from *Calomys laucha* (Rodentia: *Cricetidae*) in an Agroecosystem of central Argentina. *J Parasitol* 2011; 97(4): 676-681.
 38. Robles, M. New species of *Trichuris* (Nematode: *trichuridae*) from *Akodon montensis* Thomas, 1913, of the Paranaense forest in Argentina. *J Parasitol* 2011; 97(2): 319-327.
 39. Tung KC, Hsiao FC, Yang CLL, et al. Surveillance of endoparasitic infections and the first report of *Physaloptera sp.* and *Sarcocystis spp.* in farm rodents and shrews in central Taiwan. *J Vet Med Sci* 2009; 71 (1): 43-47.
 40. Waugh CA, Lindo JF, Foronda P, et al. Population distribution and zoonotic potential of gastrointestinal helminths of wild rats (*Rattus rattus* and *R. norvegicus*) from Jamaica. *J parasitol* 2006; 92 (5): 1014-1018.
 41. Chen D, Zhang Y, Shen H. Epidemiological survey of *Angiostrongylus cantonensis* in the west - central region of Guangdong province. *China Parasitol Res* 2011; 4: 124-127.
 42. Foronda P, Lopez - Gonzalez M, Miquel J. Finding of *Parastrongylus cantonensis* (Chen, 1935) in *Rattus rattus* in Tenerife, Canary Islands (Spain). *Acta Trop* 2010; 114(2): 123-127.
 43. Mowlavi GR, Kia EB, Mobedi I. Natural infection of the gerbil *Meriones libycus* with the metacestode of *Taenia endotheracicus* in Arak, central Iran. *J Helminthol* 2004; 78: 275-276.
 44. Asmar M, Mobedi I, Motavallian SA, et al. Study on prevalence of pathogenic parasites in rodents in Lahijan. *J Infect Trop Dis* 2004; 26: 7-10.
 45. Matthee S, Krasnov BR. Searching for generality in the patterns of parasite abundance and distribution: ectoparasites of a south African rodent, *Rhombomys pumilio*. *Intern J Parasitol* 2009; 39: 781-788.
 46. Rokni MB. The present status of human helminthic diseases in Iran. *Ann Trop Med Parasitol* 2008; 102(4): 283-295.
 47. Mowlavi GH, Mobedi I, Mamishi S, et al. *Hymenolepis diminuta* (Rudolphi, 1819) infection in a child from Iran. *Iran J Public Health* 2008; 37(2): 120-121.
 48. Chadirian E, Arfaa F. Human infection with *Hymenolepis diminuta* in villages of Minab, southern Iran. *Int J Parasitol* 1972; 2: 481-482.
 49. Nemat-Ealahi A, Moghadam GHA, Jamali R, et al. A survey on parasitic infestation (helminths and ectoparasites) of the rodents in Tabriz. *J Vet Res* 2006; 61(3): 265-268.
 50. Reperant LA, Deplazes P. Cluster of *Capillaria hepatica* infections in non-commensal rodents from the canton of Geneva, Switzerland. *Parasitol Res* 2005; 96: 340-342.