

Scotland's Rural College

Evaluation of the effect of different wheats and xylanase supplementation on performance, nutrition and energy utilisation in broiler chicks

Gonzalez-Ortiz, G; Olukosi, OA; Bedford, MR

Published in:
Animal Nutrition

DOI:
[10.1016/j.aninu.2016.06.005](https://doi.org/10.1016/j.aninu.2016.06.005)

First published: 01/07/2016

Document Version
Peer reviewed version

[Link to publication](#)

Citation for published version (APA):

Gonzalez-Ortiz, G., Olukosi, OA., & Bedford, MR. (2016). Evaluation of the effect of different wheats and xylanase supplementation on performance, nutrition and energy utilisation in broiler chicks. *Animal Nutrition*, 2(3), 3 - 7. <https://doi.org/10.1016/j.aninu.2016.06.005>

General rights

Copyright and moral rights for the publications made accessible in the public portal are retained by the authors and/or other copyright owners and it is a condition of accessing publications that users recognise and abide by the legal requirements associated with these rights.

- Users may download and print one copy of any publication from the public portal for the purpose of private study or research.
- You may not further distribute the material or use it for any profit-making activity or commercial gain
- You may freely distribute the URL identifying the publication in the public portal ?

Take down policy

If you believe that this document breaches copyright please contact us providing details, and we will remove access to the work immediately and investigate your claim.

1Evaluation of the effect of different wheats and xylanase supplementation on
2performance, nutrient and energy utilisation in broiler chicks

3

4Gemma González-Ortiz^a, Oluyinka Olukosi^b, Michael R. Bedford^a

5

6^aAB Vista, Marlborough, Wiltshire SN8 4AN, United Kingdom

7^bMonogastric Science Research Centre, Scotland's Rural College, Edinburgh EH9 3JG,
8United Kingdom

9

10

11

12

13

14* Corresponding author.

15E-mail address: mike.bedford@abvista.com

16Abstract

17The aim of this study was to evaluate the performance, nutrient utilisation and energy
18metabolism of broiler chicks fed eight different wheat samples, supplemented or not
19with xylanase. Seven-hundred sixty eight male broilers (1-day old) were distributed to
2016 experimental treatments (six replicates per treatment). The treatments were in a
21factorial arrangement with eight different wheats and two levels of xylanase (0 or
2216,000 BXU/kg). The predicted apparent metabolisable energy (AME) of the wheat
23samples ranged from 13.0 and 13.9 MJ/kg and all diets were formulated to contain the
24same amount of wheat. Body weight gain (BWG) and feed intake (FI) were measured at
2521 d, as was jejunal digesta viscosity, and feed conversion ratio (FCR) calculated. On
26day 24, one representative bird pen was selected to calculate whole body energetics. At
2721 d, three chicks per replicate were randomly allocated to metabolism cages for energy
28and nutrient utilisation determinations, and were continued on the experimental diets
29until 24-d-old. No interactions were observed for any performance response variables,
30ileal nutrient utilisation or digesta viscosity. Xylanase improved BWG and reduced FCR
31and digesta viscosity ($P < 0.05$). Wheat influenced dry matter (DM) utilisation and
32xylanase increased ileal digestible energy ($P = 0.04$). Xylanase also improved ($P <$
330.05) DM and nitrogen retention. Apparent metabolisable energy and AME corrected
34for nitrogen (AMEn) were subject to an interaction whereby wheats 2 and 6, which
35returned the lowest AME and AMEn values, responded to xylanase supplementation
36and the remainder did not. Net energy for production and the efficiency of energy use
37for production were not influenced by xylanase, but were affected by wheat ($P < 0.05$).
38Despite the significant differences between wheats with regards to their nutrient
39utilisation and energy metabolism in birds, xylanase removed this variance and resulted
40in more homogeneous performance.

41Keywords: Wheat, Near-infrared spectroscopy, Xylanase, Animal performance, Nutrient
42release, energy, Broiler chickens

431. Introduction

44 Variation in the nutritive value of wheat samples is a reflection of genetic and
45 environmental effects, and the economic impact of these variations on poultry
46 performance highlights the need for improved predictors of wheat quality (Yegani and
47 Korver 2012). This is a concern for plant breeders, farmers and animal nutritionists.
48 Thus, nutritionists need to know the nutritional requirements of commercial poultry, and
49 be able to determine or predict the nutritive value of each batch of raw material in an
50 accurate and timely manner (van Kempen and Simmins 1997).

51 The use of Near-Infrared Spectroscopy (NIRS) provides an opportunity to determine
52 the chemical composition of feedstuffs and their nutritive value before inclusion in the
53 diet (Olukosi et al., 2011; Owens et al., 2009). The information from NIRS can be used
54 to reduce or minimize nutrient imbalances in commercial rations fed to the animals.
55 However, there are potential errors associated with NIRS technology such as sample-
56 related and chosen reference method errors which can lead to high values for coefficient
57 of variation (Yegani and Korver 2012), and as a result care must be taken in establishing
58 NIRS calibration to ensure it is robust, precise and accurate. Near-Infrared Spectroscopy
59 calibrations now exist which can predict non-starch polysaccharide (NSP) and energy
60 contents of wheat. In particular xylans, is often considered an anti-nutrient in wheat, and
61 as a result variation in content of this component between wheat samples may
62 contribute to differences in nutritive value. **Xylanases are the major enzymes involved**
63 **in arabinoxylan degradation, hydrolysing the 1,4- β -D-xylosidic linkage between xylose**
64 **residues in the backbone in a random manner (Mendis et al. 2016), therefore it is**
65 **hypothesised that their supplementation in poultry feed may balance animal**
66 **performance although differences in the nutritive value of different wheat origins.** This
67 work was undertaken to determine if such a calibration **by NIRS** accurately predicts

68animal performance, and if so whether the application of an NSP-degrading xylanase
69would reduce the performance differences between samples of wheat which differ in
70NSP content (Bedford, 2000).

71

72

732. Materials and Methods

74

75All the experimental procedures received prior approval from the Scotland's Rural
76College's Animal Experiment Committee.

