

## CORRELATION BETWEEN ANGIOTENSIN-CONVERTING ENZYME INHIBITORS LIPOPHILICITY AND PROTEIN BINDING DATA

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Angiotensin-converting enzyme (ACE) inhibitors represent a significant group of drugs primarily used in the treatment of hypertension and congestive heart failure. In this research, seven ACE inhibitors (enalapril, quinapril, fosinopril, lisinopril, cilazapril, ramipril, benazepril) were studied to evaluate the relationship between their protein binding and calculated (logP values) or ultra-high performance liquid chromatography-tandem mass spectrometry (UHPLC-MS) and reversed-phase thin-layer chromatography (RP-TLC) lipophilicity data ( $\phi_0$ , CHI or  $C_0$  parameters, respectively). Their protein binding data varied from negligible (lisinopril) to 99% (fosinopril), while calculated  $\log P_{KOWWIN}$  values ranged from -0.94 (lisinopril) to 6.61 (fosinopril). The good correlations were established between protein binding values and  $\log P_{KOWWIN}$  data ( $R^2=0.7520$ ) as well as between protein binding and chromatographic hydrophobicity data,  $\phi_0$ , CHI or  $C_0$  parameters ( $R^2$  were 0.6160, 0.6242 and 0.6547, respectively). The possible application of hydrophobicity data in drugs protein binding evaluation can be of great importance in drug bioavailability. *Acta Medica Medianae* 2012;51(4):13-18.

**Key words:** angiotensin-converting enzyme inhibitors (ACE inhibitors), protein binding, lipophilicity

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### Introduction

Absorption, distribution, metabolism and elimination (ADME) properties play a critical role in clinical success of drug candidates. Lipophilicity is one of the most important properties that significantly influence drugs absorption, distribution, binding to plasma proteins and elimination due to hydrophobic interactions of the drug with biological targets and its penetration across biological membranes during transport. Lipophilic molecules exhibit better absorption, penetration into tissues and higher degree of distribution. Also, it is well-known that more lipophilic drugs exert a higher degree of protein binding in comparison to less lipophilic ones with similar properties (1-3).

The plasma protein binding (PPB) degree significantly influences drugs efficiency. The less bound drug more efficiently passes through cell membranes or diffuses. Mainly, the unbound fraction actually exhibits pharmacologic effects. It is also the fraction that may be metabolized and/or excreted (4).

Angiotensin-converting enzyme (ACE) inhibitors represent significant group of drugs widely used in

the treatment of hypertension, congestive heart failure and renal failure. They were introduced in clinical practice three decades ago and today represent the most commonly prescribed antihypertensive drugs (4-10).

According to the available literature, a number of authors investigated the relationship between lipophilicity and ACE inhibitors pharmacological activity, duration of action and absorption (11-13). In our previous studies of ACE inhibitors, we reported their lipophilicity under different chromatographic conditions (14-16), a relationship between chromatographic and *in silico* hydrophobicity parameters (17), as well as the correlation between UHPLC-MS and RP-TLC hydrophobicity data and ACE inhibitors absorption values (18). Conducting these researches, the aim of this study was to investigate the relationship between ACE inhibitors lipophilicity data (both calculated and chromatographically obtained) and their protein binding data. The main topic was to establish the approach capable for protein binding prediction of selected ACE inhibitors as well as the new synthesized drugs.

### Materials and methods

#### Materials

The following ACE inhibitors (Figure 1) were investigated:

1. enalapril maleate,
2. quinapril hydrochloride,
3. fosinopril sodium,
4. lisinopril dihydrate,
5. cilazapril monohydrate,
6. ramipril
- and 7. benazepril hydrochloride.

