



Effect of biostimulation on biodegradation of dissolved organic carbon in biological filters

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Abstract. The addition of labile organic carbon (LOC) to enhance the biodegradation rate of dissolved organic carbon (DOC) in biological columns was studied. Acetate standard solution (NaAc) and Luria Bertrani (LB) medium were used as LOC as biostimulants in glass column system used for measurements of biodegradable dissolved organic carbon (BDOC). The addition of LOC related with the increase of total DOC in sample. The concentration of BDOC increased up to 7 and 5 times and was utilized after 24 min. contact time. The biodegradation rate constant was increased at least 26 times during adaptation-biostimulation period. There was a strong positive correlation between the biodegradation rate constant and the concentration of BDOC. Biostimulation period ranged from 24 to 53 h for NaAc biostimulant and from 20 to 168 h for LB. The study has shown that LOC could be used as stimulator to enhance the biodegradation rate of DOC during biofiltration.

1 Introduction

During ozonation dissolved organic carbon (DOC) is transformed to biodegradable organic carbon (BDOC), which then is metabolized by bacteria in the biofilter (Fahmi et al., 2003; Hammes and Vital, 2008; Volk et al., 1993). The amount of BDOC fraction depends on the type of natural organic matter (NOM) present in the water (Huck, 1990; Kaplan et al., 1994). The NOM compounds have different biodegradation kinetics: (i) fast biodegradable, (ii) more resistant to biodegradation or slow biodegradable and (iii) not biodegradable (Carlson and Amy, 2001; Klevens et al., 1996; Yavich et al., 2004). About 30 % of DOC is usually removed after biofiltration (Volk et al., 2002) which are usually designed for empty bed contact time (EBCT) of less than 30 min. In Boreal regions, where the surface waters contain high concentrations of NOM or organic matter substances having a low biodegradation rate and biofilters are operated at low temperatures, NOM removal in biofilter is not effective, and can reach only 15–19 % measured as DOC, or 75 % measured as neutral fraction or fast biodegradable part of NOM (Tihomirova, 2011). The slow biodegradable part of BDOC which is not removed in the biofilter will enter the

distribution network and will be used as a substrate for bacteria (Eikebrokk et al., 2007; Tihomirova, 2011). To increase the biodegradation rate for removal of recalcitrant organic substances the addition of labile organic carbon (LOC) is a widely used practice for the remediation of contaminated soils, sediments and sewage (Brandt et al., 2003; Shimp and Pfaender, 1985; Spain et al., 1980; Wiggins and Alexander, 1988), however applicability of this approach for the removal of compounds resistant to biodegradation from drinking water during biofiltration has not yet been studied.

The aim of this paper was to evaluate the effect of addition of LOC in water to enhance the biodegradation rate of DOC during biofiltration.

Biostimulation effect was determined using bioreactor set-up used to measure BDOC (Eikebrokk et al., 2007) and developed within the EU project TECHNEAU, which yields information not only on final concentrations of BDOC but also on the degradation kinetics. The study was carried out in a laboratory scale using treated humic rich water after biofilters.

2 Materials and methods

2.1 Glassware

All glassware used in these experiments were cleaned thoroughly with a 10 % solution of potassium dichromate in concentrated sulphuric acid and rinsed with ultra pure water (Elga PureLab Ultra, Veolia Water Ltd., UK), dried and covered with aluminium septum heated for 6 h at +250 °C in order to avoid organic carbon release (Van der Kooij et al., 1982).

2.2 Reagents

Acetate stock solution (NaAc) with concentration γ (DOC) = 1 g l^{-1} , was made in a 1000 ml volumetric flask where 5.6648 g of sodium acetate trihydrate ($\text{CH}_3\text{COONa} \cdot 3\text{H}_2\text{O}$, Ultra, $\geq 99.5\%$, Fluka, Germany) was dissolved and made up to volume with water. The solution is stable at 4 °C for about 6 months.

Luria Bertani (LB) medium stock solution with concentration γ (DOC) = 7.5 g l^{-1} , was made in a 1000 ml volumetric flask where LB medium contains 10 g Peptone, 5 g Yeast extract, 10 g NaCl in 1000 ml sterile ultra pure water. Both mentioned solutions were used as LOC for the biostimulation experiments.

