

SEASONAL INCIDENCE AND HORIZONTAL DISTRIBUTION PATTERNS OF OVIPOSITION BY *Aedes aegypti* IN AN URBAN ENVIRONMENT IN TRINIDAD, WEST INDIES

DAVE D. CHADEE

Insect Vector Control Division, Ministry of Health, 3 Queen Street, St. Joseph, Trinidad, W.I.

ABSTRACT. The oviposition patterns of *Aedes aegypti* in ovitraps placed along 5 horizontal transects were monitored weekly for 52 wk (January to December 1988) in St. Joseph, Trinidad. Of the 2,550 ovitraps exposed, 270 were lost and 1,177 (52%) out of 2,280 were used by gravid females, containing 38,118 eggs. During the dry season 43% (16,265 eggs) of the eggs were collected whereas during the wet season 57% (21,853 eggs) were collected. Ovitrap exposed under eaves, under houses and in the open yard attracted similar oviposition occurrences and proportions of eggs. Inspections of the 52 properties within St. Joseph revealed that 17 (House Index = 32.7%) harbored *Ae. aegypti* pre-adult stages. The container index was 5.5%. This study revealed no evidence of behavioral changes in the container preferences of *Ae. aegypti* in St. Joseph, Trinidad, after 12 years of treating with fenthion and temephos.

INTRODUCTION

Small black jars designed for use as "ovitrap" (Fay and Eliason 1966) have become a standard surveillance tool for *Aedes aegypti* (Linn.) but the criteria for their placement at specific locations are not clear. Various workers have suggested that in urban surveys, ovitraps should be placed at ground level in backyards, near other oviposition sites, avoiding exposure to excessive rain, sunlight or wind (Fay and Eliason 1966, Jakob and Bevier 1969a).

Further guidelines were given by Jakob and Bevier (1969a) for placement of ovitraps in the field, but most workers continue to place ovitraps using other criteria, such as along transects within cities (Jakob and Bevier 1969b, Tanner 1969).

From 1976 to 1987, an intensive *Ae. aegypti* eradication program was carried out in Trinidad, involving source reduction and insecticide treatments in both domestic and peridomestic environments. McClelland (1967) suggested that such insecticide application may cause shifts in the site selection of *Ae. aegypti*, from treated to untreated and atypical oviposition sites.

The present study was conducted to determine the container preferences of *Ae. aegypti* after larvicidal applications of temephos and fenthion for 12 years, and to hopefully ascertain the most suitable sites for ovitraps under field conditions in one locality, St. Joseph, Trinidad, West Indies.

MATERIALS AND METHODS

Study area: Ovitrap surveys of *Ae. aegypti* were conducted from January to December 1988 in the small town of St. Joseph (10° 30'N, 61° 25'W). Details of the study area, meteorology and *Ae. aegypti* populations have been described by Chadee and Corbet (1987).

Ovitraping: Five sites (A-E) were selected, each site consisting of 2 houses raised on stilts and separated by an exposed yard space. At each site 10 ovitraps were placed at ground level along an east-west transect: 4 traps under the houses, 3 under eaves and 3 exposed in the yard. Each ovitrap consisted of a cylindrical, black plastic cup (13 cm height × 9 cm diam), containing 450 ml of tap water and a removable paddle (Chadee and Corbet 1987). An overflow hole, 3 cm from the top, prevented the ovitrap being filled to the brim. The location of each transect and ovitrap was coded A to E and 1 to 10, respectively, making it possible to monitor the distribution of oviposition among the different sites.

All ovitraps were inspected and serviced weekly for 52 consecutive weeks from January to December 1988. At each inspection, the code-numbered paddle was removed, the ovipot itself scrubbed (to remove any eggs), refilled with 450 ml tap water and reset with an egg-free paddle, rough-side exposed. Each paddle retrieved was placed in a plastic bag and taken to the Insect Vector Control Division (IVCD) laboratory where all mosquito eggs on them were examined under a microscope (×40), counted and identified by the characteristic chorionic pattern of *Ae. aegypti* (Pratt and Kidwell 1969). Counts of *Ae. aegypti* eggs were compared for each transect, as well as individual traps within the transect and for the 3 types of locations: under houses, under eaves and in open yards. The results were analyzed with a G-test by transforming the data into contingency tables (Sokal and Rohlf 1980) to determine location preferences and oviposition patterns.