77

782.1. Birds and experimental design

79

80A total of 768 one-day old male broiler chicks (Ross 308) obtained from a commercial
81hatchery were used in the study for two experiments to determine growth performance
82and whole-body energy metabolism (Exp. 1) and nutrient utilisation (Exp. 2) responses.
83For Exp. 1 ($n = 768$) and for Exp. 2 ($n = 288$), birds were allocated to 16 experimental
84treatments in a randomized complete block design with an 8×2 factorial arrangements
85of treatments (eight wheat samples and two levels of xylanase), having in both
86experiments six replicates per treatment. Throughout the study, feed and water were
87supplied *ad libitum* and animals were raised under controlled conditions of light and
88temperature, as breeder recommended.

89

902.1.1. Experiment 1

91

92Birds were reared up to day 24 in floor pens. All broiler chickens and feed were
93weighed on day 0 and 21 to calculate growth performance responses: body weight gain
94(BWG), feed intake (FI) and feed conversion ratio (FCR). On day 21, two chickens
95were randomly selected and euthanized by an overdose of sodium pentobarbital and
96jejunal digesta were collected for viscosity measurement. On day 24, one representative
97bird (on BW basis) per floor pen was selected and fasted prior to euthanasia to calculate
98the whole body energetics.

99

1002.1.2. Experiment 2

101

102On day 21, three chicks were randomly selected from each of the 96 floor pens and
103transferred to 96 metabolism cages (for energy and nutrient utilisation trial) where
104chickens continued to receive the corresponding diets until 24 days of age. Excreta and
105ileal digesta were collected on day 24 and pooled on a cage basis for calculation of
106nutrient utilisation.

107

1082.2. Diets and wheat selection

109

110Starter experimental diets based on wheat and soybean-meal were formulated to be
111marginally lower in metabolisable energy (ME) than Ross 208 requirements (Table 1).
112Eight wheat samples originating from Germany and United Kingdom were obtained.
113Dry matter (DM), gross energy (GE), fat, nitrogen (N), calcium (Ca) and the
114phosphorous (P) contents of wheat samples were chemically analysed and further NIRS
115analyses were performed (Tables 2 and 3). A fixed amount of each wheat (58.6%) was
116used in the formula regardless of their chemical composition. Diets were predicted to

117contain 12.8 ME MJ/kg based on assumed average wheat apparent ME (AME) 58.6%
118came from wheat grain. Control diets were supplemented with 16,000 BXU/kg of
119xylanase following supplier recommendations (Econase XT, AB Vista, Marlborough,
120UK; 160,000 BXU/g), resulting in 16 experimental diets in total. All diets contained
121phytase supplemented at 500 FTU/kg (Quantum Blue, AB Vista, Marlborough, UK;
1225000 FTU/g). Activity of xylanase and phytase were determined using the reference
123method of analysis recommended by the supplier. Titanium dioxide (0.3%) was added
124to all the diets as an indigestible marker. Feed samples were taken at the beginning and
125throughout the experimental period for DM, N, fat and GE analysis.

126

1272.3. Jejunal viscosity

128

129Approximately 1.5 g (wet weight) of the fresh jejunal digesta were analysed according
130to Bedford et al. (1991). The viscosity (expressed as centipoise units, $cP = 1/100$ dyne
131 sec/cm^2) was determined using a Brookfield DV II digital viscometer.

132

1332.4. Nutrient utilisation and total tract retention

134

135Total tract retention and ileal nutrient utilisation were calculated using the index method
136(Olukosi et al., 2007), with titanium dioxide as the indigestible marker.

137

1382.5. Net energy and nutrient accretion

139

140Net energy for production (NEp), heat production (HP) and carcass fat and protein
141accretion were determined using the comparative slaughter technique as described by

142Olukosi et al. (2008). Briefly, six birds were euthanized at day 0 without feeding and
143kept frozen prior to processing and chemical analyses. On day 24, following euthanasia
144the carcasses were frozen and ground prior to freeze drying. Gross energy, N and fat
145contents were analysed. All the calculations for NE_p, ME intake, HP as well as the
146efficiencies of energy for fat and protein retention (Fat-ER and CP-ER, respectively) are
147as described previously (Olukosi et al., 2008a). Net energy for production and HP were
148expressed per kg feed by dividing the total NE_p (MJ) or HP (MJ) by kilogram of feed
149intake.

150

1512.6. Chemical analyses

152

153Ileal digesta and excreta were analysed for DM, N, fat and GE. Dry matter was
154determined by drying the samples in a drying oven (Uniterm, Russel-Lindsey
155Engineering Ltd., Birmingham, England, UK) at 105 °C for 24 h (method 934.01;
156AOAC, 2006). Total N content was determined by the combustion method (method
157968.06; AOAC, 2006). Gross energy was determined in an adiabatic oxygen bomb
158calorimeter (model 6200; Parr Instruments, Moline, IL) using benzoic acid as an
159internal standard. Titanium concentration in samples of diets and ileal digesta was
160determined using the method of Short et al. (1996).

161

1622.7. Statistical analyses

163

164Pen served as the experimental unit for FI, BWG and FCR, and cages as experimental
165unit for nutrient utilisation, jejunal viscosity, net energy and nutrient accretion. Data
166were analysed using the PROC MIXED command of SAS (SAS Inst. Inc., Cary, NC).

167When effects were found to be significant, treatment means were separated using
168Tukey's Highly Significant Difference test. Statistical significance was accepted at $P <$
1690.05 and trends were discussed at $P < 0.10$.

170

1713. Results

172

1733.1. Wheat nutritive value by NIRS and chemical analyses

174

175The chemical analysis of the wheat samples indicates that they are all very similar in
176chemical composition and GE (Table 2). The predicted nutritive values by NIRS
177showed slightly more variability in nutrient composition between wheat varieties (Table
1783), but remained close to expected average values. The predicted GE was
179underestimated while the predicted fat content was higher than chemically analysed.
180The predicted AME of wheat varieties ranged from 13.0 to 13.9 MJ/kg (CV < 2%).
181There was a great deal of variability (CV > 10%) in the predicted contents for crude
182protein, acid detergent fibre, β -glucan, lignin and total non-starch polysaccharides, but
183low variability (CV < 8%) in all other analysed chemical components, including amino
184acids.

