Influence of ionizing radiation on Cobra (Naja haje) and Cerastes cerastes venoms: Toxicological and immunological aspects.

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Abstract
The effect of gamma irradiation (15 KGy) on the Cobra Naja haje and Cerastes cerastes venoms toxicity and immunogenicity was evaluated. Irradiated venoms were at least 28.1% less toxic than non-irradiated venoms. However the antigenic response was not changed as judged by the capacity of irradiated venoms to react with polyvalent antivenom horse serum. The immunodiffusion method showed identity between irradiated and non-irradiated samples.

The effect of gamma radiation on some venom enzymes and venoms profile was studied. Proteolytic, phospholipase A₂ and hemorrhagic activities were inhibited by irradiation. Irradiation with gamma rays resulted in structural changes in the venoms as indicated by the changes in their molecular weights.

The results indicated that irradiation of toxic protein -using suitable dose- can promote significant detoxification but still retaining many of the original antigenic and immunological properties of native venom.

Introduction
Venoms in general are weakly immunogenic, yet fairly toxic (Magalhaes, 1986). This causes problems because serotherapy is the treatment of choice in snake bite-accidents and horse antisera availability is dependent upon venom immunogenicity. To improve antisera production and extend the useful life of immunized horses much effort has been devoted to decrease chronic venom toxicity. Several techniques have been used to detoxify venoms such as mixing the venom with carboxymethylcellulose (Moroz et al.,1963), irradiation by gamma rays (Baride et al., 1980), adding formaldehyde, (Costa et al., 1985), controlled iodination (Daniel et al., 1987) and encapsulation of purified toxins in liposomes (Freitas and Freszard, 1997).

One method that has been shown to be effective for attenuating venom toxicity and maintaining immunogenicity is gamma radiation (Baride et al., 1980; Hati et al.,1990; Shaban, 1990; Mandal et al., 1993, Nascimento et al., 1996).

Proteins irradiated either in the dry state or in solution undergo chemical and physicochemical changes that can alter their primary, secondary or tertiary structures, while keeping many of their native immunological properties intact (Shalka and Antoni, 1970). Lauhatirananda et al., (1970) observed that several functions of a protein have different radio-sensitivity and that its antigenic properties are the most radioresistant. Hati et al., 1990 showed that Russell viper venom was detoxified by gamma radiation (100 Krad or 200 Krad) and used for active immunization of rabbits. Shaban (1990) found that irradiation of Androctonus amoreuxia scorpion venom with gamma rays in dry state (15 and 30 KGy) decreased its lethal and toxic activity while retaining its antigenicity. Therefore, irradiation of toxic proteins can promote significant detoxification, but still retain many of the original antigenic and immunological properties of native venom.

Snake venoms are complex mixtures which may show qualitative differences in their composition not only between species but also from individual to another (Tu,1977). Cerastes cerastes and Naja haje snakes one belong to family Viperidae and
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family Elemedea, respectively. Cerastes cerastes is mainly vasculotoxic in action while Naga hage is mainly neurotoxic.

In this study we describe the effects of a dose of 15KGY gamma radiation on Cerastas cerastes and Naga hage whole venom to produce detoxified venom with large amount of non-toxic but still immunogenic aggregate with an emphasis on subsequent biological properties and immunogenicity of the irradiated venoms.

Materials and methods:

Venom:
Venoms of Naja haje (Egyptian Cobra) and Cerastes cerastes were pooled and obtained from the venom serpentariums, Faculty of Medecine, Ain Shams University. The venoms were obtained by milking healthy snakes, dried and kept in desiccator till used.

Antivenin:
Egyptian polyvalent antivenin prepared against Naja haje, Cerastes cerastes, Cerastes vipera and Naja nigrocoulos obtained as a gift from the Egyptian Organization of Biological products and Vaccines, Agouza, Cairo, Egypt, was used. The lyophilized polyvalent antivenin produced in horses was kept at 4ºC and reconstituted to 10ml with distilled water before use.

Irradiation facilities:
The venoms were irradiated with 15KGY gamma rays in the National Center for Research and Radiation Technology, Cairo, Egypt, using cobalt-60 gamma cell 220, manufactured by the atomic energy of Canada (AECL). The radiation dose rate was 1.4 Rad per second .

In this study, a saline solution of Naja haje and Cerastes cerastes snake venom, were subjected to integral radiation dose level (15KGY). Non-irradiated samples of venoms were used as control.

Animals:
Male Swiss albino mice weighing between 20-25 gm were used in this study; animals were maintained under standard conditions of food and given standard food add libitum.