Container preferences: In a general survey of the town, 52 house properties were inspected weekly by 6 mosquito evaluators to determine the numbers, types, positions and conditions of containers that might serve as breeding sites.

Table 1. Monthly rainfall and oviposition patterns of *Aedes aegypti* collected from St. Joseph, Trinidad, in 1988.

Month	No. ovitraps positive/set	%	No. eggs recovered	Rainfall (mm)
<i>Dry season (1988)</i>				
December	169/200	85	6,712	48.4
January	19/200	10	505	37.2
February	133/250	53	4,789	4.1
March	75/200	38	1,754	8.5
April	100/200	50	2,505	69.7
Sub-total	496/1,050	47	16,265	167.9
<i>Wet season</i>				
May	68/250	27	1,560	287.2
June	66/200	33	1,935	276.7
July	92/200	46	2,555	286.1
August	156/200	78	5,572	252.6
September	65/200	33	2,209	488.4
October	98/200	49	2,619	216.7
November	136/250	54	5,403	191.2
Sub-total	681/1,500	45	21,853	1,998.9
Total	1,177/2,550	46	38,118	2,166.8

Table 2. Oviposition of *Ae. aegypti* in the yard, under eaves and under houses in St. Joseph, Trinidad (January to December 1988).

	Localities			
	Yard	Under eaves	Under house	Total
No. traps visited	765	765	1,020	2,550*
No. of positive traps	376	349	452	1,177
% positive	49	46	44	46
Total no. of eggs laid	11,391	11,591	15,136	38,118
Mean no. of eggs/positive ovitrap	30	33	33	32
% of total eggs collected	30	30.4	39.7	100.0

* 270 traps lost.

Table 3. Locations within premises of *Aedes aegypti* preferred containers in St. Joseph, Trinidad, in 1988.

Types of containers	No. of containers	No. positive	%	Locations	
				Under eaves	Yard
Drums	132	38	29	25	13
Tanks and cisterns	33	9	27	7	2
Buckets and tubs	107	0	0	0	0
Tires	100	0	0	0	0
Small miscellaneous containers*	518	2	0.4	0	2
Total	890	49	5.5	32	17

* Cans, pans, tins, etc.

Samples of larvae and pupae from all positive containers were taken to the IVCD laboratory for identification.

RESULTS

During 1988, the monthly number of eggs obtained from the exposed ovitraps ranged from 505 to 6,712, with a mean of 3,177 eggs per month (Table 1). Rainfall totalled 2,167 mm, averaging 34 mm/month in the dry season (De-

ember-April) and 286 mm/month in the wet season (May-November) (Table 1).

During 2,550 ovitrap-weeks along 5 transects, 270 ovitraps were lost. A total of 38,118 *Ae. aegypti* eggs was found in 1,177/2,280 (52%) of ovitraps. The seasonal pattern of oviposition fluctuated from 9.5% of ovitraps positive in January to 85% of ovitraps positive in December and with a significant difference ($G = 54.8$; d.f. = 2; $P < 0.001$) in the oviposition rates between the wet and dry seasons (Table 1). No significant

differences were detected between the 5 ovitrapping transects ($G = 1.03$) and similar population densities of *Ae. aegypti* were present at all ovitrapping transects.

Table 2 summarizes the ovitrap results. All the mosquito eggs collected were identified as *Ae. aegypti*. No significant ($G = 0.41$) differences were detected in the number of eggs or occurrence of positive ovitraps in the open yard, under eaves or under houses [G-test by transforming the data into 3×3 contingency tables (Sokal and Rohlf 1980)]. The mean number of *Ae. aegypti* was 30–33 eggs per positive ovitrap during both the wet and dry seasons (Table 2).