185

1863.2. Feed enzyme activity, growth performance and jejunal viscosity

187

188Enzyme activities in feed samples were close to expected (16,038 BXU/kg average
189value analysed in all the xylanase-supplemented diets). No interactions were observed
190in any of the performance parameters measured (Table 4). There were no effects of
191wheat on performance or jejunal digesta viscosity. Nevertheless, improvements in

192performance were observed when xylanase was supplemented, regardless of wheat
193sample. Xylanase application resulted in a near significant 20 g ($P = 0.077$) increase in
194BWG. Although FI was not influenced by xylanase, FCR was significantly improved by
195four points better (1.33 vs. 1.37, xylanase vs. control, respectively; $P = 0.003$). Xylanase
196supplementation also reduced viscosity of jejunal digesta (3.32 vs. 2.34 cP, for control
197and xylanase supplemented diets, respectively; $P < 0.001$). In the diets without xylanase
198supplementation, there were low and non-significant correlations between nutrient
199content of the wheats and bird FCR (Table 5). For the birds receiving xylanase
200supplemented diets, FCR was positively correlated with the analysed P and the
201predicted contents by NIRS of NDF, total and soluble AX as well as insoluble NSP. In
202addition, FCR was positively correlated with the analysed fat content.

203

2043.3. Nutrient utilisation and total tract retention

205

206No interactions between the main factors were observed for any of the ileal nutrient
207utilisation results (Table 6). The DM utilisation of wheat 3 was significantly lower
208compared with wheats 6, 7 and 8 ($P < 0.05$), whereas wheats 1, 2, 4 and 5 had
209intermediate values. Wheats 7 and 8 had greater energy utilisation ($P < 0.001$) compared
210with wheats 1, 2, 3 and 5, whereas wheats 4 and 6 were in between. Xylanase
211supplementation increased ileal utilisation of energy (IDE) measured as MJ/kg ($P =$
2120.04), regardless of wheat. Ileal N utilisation tended to be influenced by wheat ($P =$
2130.06), and was not influenced by xylanase supplementation.

214There were significant interactions of the main factors for all total tract measurements
215($P < 0.001$). Xylanase supplementation improved the retention of DM and N as well as
216AME and AMEn for diets based on wheats 2 and 6. For those diets based on wheats 3,

2174, 5 and 8 xylanase, inclusion led to no effect or marginally lower results in total tract
218retention of N, AME and AMEn.

219

2203.4. Net energy and nutrient accretion

221

222There were no interactions between wheat and xylanase for any energy utilisation and
223efficiency responses, except for HP and Kre-protein, and no xylanase effect on any of
224the responses (Table 8). Net energy for production and K_{RE} were greater ($P < 0.05$) for
225wheat 2 compared with wheats 4, 5 and 7, but similar, although numerically higher, than
226the other wheats. Energy retained as protein was greater ($P < 0.05$) for wheats 3, 4 and 5
227compared with wheats 7 and 8. Energy retained as fat and Kre-fat was greater ($P < 0.05$)
228for wheat 2 than wheats 1, 3, 4 and 5. The interaction observed for HP ($P = 0.02$) was
229explained by xylanase supplementation increasing HP when birds were fed wheats 2
230and 6 (data not shown), but decreased HP for wheat 8, with no effect observed for the
231remaining wheats. The interaction noted for Kre-protein ($P = 0.006$; data not shown)
232was due to xylanase addition resulting in birds fed wheats 3 and 8 supplemented being
233more efficient in protein accretion, whereas it was reduced for wheats 2 and 6, with no
234effect on the remaining wheats.

235

2364. Discussion

237

238It is well known that wheats, even of the same variety, can vary in both chemical
239composition and nutritive value (Theander et al., 1989). The current study investigated
240the effect of wheat sample and xylanase supplementation on the performance of broilers
241fed starter diets. In spite of the variability found between wheats in both the analysed

242chemical composition and that predicted by NIRS, performance was not affected.
243Nevertheless, supplementation with xylanase improved BWG, FCR and reduced digesta
244viscosity, as has been shown in numerous studies (Olukosi et al., 2007; Wu et al., 2004;
245Zyla et al., 1999). Arabinoxylan is the main NSP in cereals, representing about 60-70%
246in the cell wall endosperm cells an aleurone layer. Although AX from different sources
247differs in their substitution along the xylan backbone, a general structure can be
248assigned for AX: a backbone of β -(1,4)-linked xylose residues, which are substituted
249with arabinose residues on the C(O)-2 and/or C(O)-3 position and phenolic acids can be
250linked on the C(O)-5 position of arabinose. The structure of AX leads to high water
251holding capacity in the gastrointestinal tract resulting in high viscosity, and as a
252consequence animal production process is less efficient. Xylanases cleave AX by
253internally hydrolysing the β -1,4- β -D-xylosidic linkage between xylose residues giving
254small fragments of oligosaccharides with high or low degree of substitution (Mendis et
255al. 2016). The successful exposure of xylanase to different wheats with variations in the
256level content of soluble NSP makes them a feasible choice to mitigate the negative
257effects of arabinoxylan (AX) in monogastrics (Bedford 2000).

258

259Feed conversion ratio was not correlated with any of the analysed or predicted
260composition values of wheat without xylanase, but those supplemented with the enzyme
261had an unexpected positive correlation with the predicted contents of fat and fibre
262components (NDF, AX, soluble AX and total insoluble NSP). These findings are
263puzzling and suggest that the presence of more fibre (substrate) when the enzyme is
264present resulted in poorer performance but that the presence of this fibre in the absence
265of the enzyme was, if anything, marginally beneficial. Scott et al. (1999) found a

266significant relationship between predicted AME and FCR in wheat based diets with
267enzymes ($r = -0.46$), similarly found in this study ($r = -0.45$).