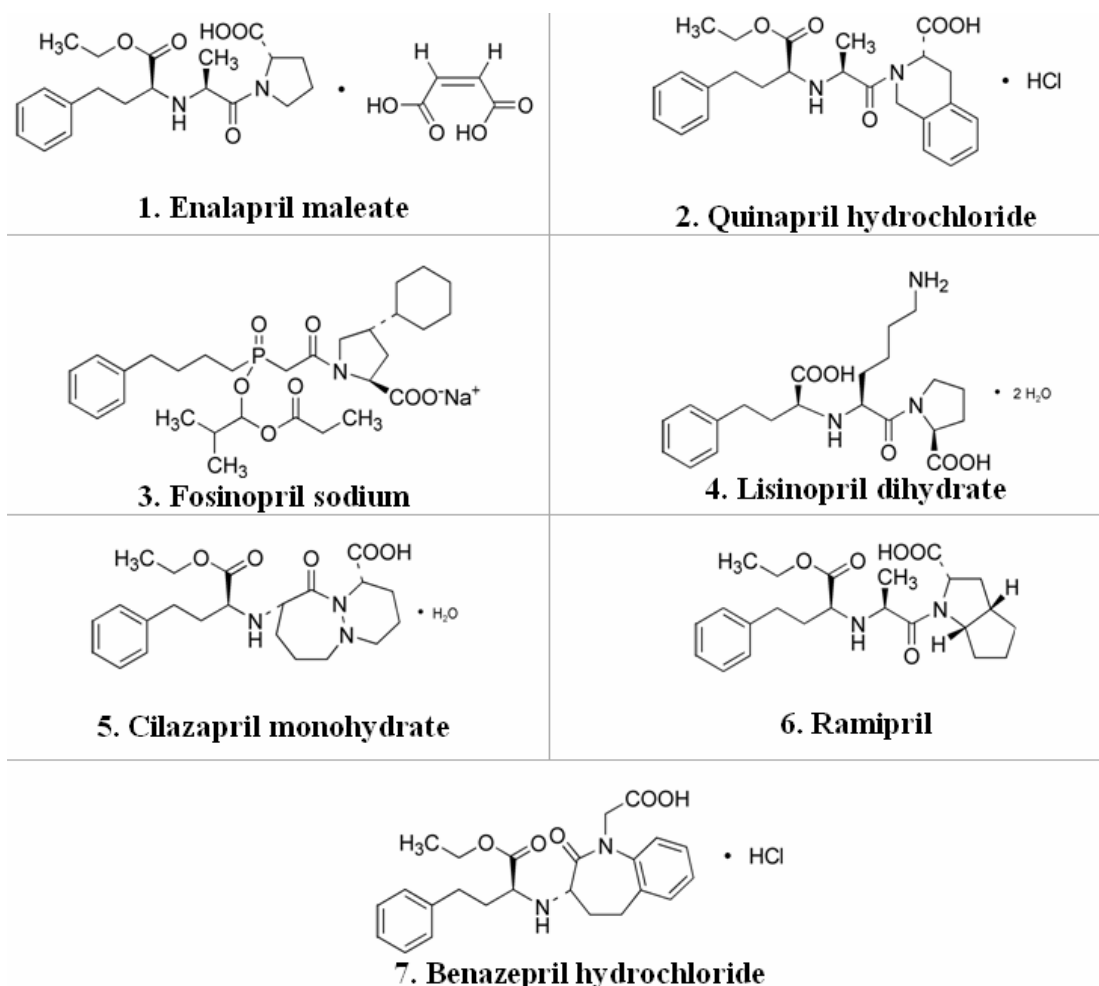


Figure 1. The chemical structure of investigated ACE inhibitors

## Methods

The chromatographic conditions, ultra-high performance liquid chromatography-tandem mass spectrometry (UHPLC-MS) and reversed phase thin-layer chromatography (RP-TLC) for ACE inhibitors examination were reported previously (18). The UHPLC-MS investigations were carried out, as previously described (18) on a UHPLC-MS/MS system consisting of a Thermo ACCELA UHPLC (Thermo Scientific, Waltham, Massachusetts, USA) coupled to a triple quad Mass Spectrometer Thermo TSQ Quantum Access Max (Thermo Scientific, Waltham, Massachusetts, USA) with a heated electrospray ionization (HESI) interface. Samples were placed in thermostated autosampler at 4°C. A 10  $\mu$ l samples were injected onto a Thermo Hypersil Gold column (1.9 $\mu$ m, 50x2.1mm) and eluted at a temperature of 25°C and a flow rate of 300 $\mu$ l min<sup>-1</sup> by the use of binary solvent system (mobile phase A was 0.1% CH<sub>3</sub>COONH<sub>4</sub> aqueous solution (pH=6.85), while mobile phase B was methanol) (18). The RP-TLC examinations were performed, as previously described on RP-18 silica gel plates with water-methanol binary solvent system (18).

Reagents: ammonium acetate (Analytika, Ltd., Prague, Czech Republic), methanol (LC-MS Chromasolv, Sigma-Aldrich Steinheim, Germany) and deionized water (Gen Pure Ultrapure, Germany) were used throughout. Uracil (Merck, Darmstadt, Germany) was used for dead time determination (18).

The Excel 2003 from Microsoft Office and Origin 7.0 PRO (Origin Lab Corporation, USA) were used to perform the statistical analysis of the regression.

## Results

In this research, seven ACE inhibitors (enalapril, quinapril, fosinopril, lisinopril, cilazapril, ramipril, benazepril) were studied to evaluate the relationship between their protein binding and calculated (logP values) or ultra-high performance liquid chromatography-tandem mass spectrometry (UHPLC-MS) and