Gel chromatography (filtration) technique:
The molecular weights of the irradiated and non-irradiated venoms were determined in the National Research Center, Cairo, Egypt, by the method of Andraws, (1964). 8.0 gm of Sephadex G-200 was used for packing the column (90x1.6cm) and previously equilibrated with 50 Mtris HCL buffer, pH 7.5. Samples containing 10 mg/ml of venoms were applied separately to the same column and developed using the same equilibrating buffer at a flow rate of 20 ml/hr; fractions of 6 ml elution volume were collected. Column fractions were monitored for protein at 280 nm.

The column was calibrated by five known molecular weight proteins as standards, cytochrome C 12.4 KDa carbonic anhydrase 29 KDa , bovin serum albumn 66 KDa , alcohol dehydrogenase 150 KDa , and B . Amylase 200 KDa . Dextran blue (2000000) was used to determine the void volume (V0). A calibration curve was constructed by plotting log molecular weight versus Ve/V0 where Ve is the elution volume. The molecular weight of each peak was estimated from the standard curve.

Double immunodiffusion technique:
Immunodiffusion experiments were carried out as described by Ouchterlony (1948). They were carried out using 1.3 Nobel Agar (Difeco. Lab. Detroit, Mich.) in 0.9% NaCl solution , sodium azide was added in a concentration of 0.05%, to retard bacterial growth. The wells were filled with 20 μl volumes. The venom samples were added in the peripheral wells while the antivenin was added in the central well. After developing of the precipitation bands (48 hr) the plates were washed for 24 hr in saline, dried and stained.

Biological activities of venoms:
Lethality assay:
Toxicity of venom Naja haje and Cerastes cerastes was established before
and after 15 KGy gamma irradiation. Lethality was determined by giving the venom intra peritoneally (in different doses) to Swiss albino mice. The LD50 was calculated by the method of Spearman karber (WHO 1981) using six dosage levels. Six mice were used for each dose.

Assay of proteolytic activity:
Proteolytic activity of non-irradiated and 15 KGy gamma irradiated venoms, was determined with casein (B.D.H.) according to Labib et al. (1980); (EC 3.4.22.17). Mixtures of 1.0 ml of 1% substrate solution (casein in 0.02N NaOH, 0.5 ml of 0.1 MTris (hydroxy methyl) aminomethane- HCl buffer (pH 7.2) and 0.5 ml of a dilution of the venom were incubated at 37º C for 60 min followed by shaking with 2.0 ml of 10% trichloracetic acid (TCA) solution. After standing for 60 min the mixtures were clarified by centrifugation or filtered (using whatmann filter paper) and the absorbance at 280 nm (A280) of the supernatants or filtrates were determined against zero time tests, which were prepared similarly except that TCA was added before the venom.

Determination of hemorrhagic activity:
Hemorrhagic activity of non-irradiated and 15 KGy gamma irradiated Cerastes cerastes venom was assayed in the skin (intradermally, i.d.) according to Borkow et al. (1993). The minimum hemorrhagic dose (MHD) defined as the amount of venom that produces a hemorrhagic spot of about 1 cm in diameter after 2 hr) was determined.

Intradermal observations were carried out after injecting samples of 0.1 ml of non-irradiated and 15 KGy irradiated venom containing 5 MHD into the back skin of six mice. Two hours later the mice were killed and the inner surface of skin was examined.

Determination of phospholipase activity:
Phospholipase A2 activity of both non-irradiated and 15 KGy gamma irradiated venoms (Naja haje, Cerastes cerastes), was assayed according to the method of Desnuelle et al.(1955) and Nieuwenhuizen et al. (1974). Lecithin was used as substrate, and was solubilised either with sodium cholate or Triton X-100. The substrate solution was prepared by stirring lecithin at a final concentration of 3.5 mM in 7 mM triton X-100, 100mM NaCl, 0.055 mM phenol red and 10 mM CaCl2, also 10mM NaOH was used as titrant. 1.5ml of this mixture was introduced into titration cuvette and incubated at 30º C for 15min. NaOH was added to give absorbance of 1.8 at 558 nm in a 1cm path length cuvette. The reaction was started by adding 20µl of venom solution at different concentrations and the changes in optical density (O.D.) at 5,10 and 15 min, were recorded. The phospholipase A2 activity was determined as corresponding to the decrease in O.D.