268

269Non-starch polysaccharide degrading enzymes reduce digesta viscosity in the animal by
270shortening the molecular weight of NSP and also partly remove the nutrient
271encapsulation effect of the cell wall components and, as a consequence, nutrient
272absorption is promoted and growth performance maximized (Masey O'Neill et al.,
2732012, 2014a,b; Persia et al., 2002). In this study the measured intestinal viscosity of all
274samples was extremely low in comparison with the literature, which suggests that the
275wheat samples employed were not particularly challenging from a viscosity viewpoint.
276In a similar study, xylanase supplementation improved performance in broilers fed
277different Chinese maize samples varying in chemical composition, improving the
278homogeneity in animal flocks with NSP-ases addition (Masey O'Neill et al., 2012).

279

280Some studies have reported improved performance and energy utilisation when NSP-
281enzymes are used in diets based on wheat, rye, barley (Bedford and Morgan, 1996;
282Bedford and Schulze, 1998) or maize (Masey O'Neill et al., 2012), but other studies
283have only shown improvements in animal performance without changes in nutrient
284utilisation (Hong et al., 2002; Wu et al., 2004). Wheat sample influenced ileal DM, N,
285energy utilisation and IDE. In particular, wheats 6, 7 and 8 were particularly good
286samples and this coincided with wheats 7 and 8 having the lowest viscosity. On the
287other hand, wheats 6, 7 and 8 also had lower contents of N (and predicted crude
288protein), acid detergent fiber, AX and NSP compared with the other wheats. Xylanase
289use increased energy utilisation and AME. Aside from the effect of reducing viscosity,
290there may be an additional benefit of increasing the permeability of the aleurone layer.

291This may enhance contact with digestive enzymes and their substrates, for better
292nutrient utilisation (Parkkonen et al., 1997).

293

294The interaction of the main factors for all total tract measures of nutrient utilisation was
295significant. This was mostly due to the large responses of wheats 2 and 6 to xylanase
296addition due to their comparatively low nutrient utilisation in the absence of enzyme.
297Feed conversion ratio and measured AME significantly correlated in both wheats,
298suggesting the added benefit ($r = -0.65$, $P = 0.023$ and $r = -0.49$, $P = 0.11$, respectively;
299data not shown). This observation implies that xylanase may have greater effects in
300poorer quality samples, elevating their nutritive value and thus reducing the variability
301between samples. Nonetheless, none of the results from the chemical analysis or NIRS
302predictions suggested that these two samples may have had a poorer nutritive value than
303the others. In this regard, it is noteworthy the low correlation between the predicted
304AME and the measured AME ($r = -0.16$; $P = 0.13$) suggesting the limited capacity of
305NIR to predict animal performance (data not shown). Wheats 3, 4, 5 and 8 had higher
306nutrient utilisation in the absence of enzyme, which may be due to the presence of
307endoxyylanase in the outer layer of wheat (Cleemput et al., 1997), being responsible for
308part of the degradation of AX (Dornez et al., 2006), or lesser content of xylanase
309inhibitors or both.

310

311The response of broilers to dietary intervention in general and enzyme supplementation
312in particular is usually measured using growth performance responses or ileal nutrient
313utilisation and total tract nutrient retention. These can be adequate for measuring gross
314efficiency of nutrient utilisation, but to further characterize the efficiency of nutrient
315utilisation it is important to delineate the weight gain into the composition of gain, i.e.,

316protein or fat, especially because of the differences in the efficiency with which these
317nutrients are deposited (Olukosi and Adeola, 2008). Net energy for production can be
318used as a more sensitive measure of energy utilisation by chickens receiving exogenous
319enzymes because it takes into account the efficiency of utilisation of ME for growth
320(Bhuiyan and Iji, 2015; Pirgozliev and Rose, 1999; Olukosi and Adeola, 2008; Olukosi
321et al., 2008a). Net energy for production is not only dependent on body weight but also
322on the amount of energy deposited in the carcass, which is an indication of how
323effectively the enzyme used facilitated energy utilisation. Net energy for production and
324 K_{RE} were not influenced by xylanase supplementation, but wheat sample did. Wheat
325samples 2, 7 and 8 presented better indices of energy utilisation, which may be related
326to the fact that they have the lowest viscosities compared with the other wheats. Heat
327production and Kre-Protein varied depending on wheat and xylanase inclusion.

328

329Interestingly, xylanase supplementation of wheats 2 and 6 increased total tract AME
330retention, Nep and HP but reduced K_{RE} , Kre-CP and Kre-Fat and the efficiency of
331energy use for protein and fat accretion, as has been demonstrated previously (Bhuiyan
332and Iji, 2015; Daskiran et al., 2004; Olukosi and Adeola, 2008; Olukosi et al., 2008a). In
333the current study, the comparison between animal performance and energy utilisation
334must be considered with caution as only one bird from each replicate was selected. The
335extrapolation of the performance data derived from eight animals per replicate may thus
336have some mis-alignment with the energy utilisation results obtained from one
337individual bird.

338

339The utilisation of ME was more efficient for energy deposition and less for protein and
340fat. Nevertheless the efficiency of protein accretion was almost two-fold that of fat

341 accretion, which was similarly shown by previous authors (Olukosi and Adeola, 2008;
342 Olukosi et al., 2008b). The genetics and age of birds are important factors (Leeson and
343 Summers, 1997; Lopez and Leeson, 2005). The higher proportion and retention of
344 protein than fat is likely because the young broiler chicks were still actively growing
345 and have not reached the stage at which fat deposition can overtake protein deposition
346 (Bregendahl et al., 2002; Sanz et al., 2000).

347

348 5. Conclusion

349

350 Under the current experimental conditions xylanase supplementation may compensate
351 for the poorer nutritive value of some wheats, enabling more homogenous broiler chick
352 performance. Unfortunately the predicted nutrient composition by NIR did not predict
353 accurately animal performance, and moreover taken together the predicted nutrient and
354 chemically determined contents of the wheats used in this study did not allow for
355 accurate ranking of the samples prior to feeding, which may relate to the very low
356 viscosity of the wheat samples employed. In this regard, the use of the xylanase as an
357 insurance policy is justified.

358

359 Acknowledgments

360

361 The authors acknowledge the help of Derek Brown and Irene Yuill of the Avian Science
362 Research Centre, Scotland's Rural College, Auchincruive, Ayr, for the care of the birds
363 used in the study. SRUC is supported by the Scottish Government.