To avoid inorganic nutrient limitation all samples (100 ml) were supplemented with 100 μl solution of inorganic nutrients. The solution was prepared by dissolving 4.55 g $(\text{NH}_4)_2\text{SO}_4$, 0.2 g KH_2PO_4 , 0.1 g $\text{MgSO}_4 \cdot 7\text{H}_2\text{O}$, 0.1 g $\text{CaCl}_2 \cdot 2\text{H}_2\text{O}$ and 0.2 g NaCl in sterile ultra pure water (1000 ml) (Miettinen et al., 1999).

2.3 DOC determination

The concentrations of DOC were measured with a TOC-5000A Analyzer (Shimadzu Corporation, Kyoto, Japan) according to European Standard EN 1484:1997. The 0.45 μm pore size membrane filters (Millipore Corporation, USA or Sartorius AG, Germany) used for DOC measurements were carefully rinsed, first with ultra pure water and then with the water sample. The blank and control solution were analyzed with each series of DOC sample in order to verify the accuracy of the results obtained by the method. Every DOC sample was tested in duplicate and the mean values were calculated (instrumental relative standard deviation (CV) $\leq 2\%$).

2.4 Study site

The experiments were done using water samples taken after passing through biologically activated carbon (BAC) filters from the surface water treatment plant (WTP) in Riga, Latvia. The raw water in River Daugava contains $15.34 \pm 3.84 \text{ mg l}^{-1}$ of DOC and is treated conventionally by coagulation-sedimentation and filtration in rapid sand filters.

Then the water is ozonated and filtrated through the BAC filters. Water after biofilter contains $5.33 \pm 1.45 \text{ mg l}^{-1}$ of DOC (Tihomirova et al., 2010).

The BAC samples were taken without strict frequency over the period of 1 yr. In total 30 experiments were carried out in this study.

2.5 Experimental design and sampling procedure

All water samples (2000 ml) from BAC filtration were collected in glass bottles completely filled with the sample and subsamples (50 ml) from glass column system were collected in sterile glass bottles and stored in a refrigerator at temperature in the range of 2 to 5 °C, before the analyses were done.

To evaluate the effect of addition of LOC on biodegradation the BDOC experimental set-up contains several chromatography glass columns with different height (H , from 5 to 25 cm) (Chromaflex, USA) coupled in-series system (Fig. 1), which was filled with glass carrier beads ($\varnothing = 6 \text{ mm}$, specific surface area = $3.76 \text{ cm}^2 \text{ g}^{-1}$) as a support media for bacteria (Eikebrokk et al., 2007) was used.

The EBCTs in system was 272 min. The samples were continuously pumped upward the columns using a peristaltic pump (Masterflex L/S, Cole-Parmer, USA). An optimal flow rate of 2–5 ml min^{-1} was used. The biodegradation kinetics of the sample was measured using the intermediate samples (i.e. EBCTs with 8 min intervals).

The biomass in all the columns was kept constant by the homogenization after each experiment. The glass beads were removed from the columns and homogenized by shaking for 24 h and reused after. The biomass concentration in all columns was $5.23 \times 10^{11} \text{ cell cm}^{-2}$ recalculated from adenosine 5-triphosphate (ATP) measurements, according to Magic-Knezev and Van der Kooij (2004).

2.6 Determination of biomass concentration

The concentration of biomass in the column system was measured as the concentration of ATP. The total ATP concentration was determined as described by Berney et al. (2006) using the Promega Bac Titer-Glo Microbial Cell Viability Assay (Promega Corporation, USA) and the calculations were based on the standard curve made with known ATP standard dilutions (Promega Corporation, USA) in sterile ultra pure water. The measurements of ATP were made in the solution obtained from 5 g of glass beads from each column collected in a sterile plastic tube filled with 25 μl ultra pure water and treated by sonification for 2 min with 40 % amplitude or 115 J of energy (Ultra Sonic processor, Cole Parmer, USA) and in the effluent water after each column during the sampling process for DOC samples. The bacterial ATP was calculated by subtracting the extracellular ATP from the total ATP (Hammes and Vital, 2008). The luminescence was measured as an integral over 10 s in relative light units (RLU)