reversed-phase thin-layer chromatography (RP-TLC) determined lipophilicity data ( $\phi_0$ , CHI or C<sub>0</sub> parameters, respectively). Their protein binding data vary from negligible (lisinopril) to 99% (fosinopeil), while calculated logP<sub>KOWWIN</sub> values ranged from -0.94 (lisinopril) to 6.61 (fosinopril).

Table 1. The lipophilicity and protein binding data of selected ACE inhibitors

ACE inhibitors	$\log P_{O/W}$	$\log P_{KOWWIN}$	Protein binding %
Enalapril	2.45	2.45	55
Quinapril	3.72	3.72	97
Fosinopril	6.61	6.61	99
Lisinopril	-1.22	-0.94	0
Cilazapril	/	2.27	24
Ramipril	3.32	3.32	73
Benazepril	3.50	3.50	94

Table 2. The hydrophobicity parameters of investigated ACE inhibitors obtained in UHPLC-MS and RP-TLC

ACE inhibitors	UHPLC-MS		RP-TLC
	$\varphi_0$	CHI	$C_0$
Enalapril	0.533	0.537	0.596
Quinapril	0.642	0.637	0.673
Fosinopril	0.773	0.800	0.802
Lisinopril	0.201	0.213	0.429
Cilazapril	0.636	0.620	0.655
Ramipril	0.615	0.610	0.660
Benazepril	0.623	0.606	0.670

The ACE inhibitors PPB data were (Table 1) were collected from relevant references (4).

The experimentally determined  $\log P_{O/W}$  values (Table 1) of examined ACE inhibitors were obtained from Clarke's Analysis of drugs and Poisons (19). The software packages Molinspiration (20), Virtual Computational Chemistry Laboratory (21) and CS Chem Office, version 7.0 (22) were used to calculate different ACE inhibitors lipophilicity descriptors, (in silico hydrophobicity parameters) computed  $\log P$  values.

The ACE inhibitors hydrophobicity parameters,  $\varphi_0$ , CHI, and  $C_0$  values (Table 2) were chromatographically obtained in UHPLC-MS and RP-TLC investigations (18).

## Discussion

This study included the most often prescribed ACE inhibitors. In the first stage the selection of appropriate  $\log P$  values was evaluated. The applied software packages (20-22) could calculate different  $\log P$  values (milogP, AlogP, AClogP, XlogP,  $\log P_{KOWWIN}$ ) of investigated ACE inhibitors.