Results

Gel filtration on Sephadex G-200:
The result of the gel filtration on Sephadex G-200 are presented in (Fig.1). The figure shows that each of the non-irradiated and irradiated crude Naja haje snake venom is separated into three peaks. However, the intensity of the peaks of irradiated samples were lower than the non-irradiated which indicates a break in the chemical bonds of the irradiated venom molecule. The logarithm of each known molecular weight was plotted against the elution volume (fig.2 standard curve). The molecular weights of all the samples were estimated from the standard curve as shown in (Fig.2).

Also the results indicate that the value of the molecular weights of the non-irradiated Naja haje venom peaks were 14, 33 and 123KDa but the molecular weight of the irradiated Naja haje venom tends to progressive shift towards lower molecular weights (11,21 and 110 KDa) than, the non-irradiated Naja haje venom (Fig. 2).

The result of the non-irradiated and irradiated Cerastes cerastes snake venom results are presented in (Figs.3&4). It is clear that the non-irradiated samples had three peaks with molecular weights of 18, 37 and 110 KDa but the irradiated venom had only two peaks (33 and 110 KDa) as the smaller molecular weight disappeared.
The absorption of the irradiated Cerastes cerastes venom was lower than the non-irradiated one.

**Double immunodiffusion:**

The results of the double immunodiffusion test of non-irradiated and 15 KGy gamma irradiated venoms against a commercial polyvalent Egyptian antivenin, all showed similar patterns; the visible lines obtained in the immunodiffusion reactions were identical and join smoothly at the corners, indicating that there was no change in antigenic determinents (Fig.5 A,B).

**Biological activities of venoms:**

**Lethality assay:**

Toxicity assays for the venoms subjected to 15 KGy gamma irradiation are shown in Table 1.

The i.p. LD50 for non-irradiated Naja haje and Cerastes cerastes venom were estimated to be 5 μg and 40μg/20gm mouse, respectively. On the other hand, after 15KGy gamma irradiation of the venom samples, the lethality was affected. Naja haje and Cerastes cerastes were at least 28.1% and 30.8% less toxic than the native one.

Table 1: Toxic activity of non-irradiated and 15 KGy irradiated venoms.

<table>
<thead>
<tr>
<th>Samples</th>
<th>LD50 (μg/ g)</th>
<th>Relative toxicity</th>
</tr>
</thead>
<tbody>
<tr>
<td>Non-irradiated Naga hage venom</td>
<td>0.251</td>
<td>100%</td>
</tr>
<tr>
<td>Irradiated Naga hage venom</td>
<td>0.891</td>
<td>28.1%</td>
</tr>
<tr>
<td>Non-irradiated Cerastes cerastes venom</td>
<td>2.00</td>
<td>100%</td>
</tr>
<tr>
<td>Irradiated Cerastes cerastes venom</td>
<td>6.50</td>
<td>30.8%</td>
</tr>
</tbody>
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![Fig (1): Elution profile of Naja haje snake venom on Sephadex G-200 column](image)
Fig (2): The logarithm of each of known molecular weight (•), irradiated Naje haje venom (0) and the unirradiated venom (+), versus their elution volume.

Fig (3): Elution profile of Cerastes cerastes snake venom on Sephadex G-200 column.

Fig (4): The logarithm of each of known molecular weight (•), and molecular weight of the irradiated (0) and unirradiated (+) Cerastes cerastes snake venom, versus their elution volume.
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Fig.5: A- Immunodiffusion reaction of horse serum polyvalent antivenin (in the central well) with non-irradiated and 15KGy gamma irradiated Naja haje venom (20mg/ml). S= saline Ve1= non-irradiated venom Ve2= 15KGy irradiated venom A=Antivenin

B- Immunodiffusion reaction of horse serum polyvalent antivenin (in the central well) with non-irradiated and 15KGy gamma irradiated Cerastes cerastes venom (20mg/ml) S- saline Ve3= non-irradiated venom Ve4= 15 KGy irradiated venom A=Antivenin

Hemorrhagic activity:

Table 2: shows the inhibition of hemorrhagic activity of Cerastes cerastes venom after 15 KGy gamma irradiation.

<table>
<thead>
<tr>
<th>Venom</th>
<th>Hemorrhagic spot area Cm²</th>
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<tr>
<td>Non-irradiated Cerastes cerastes venom</td>
<td>5.5± 0.45</td>
</tr>
<tr>
<td>15 KGy irradiated Cerastes cerastes</td>
<td>3.8± 0.35</td>
</tr>
</tbody>
</table>

Non-irradiated and 15KGy irradiated whole venom (2.5µg : 5 MHD) were injected i.d. After 2 hr the mice (n=6) were killed and the hemorrhagic spots were measured. Each mean represents the mean of 6 observation (n=6) ± SE. The difference between non-irradiated and 15KGy gamma irradiated venom is significant (P<0.05). Hemorrhagic activity was strongly inhibited at 15 KGy radiation dose.
Proteolytic activity:
Figure 6 A & 6B represents the inhibition in the proteolytic activity by 15 KGy gamma irradiation of Cerastus cerastes and Naja haje venoms respectively.

