



Open Archive TOULOUSE Archive Ouverte (OATAO)

OATAO is an open access repository that collects the work of Toulouse researchers and makes it freely available over the web where possible.

This is an author-deposited version published in : <http://oatao.univ-toulouse.fr/>
Eprints ID : 11430

To link to this article : DOI:10.1371/journal.pone.0084203
<http://dx.doi.org/10.1371/journal.pone.0084203>

To cite this version Zouine, Mohamed and Fu, Yongyao and Chateigner-Boutin, Anne-Laure and Mila, Isabelle and Frasse, Pierre and Wang, Hua and Audran-Delalande, Corinne and Roustan, Jean-Paul and Bouzayen, Mondhe *Characterization of the Tomato ARF Gene Family Uncovers a Multi-Levels Post-Transcriptional Regulation Including Alternative Splicing*. (2014) PLoS ONE, vol. 9 (n° 1). ISSN 1932-6203

Any correspondence concerning this service should be sent to the repository administrator: staff-oatao@listes-diff.inp-toulouse.fr

Characterization of the Tomato *ARF* Gene Family Uncovers a Multi-Levels Post-Transcriptional Regulation Including Alternative Splicing

Mohamed Zouine^{1,2*§}, Yongyao Fu^{1,2§}, Anne-Laure Chateigner-Boutin^{1,2}, Isabelle Mila^{1,2}, Pierre Frasse^{1,2}, Hua Wang^{1,2}, Corinne Audran^{1,2}, Jean-Paul Roustan^{1,2}, Mondher Bouzayan^{1,2*}

¹ Université de Toulouse, INP-ENSA Toulouse, Génomique et Biotechnologie des Fruits, Castanet-Tolosan, France, ² INRA, Génomique et Biotechnologie des Fruits, Auzeville, France

Abstract

Background: The phytohormone auxin is involved in a wide range of developmental processes and auxin signaling is known to modulate the expression of target genes via two types of transcriptional regulators, namely, Aux/IAA and Auxin Response Factors (ARF). ARFs play a major role in transcriptional activation or repression through direct binding to the promoter of auxin-responsive genes. The present study aims at gaining better insight on distinctive structural and functional features among ARF proteins.

Results: Building on the most updated tomato (*Solanum lycopersicon*) reference genome sequence, a comprehensive set of ARF genes was identified, extending the total number of family members to 22. Upon correction of structural annotation inconsistencies, renaming the tomato ARF family members provided a consensus nomenclature for all ARF genes across plant species. *In silico* search predicted the presence of putative target site for small interfering RNAs within twelve *SI-ARFs* while sequence analysis of the 5'-leader sequences revealed the presence of potential small uORF regulatory elements. Functional characterization carried out by transactivation assay partitioned tomato ARFs into repressors and activators of auxin-dependent gene transcription. Expression studies identified tomato ARFs potentially involved in the fruit set process. Genome-wide expression profiling using RNA-seq revealed that at least one third of the gene family members display alternative splicing mode of regulation during the flower to fruit transition. Moreover, the regulation of several tomato ARF genes by both ethylene and auxin, suggests their potential contribution to the convergence mechanism between the signaling pathways of these two hormones.

Conclusion: All together, the data bring new insight on the complexity of the expression control of *SI-ARF* genes at the transcriptional and post-transcriptional levels supporting the hypothesis that these transcriptional mediators might represent one of the main components that enable auxin to regulate a wide range of physiological processes in a highly specific and coordinated manner.

Introduction

The plant hormone auxin, indole-3-acetic acid (IAA), is a simple signaling molecule that plays a critical role in plant development and growth. This phytohormone regulates cell division and cell elongation and exerts pleiotropic effects on a wide range of developmental processes including organ differentiation, embryogenesis, lateral root initiation, apical dominance, gravitropism and phototropism, leaf elongation, shoot architecture and fruit development [1,2,3,4,5]. A critical move towards understanding the mechanisms underlying auxin action [6] happened when it was shown that the hormone coordinates plant development essentially through transcriptional regulation of genes such as *Aux/*

IAA, *Gretchen Hagen3* (*GH3*), *Small Auxin Up RNA* (*SAUR*) and *Auxin Response Factor* (*ARF*). It was subsequently found that these so-called early auxin-responsive genes contain in their promoter one or more copies of a conserved motif, TGTCTC or its variants, known as the auxin-responsive element (AuxRE) [7]. Experimental evidences were then provided showing that transcription factors from the ARF type specifically bind to this AuxRE to mediate the transcription of auxin responsive genes [8]. The components of the pathway linking auxin perception to gene expression are now well established indicating that ubiquitination of Aux/IAA proteins by the TIR1/AFB subunit of the SCF^{TIR1/AFB} ubiquitin ligase leads to their degradation by the 26S proteasome thus releasing the Aux/IAA-mediated inhibition of ARFs which allows these

transcription factors to modulate the expression of their target genes [9].

Three types of transcriptional regulators are required for the control of auxin-responsive genes, Auxin Response Factors (ARFs), Aux/IAAs and Topless (TPLs) [10,11]. Members of the Aux/IAA and TPL families have been reported to function as repressors of auxin-induced gene expression [10,12,13,14]. An increasing number of studies demonstrate the critical role of ARFs in a variety of developmental processes, such as embryo patterning [15,16], leaf expansion and senescence [17,18,19], lateral root growth [18,20,21], floral organ abscission and petal growth [19,22], fruit set and development [23,24,25,26], apical hook formation [27], and various responses to environmental stimuli [28]. In addition, *ARF* genes are involved in the cross-talk between auxin and other hormones like gibberellins [29], ethylene [30], ABA [31] and brassinosteroid signaling [32]. A typical ARF protein consists of a conserved N-terminal B3-type DNA Binding Domain (DBD) that regulates the expression of early auxin response genes, a variable middle region (MR) that function as a transcriptional activation or repression domain (AD or RD), and a conserved C-terminal dimerization domain (CTD) that contributes to the formation of either ARF/ARF homo- and heterodimers or ARF/Aux-IAA hetero-dimers [8,33,34]. The amino acid composition of MRs, located between the DBD and CTD, showed that AD types are rich in glutamine(Q), serine (S), and leucine (L) residues while RD types are rich in proline (P), serine (S), threonine (T), and glycine (G) residues [33,35].

Since the cloning of the first *AtARF1* from Arabidopsis, 22 members of this family, distributed over all five chromosomes, have been identified [33]. The functional characterization of *AtARF* genes was revealed by mutant analysis approach. For instance, *arf1* and *arf2* T-DNA insertion mutations indicated that ARF2 regulates leaf senescence [17] and floral organ abscission [19]. The *arf7/arf19* double mutant had stronger auxin resistance than the single mutant and displayed phenotypes not seen in the single mutant [30]. ARF8 was reported to regulate fertilization and fruit development, and *arf8-4* mutation results in the uncoupling of fruit development from pollination and fertilization giving rise to parthenocarpic fruit [23], while flowers in *arf6/arf8* double mutant are arrested as infertile closed buds with short petals, short stamen filaments, undehiscent anthers and immature gynoecia [36]. In tomato, recent studies have shown the involvement of *ARF* genes in fruit set, development, ripening and fruit quality [3,4,5,24,25,26,37]. Because of these findings, members of this gene family are becoming one of the main targets towards improving fruit traits in tomato and more broadly in fleshy fruits.

Studies using different species have indicated a total of 25 ARF genes in rice (*Oryza sativa*), 39 ARF genes in *Populus trichocarpa*, 24 ARF genes in sorghum (*Sorghum vulgare*) and 31 ARF genes in maize [38,39,40,41]. Though 21 *ARF* genes have been previously identified in the tomato (*Solanum lycopersicum*), yet, the list was incomplete and some the family members were either misannotated or suffered structural inconsistency due to the lack at that time of a high quality assembled tomato genome sequence [42,43]. The present study, while comprehensively revising the entire *ARF* gene family in tomato, brings new insight on the complexity of their expression control at the post-transcriptional level. The distinctive spatio-temporal pattern of expression of tomato *ARF* genes and their differential responsiveness to auxin and ethylene lay the foundation for a deeper functional characterization of these transcriptional mediators.