The absolute calculated  $\log P$  values were significantly different. In our previous paper the relationships between all collected  $\log P$  values were studied. The selection of appropriate  $\log P$  values was estimated on the basis of their agreement with experimentally determined partition coefficients  $\log P_{O/W}$  values (19). The  $\log P_{KOWWIN}$  values were selected for this study due to its best correlation ( $R^2=0.999$ ) with the experimentally obtained ones  $\log P_{O/W}$  values (17). The  $\log P_{KOWWIN}$  values were also selected, since the best correlations were obtained between these values and chromatographically obtained

hydrophobicity parameters,  $C_0$  (RP-TLC) as well as  $\varphi_0$  or CHI (UHPLC-MS) (18).

In the next stage of this study, the relationship between calculated lipophilicity ( $\log P_{KOWWIN}$  values) and PPB data of examined ACE inhibitors was investigated and the following correlation was obtained:

$$\log P_{KOWWIN} = 0.0501(\pm 0.0129) \text{PPB} - 0.1734(\pm 0.9351) \dots (1)$$

$$n=7, R^2=0.7520$$

The good correlation (Figure 2) was obtained as proposed (the range of  $R^2$  0.49–0.79) in literature (23).

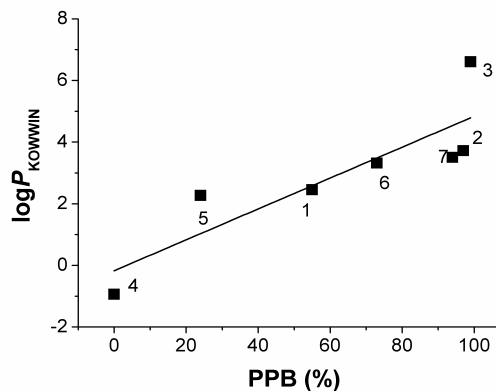


Figure 2. The relationships between protein binding (%) and  $\log P_{KOWWIN}$  values ( $R^2=0.7520$ ). The numbers denote substances used for linear regression

As a final point, the possible correlations between chromatographically (UHPLC-MS or RP-TLC) obtained hydrophobicity parameters ( $\varphi_0$ , CHI or  $C_0$ ) and protein binding data were studied.

$\varphi_0=0.0036(\pm 0.0013)PPB+0.3461(\pm 0.0929)\dots(2)$   
 $n=7, R^2=0.6160$   
 $CHI=0.0036(\pm 0.0013)PPB+0.3454(\pm 0.0915)\dots(3)$   
 $n=7, R^2=0.6241$   
 $C_0=0.0023(\pm 0.0007)PPB+0.4934(\pm 0.0550)\dots(4)$   
 $n=7, R^2=0.6547$

All the correlations obtained can be considered as good, with acceptable F values due to limited number of compounds. The lipophilicity data both calculated as well as obtained with different chromatographic techniques, are capable of evaluating ACE inhibitors protein binding. However, the best relationship was observed between protein binding data and calculated  $\log P_{KOWWIN}$  values. The best correlation was found between in silico hydrophobicity parameters and protein binding data, thus confirming the calculation of  $\log P$  as high-throughput screening technique for the evaluation of selected compounds protein binding degree.

It is generally accepted that increase in drugs lipophilicity led to increase of their absorption, distribution, activity and duration of action, but also to increase of drugs protein binding. Since high protein binding degree may cause decrease of drugs action and activity, the good balance between lipophilicity and protein

binding should be established, especially in new synthesized drugs, for the patients' benefits.

### Conclusion

The discovery of new pharmacologically active substances and drugs modeling led to necessity of predicting drugs properties and its ADME data.