Phospholipase activity:
Figure 7 and 8 represent the decrease in absorbance at 558 nm of phenol red as a function of the incubation time with different concentrations of non-irradiated and 15KGy gamma irradiated venoms (Naja haje and Cerastes cerates).
The rapid change of absorbance obtained just after adding the aqueous solution of venoms (less than 20μl) to the medium is due to phospholipase activity.
It is shown that 15 KGy gamma irradiation inhibits the phospholipase activity of both venom.

Fig 6 : Effect of 15KGy gamma irradiation on venom protease activity using casein as the substrate.
A- Cerastes cerastes venom
B- Naja haje venom
Fig 7: Decrease in absorbance on phenol red as a function of time for non-irradiated and 15KGY gamma irradiated Naja haje venom:
The change in absorbance at 558 nm of medium containing 3.5 nM lecithin, 4mM cholate, 100nM NaCl, 10nM CaCl2 0.055 nM phenol red and the different concentrations of Naja haje venom was followed as a function of time at room temperature in a 1 cm path length cuvette. The phospholipase solution in water was added in a volume smaller than 20 μl. Recorder tracing represents superimposable data from 4 different solutions.
Fig 8: Decrease in absorbance on phenol red as a function of time for non-irradiated and 15KGY gamma irradiated Cerastes cerastes venom:
The change in absorbance at 558 nm of medium containing 3.5 nM lecithin, 4mM cholate, 100nM NaCl, 10nM CaCl2 0.nM phenol red and the different concentrations of Cerastes cerastes venom was followed as a function of time at room temperature in a 1 cm path length cuvette. The phospholipase solution in water was added in a volume smaller than 20 μl. Recorder tracing represents super imposable data from 4 different solutions.
Discussion

In the present study, the 15KGy irradiated Naja haje and Cerastus cerastus venoms were 28.1% and 30.8% less toxic respectively than the native ones. However, the antigenic response was not changed as judged by the capacity of irradiated venoms to react with a polyvalent antivenom horse serum. The immunodiffusion method showed identity between irradiated and non-irradiated samples. Irradiation with gamma rays resulted in structural changes in the venoms as indicated by the changes in their molecular weights. Proteolytic and phospholipase activity of both venoms were inhibited after 15KGy gamma irradiation, and the hemorrhagic activity of Cerastus cerastus venom was also highly inhibited.

Our result agrees with Murata et al. (1991) who showed that irradiation with 2000Gy crude venom of c.d. terrificus was the ideal irradiation dose, promoting venom detoxification with maintenance of its immunogenicity. After Nascimento et al. (1996) irradiation of crotoxin by X-rays from Co (2000 Gy) results in aggregation and the generation of lower mol wt. breakdown products. The largest aggregates are greatly reduced in their ability to promote creatine kinase (ck) release when injected into muscle tissue, are largely devoid of phospholipase activity and are actually non-toxic to mice relative to the native toxin. Aggregates separated by filtration retain at least part of their higher ordered structure, based on their reactivity with monoclonal antibodies known to rest with conformational epitopes in native crotoxin. Also Hati et al. (1990) studied the effect of gamma radiation on some enzymes and venom protein profiles. Phosphodiesterase, protease and hyaluronidase were inhibited by radiation though phospholipase A activity remained unaffected. Glissa et al. (1999) used whole C. durissus terrificus venom irradiated with incremental doses of gamma rays to increase the amount of a toxic but still immunogenic aggregate. According to Boni Mitake et al. (2001) crotamine was purified from C. d. terrificus venom by Sephadax G-100 gel filtration followed by ion exchanger Somatography, and irradiated at 2mg/ml in 0.15 MNaCl with 2.0KGy gamma radiation, the native and irradiated toxins were evaluated as regards structure and toxic activity (LD50). Irradiation did not change the protein concentration, the electrophoretic profile or the primary structure of the protein although differences were shown by spectroscopic techniques. Gamma radiation reduced crotamine toxicity by 48.3% but did not eliminate it.

Snake venoms are complex mixtures of toxins and enzymes, and there pharmacologic effects result from both the direct effect of the toxin and the indirect effects of enzymes, such as by liberation of autopharmacologic substances such as bradykinin and histamine (Lee, 1979). Cobra toxin was shown to consist of a single peptide chain cross linked intramolecularly by four disulfide bonds that were essential for toxicity (Yang 1970).