Results

Genome-wide search for tomato ARF genes

Comprehensive identification of the *ARF* gene family members in the tomato was achieved using all ARF proteins previously reported from Arabidopsis and other plant species in BLAST queries on the recently published tomato genome sequence (SL2.40 genome sequence and iTAG2.30 whole protein sequences). Twenty four significant hits corresponding to non-redundant putative *Sl-ARF* genes were identified. PCR amplification of full length coding sequences (CDS) revealed two structural annotation inconsistencies reducing the total number of ARFs in the tomato genome to 22 (Table 1). Indeed, four sequences previously annotated as distinct *ARF* genes in iTAG2.30 corresponded to C-terminal or N-terminal parts of two ARF proteins (Solyc12g006340/Solyc00g196060; Solyc11g013480/Solyc11g013470). The mapping of tomato RNA-Seq data allowed to further improve the annotation of tomato *ARFs* by identifying the 3' and/or 5' UTR regions for 13 *Sl-ARF* genes (Table 1). All *Sl-ARF* proteins were found to contain a typical DBD domain (Figure 1A) as revealed by the Pfam analysis tool (<http://pfam.sanger.ac.uk/>). The molecular weight of the deduced *Sl-ARF* proteins showed a large variation ranging from 68 to 126 kDa (Table 1). Of particular note, *Sl-ARF6B* contains a premature stop codon in the region corresponding to the DBD domain and is therefore likely to be a pseudo-gene whose expression at the protein level is not expected (data not shown). Using cNLS Mapper, nuclear localisation signals (NLS) were also identified in all of *Sl-ARFs* (data not shown).

Building on the available tomato genome assembly sequence, the mapping of *Sl-ARF* genes revealed that *Sl-ARF* family members are distributed among the 12 tomato chromosomes. Chromosome 7 and 11 are found to harbor three *ARF* genes each; chromosome 1, 2, 3, 5, 8 and 12 bear two *ARFs*, while each of chromosome 4, 6, 9 and 10 contains only a single *ARF* gene (Figure S1 in File S1). Unlike the situation prevailing in Arabidopsis, there is no evidence for tandem or segmental duplication events involving members of the tomato ARF family.

Phylogenetic relationship and consensus nomenclature for *Sl-ARFs*

To explore phylogenetic relationship among ARF proteins in largely distributed land plant species, a phylogenetic tree (Figure 1B) was constructed that included ARF family members from tomato, *Arabidopsis*, potato, grape and rice. The phylogenetic distribution revealed that *ARF* genes group into four major classes named Class I, II, III and IV. ARFs predicted to function as transcriptional activators, based on the presence the Q-rich activation domain in their middle region, belong to sub-class IIa (*Sl-ARF5*, *Sl-ARF6A*, *Sl-ARF7A*, *Sl-ARF7B*, *Sl-ARF8A*, *Sl-ARF8B* and *Sl-ARF19*) while ARFs from the remaining classes (Ia, Iib, III and IV) all harbor a repression domain in the middle region and are consequently predicted to function as transcriptional repressors.

Compared to Arabidopsis which contains 23 members, the size of the tomato *Sl-ARF* gene family is slightly contracted to 22 members. In order to reach a consensual nomenclature for *ARF* genes across species, the tomato members of this gene family were renamed, based on phylogenetic relationship and according to the numbering of the closest Arabidopsis homolog. While complying with the most complete classification available in Arabidopsis [33], the proposed nomenclature better clarifies the correspondence between ARF subclasses in plant species. Noteworthy, sub-class Ib which has no representative in the tomato, contains 7 members in

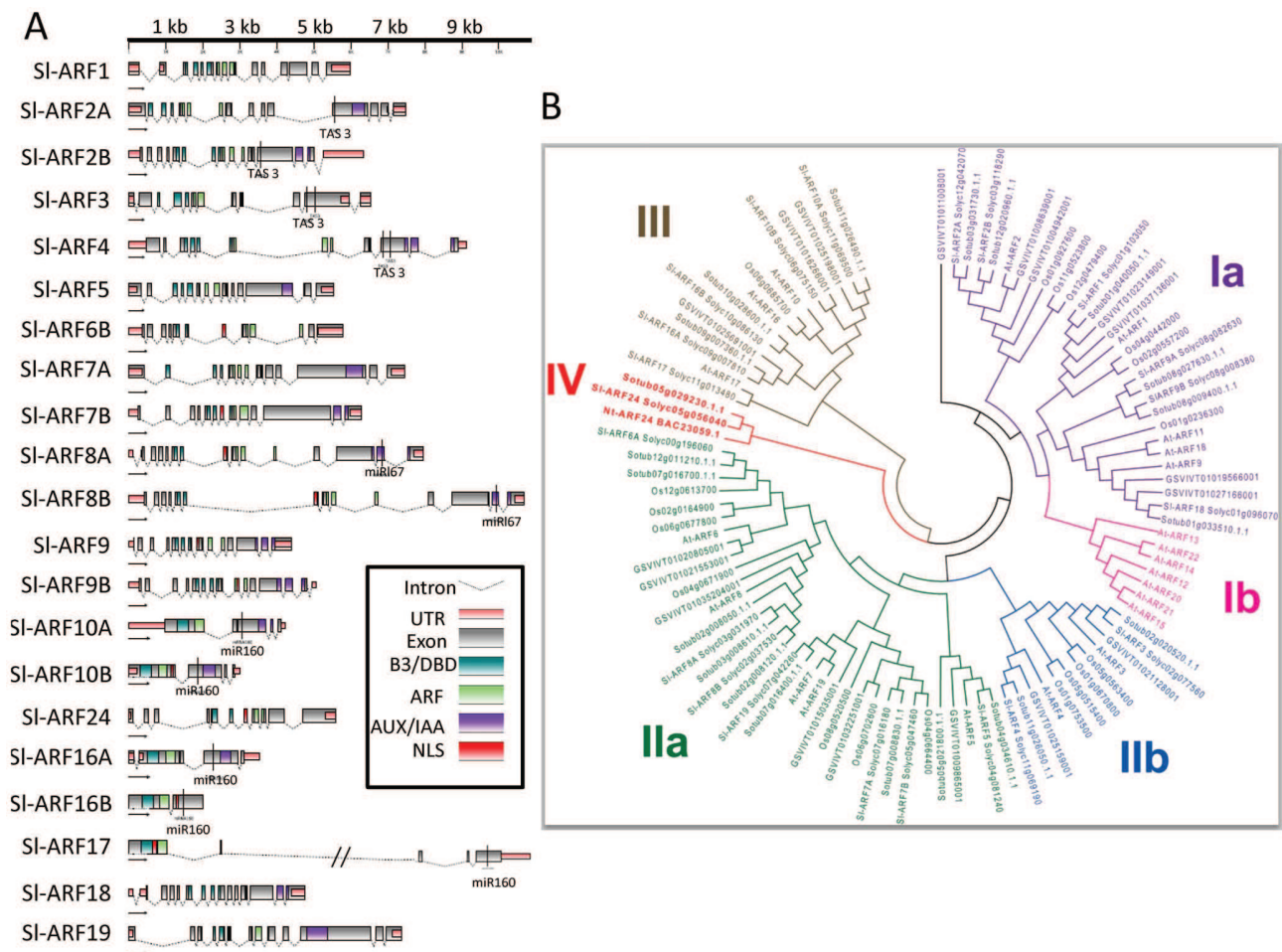


Figure 1. The ARF family structures in tomato and phylogenetic relationship between rice, potato, tomato, grape and Arabidopsis. (A) The generic structures of SI-ARF family except *SI-ARF6A*. The gene size (kb) is indicated in the upper panel. The domain of SI-ARF gene is indicated by different colours. The marker in SI-ARF family shows *SI-ARF2A*, *2B*, *3* and *4* genes are spliced by TAS3, *SI-ARF8A* and *8B* spliced by miR167, and *SI-ARF10A*, *10B*, *16A*, *16B* and *17* spliced by miR160. (B) The unrooted tree was generated using MEGA4 program by neighbor-joining method. Bootstrap values (above 50%) from 1000 replicates are indicated at each branch. All SI-ARFs contain a DBD (brown). Most of the SI-ARF proteins except SI-ARF3, 10, 24, 16 and 17 contain a carboxy-terminal domain related to the domains III and IV found in the Aux/IAA proteins (blue). SI-ARF5, 6A, 7, 8A, 8B, 19 contains a middle region that corresponds to the predicted activation domain (green) found in some AtARFs. The remaining SI-ARFs contains a predicted repression domain (red). SI-ARF-6B and AtARF23 contain only a truncated DBD (B3 domain). doi:10.1371/journal.pone.0084203.g001