364

365 Conflict of interests

366

367 All authors declare no conflict of interests.

368

369 References

370

371 Bedford MR. Exogenous enzymes in monogastric nutrition — their current value and

372 future benefits. *Anim Feed Sci Technol* 2000 Jul;86(1-2):1–13.

373 Bedford MR, Classen HL, Campbell GL. The effect of pelleting, salt, and pentosanase

374 on the viscosity of intestinal contents and the performance of broilers fed rye. *Poult*

375 *Sci* 1991;70(7):1571–7.

376 Bedford MR, Morgan AJ. The use of enzymes in poultry diets. *Worlds Poult Sci J*

377 1996;52(1):61–8.

378 Bedford MR, Schulze H. Exogenous enzymes for pigs and poultry. *Nutr Res Rev*

379 1998;11(1):91–114.

380 Bhuiyan MM, Iji PA. Energy Value of Cassava Products in Broiler Chicken Diets with

381 or without Enzyme Supplementation. *Asian-Australasian J Anim Sci*

382 2015;28(9):1317–26.

383 Bregendahl K, Sell J, Zimmerman D. Effect of low-protein diets on growth performance

384 and body composition of broiler chicks. *Poult Sci* 2002;81(8):1156–67.

385 Cleemput G, Van Laere K, Hessing M, Van Leuven F, Torrekens S, Delcour JA.

386 Identification and characterization of a novel arabinoxylanase from wheat flour.

387 *Plant Physiol* 1997;115(4):1619–27.

388 Daskiran M, Teeter RG, Fodge D, Hsiao HY. An evaluation of endo-beta-D-mannanase

389 (Hemicell) effects on broiler performance and energy use in diets varying in beta-

390 mannan content. *Poult Sci* 2004;83(4):662–8.

391Dornez E, Joye IJ, Gebruers K, Delcour JA, Courtin CM. Wheat-kernel-associated
392 endoxylanases consist of a majority of microbial and a minority of wheat
393 endogenous endoxylanases. *J Agric Food Chem* 2006;54(11):4028–34.

394Hong D, Burrows H, Adeola O. Addition of enzyme to starter and grower diets for
395 ducks. *Poult Sci* 2002;81(12):1842–9.

396van Kempen TAT., Simmins P. Near-infrared reflectance spectroscopy in precision feed
397 formulation. *J Appl Poult Res* 1997;6:471–7.

398Leeson S, Summers J. Feeding programs for broilers. In: Books U, editor. *Commer*
399 *Poult Nutr*. 2nd ed. Guelph, Canada; 1997 p. 207–54.

400Lopez G, Leeson S. Utilization of metabolizable energy by young broilers and birds of
401 intermediate growth rate. *Poult Sci* 2005;84(7):1069–76.

402Masey O’Neill H V, Liu N, Wang JP, Diallo A, Hill S. Effect of xylanase on
403 performance and apparent metabolisable energy in starter broilers fed diets
404 containing one maize variety harvested in different regions of china. *Asian-*
405 *Australasian J Anim Sci* 2012;25(4):515–23.

406Masey O’Neill H V, Smith JA, Bedford MR. Multicarbohydase Enzymes for Non-
407 ruminants. *Asian-Australasian J Anim Sci* 2014;27(2):290–301.

408Masey-O’Neill H V, Singh M, Cowieson AJ. Effects of exogenous xylanase on
409 performance, nutrient digestibility, volatile fatty acid production and digestive tract
410 thermal profiles of broilers fed on wheat- or maize-based diet. *Br Poult Sci*
411 2014;55(3):351–9.

412Mendis M, Leclerc E, Simsek S. Arabinoxylans, gut microbiota and immunity.
413 *Carbohydr Polym* 2016;139:159–66.

414Olukosi O, Bedford M, Adeola O. Xylanase in diets for growing pigs and broiler chicks.
415 *Can J Anim Sci* 2007;87:227–35.

416Olukosi O, Cowieson A, Adeola O. Energy utilization and growth performance of
417 broilers receiving diets supplemented with enzymes containing carbohydrase or
418 phytase activity individually or in combination. *Br J Nutr* 2008a;99:682–90.

419Olukosi O, Paton N, Kempen T, Adeola O. Short Communication: An investigation of
420 the use of near infrared reflectance spectroscopy to predict the energy value of
421 meat and bone meal for swine. *Can J Anim Sci* 2011;91(3):405–9.

422Olukosi OA, Adeola O. Whole Body Nutrient Accretion, Growth Performance and Total
423 Tract Nutrient Retention Responses of Broilers to Supplementation of Xylanase
424 and Phytase Individually or in Combination in Wheat-Soybean Meal Based Diets. *J*
425 *Poult Sci* 2008;45(3):192–8.

426Olukosi OA, Cowieson AJ, Adeola O. Influence of enzyme supplementation of maize-
427 soyabean meal diets on carcass composition, whole-body nutrient accretion and
428 total tract nutrient retention of broilers. *Br Poult Sci* 2008b;49(4):436–45.

429Owens B, McCann MEE, McCracken KJ, Park RS. Prediction of wheat chemical and
430 physical characteristics and nutritive value by near-infrared reflectance
431 spectroscopy. *Br Poult Sci* 2009;50(1):103–22.

432Parkkonen T, Tervilä-Wilo A, Hopekoski-Nurminen M, Morgan A, Poutanen K, Autio
433 K. Changes in wheat micro structure following in vitro digestion. *Acta Agric Scand*
434 *Sect B - Soil Plant Sci* 1997;47(1):43–7.

435Persia M, Dehority B, Liburn M. The effects of enzyme supplementation of corn-and
436 wheat-based diets on nutrient digestion and cecal microbial populations in turkeys.
437 *J Appl Poult Res* 2002;11:134–45.

438Pirgozliev V, Rose SP. Net energy systems for poultry feeds: A quantitative review.
439 *Worlds Poult Sci J* 1999;55(1):23–36.