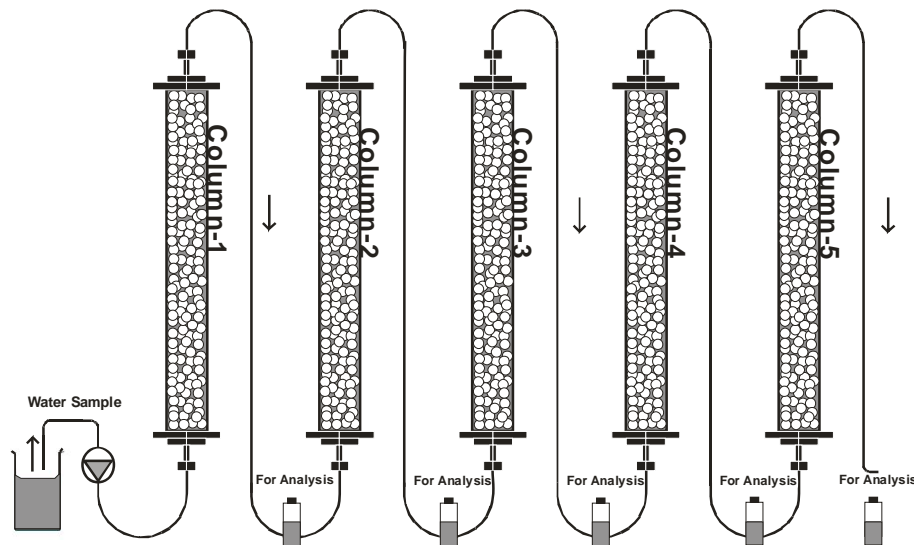


Figure 1. BDOC experimental set-up (adapted from Tihomirova, 2011).

using a luminometer (Hygiene International, Pi-102, Germany). All the samples were measured in triplicate.

2.7 Calculations

The k degradation rate was obtained by fitting the experimental data to the exponential function and expressed as first order kinetic constant (min^{-1}). The equation was fitted separately to data for period in which the minimum concentration of DOC or maximum concentration of BDOC was reached. The regression coefficients (R^2) for exponential curve and Pearson criteria (P) were used (Microsoft Office Excel, 2003). To compare the degradation rate values of statistically significant assays the Moment correlation coefficient (r) was used (Fower et al., 1998).

3 Results

To stimulate biodegradation rate of slowly degradable part of DOC in drinking water treatment the effect of addition of LOC (NaAc or LB) in water was tested in the glass column system.

The average concentration of BDOC of water sample from BAC without addition of LOC was only $0.49 \pm 0.29 \text{ mg l}^{-1}$ ($n = 14$) or 7% of DOC_{BAC} in this study. The biodegradation rates constant (k) of BDOC in BAC water samples with NaAc and LB as biostimulants were higher (about 4.16×10^{-2} and $2.57 \times 10^{-2} \text{ min}^{-1}$, respectively), whereas the biodegradation rate in the sample without biostimulant was one order of magnitude lower ($0.10 \times 10^{-2} \text{ min}^{-1}$), see Fig. 2. The biodegradation rates in the samples with NaAc and LB as biostimulant were at least 26 times higher compared with the BAC sample without biostimulant.

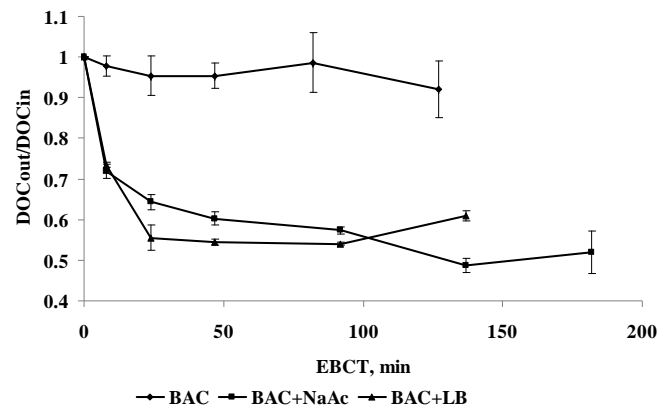


Figure 2. Average DOC changes versus EBCT (adapted from Tihomirova, 2011). Legends: water sample after biofilters from Dau-gava WTP (BAC, $n = 14$); water sample after biofilters from Dau-gava WTP with biostimulant sodium acetate after 30h feeding (BAC+NaAc; $n = 3$) and with Luria Bertrani broth after 48 h feeding (BAC+LB; $n = 3$), respectively.