Arabidopsis that are likely to derive from multiple duplications of At-ARF13 which has no ortholog in any of the plant species tested in the present study. A distinctive feature of the tomato ARF family is the expanded size of the activators' sub-class (IIa) which represents 36.5% of the ARF genes whereas the activators only account for 21.7% of Arabidopsis ARFs. Another specific feature of the tomato ARF family is the presence of *SI-ARF24* (sub-class IV) that is not found out of the *Solanaceae* family. Interestingly, this presumably *Solanaceae*-specific gene encodes an ARF protein that lacks domain III and IV involved in protein/protein interactions and required for the binding to Aux/IAA proteins. Likewise, SI-ARF3, SI-ARF16B and SI-ARF17 are also deprived of domain III and IV necessary for interaction with Aux/IAs (Figure 1A and Figure S2 in File S1). It is therefore likely that these SI-ARFs escape the classical mechanism underlying auxin signaling which implies the sequestration of ARF proteins through interaction with Aux/IAs.

Predicted siRNA-mediated degradation and multiple upstream ORFs in the 5' leader sequences of tomato ARF transcripts

ARF genes have been already reported to undergo post-transcriptional regulation involving small interfering RNAs. *In silico* analysis at the RNA level predicted that 12 out of the 22 tomato SI-ARFs have a putative target site for small interfering RNAs (Figure 1A). That is, SI-ARF2A, SI-ARF2B, SI-ARF3 and SI-ARF4 are predicted to be potentially targeted by TAS3; SI-ARF6A, SI-ARF8A and SI-ARF8B by miR167; and SI-ARF10A, SI-ARF10B, SI-ARF16A, SI-ARF16B and SI-ARF17 by miR160.

The uORFs are elements found in the 5'-leader sequences of specific mRNAs that modulate the translation of downstream ORFs by ribosomal stalling and inefficient re-initiation or by affecting transcript accumulation through nonsense-mediated mRNA decay pathway. Among the 19 *SI-ARFs* for which the 5' leader sequences are available in iTAG2.30 (8 members) or identified in this study (11 members), uORFs were predicted for 17

Table 1. *Sl-ARF* gene family in tomato.

| Generic name ^a | Alias ^b | CDS length ^c | Str | Chr | Domains ^d | Name in Wu et al. ^e | Improvement ^f | New location ^g |
|---------------------------|---|-------------------------|-----|------|--------------------------------|--|--------------------------|---------------------------|
| Sl-ARF1 | Solyc01g103050 | 1965 | + | 1 | B3,ARF,AUX/IAA,SPL-Rich RD | SIARF1(HM061154.1) | - | |
| Sl-ARF2A | Solyc03g118290 | 2541 | + | 3 | B3,ARF,AUX/IAA,SPL-Rich RD | SIARF2(DQ340255.1) | - | |
| Sl-ARF2B | Solyc12g042070 | 2490 | - | 12 | B3,ARF,AUX/IAA,SPL-Rich RD | SIARF11(HM143940.1) | 5',3' UTR | 42538600..42544937 |
| Sl-ARF3 | Solyc02g077560 | 2244 | + | 2 | B3,ARF,SL/G-Rich RD | SIARF3(DQ340254.1) | - | |
| Sl-ARF4 | Solyc11g069190 | 2436 | - | 11 | B3,ARF,AUX/IAA,SPL-Rich RD | SIARF4(DQ340259.1) | 5',3' UTR | 50900912..50910023 |
| Sl-ARF5 | Solyc04g081240 | 2793 | - | 4 | B3,ARF,AUX/IAA,DSL-Rich AD | SIARF5(HM195248.1) | - | |
| Sl-ARF6A | Solyc12g006340(Nter); Solyc00g196060(Cter) | 2643 | -/+ | 12/0 | B3,ARF,AUX/IAA, QSL-Rich AD | SIARF6(HM594684.1) | 5' UTR, CDS | 857256..859656(Nter) |
| Sl-ARF6B | Solyc07g043620 | 2673 | - | 7 | B3,ARF,QSL-Rich AD | SIARF6-1(NM_001247611.1) | 5' UTR | 54884781..54890560 |
| Sl-ARF7A | Solyc07g016180 | 3339 | - | 7 | B3,ARF,AUX/IAA,DSL-Rich AD | SIARF19(HM130544.1) | - | |
| Sl-ARF7B | Solyc05g047460 | 3294 | - | 5 | B3,ARF,AUX/IAA,DSL-Rich AD | SIARF19-1(HM565130.1) | 5' UTR | 58050744..58057040 |
| Sl-ARF8A | Solyc03g031970 | 2535 | + | 3 | B3,ARF,AUX/IAA,DSL-Rich AD | SIARF8-1(HM560979.1) | 5' UTR | 8739535..8747501 |
| Sl-ARF8B | Solyc02g037530 | 2529 | + | 2 | B3,ARF,AUX/IAA,DSL-Rich AD | SIARF8(EF66734F2.1) | 5' UTR | 21756022..21766699 |
| Sl-ARF9A | Solyc08g082630 | 1977 | + | 8 | B3,ARF,AUX/IAA,SPL-Rich RD | SIARF9(HM037250.1) | 5' UTR | 62527409..62531812 |
| Sl-ARF9B | Solyc08g008380 | 2052 | + | 8 | B3,ARF,AUX/IAA,SPL-Rich RD | SIARF12(HM565127.1) | 5' UTR | 2807931..2812983 |
| Sl-ARF10A | Solyc11g069500 | 2100 | - | 11 | B3,ARF,AUX/IAA,SL/G-Rich RD | SIARF10(HM143941.1) | 5',3' UTR | 51188434..51192539 |
| Sl-ARF10B | Solyc06g075150 | 2016 | + | 6 | B3,ARF,AUX/IAA,SL/G-Rich RD | SIARF16(HM195247.1) | 3' UTR | 43020594..43023604 |
| Sl-ARF24 | Solyc05g056040 | 1953 | - | 5 | B3,ARF,SPL-Rich RD | SIARF13(HM565128.1); SIARF13-1(HM565129.1) | - | |
| Sl-ARF16A | Solyc09g007810 | 2085 | - | 9 | B3,ARF,AUX/IAA,SL/G-Rich RD | No | 3' UTR | 1332230..1335760 |
| Sl-ARF16B | Solyc10g086130 | 1896 | - | 10 | B3,ARF,SL/G-Rich RD | SIARF16(NM_001247861.1) | - | |
| Sl-ARF17 | Solyc11g013480(Nter); Solyc11g013470(Cter) | 1869 | - | 11 | B3,ARF,SL/G-Rich RD | SIARF17(HQ456923) | 3' UTR, CDS | 6495469..6511349 |
| Sl-ARF18 | Solyc01g096070 | 2058 | + | 1 | ARF,AUX/IAA,SPL-Rich RD | No | 5' UTR | 78941268..78946012 |
| Sl-ARF19 | Solyc07g042260 | 3357 | - | 7 | B3,ARF,AUX/IAA,DSL-Rich AD | SIARF7(EF121545.1) | - | |

^aSl-ARF gene names

^bthe alias of each *ARF* gene in iTAG2.30 genome annotation

^cLength of the corresponding Coding Sequence (CDS) in base pairs.

^dConserved Domains found in PFAM database; B3 means DNA binding domain, ARF means Auxin response Factor conserved domain, AUX/IAA means AUX/IAA dimerization domain, AD means transcriptional activation domain, RD means transcriptional repression domain.