440Sanz M, Lopez-Bote CJ, Menoyo D, Bautista JM. Abdominal fat deposition and fatty

441 acid synthesis are lower and beta-oxidation is higher in broiler chickens fed diets
442 containing unsaturated rather than saturated fat. *J Nutr* 2000;130(12):3034–7.

443 Scott TA, Silversides FG, Classen HL, Swift ML, Bedford MR. Prediction of the
444 performance of broiler chicks from apparent metabolizable energy and protein
445 digestibility values obtained using a broiler chick bioassay. *Can J Anim Sci*
446 1999;79(1):59–64.

447 Short FJ, Gorton P, Wiseman J, Boorman KN. Determination of titanium dioxide added
448 as an inert marker in chicken digestibility studies. *Anim Feed Sci Technol*
449 1996;59(4):215–21.

450 Theander O, Westerlund E, Åman P, Graham H. Plant cell walls and monogastric diets.
451 *Anim Feed Sci Technol* 1989;23(1-3):205–25.

452 Wu Y, Ravindran V, Thomas D, Birtles M, Hendriks W. Influence of method of whole
453 wheat inclusion and xylanase supplementation on the performance, apparent
454 metabolisable energy, digestive tract measurements and gut morphology of
455 broilers. *Br Poult Sci* 2004;45:385–94.

456 Yegani M, Korver D. Review: Prediction of variation in energetic value of wheat for
457 poultry. *Can J Anim Sci* 2012;92:261–73.

458 Zyla K, Gogol D, Koreleski J, Swiatkiewicz S, Ledoux DR. Simultaneous application of
459 phytase and xylanase to broiler feeds based on wheat: in vitro measurements of
460 phosphorus and pentose release from wheats and wheat-based feeds. *J Sci Food*
461 *Agric* 1999;79:1832–40.

462

464Table 1 Ingredient and calculated composition as-fed of the experimental diets

Item	Control	+ Xylanase
Ingredient, g/kg		
Wheat - feed	585	585
Soybean meal 48	325	325
Soy oil	44.4	44.4
Salt	3.00	3.00
Sodium bicarbonate	1.87	1.87
DL-methionine	2.99	2.99
Lysine HCl	2.46	2.46
Threonine	0.77	0.77
Limestone	7.86	7.86
Dicalcium phosphate	15.5	15.5
Vitamin premix ¹	4.90	4.90
Phytase ²	+	+
Xylanase ³	-	+
Calculated nutrient composition, %		
Crude protein	22.4	22.4
Ca	0.90	0.90
P	0.74	0.74
Available phosphorous	0.45	0.45
Fat	5.72	5.72
Fibre	2.55	2.55
Met	0.62	0.62
Cys	0.38	0.38
Met + Cys	1.00	1.00
Lys	1.35	1.35
His	0.55	0.55
Trp	0.28	0.28
Thr	0.88	0.88
Arg	1.45	1.45
Ile	0.92	0.92
Leu	1.64	1.64
Phe	1.05	1.05
Val	1.00	1.00
AME, MJ/kg	12.8	12.8

465AME = apparent metabolisable energy.

466¹ Vitamin/mineral premix supply per kilogram of diet: vitamin A, 16,000 IU; vitamin

467D₃, 3000 IU; vitamin E, 25 IU; vitamin B₁, 3 mg; vitamin B₂, 10 mg; vitamin B₆, 3 mg;

468vitamin B₁₂, 15 µg; nicotinic acid, 60 mg; pantothenic acid, 14.7 mg; folic acid, 1.5 mg;

469biotin, 125 µg; choline chloride, 25 mg; Fe as iron sulfate, 20 mg; Cu as copper sulfate, 47010 mg; Mn as manganous oxide, 100 mg; Co as cobalt oxide, 1.0 mg; Zn as zinc oxide, 47182.222 mg; I as potassium iodide, 1 mg; Se as sodium selenite, 0.2 mg; and Mo as 472molybdenum oxide, 0.5 mg.

473² Quantum Blue 5G, AB Vista, Marlborough, UK; 5000 FTU/g.

474³ Econase XT 25P, AB Vista, Marlborough, UK; 160,000 BXU/g.

475Table 2 Analysed nutrient composition and coefficient of variation (CV) of the wheat

476samples

Item	Wheat samples								CV
	1	2	3	4	5	6	7	8	
Gross energy, MJ/kg	18.0	18.1	18.1	18.0	18.2	17.9	18.0	18.1	<1
Viscosity, cP	10.5	8.50	12.8	13.0	11.3	11.2	7.60	7.80	21
Dry matter, %	87.2	87.4	87.8	87.5	87.1	87.2	88.6	87.6	<1
Fat, %	1.49	1.37	1.48	1.37	1.26	1.15	1.24	1.94	17
Nitrogen, %	2.22	1.88	2.37	2.10	2.02	1.79	1.55	1.79	13
Calcium, %	0.03	0.03	0.03	0.03	0.03	0.02	0.02	0.05	31
Phosphorous, %	0.28	0.32	0.34	0.33	0.38	0.29	0.27	0.33	11
Phytic acid, %	0.75	0.77	0.64	0.72	0.81	0.92	0.53	0.53	19

477

478

479Table 3 Nutrient composition predicted by near-infrared spectroscopy (NIRS) and
480coefficients of variation (CV) of the wheat samples