Traditionally the concentrations of BDOC analysed with dynamic method in glass column system with attached biofilm were calculated as the difference in concentration of DOC between the inlet water sample in column system and the effluent water sample with lowest concentration of DOC, namely DOC_{BAC} and DOC_{min} (Fig. 3) based on the definitions developed by other researchers (Fahmi et al., 2003; Hammes and Vital, 2008; Volk et al., 1993; Ribas et al., 1991 and Yavich et al., 2004) and in this study called BDOC_{BAC} . The total concentration of BDOC (namely $\text{BDOC}_{\text{total}}$) which accounts for both parts – quantity of biodegradable part in sample (DOC_{BAC}) and LOC (DOC_{LOC}) was not considered in this paper since it was not comparable with measurements

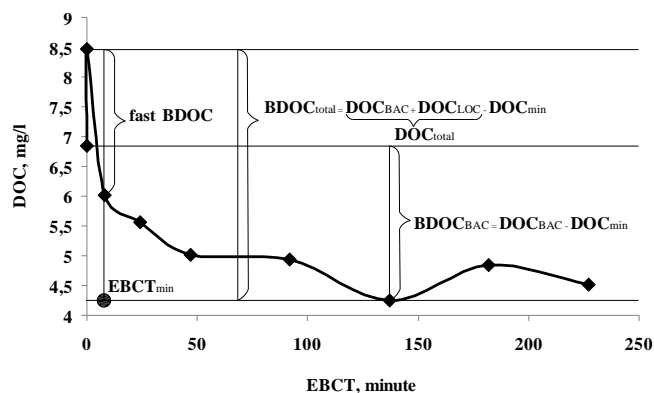


Figure 3. Principle of BDOC quantification shown on an example of 30 h feeding of BAC water sample supplemented with NaAc as a biostimulant.

of BDOC in WTP. The initial concentration of substrate in the BAC sample (DOC_{BAC}) for the series of experiments with NaAc and LB was 5.87 ± 0.96 ($n = 9$) and 4.73 ± 0.19 ($n = 7$) $mg\ l^{-1}$, accordingly. For given DOC measurement this is variation of DOC measurements in inflow to the column during the all experiments due to natural variation of DOC in the BAC samples. The concentration of dose of biostimulant (DOC_{LOC}) of NaAc and LB was 1.81 ± 0.36 ($n = 9$) and 1.25 ± 0.25 ($n = 7$) $mg\ l^{-1}$, respectively.

The period of biodegradation time when $BDOC_{BAC}$ was $< 15\%$ in each water sample supplemented with LOC can be named an adaptation period of biomass (Tihomirova et al., 2012). During the adaptation period a decrease of minimal EBCT was observed (Table 1). The results showed that during the experiment the biodegradation rate constant using biostimulants NaAc and LB increased up to 41 and 26 times, respectively, and this time interval can be called the biostimulation period. The maximum concentration of BDOC and the maximum biodegradation rate was reached after 51 and 48 h feeding with NaAc and LB, respectively (Table 1). The biostimulation period with NaAc was accomplished after 53 h, after which both $BDOC_{BAC}$ and biodegradation rate decreased. In water samples supplemented with LB the biostimulation period was accomplished after 168 h. The removal efficiency of $BDOC_{BAC}$ reached up to 49 % and 37 % at maximal biodegradation rate with both NaAc and LB, respectively which is significantly higher result compared with sample after BAC filters (Fig. 2). There was a strong positive correlation between biodegradation rate constant and $BDOC_{BAC}$ ($r = 0.63$ for NaAc; $r = 0.65$ for LB; $P = 0.6$ for both biostimulants). The biodegradation process can be divided in two periods – adaptation or coadaptation (20–24 h with biostimulants in this study) and biostimulation period, which was limited from 24 to 53 h for NaAc and from 20 to 168 h for LB biostimulant.

The experiments with BAC samples and NaAc and LB biostimulants showed that after the adaptation phase the fast

degradable part increased to 28.9 and 29.3 % of DOC, the slowly degradable increased to 20.9 and 10.7 % and the non-degradable part decreased to 50.2 and 60 %, respectively (data not shown). The BDOC was 50 and 40 % of DOC_{total} , respectively.