^eCorresponding names in Wu et al.; accession numbers are in the parentheses.

^fGene Model modification type: UTR means that the UTR sequence have been identified and annotated, CDS means that the coding sequence have been corrected.

^gNew locations in the tomato genome version S12.40 taking into account the manual curation of the previous gene annotation in iTAG2.30

doi:10.1371/journal.pone.0084203.t001

genes, ranging from 1 to 52 amino acids in size with four genes (SI-ARF2A, SI-ARF5, SI-ARF10A and SI-ARF16A) having five or more uORFs (Table S1 in File S1). The average number of uORF per SI-ARF gene is similar in tomato (2.8/leader) and Arabidopsis (3.3/leader), indicating that tomato ARFs are suitable candidates to be regulated through translational uORFs depending mechanism

Transcriptional activation and repression activities of tomato ARFs

To characterize the capacity of tomato ARF proteins to *in vivo* activate or repress gene transcription, tobacco cells were co-transfected with an effector construct expressing the full-length coding sequence of SI-ARFs and a reporter construct carrying the auxin-responsive *DR5* promoter fused to *GFP* coding sequence [44]. *DR5* is a synthetic auxin-responsive promoter made of 9 inverted repeats of the conserved Auxin-Responsive Element, the so-called TGTCTC box, fused to a *CaMV35S* minimal promoter. The *DR5*-driven GFP chimeric gene showed low basal activity which was induced up to 5-fold by exogenous auxin treatment (Figure 2). Co-transfection of tobacco protoplasts with the *DR5::GFP* reporter construct and effector plasmids expressing either *SI-ARF1*, *SI-ARF2A*, *SI-ARF2B*, *SI-ARF3*, *SI-ARF4*, *SI-ARF9A*, *SI-ARF10A* or *SI-ARF17* coding sequences, resulted in repression of the auxin-induced expression of the reporter gene (Figure 2). By contrast, co-transfection with effector constructs expressing *SI-ARF5*, *SI-ARF6A*, *SI-ARF7*, *SI-ARF8B* or *SI-ARF19* enhanced slightly the auxin-induced expression of the reporter gene. Noteworthy, with the exception of *SI-ARF6A* and *SI-ARF7*, these activator ARFs were unable to enhance the basal activity of the *DR5* promoter in the absence of auxin treatment (Figure 2) suggesting that most ARFs require the input of an active auxin signalling for transcriptional activation of target genes.

Expression of SI-ARF genes in different tomato tissues

To gain clues on the physiological function of tomato ARFs, the spatio-temporal expression of individual members of the gene

family was examined at the transcriptional level using qRT-PCR. Transcript accumulation could be assessed for 15 *ARF* genes in different tissues including root, stem, leaves, flower and fruit at various developmental stages. For the remaining 7 tomato *ARF* genes, transcript detection was unsuccessful in any of the samples tested suggesting their extremely low expression in these tissues. The data indicate that the expression of *ARF* genes is ubiquitous in all tissues with most genes being expressed in reproductive tissues suggesting their putative role in flower and fruit development (Figure 3). Heatmap representation (Figure 4) allowed the clustering of tomato *ARFs* into two main groups based on their expression pattern: group I (*SI-ARF1*, *SI-ARF2A*, *SI-ARF2B*, *SI-ARF4*, *SI-ARF7A*, *SI-ARF6B* and *SI-ARF18*) are genes preferentially expressed in roots and group II *SI-ARFs* in the areal part of the plant.

SI-ARF6B displays a very low expression in all tomato tissues analyzed and the corresponding CT values showed high variability among repeats making these *SI-ARF6B* expression data not meaningful. Therefore, they were not included in Figure 3 but retained for the heat map (Figure 4) in despite of the variability between the repeats in order to give a general idea about its expression in different tissues.

Auxin and ethylene regulation of SI-ARF genes

Screening for *cis*-acting elements corresponding to Auxin Response Elements (AuxRE) within the promoter regions using the PLACE database (<http://www.dna.affrc.go.jp/PLACE/signalscan.html>) identified conserved (TGTCTC) and degenerate (TGTCCC) motifs in most tomato ARF promoters. In addition to these AuxRE, *SI-ARF* promoters contain conserved Ethylene-Response motifs, the so-called ERELEE4 motif found in the promoter of tomato *E4* gene (AWTTCAAA) (Table S2 in File S1). The presence of these *cis*-regulatory elements suggests a potential regulation of *ARF* genes by both auxin and ethylene. To test the responsiveness of tomato *ARF* genes to both hormones, transcript accumulation was assessed by qRT-PCR in seedlings treated with auxin or ethylene. All *SI-ARFs* were found to be auxin-responsive after 2 hour treatment (Figure 5A), with *SI-ARF4*, *SI-ARF5* and

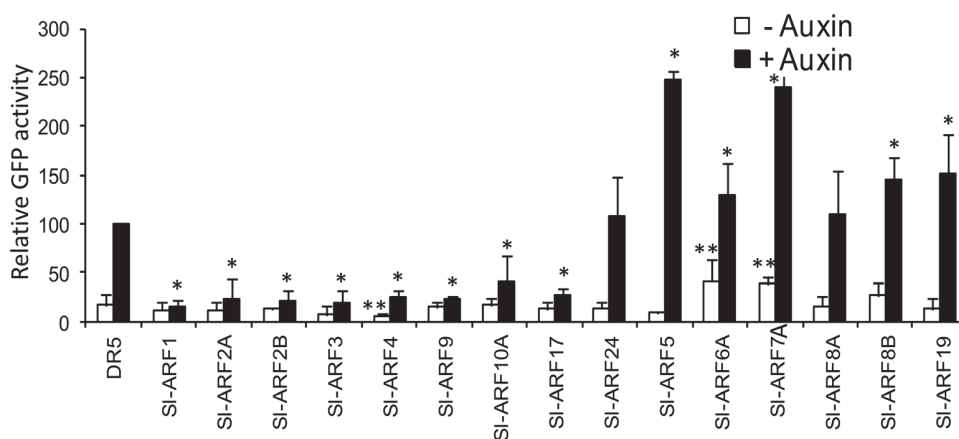


Figure 2. SI-ARF factors differentially regulate the expression of reporter genes driven by synthetic and native auxin-responsive promoters. SI-ARF factors were challenged with a synthetic auxin-responsive promoter called *DR5*, consisting of seven tandem copies of the AuxREtgtctc element. A transient expression using a single cell system was performed to measure the reporter gene activity. The fluorescence was measured by flux cytometry. Because of the very low basal activity of the *DR5* promoter without auxin treatment, the auxin inducible fluorescence obtained by co-transformation with the promoter fused to the reporter gene and with the empty vector was standardized to 100 and taken as reference. Biological triplicates were averaged and analysed statistically using Student's t-test at ($P < 0.05$). (*) indicates significant changes corresponding to co-transformation with effector SI-ARF and reporter *DR5*-GFP constructs compared to basal activity of *DR5* promoter in the absence of auxin treatment. (**) indicates significant changes for the same experiment carried out in the presence of auxin. Bars indicate the SEM. doi:10.1371/journal.pone.0084203.g002