Item	Wheat samples								CV
	1	2	3	4	5	6	7	8	
Energy, MJ/kg and ether extract, %									
GE	16.5	16.5	16.6	16.6	16.6	16.4	16.3	16.4	<1
AME	13.4	13.4	13.2	13.0	13.3	13.6	13.9	13.4	2
Fat	2.10	2.23	2.16	2.40	2.28	2.25	1.95	2.13	6
Fibre									
NDF	15.1	16.4	16.1	19.0	17.2	16.1	14.0	16.1	9
ADF	2.44	2.82	2.94	3.94	3.15	2.99	2.11	3.02	18
Lignin	0.72	0.97	0.89	1.15	1.06	1.02	0.95	0.94	13
AX	7.66	8.10	7.97	9.19	8.48	7.83	7.30	8.02	7
Soluble AX	0.56	0.61	0.57	0.62	0.63	0.59	0.58	0.57	4
β -glucan	1.21	1.66	1.66	2.36	1.88	1.63	1.78	1.90	18
Total insoluble NSP	10.2	11.4	11.1	13.2	12.1	11.1	10.4	11.3	8
Total soluble NSP	1.84	2.47	2.36	3.23	2.75	2.43	2.56	2.64	15
Protein,% and amino acid profile, g/100 g CP									
CP	13.4	11.44	13.6	11.7	11.9	10.6	8.36	10.2	15
Lysine	3.01	3.35	2.96	3.42	3.26	3.18	3.07	3.19	5
Methionine	1.56	1.68	1.60	1.65	1.69	1.70	1.78	1.67	4
Leucine	7.37	6.64	7.07	6.47	6.43	6.98	7.22	7.10	5
Threonine	3.30	3.42	3.25	3.42	3.34	3.41	3.41	3.42	2
Tryptophan	1.16	1.21	1.16	1.21	1.19	1.22	1.30	1.27	4
Tyrosine	3.21	3.13	3.20	3.09	3.12	3.23	3.28	3.23	2
Valine	4.75	4.91	4.76	4.96	4.86	4.90	4.99	4.95	2
Phenylalanine	4.43	4.39	4.55	4.53	4.47	4.44	4.53	4.50	1
Histidine	2.57	2.62	2.59	2.61	2.63	2.62	2.60	2.58	<1
Isoleucine	3.48	3.43	3.53	3.45	3.46	3.47	3.46	3.46	<1
Arginine	5.18	5.43	5.09	5.52	5.33	5.07	4.54	4.99	6
Alanine	4.28	3.76	4.11	3.89	3.67	4.04	4.33	4.22	6
Asparagine	6.06	6.22	5.79	6.27	5.98	6.04	5.59	6.00	4
Cysteine	2.19	2.34	2.20	2.27	2.34	2.33	2.40	2.32	3
Glutamine	24.9	23.5	26.0	23.2	24.7	24.1	24.0	23.4	4
Glycine	3.99	4.27	3.98	4.19	4.23	4.22	4.19	4.15	3
Proline	9.23	8.99	9.78	9.23	9.39	9.28	9.91	9.40	3
Serine	4.88	4.83	4.83	4.70	4.79	4.84	4.81	4.78	1

481GE= gross energy; AME = apparent metabolisable energy; NDF = neutral detergent

482fibre; ADF = acid detergent fibre; AX = arabinoxylan; NSP = non-starch

483polysaccharides; CP = crude protein.

484

485

486

487

488

489

490

491

492

493

494

495

496

497

498

499

500

501

502

503

504

505

49

50

507 Table 4 Animal performance and jejunal digesta viscosity¹

Item	Weight gain, g/bird	Feed intake, g/bird	Feed conversion ratio, g/g	Jejunal viscosity, cP
Wheat effect				
1	824	1105	1.341	2.81
2	816	1103	1.353	3.13
3	820	1087	1.329	2.89
4	817	1097	1.345	3.01
5	781	1087	1.394	2.77
6	791	1079	1.369	2.94
7	787	1049	1.341	2.40
8	791	1065	1.349	2.67
SEM	16	16	0.019	0.06
Xylanase effect				
0 BXU/kg	793 ^b	1088	1.373 ^a	3.32 ^a
16,000 BXU/kg	813 ^a	1079	1.331 ^b	2.34 ^b
SEM	8	8	0.010	0.03
<i>P</i> -value				
Wheat	0.465	0.192	0.367	0.380
Xylanase	0.072	0.496	0.003	<0.001
Interaction	0.951	0.950	0.894	0.845

508^{a,b} Means in the same column with different letters differ at $P < 0.05$.

509¹ Mean values for six replicate cages with eight broilers per replicate cage.

510 Table 5 Correlation of feed conversion ratio (FCR) with the analysed chemical
 511 composition and the predicted values by near-infrared spectroscopy (NIRS) of wheat in
 512 diets supplemented with or without xylanase

Item	Pearson's Correlation	
	coefficients with FCR	
	Without xylanase	With xylanase
Analysed composition		
GE	-0.27	0.38
Fat	0.14	-0.26
Nitrogen	-0.47	0.07
Calcium	-0.43	0.36
Phosphorous	-0.35	0.70*
NIRS predicted composition		
CP	0.06	0.07
Fat	-0.27	0.68*
GE	-0.09	0.49
AME	0.53	-0.45
ADF	-0.34	0.63
NDF	-0.31	0.69*
Total AX	-0.36	0.73*
Soluble AX	0.25	0.85*
β-glucan	-0.34	0.55
Lignin	0.07	0.62
Total insoluble NSP	-0.26	0.74*
Total soluble NSP	-0.23	0.60

513 GE = gross energy; NIRS = near-infrared spectroscopy; CP = crude protein; AME =
 514 apparent metabolisable energy; ADF = acid detergent fibre; NDF = neutral detergent
 515 fibre; AX = arabinoxylan; NSP = non-starch polysaccharides.

516 * $P < 0.05$

517Table 6 Ileal nutrient utilisation of nutrients¹

Item	Dry matter, %	Nitrogen, %	Energy, %	IDE, MJ/kg
Wheat effect				
1	68.0 ^{bc}	78.0	70.8 ^{bc}	13.2 ^b
2	66.9 ^{bc}	74.8	70.0 ^c	13.1 ^b
3	65.2 ^c	74.4	69.5 ^c	13.2 ^b
4	68.8 ^{bc}	78.2	72.0 ^{abc}	13.7 ^{ab}
5	66.9 ^{bc}	75.9	69.6 ^c	13.0 ^b
6	70.2 ^{ab}	78.1	73.2 ^{abc}	13.8 ^{ab}
7	70.4 ^{ab}	79.4	73.9 ^{ab}	14.0 ^a
8	73.0 ^a	79.7	76.0 ^a	14.4 ^a
SEM	1.47	1.42	1.47	0.28
Xylanase effect				
0 BXU/kg	67.8	76.9	70.9 ^b	13.35 ^b
16,000 BXU/kg	69.5	77.8	72.9 ^a	13.77 ^a
SEM	0.74	0.71	0.74	0.14
<i>P</i> -value				
Wheat	0.012	0.062	0.019	0.004
Xylanase	0.111	0.205	0.057	0.039
Interaction	0.550	0.104	0.571	0.577

518IDE = ileal utilization of energy.