It is known that most proteolytic and hemorrhagic components so far isolated from snake venoms are proteins with relatively high molecular weights (usually more than 20000). Also snake venom phospholipases mol wt. are high (range from 8500 to 36000) (Tu, 1977 & 1982). That is why snake venoms are highly affected by radiation which inhibits their toxic activity.

It was well known that the doses of the order of Kilograys used in detoxification of the venom influenced both the physio-chemical properties and the biological activity of macromolecules, when irradiated in aqueous media or even in the solid state (Antoni, 1973; Hayes, 2001) since there is a close interrelationship between the structure and the biological activity of macromolecules, some alteration appeared to be the most possible explanation for the radiation effects.

As regards the effect of radiation on proteins, the amino group is the most radiosensitive portion of an amino acid. However, in the formation of a protein, this groups linked to a carboxyl group and it is, therefore, not easily removed from the molecule. Similarly, the carboxyl group is
no larger available for the reaction. Thus, the side-chains are the most radiosensitive portion of a protein molecule. The specific changes which occur in the side-chains depend on their chemical composition. A hydrogen atom may be removed or a break may occur between, almost, many of the atoms in the chain (Cassarett, 1968).

Loss of function of protein by irradiation, is not usually due to breaking peptide bonds, or otherwise, disrupting the primary skeletal structure of the peptide chain. It may result from a change in a critical side chain or from a break in the hydrogen or disulfide bonds which in turn, can result in a disorganization of the internal relationships of side chain groups, or an exposure of amino-acid groups, resulting in a change in biological activity (Hayes, 2001).

Souza –Filho et al. (1992) found that crotoxin, the major neurotoxin of the South American rattlesnake was irradiated with a Co-60 gamma source at doses of 250, 500, 1000,1500 and 2000Gy. Irradiated crotoxin was analysed for free SH-groups, protein concentration, electrophoretic profile, 50% lethal dose (LD50) in mice, and antigenicity against crotalic antiserum by diffusion immuno-assay. The dose of 1000Gy cleaved 0.95 disulfide bridges/mol and 1500Gy cleaved 1.42 bridges/mol, indicating the importance of disulfide bond integrity for inducing toxicity.

It seems most probable that, the radiation, has affected the three dimensional configuration of the molecule, causing it to unfold and make the disulfide bond available for reactions. Since the venom was irradiated in solution state, the changes have been attributed largely to indirect action of the radiation on the protein molecules. Such an indirect action in an aqueous system is mainly through short-lived radicals such as OH, H, HO2 and free electrons and molecular systems such as H2O2 and O2 (Alvin and Edvin, 1964).

Thus, our results suggest that the treatment with gamma radiation is a suitable way to detoxify Naja haje as well as Cerasus cerasus venom without affecting their immunogenicity provided that a proper dose is used and, therefore, proper toxoid could be obtained.

References


تأثير الأشعة المؤينة على الخواص السمية و المناعية لسم الكوbra المصرية و سم الحية سراستس سراستس

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تم في هذا العمل دراسة تأثير أشعة جاما بجرعة 15 كيلو جرمي على كل من السم و خصائص تكوين أجسام مضادة للسم لكل من سم الكوبرا المصرية و سم الحية سراستس سراستس لذا ظهرت الدراسة أن تعرض كل من سم الناغج هاج و سراستس سراستس لأشعة جاما في الجرعة 15 كيلو جرمي (و هما في الحالة السائبة) يقلل من خاصيتها السمية القاتلة مع احتفظهما بالخواص المناعية و في تجارب الانتشار المزدوج كانت هناك تفاعلات مماثلة لسم غير المشع و المشع مع المصل المضاد و تبين من الفصل الغروي الكروماتوجرافي (سيفادكس 200) تغير الوزن الجزيئي للسمين بعد التشغيع كما تم دراسة تأثير الإشعاع الجامى على النشاط الأنزيمي لهذه السموم و وجد أن إنزيم الفوسفوليبا يز ء 2 , و البروتياز يقل فاعاليتهم بعد تعرض السم لهذه الجرعة من الإشعاع كما وجد أن نشاط النزفي لسم سراستس سراستس بصلورة محتوية بعد التعرض للإشعاع و من هذه النتائج ثبت أن تعرض البروتينات السامة للإشعاع الجامى في جرعات مناسبة يساعد على إزالة سميتها مع الاحتفاظ بالعديد من خصوصها السمية.