In the 52 house premises survey, 890 containers were identified as potential breeding sites for *Ae. aegypti*, mainly drums (14%), tubs and buckets (12%), tires (11%), cisterns and tanks (4%) and small miscellaneous containers. The House index was $17/52 = 32.7\%$ positive, with 49 positive containers (Container index 5.5%). Although potential breeding sites were also found on the periphery of house lots, only *Culex quinquefasciatus* Say immatures were collected.

Drums represented the most favored container used by *Ae. aegypti* with 38/49 (78%) being positive. However, of the 132 drums found, 93 and 39 were found in the yard and under eaves, respectively. Of these, 13 (35%) were found positive in the yard and 25 (65%) under the eaves. The combined results show that 38/132 (29%) of drums were positive (Table 3).

DISCUSSION

Gravid *Ae. aegypti* search extensively within premises for oviposition sites and their flight is not limited to within houses but also covers open spaces (yards) in Trinidad. Similar dispersion patterns have been recorded during *Ae. aegypti* mark-release-recapture studies showing a relatively short flight range of 23–50 m (Morlan and Hayes 1958, Nayar 1981).

In Trinidad, the *Ae. aegypti* population is domestic in habit and genetically heterogenous (Wallis et al. 1984). After 12 years of insecticide applications, there is no evidence of major changes in the *Ae. aegypti* populations in St. Joseph, Trinidad, since, all of the positive ovitraps and artificial containers (drums, cans, tyres) were found within urban premises. A similar pattern has been reported in Puerto Rico (Moore 1983).

The number of eggs laid per positive ovitrap during any one week was highly variable, 4–337 eggs in ovitraps under eaves, 1–261 eggs in ovitraps under houses and 1–240 eggs in ovitraps in the yard. But, despite the differences in the

physical locations of these ovitraps, the average number of eggs laid per positive ovitrap per week remained almost the same during both seasons, ranging from 30 to 33. A similar average number of eggs per positive ovitrap was found in Texas, despite variations in the total numbers of eggs collected per month (Micks and Moon 1980). Similar numbers of eggs (less than 30 per positive ovitrap) were collected in ovitraps exposed for 2-h intervals during diel oviposition periodicity studies conducted in Trinidad (see Fig. 5 in Chadee and Corbet 1987).

Evidently females laid only a part of their full egg complement in any given ovitrap, since the fecundity of a nulliparous *Ae. aegypti* ranges from 60 to 120 eggs (Christophers 1960, Gillett 1962), which is far more than the mean number of eggs/ovitrap/week. Our data indicate that *Ae. aegypti* females oviposit in multiple sites and each ovitrap probably received eggs from more than one female.

In St. Joseph, the equivalent use of ovitraps in each of the 3 sampling situations suggests that environmental factors such as the exposure to direct sunlight is irrelevant to the placement of ovitraps. Corbet and Chadee (1990) similarly showed no effect of sunlight on oviposition, in that no difference was detected in oviposition patterns in traps placed in sites facing east or west. These findings are probably related to the fact that peak oviposition occurs mainly at 600–800 and 1600–1800 hours (Chadee and Corbet 1987). Even so ovitrap placement in partial shade, as proposed by Fay and Eliason (1966), Jakob and Bevier (1969a) and others, has the advantage of reducing desiccation.

The difference in oviposition levels found between drums and ovitraps exposed to the sun and in the shade may have been due not to light levels *per se* but perhaps to differences in temperature and humidity, in view of the different material (steel vs. plastic) and volume (208 liters vs. 450 ml) of the drums as opposed to ovitraps.

The results of the present study raise important questions regarding the positioning of ovitraps for adequate surveillance of *Ae. aegypti*. Tonn et al. (1969) and Macdonald and Rajapaksa (1972) working in Thailand and Borneo, respectively, concentrated on between-house studies and did not take the positioning of containers in and around houses into consideration, though the latter suggested that such information was desirable.