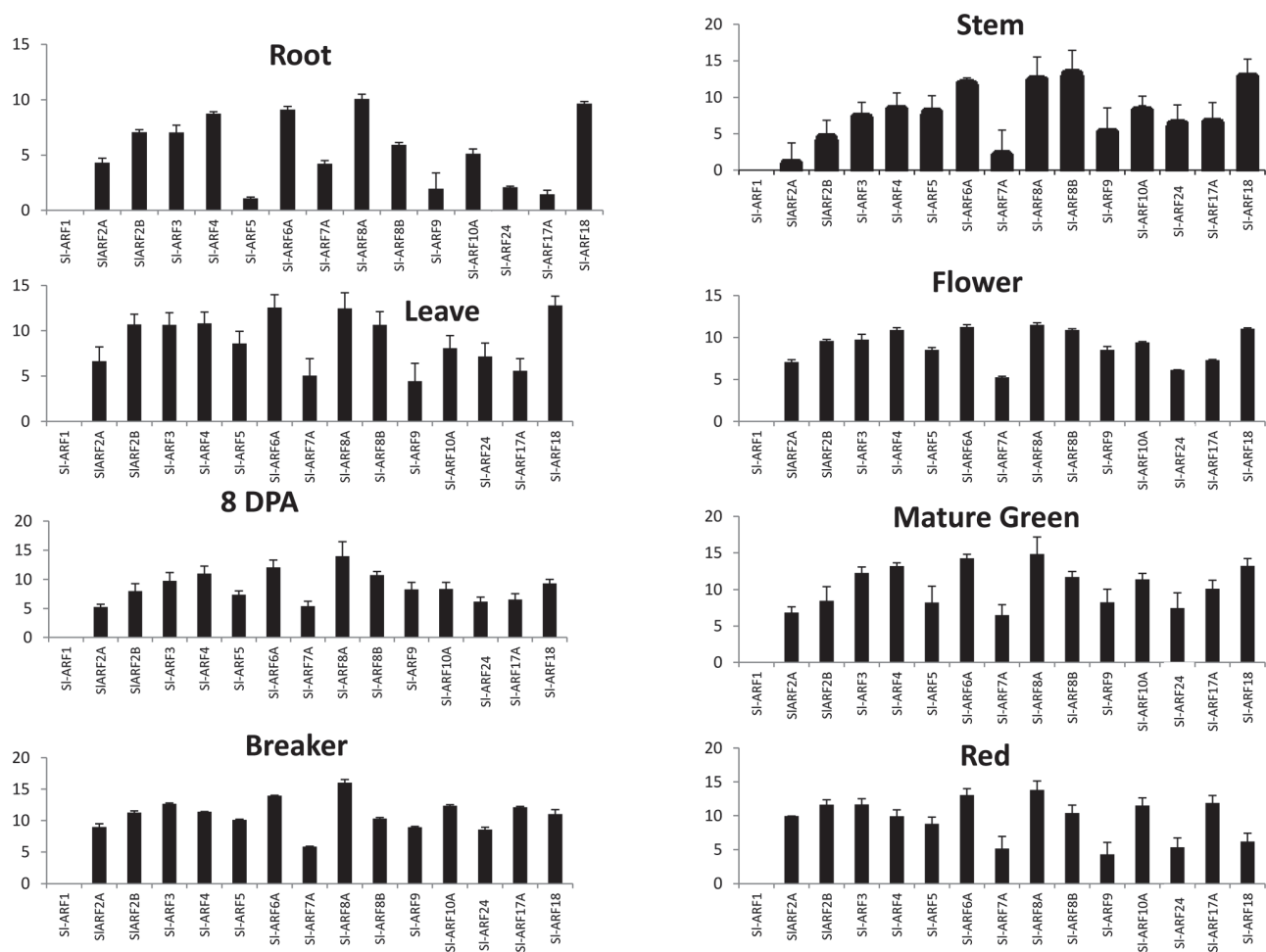


Figure 3. Real-time PCR expression profiles of individual *SI-ARF* genes. Total of 15 *SI-ARF* genes were performed in different tomato organs (root, stem, leaf, flower, 8DPA, Mature Green, Breaker and Red). X-axis represents different *SI-ARF* genes, while Y-axis represents three relative expressions of those genes. 8DPA: 8 days after pollination, Mature Green, Breaker and Red represent different stage of the fruit development. doi:10.1371/journal.pone.0084203.g003

SI-ARF2A showing the highest up-regulation whereas *SI-ARF1*, *SI-ARF7* *SI-ARF10* displayed the most significant down-regulation. On the other hand, the expression of *SI-ARF2B*, *SI-ARF5* and *SI-ARF9A* showed strong up-regulation (more than four folds increase) when treated 5 hours with ethylene (Figure 5B). Of particular interest, *SI-ARF5* is strongly up-regulated by both hormones and may therefore be involved in mediating responses to both hormones.

Expression of *SI-ARF* genes during tomato fruit set

The expression of a high number of *SI-ARFs* in reproductive tissues (Figure 3 and 4) along with the previously reported role of auxin in controlling the fruit set process, prompted us to investigate the expression of *SI-ARF* genes during the flower-to-fruit transition. To determine the expression dynamics throughout the fruit set process, transcript accumulation of tomato *ARFs* was monitored by RNA-seq approach at flower buds, anthesis and post-anthesis stages (young fruit at 4 DPA). For each stage, RNA libraries were generated from three independent biological replicates and subjected to Illumina mRNA-Seq technology sequencing (Data deposited at NCBI SRA database under accession number SRP029978). Reads were then mapped on the

tomato genome sequence and read counts were determined as described in Maza et al. 2013 [45]. The data indicate that most *SI-ARFs* undergo a strong change in their expression associated with the flower-to-fruit transition (Figure 6). Three groups could be discriminated based on RNA counts distribution during the fruit set process. Group 1 corresponds to *SI-ARFs* whose expression increased following pollination, Group 2 to *ARFs* with unchanged expression and Group 3 to *SI-ARFs* displaying decreased expression following pollination (Figure 6).

SI-ARF transcripts undergo intense alternative splicing during tomato fruit set

Closer analysis of the mapping of RNA-seq data on the gene models revealed possible alternative splicing regulation during fruit set for 30% of *SI-ARF* genes. *SI-ARF2B* and *ARF19* shows one possible alternative splicing occurring at intron 11 and intron 1, respectively (Figure 6A and Figures S3.1 in File S1). *SI-ARF3* and *SI-ARF4* could putatively give rise to two alternative splicing events at introns 7 and 9, and at introns 6 and 10, respectively. Three possible alternative splicings were found at introns 3, 6 and 10 in *SI-ARF8A* and at introns 9, 11 and 13 in *SI-ARF8B*. Finally, *SI-ARF24* offers up to four alternative splicing possibilities at introns

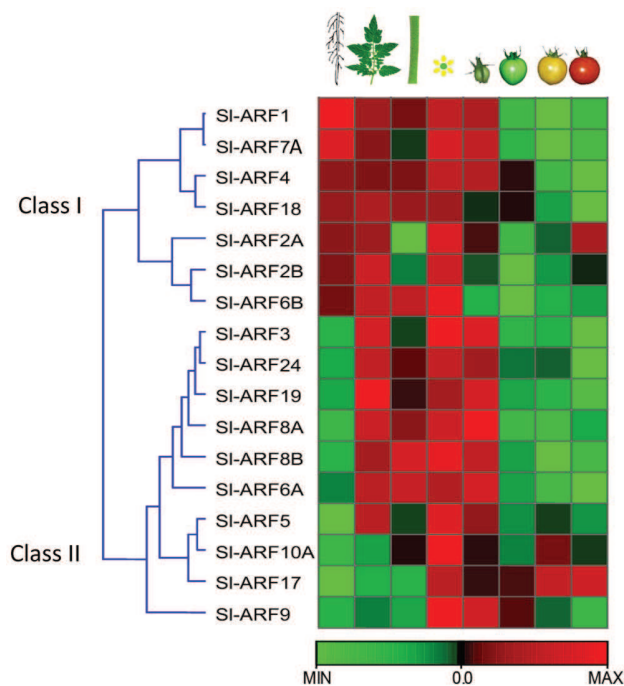


Figure 4. Heatmap showing *SI-ARF* gene expression in different tomato tissues. Changes in RNA accumulation in different tomato tissues (Roots, Leaves, Stems, Flowers, Early Immature Green (8 DPA), Mature Green, Breaker, Red (Breaker + 7 days) as schematically depicted above the displayed array data, are shown relative to the RNA accumulation levels in roots. Levels of down expression (green) or up expression (red) are shown on a log₂ scale from the high to the low expression of each *SI-ARF* gene.
doi:10.1371/journal.pone.0084203.g004

1, 3, 6 and 10 (Figures S3.1–6 in File S1). In all cases, the detected *SI-ARF* splice variants resulted in a frame shift within the coding region that generates a premature stop codon. To further validate the occurrence of the alternative splicing forms and assess the relative levels of the various splice variants, a semi quantitative PCR approach was conducted. To this purpose, two pairs of primers were designed, one aiming to specifically amplify the retained intron fragment while the second pair was designed in the margins of the two exons framing the retained intron. A PCR product with the expected size was detected for all genes confirming the presence of the splice variant in each RNA extraction (Figure 7B). Interestingly, the data indicate that the abundance of the *SI-ARF8B_int11* transcript variant decreases dramatically in young fruits whereas the global expression of the corresponding *SI-ARF8B* gene increases significantly. This finding suggests that the down-regulation of the *SI-ARF8B_int11* transcript variant may potentially play a role in the regulation of the flower to fruit transition. By contrast, increased accumulation of the *SI-ARF19_int1* was observed concomitant to the transition from flower to fruit. Taking together, these data uncover a potential role for alternative splicing in regulating the expression of tomato *ARF*s during the fruit set process.