4 Discussions

Biostimulation approach was based on the hypothesis that the biodegradability in water samples is higher than the average concentration of BDOC measured at the effluent from BAC filters in WTP. This was concluded by Tihomirova (2011) over regular three year monitoring period of DOC and BDOC in WTP. The concentration of BDOC decreased from $1.47 \pm 1.05\ mg\ l^{-1}$ in the raw water to $0.59 \pm 0.51\ mg\ l^{-1}$ (10 % and 9 % of the DOC, respectively) after rapid filter and increased to $0.82 \pm 0.38\ mg\ l^{-1}$ (15 %) after BAC filtration. High error range might be attributed to the fluctuation of raw water and treated water quality. Consequently, biological stability of water samples increased during coagulation-sedimentation and rapid sand filtration and ozonation, but it decreased again after BAC filters. DOC was higher in the river water than in the drinking water, but the ratio of BDOC/DOC (%) was higher in the drinking water (15 %). This indicate that after ozonation concentration of biodegradable organic compounds increased, but removal of biodegradable organic matter in biofilters or the production of biologically stable water is not efficient. Results of this study showed that average BDOC in BAC samples ($n = 14$) was 7 % only.

The addition of LOC was related to time of adaptation of biomass in glass column system which for NaAc and LB biostimulants was 24 and 20 h, respectively (Table 1) (Tihomirova et al., 2012), the time interval during which the BDOC value was negligible or biodegradation is not detected (Wiggins and Alexander, 1988). As shown previously (Yavich et al., 2004), the addition of a small amount of biostimulant resulted in an increase in BDOC concentration and a sharper decrease in “fast” BDOC.

As shown in this study for samples taken at Daugava WTP the biodegradation rate constant k of samples containing NaAc and LB as biostimulants increased up to 26 times compared with the BAC sample without biostimulant. Thus the biodegradation rates can be enhanced by using biostimulants.

According to obtained measurements in this study and recalculation biomass concentration was $806\ ng\ ATP\ per\ cm^3$ in experimental system. As shown previously (Magic-Knezev and Van der Kooij, 2004), ATP concentration can range from 25 to $5000\ ng\ ATP\ per\ cm^3$ in biofilters at different WTP. The results (Table 1) showed that biomass concentration detached from biofilter when biodegradation process was completed and decrease of biodegradation rate constant. Recalculated total biomass amount in system

Table 1. Biodegradation of substrate in glass column system feed using water samples supplemented with labile organic carbon depending on the adaptation time of the experimental system.

Adaptation time, h	BAC+NaAc				BAC+LB			
	EBCT _{min} , min	$k \times 10^{-2}$, min ⁻¹	BDOC _{BAC} , %	ATP, cells $\times 10^8$ ml ⁻¹	EBCT _{min} , min	$k \times 10^{-2}$, min ⁻¹	BDOC _{BAC} , %	ATP, cells $\times 10^8$ ml ⁻¹
0	> 272	0	0	60	> 272	0.31	0.4	394
4	> 272	0	0	26	> 272	0.41	2	412
20	–	–	–	–	137	0.33	20	333
24	272	0.18	27	52	–	–	–	–
30	137	0.19	38	65	–	–	–	–
44	–	–	–	–	47	1.30	27	291
48	24	3.22	34	65	24	2.57	37	419
51	24	4.16	49	49	–	–	–	–
53	8	3.70	20	65	–	–	–	–
168	–	–	–	–	272	0.18	30	552
192	47	0.53	6	105	227	0.11	10	656
240	92	0.25	1	1602	–	–	–	–

Legends: the minimal EBCT (EBCT_{min}), biodegradation rate constant (k), concentration of BDOC (%) and biomass concentration (determined as ATP, cell per ml) in effluent are shown.

was 405 170 ng ATP, and max detached biomass amount –33 640 ng ATP, what is 8 % of total biomass concentration in system. There was not significant correlation between cell concentration in the effluent and the ATP concentrations in the biofilter. These results showed that bacterial release into the effluent sample may be due to substrate concentration limitation after biodegradation.