ACKNOWLEDGMENTS

I wish to thank R. Paul, Specialist Medical Officer, Insect Vector Control Division, Ministry of Health for valued help. In addition, I also

thank R. Ganesh, R. Sedino, N. Andalcio, A. Lakhan and L. Fernando for laboratory and field assistance. Special thanks are due to J. M. Sutherland for reviewing the manuscript.

REFERENCES CITED

- Chadee, D. D. and P. S. Corbet. 1987. Seasonal incidence and diel patterns of oviposition in the field of the mosquito, *Aedes aegypti* (L.) (Diptera: Culicidae) in Trinidad, West Indies: a preliminary study. *Ann. Trop. Med. Parasitol.* 81:151-161.
- Christophers, R. S. 1960. *Aedes aegypti* (L.) the yellow fever mosquito. Its life history, bionomics and structure. Cambridge Univ. Press.
- Corbet, P. S. and D. D. Chadee. 1990. Incidence and diel pattern of oviposition outdoors of the mosquito, *Aedes aegypti* (L.) (Diptera: Culicidae) in Trinidad, West Indies in relation to solar aspect. *Ann. Trop. Med. Parasitol.* 84:63-78.
- Fay, R. W. and D. A. Eliason. 1966. A preferred oviposition site as a surveillance method for *Aedes aegypti*. *Mosq. News* 26:531-535.
- Gillett, J. D. 1962. Contributions to the oviposition cycle by individual mosquitoes in a population. *J. Insect Physiol.* 8:665-681.
- Jakob, W. L. and G. A. Bevier. 1969a. Application of ovitraps in the U.S. *Aedes aegypti* eradication program. *Mosq. News* 29:55-62.
- Jakob, W. L. and G. A. Bevier. 1969b. Evaluation of ovitraps in the U.S. *Aedes aegypti* eradication program. *Mosq. News* 29:650-653.
- Macdonald, W. W. and N. Rajapaksa. 1972. A survey of the distribution and relative prevalence of *Aedes aegypti* in Sabah, Brunei, and Sarawak. *Bull. W.H.O.* 46:203-209.
- McClelland, G. A. H. 1967. Problems of interaction and density in mosquitoes in relation to *Aedes aegypti* eradication. *Proc. Calif. Mosq. Control Assoc.* 35:94-95.
- Micks, D. W. and W. B. Moon. 1980. *Aedes aegypti* in a Texas coastal county as an index of dengue fever receptivity and control. *Am. J. Trop. Med. Hyg.* 29:1382-1388.
- Moore, C. G. 1983. Habitat differences among container-breeding mosquitoes in Western Puerto Rico (Diptera: Culicidae). *Pan-Pacific Entomol.* 59:218-228.
- Morlan, H. B. and R. O. Hayes. 1958. Urban dispersal and activity of *Aedes aegypti*. *Mosq. News* 18:137-144.
- Nayar, J. K. 1981. *Aedes aegypti* (L.) (Diptera: Culicidae): observations on dispersal, survival, insemination, ovarian development and oviposition characteristics of a Florida population. *J. Fla. Anti-Mosq. Assoc.* 52:24-40.
- Pratt, H. D. and A. S. Kidwell. 1969. Eggs of mosquitoes found in *Aedes aegypti* oviposition traps. *Mosq. News* 29:545-548.
- Sokal, R. R. and F. J. Rohlf. 1980. *Biometry. The principles and practice of statistics in biological research.* (2nd edition). Freeman, San Francisco.
- Tanner, G. D. 1969. Oviposition traps and population sampling for the distribution of *Aedes aegypti* (L.). *Mosq. News* 29:116-121.
- Tonn, R. J., P. M. Sheppard, W. W. Macdonald and Y. H. Bang. 1969. Replicate surveys of larval habitats of *Aedes aegypti* in relation to Dengue Haemorrhagic fever in Bangkok, Thailand. *Bull. W.H.O.* 40:819-829.
- Wallis, G. P., W. J. Tabachnik and J. R. Powell. 1984. Genetic heterogeneity among Caribbean populations of *Aedes aegypti*. *Am. J. Trop. Med. Hyg.* 33:492-498.