Discussion

Being down-stream components of auxin signalling pathway, *ARF*s likely contribute to the specificity of the hormone responses. Hence, the functional characterization of these transcriptional mediators is essential towards understanding the mechanisms by

which auxin triggers appropriate growth and developmental responses in a timely and tissue-specific manner. To better define the role of *ARF*s in mediating specific auxin responses, the present study brings a complete picture on the main structural features of the tomato *ARF* gene family. Identification of tomato *ARF*s has been already described but this attempt built on a draft tomato genome sequence and ESTs and could therefore not be comprehensive [42,43]. The present work takes advantage of the most updated tomato reference genome sequence [46] to isolate the complete *ARF* family members and perform functional analysis and expression profiling of these transcriptional regulators. Using these extended resources, the list of tomato *ARF*s has been enlarged to 22 members and manual annotation based on deep RNA-Seq data, allowed the curation of some structural annotation inconsistencies as well as the identification of the 3' and 5' UTR regions for more than 50% of the *SI-ARF* gene family. The tomato members of the *ARF* family were renamed according to the numbering of the closest Arabidopsis homolog, which provides a consensus nomenclature for *ARF* genes across plant species. In this way, the proposed nomenclature better clarifies the correspondence between *ARF* subclasses in various plant species. The phylogenetic approach applied on a well distributed set of plant *ARF*s allowed to identify a specific sub-class (sub-class IV) that is absent out of the *Solanaceae* family. Interestingly, this sub-class contains a specific gene, *SI-ARF24*, encoding a putative *ARF* protein that lacks the two protein/protein interaction domains, known as domain III and IV and required for the binding to Aux/IAA proteins. It is therefore likely that *SI-ARF24* escapes the classical mechanism underlying auxin signaling which implies the sequestration of *ARF* proteins through interaction with Aux/IAAs.

As a preliminary approach towards functional characterization of members of the tomato *ARF* family, the present study describes their expression pattern, their post-transcriptional regulation and their ability to activate or repress transcriptional activity on synthetic or native auxin-responsive promoters. Transactivation assays revealed that 36% of tomato *ARF*s are strong repressors of transcriptional activity while only 22% are transcriptional activators. The repressor/activator ratio among *ARF*s is more than twice higher in tomato (3.6) compared to Arabidopsis (1.7), yet, it remains to be elucidated whether this feature may account for differences in developmental and growth behaviour between the two species. In contrast to repressor *ARF*s, most activator *SI-ARF*s promote transcription of target genes only upon exogenous auxin treatment thus suggesting that activator *ARF*s require some input from a highly activated auxin signalling pathway in order to potentiate transcriptional activity. It is conceivable that when the auxin level is low, the amount of Aux/IAA proteins available is sufficient to block *ARF*s at the protein level thus preventing these latter from activating the transcription of the target genes. In this perspective, it has to be postulated that Aux/IAAs are present in excess in the cell when the tissue is not subjected to auxin treatment.

The spatio-temporal pattern of expression indicated that all *SI-ARF* genes are expressed in flower and fruit suggesting a putative important role in reproductive tissue development. The shift from the static flower ovary to fast-growing young fruit is a phenomenon known as fruit set and auxin has been shown to play a crucial role in controlling this developmental process [47,48] representing an important step in the development of all sexually reproducing higher plants. Adding to the primary role of Aux/IAAs in triggering the fruit set process previously reported [3,49], the present study reveals the potential active role of a number of *SI-ARF*s during this process based on genome-wide transcriptomic profiling of the flower to fruit transition. The expression of 12

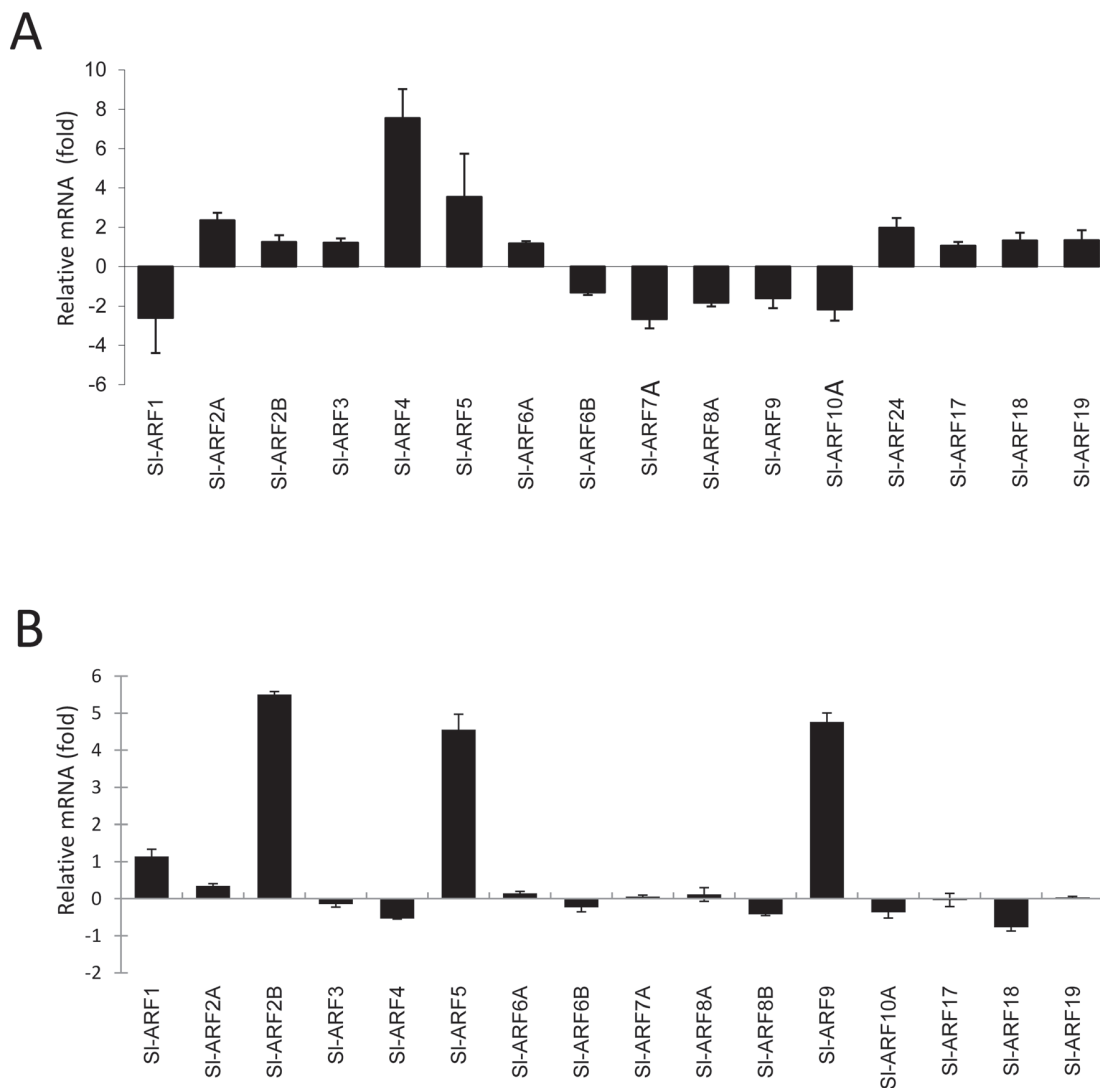


Figure 5. The expression of *SI-ARF* family genes in response to auxin and ethylene. (A) Auxin induction of *SI-ARF* genes on light grown seedlings. Quantitative RT-PCR of *SI-ARF* transcripts in RNA samples extracted from 12-day-old tomato seedlings soaked in liquid MS medium with 10 μ M IAA for 2 hours. $\Delta\Delta$ CT refers to the fold of difference in *SI-ARF* expression to the untreated seedlings. The *SAUR* gene was used as control to validate the auxin treatment.(B) Ethylene regulation of *SI-ARF* genes on dark grown seedlings. Quantitative RT-PCR of *SI-ARF* transcripts in RNA samples extracted from 5-days dark-grown tomato seedlings treated 5 hours with ethylene (50 μ L/L). $\Delta\Delta$ CT refers to fold differences in *SI-ARF* expression relative to untreated seedlings. The *E4* gene was used as control for efficient ethylene treatment. doi:10.1371/journal.pone.0084203.g005