The rate of biodegradation is highly variable and depends on many parameters including chemical structure and concentration of the organic substance in water samples, applied oxidant (ozone, chlorine) dose, temperature, salinity, pH, concentration of oxygen, and inorganic nutrients and concentration of active microorganisms (Becker et al., 2006; Brandt et al., 2003, 2004; Kulp et al., 2007; Sanchez et al., 2007; Spain et al., 1980; Spain and Veld, 1983; Steiner and Sauer, 2001; Swindoll et al., 1988). Some organic compounds, especially aromatic compounds are rather resistant to natural biodegradation and biodegradation of a compound of the mixture can be strongly influenced by the presence of other components in the mixture (Tsai and Juang, 2006). It has been shown previously that the addition of LOC results in an increase of the concentration of easy degradable part of mixture and the stimulated biodegradation results in decrease in minimum EBCT compared to that without biostimulation (Spain et al., 1980; Yavich et al., 2004). The biodegradable DOC was utilized after short contact time or at the top of the biofilter (Moll et al., 1998). Direct addition of a nutrient can stimulate the activity of microbial communities (Macbeth et al., 2004) and the presence of easy degradable carbon sources can enhance the biodegradation of more persistent compounds (Brandt et al., 2003; Shimp and Pfaender, 1987).

Method of co-adaptation and its impact on the biodegradation of different chemicals has been used to degrade resistant chemical pollutions in different environmental settings (Schmidt and Alexander, 1985; Shimp and Pfaender, 1987; Wiggins and Alexander, 1988). Biostimulation method described in this study can also be compared with the other (e.g. phosphorus dosing into biofilters to stimulate the activity of the biomass, Rubulis, 2006).

Addition of biostimulant increased BDOC_{total} in influent water; however increase in the total consumption of BDOC allowed to reduce both BDOC added as biostimulant and native BDOC in the sample. In this study biostimulants (NaAc or LB) are easy convertible substrate for bacterial biomass. In these experiments NaAc was used as single carbon source and LB medium as multicarbon source. Results obtained in this study showed that BDOC concentration increased significantly and was utilized after 24 min contact time due to biostimulation comparing with sample without biostimulant.

This study showed that the biofiltration systems of the WTPs in Boreal regions treating humic rich water can be improved and with biostimulation approach biodegradation of humic rich water can reach up to 29 %, which is very close to the theoretical –30 % as DOC (Volk et al., 2002). LOC addition stimulated biodegradation rate of slowly biodegradable organic matter. The biomass consumed easy degradable substrate and the adaptation period contributed to that slowly degradable organic matter was used faster as a secondary substrate. From the results it can be concluded that LOC can be used as a stimulator of biodegradation which would allow to effective remove of slowly degradable organic matter within EBCT which is usually about 30 min. The biostimulation period is the period limited in time when

BDOC and the biodegradation rate constant substantially increase.

This study demonstrates principle of biostimulation applicability for enhanced DOC removal in drinking water technology. Before implementation in real plant the technology should be engineered to avoid both problems with release of biomass and other problems, which can related to optimal condition for filter operations such as application of automatic control of biomass and substrate dosing. In practice the optimal concentration of LOC would be depended from the DOC concentration in influent water. LOC work as stimulator, and to avoid the increase the BDOC in system after biofilter, LOC concentration should be verified for every individual plant.

5 Conclusions

From the obtained results it can be concluded that:

- The addition of LOC was related to the increase of the DOC_{total} . At the same time $BDOC_{total}$ increased significantly (up to 7 and 5 times higher with NaAc and LB as biostimulants, respectively) and it was utilized after 24 min contact time due to biostimulation.
- The study demonstrates that biodegradation rates can be increased by using a biostimulant. The biodegradation rates of BDOC in BAC water samples with NaAc and LB as biostimulants were 4.16×10^{-2} and $2.57 \times 10^{-2} \text{ min}^{-1}$ whereas the degradation rate in the sample without biostimulant was $0.10 \times 10^{-2} \text{ min}^{-1}$.
- There was a strong positive correlation between the constant of biodegradation rate and $BDOC_{BAC}$ ($r = 0.63$ for NaAc; $r = 0.65$ for LB; $P = 0.6$ for both biostimulants).
- The adaptation time for the mixture of sample and biostimulant was 20 and 24 h for LB and NaAc, respectively.
- The biostimulation period for NaAc and LB biostimulants was from 24 to 53 h, and from 20 to 168 h, respectively.
- The biostimulation period was accomplished with increasing EBCT, decreasing BDOC and biomass detachment.

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