It was established that all ACE inhibitors lipophilicity data (calculated hydrophobicity parameters  $\log P_{KOWWIN}$  as well as chromatographically obtained parameters  $C_0$ ,  $\varphi_0$  and CHI) correlate well with protein binding values. The proposed model based on hydrophobicity data, calculated or chromatographically obtained, is capable of evaluating ACE inhibitors protein binding values. The possible application of computed  $\log P$  values in drugs protein binding evaluation is of great importance in drug research.

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## References

1. Kaliszan R. QSPR: Quantitative structure-(chromatographic) retention relationships. *Chem Rev* 2007; 107(7): 3212-46. [[CrossRef](#)] [[PubMed](#)]
2. Di L, Kernsy EH. Profiling drug - like properties in discovery research. *Curr Opin Chem Biol* 2003; 7(3): 402-8. [[CrossRef](#)] [[PubMed](#)]
3. Hartmann T, Schmitt J. Lipophilicity – beyond octanol/water: a short comparison of modern technologies. *Drug Discov Today Techn* 2004; 1(4): 431-9. [[CrossRef](#)]
4. Lemke TL, Williams DA, editors. *The Foye's Principles of Medicinal Chemistry*. 6th ed. Philadelphia: Wolters Kluwer, Lippincott Williams & Wilkins; 2008.
5. Giverhaug T, Falck A, Eriksen BO. Effectiveness of antihypertensive treatment in chronic renal failure: to what extent and with which drugs do patients treated by nephrologists achieve the recommended blood pressure? *J Hum Hypertens*. 2004; 18(9): 649-54. [[CrossRef](#)] [[PubMed](#)]
6. Lopez-Sendon J, Swedberg K, McMurray J, Tamargo J, Maggioni AP, Dargie H, et al. Expert consensus document on angiotensin converting enzyme inhibitors in cardiovascular disease. The Task Force on ACE-inhibitors of European Society of Cardiology. *Eur Heart J* 2004; 25(16): 1454-70. [[CrossRef](#)] [[PubMed](#)]
7. Jafar TH, Schmid CH, Landa M, Giatras I, Toto R, Remuzzi G, et al. Angiotensin-converting enzyme inhibitors and progression of nondiabetic renal disease. A meta analysis of patient-level data. *Ann Intern Med* 2001; 135(2): 73 –87. [[PubMed](#)]
8. Penno G, Chaturvedi N, Talmud PJ, Cotroneo P, Manto A, Nannipieri M, et al. Effect of angiotensin-converting enzyme (ACE) gene polymorphism on progression of renal disease and the influence of ACE inhibition in IDDM patients: Findings from the EUCLID randomized controlled trial. *Diabetes* 1998; 47(9): 1507–11. [[CrossRef](#)] [[PubMed](#)]
9. Ruster C, Wolf G. Renin-angiotensin-aldosterone system and progression of renal disease. *J Am Soc Nephrol* 2006; 17(11): 2985-91. [[CrossRef](#)] [[PubMed](#)]
10. Johnson CA, Simmons WD. *Dialysis of Drugs*. Madison, Wisconsin: University of Wisconsin; 2002.
11. Ranadive SA, Chen AX, Serajuddin TM. Relative lipophilicities and structural – pharmacological considerations of various angiotensin-converting enzyme (ACE) inhibitors. *Pharm Res* 1992; 9(11): 1480-6. [[CrossRef](#)] [[PubMed](#)]
12. Zannad F. Duration of Action of Angiotensin Converting Enzyme Inhibitors. *Am J Hipertens* 1995; 8(10): 75S-81S. [[CrossRef](#)] [[PubMed](#)]
13. Kim JS, Oberle RL, Krummel DA, Dressman JB, Fleisher D. Absorption of ACE-inhibitors from small intestine and colon. *J Pharm Sci* 1994; 83: 1350-6. [[CrossRef](#)] [[PubMed](#)]
14. Odović J, Stojimirović B, Aleksić M, Milojković-Opsenica D, Tešić Ž. Reversed-phase thin-layer chromatography of some angiotensin converting enzyme (ACE) inhibitors and their active metabolites. *J Serb Chem Soc* 2006; 71: 621-8. [[CrossRef](#)]
15. Odović J, Stojimirović B, Aleksić M, Milojković-Opsenica D, Tešić Ž. Examination of the hydrophobicity of ACE inhibitors and their active metabolites by salting-out thin-layer chromatography. *J Planar Chromatogr* 2005; 18: 102-7. [[CrossRef](#)]
16. Odović J, Aleksić MB, Stojimirović B, Milojković-Opsenica D, Tešić Ž. Normal-phase thin-layer chromatography of some ACE inhibitors and their metabolites. *J Serb Chem Soc* 2009; 74(6): 677-88. [[CrossRef](#)]
17. Odović JV, Markovic BD, Trbojević-Stanković JB, Vladimirov SM, Karljiković-Rajić KD. Evaluation of ACE inhibitors lipophilicity using *in silico* and chromatographically obtained hydrophobicity parameters. *Hem Ind*. In press 2013. [[CrossRef](#)] [[PubMed](#)]
18. Odovic JV, Markovic BD, Injac RD, Vladimirov SM, Karljikovic-Rajic KD. Correlation between ultra-high performance liquid chromatography–tandem mass spectrometry and reversed-phase thin-layer chromatography hydrophobicity data for evaluation of angiotensin-converting enzyme inhibitors absorption. *J Chromatogr A* 2012; 1258: 94-100. [[CrossRef](#)] [[PubMed](#)]
19. Moffat AC, Osselton MD, Widdop B, editors. *Clarke's Analysis of Drugs and Poisons*. 4th ed. London: Pharmaceutical Press; 2011.
20. Molinspiration software or free molecular property calculation services. Available from URL: [www.molinspiration.com](http://www.molinspiration.com)
21. Tetko IV. Virtual Computational Chemistry Laboratory. Available from URL: [www.vcclab.org](http://www.vcclab.org)
22. CS Chem Office, Version 7.0. Cambridge, MA, U.S.A: Cambridge Soft Corporation; 2001.
23. Asuero AG, Sayago A, Gonzalez AG. The correlation coefficient: An overview. *Crit Rev Anal Chem* 2006; 36: 41-59. [[CrossRef](#)]