members of the gene family sharply increases upon pollination/fertilization, while the expression of a fewer number of SI-ARF genes peaks at anthesis and then dramatically declines at post-pollination stage. Given the role of auxin signaling in the fruit set process [48,50], the dynamics of the expression pattern of these SI-ARFs is indicative of their putative involvement in mediating auxin responses during the flower-to-fruit transition. This is consistent with the prominent role reported for *SI-ARF8A* and *SI-ARF7* (referred as SI-ARF19 in the present study) during fruit set and parthenocarp in Arabidopsis and tomato, respectively [24,26]. Of particular interest, *SI-ARF8A* shows the most dramatic rise in expression at post-anthesis stage which may designate this ARF among all family members as the main actor of the fruit set process.

The data indicate that tomato ARFs are subject to multi-levels post-transcriptional regulation of their expression. In line with

Arabidopsis ARFs [51,52,53], it is shown here that 11 out of the 22 tomato *ARF* genes are potentially regulated by siRNAs. Moreover, the direct evidence for active alternative splicing described here uncover a new layer of complexity in the post-transcriptional regulation of ARF genes in the tomato. This mode of regulation may account for a significant part of the control of ARF expression in developmental processes such as fruit set in the tomato as indicated by the abundance of some transcript splice variants concomitant to the flower to fruit transition. An additional mean towards controlling ARF expression in the tomato may also take place at the translational level via upstream ORFs (uORFs) that have been predicted in most members of *ARF* genes. This mode of regulation has been first suggested in Arabidopsis where *in silico* search revealed an enrichment of uORFs in the ARF 5'-leader sequences that is not seen in other auxin-related genes such *Aux/IAA*, *YUCCA*, *TIR1* auxin receptors homologs and *PIN* family of

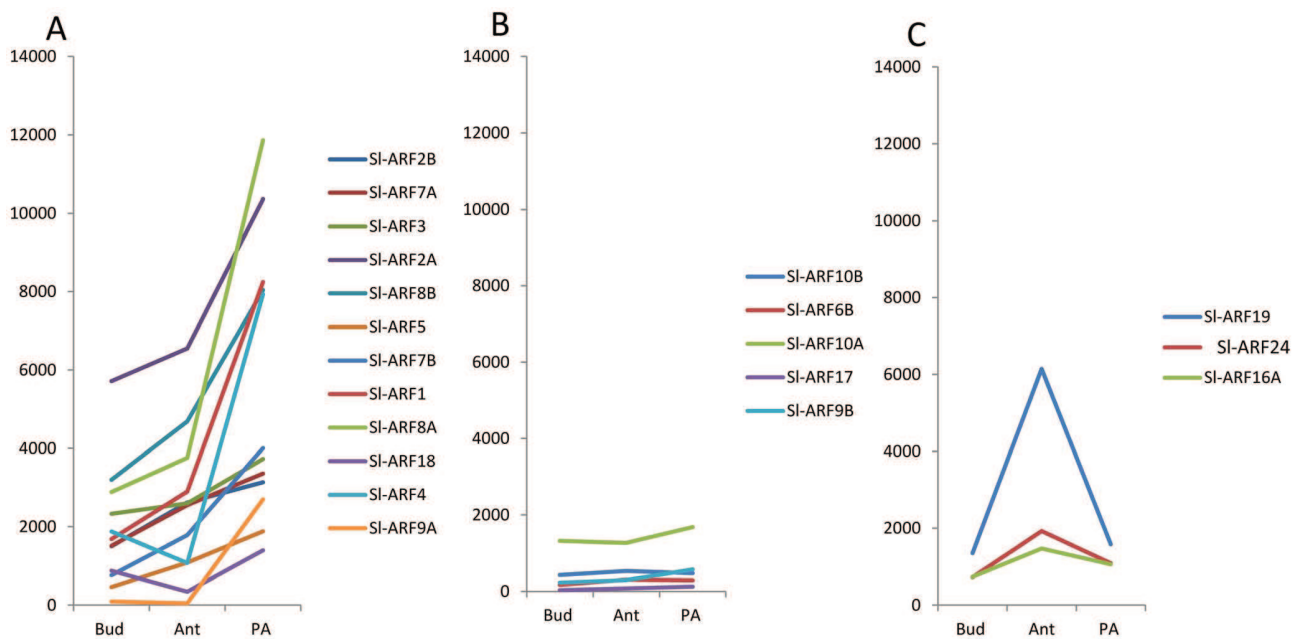


Figure 6. The expression profile of *SI-ARF* family genes in tomato fruit set. (A) 12 *SI-ARF* genes are over-expressed after pollination and fertilization (4DPA), which are SI-ARF9A, 4, 18, 8A, 1, 7B, 5, 8B, 2A, 3, 7A and 2B genes in turn according to the log change of P/A (Post-anthesis/Anthesis). (B) 5 *SI-ARF* genes keep stable expression from flower bud to post-anthesis, including SI-ARF10A, 10B, 6B, 9B, 17 genes. (C) 3 *SI-ARF* genes are up-regulated from flower bud to anthesis and down-regulated after pollination and fertilization (4DPA), including SI-ARF24, 19, and 16A genes. The expression values are taken from RNA-sequencing data and the colors represent different *SI-ARF* genes. doi:10.1371/journal.pone.0084203.g006

auxin transporters [54]. Subsequently, translational control of *AtARFs* by upstream ORF (uORFs) has been proposed as a regulatory mechanism required in modulating auxin responses during plant development [55]. Though direct experimental evidence is still lacking, tomato ARFs may also undergo the same mode of regulation.

In addition of being auxin-responsive, the expression of some *SI-ARFs* was found to be regulated by ethylene. The presence of auxin and ethylene *cis*-regulatory elements in the promoter region of a number of *SI-ARFs*, supports the potential regulation of *ARF* genes by both auxin and ethylene and suggests that these transcription factors have the ability to mediate both auxin and ethylene responses. In support to this hypothesis, Arabidopsis *ARF19* has been shown to be inducible by ethylene and has been reported to contribute to ethylene sensitivity through a cross-talk between auxin and ethylene signalling [27,30]. Also, *ARF2* has been shown to regulate the hook curvature of etiolated Arabidopsis seedlings, a typical ethylene response [27]. Taking together, these data suggest that ARFs may act at the crossroads of auxin and ethylene signaling.

Altogether, the data provide molecular clues on how ARFs can contribute to the specificity and selectivity of auxin responses through (i) structural features, (ii) differential expression of family members at the tissue and organ levels and, (iii) ability to negatively or positively impact transcriptional activity of target genes. The auxin and ethylene regulation of some ARF members suggest their specific role in the multi-hormonal cross-talks. The regulation of the expression of *ARFs* by alternative splicing during fruit set provides new insight into the complexity of regulation of these genes at the post-transcriptional level.