519^{a-c} Means in the same column with different letters differ at $P < 0.05$.

520¹Mean values for six replicate cages with eight broilers per replicate cage.

521

522

523

524 Table 7 Total tract retention of nutrients¹

Wheat & Xylanase effect		Dry matter, %	Nitrogen, %	AME, MJ/kg	AMEn, MJ/kg
Wheat	Xylanase, BXU/kg				
1	0	69.4 ^{cde}	62.9 ^{de}	13.5 ^{ef}	13.0 ^e
1	16,000	71.2 ^{bc}	60.7 ^{ab}	14.1 ^{cde}	13.5 ^{cd}
2	0	65.4 ^g	57.9 ^f	12.9 ^h	12.4 ^g
2	16,000	73.4 ^a	65.7 ^a	14.5 ^{ab}	14.0 ^{ab}
3	0	68.6 ^{de}	58.1 ^{cd}	13.8 ^{fg}	13.2 ^e
3	16,000	68.4 ^e	57.4 ^{cd}	13.8 ^{fg}	13.2 ^{ef}
4	0	71.5 ^{abc}	63.3 ^{abc}	14.2 ^{abc}	13.8 ^{abc}
4	16,000	70.6 ^{cd}	62.7 ^{cd}	14.1 ^{cd}	13.6 ^{cd}
5	0	69.6 ^{cde}	64.6 ^e	13.5 ^{def}	13.0 ^{de}
5	16,000	66.1 ^{fg}	57.7 ^{ef}	13.0 ^{gh}	12.4 ^{fg}
6	0	65.2 ^g	55.1 ^{ef}	13.0 ^h	12.4 ^g
6	16,000	73.1 ^{ab}	66.9 ^{abc}	14.4 ^{ab}	13.9 ^{ab}
7	0	68.0 ^{ef}	62.0 ^{ef}	13.6 ^{fg}	13.1 ^{ef}
7	16,000	69.9 ^{cde}	65.3 ^e	14.1 ^{ef}	13.7 ^e
8	0	72.8 ^{ab}	65.8 ^{abc}	14.5 ^a	14.0 ^a
8	16,000	71.4 ^{abc}	64.1 ^{abc}	14.2 ^{bc}	13.7 ^{bc}
Pooled SEM		0.73	0.79	0.12	0.12
P-value					
Wheat		<0.001	<0.001	<0.001	<0.001
Xylanase		<0.001	<0.001	<0.001	<0.001
Interaction		<0.001	<0.001	<0.001	<0.001

525 AME = apparent metabolisable energy; AMEn = AME corrected for nitrogen ;

526^{a-e} Different letters mean significant differences between treatments, highlighting the

527 statistical interaction between main factors wheat x xylanase ($P < 0.05$).

528¹ Mean values for six replicate cages with three broilers per replicate cage.

529

530

531

532

533

534Table 8 Energy utilisation, energy retained and efficiencies of energy use¹

Item	Energy utilisation, MJ/kg		Energy retained, MJ/kg		Efficiencies of energy use for energy, protein and fat retention accretion		
	Nep ²	HP ³	Protein-ER ⁴	Fat-ER ⁵	K _{RE} ⁶	Kre-Protein ⁷	Kre-Fat ⁸
Wheat effect							
1	5.59 ^{abc}	6.84 ^{bc}	3.92 ^{ab}	2.20 ^{bc}	0.45 ^{bc}	0.275 ^{ab}	0.154 ^{abcd}
2	5.94 ^a	6.35 ^d	3.89 ^{ab}	2.48 ^a	0.48 ^a	0.275 ^{ab}	0.175 ^a
3	5.53 ^{abc}	6.54 ^{cd}	3.94 ^a	2.04 ^c	0.46 ^{ab}	0.289 ^a	0.149 ^{bcd}
4	5.53 ^c	7.24 ^a	3.95 ^a	2.04 ^c	0.43 ^c	0.269 ^{bc}	0.139 ^d
5	5.32 ^c	6.74 ^{bc}	3.96 ^a	2.05 ^c	0.44 ^{bc}	0.277 ^{ab}	0.143 ^{cd}
6	5.64 ^{abc}	6.60 ^{bcd}	3.89 ^{ab}	2.24 ^{abc}	0.46 ^{ab}	0.275 ^{ab}	0.158 ^{abcd}
7	5.72 ^{bc}	6.44 ^{bcd}	3.83 ^b	2.25 ^{abc}	0.47 ^b	0.278 ^{ab}	0.163 ^{ab}
8	5.77 ^{ab}	7.22 ^{ab}	3.84 ^b	2.41 ^{ab}	0.44 ^{bc}	0.258 ^c	0.162 ^{abc}
SEM	0.135	0.155	0.031	0.098	0.011	0.0050	0.0078
Xylanase effect							
0 BXU/kg	5.54	6.68	3.92	2.18	0.454	0.275	0.153
16,000 BXU/kg	5.72	6.81	3.89	2.24	0.457	0.274	0.158
SEM	0.068	0.077	0.016	0.049	0.005	0.0025	0.0039
P-value							
Wheat	0.029	0.001	0.032	0.008	0.012	0.017	0.029
Xylanase	0.950	0.060	0.135	0.373	0.327	0.869	0.354

Interaction	0.198	0.018	0.429	0.922	0.284	0.006	0.984
-------------	-------	-------	-------	-------	-------	-------	-------

535¹Mean values for six replicate cages with one broiler per replicate cage.

536² NEp - net energy for production.

537³ HP - heat production.

538⁴ Protein-ER - energy retained as protein.

539⁵ Fat ER - energy retained as fat.

540⁶ K_{RE} - efficiency of energy use for production.

541⁷ Kre-Protein - efficiency of energy use for protein accretion.

542⁸ Kre-Fat - efficiency of energy use for fat accretion.

543^{a-d} Means in the same column with different letters differ at $P < 0.05$.