## ZAVISNOST LIPOFILNOSTI I VEZIVANJA ZA PROTEINE PLAZME INHIBITORA ANGIOTENZIN KONVERTUJUĆEG ENZIMA

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Inhibitori angiotenzin konvertujućeg enzima (ACE inhibitori) predstavljaju veliku grupu lekova koji nalaze primenu u lečenju hipertenzije. U ovom radu analizirano je sedam ACE inhibitora (enalapril, kvinapril, fosinopril, lizinopril, cilazapril, ramipril i benazepril) kako bi se ispitala zavisnost između njihovog vezivanja za proteine plazme i lipofilnosti. Korelisane su vrednosti izračunatih ( $\log P_{KOWWIN}$ ) ili hromatografski (UHPLC-MS i RP-TLC) dobijenih ( $\varphi_0$ , CHI ili  $C_0$ ) hidrofobnih parametara. Procenat vezivanja za proteine plazme ispitivanih ACE inhibitora kretao se u opsegu od 0% do 99%, dok su vrednosti izračunatih  $\log P_{KOWWIN}$  vrednosti iznosile od -0.94 do 6.61. Dobijene su zadovoljavajuće korelacije između vrednosti vezivanja ACE inhibitora za proteine plazme i izračunatih  $\log P_{KOWWIN}$  vrednosti ( $R^2=0,7520$ ) kao i hromatografski dobijenih parametara hidrofobnosti,  $\varphi_0$ , CHI,  $C_0$  ( $R^2$ : 0,6160; 0,6242; 0,6547). *Acta Medica Medianae 2012;51(4):13-18.*

**Ključne reči:** inhibitori angiotenzin konvertujućeg enzima (ACE inhibitori), vezivanje za proteine plazme, lipofilnost