Materials and Methods

Plant material and growth conditions

Tomato seeds (*Solanum lycopersicum* cv MicroTom or Ailsa Craig) were sterilized, rinsed in sterile water and sown in recipient Magenta vessels containing 50 mL of 50% Murashige and Skoog (MS) culture medium added with R3 vitamin (0.5 mg L⁻¹ thiamine, 0.25 mg L⁻¹ nicotinic acid and 0.5 mg L⁻¹ pyridoxine), 1.5% (w/v) sucrose and 0.8% (w/v) agar, pH 5.9. Plants were grown under standard greenhouse conditions. The culture chamber rooms are set as follows: 14-h-day/10-h-night cycle, 25/20°C day/night temperature, 80% hygrometry, 250 μmol m⁻²s⁻¹ intense luminosity.

In silico Identification of the tomato ARFs

All the ARF gene sequences (ITAG2.3_gene_models.gff3) are download from the Sol Genomics Network (<http://solgenomics.net/>), and analyzed in Notepad++ software. The NLS location was searched using cNLS Mapper (http://nls-mapper.iab.keio.ac.jp/cgi-bin/NLS_Mapper_form.cgi). All the obtained sequences were sorted for the unique sequences and these were further used for B3, AUX_RESP, and Aux/IAA domain search using InterProScan Sequence Search (<http://www.ebi.ac.uk/Tools/pfa/iprscan/>). The UTR of SI-ARFs were found by two steps, first, the whole tomato genome and SI-ARF gene structures (ITAG2.3_gene_models.gff3) were loaded into the Java, and then, the complete cDNA sequences from RNA-Seq data including three stages (flower bud, anthesis and post-anthesis) were blast with SI-ARF gene structures to identify the final 5' or 3' UTRs in SI-ARFs. The miRNA location on the SI-ARFs were searched depend on the GBF data (<http://tata.toulouse.inra.fr/gbf/blast/blast.html>) and SGN Blast tools. Taken together, all of the SI-ARF family structures were

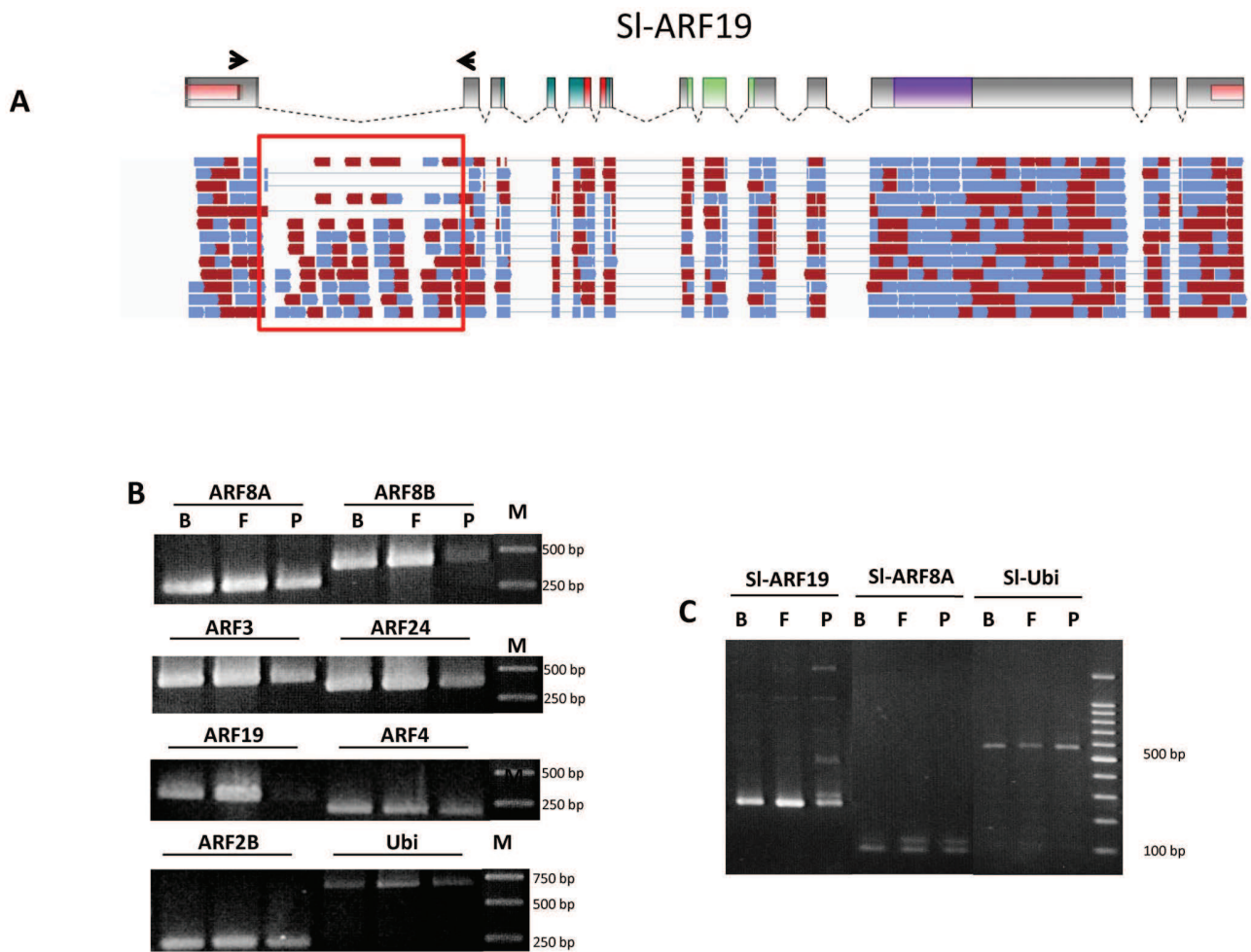


Figure 7. The ARF family genes showed alternative splicing mode of regulation in tomato fruit set. (A) RNA-seq reads generated during the fruit-set and mapped on *Sl-ARF19* gene structure showing one alternative splicing that can be generated in the Intron 1. Reads are represented by red and blue rod arrows (B) The RT-PCR was carried out using pairs of primers designed within the introns of 7 *Sl-ARF* genes highlighted in Figures S3.1 to S3.6 in File S1, such as *Sl-ARF8A_Intron 6*, *Sl-ARF8B_Intron 11*, *Sl-ARF3_Intron 9*, *Sl-ARF24_Intron 3*, *Sl-ARF19_Intron 1*, *Sl-ARF4_Intron 6* and *Sl-ARF2B_Intron 11*. The ubiquitin gene was used as the reference. (C) The RT-PCR was performed using pairs of primers nested in the two exons encompassing the intron of target *Sl-ARF* genes, such as Exon1-Exon2 in *Sl-ARF19* and Exon6-Exon7 in *Sl-ARF8A*. The cDNAs generated from flower bud (B), flower at anthesis (F) and young fruit 4 days post-pollination (P) tissues were used as the template. The ubiquitin was used as the reference. doi:10.1371/journal.pone.0084203.g007

drawn by Fancy Gene v1.4 (<http://host13.bioinfo3.ifom-ieo-campus.it/fancygene/>) with manual correction.

Transient Expression Using a Single Cell System

Protoplasts were obtained from suspension-cultured tobacco (*Nicotianatabacum*) BY-2 cells and transfected by a modified polyethylene glycol method as described by Abel and Theologis [56]. For nuclear localization of the selected ARF fusion proteins, the coding sequence of genes were cloned as a C-terminal fusion in frame with GFP under the control of the 35S CaMV, a cauliflower mosaic virus promoter. Transfected protoplasts were incubated for 16 h at 25°C and analysed for GFP fluorescence by confocal microscopy. For co-transfection assays, aliquots of protoplasts (0.5×10^6) were transformed either with 10 µg of the reporter vector alone containing the promoter fused to the GFP reporter gene or in combination with 10 µg of ARF constructs as the effector plasmid. Transformation assays were performed in three independent replicates. After 16 h, GFP expression was analyzed and quantified by flow cytometry (FACS Calibur II instrument,

BD Biosciences, San Jose, CA) on the flow cytometry platform, IRF31, Inserm, Toulouse and cell sorting platform, INSERM UPS UMR 1048, Toulouse RIO imaging platform. Data were analyzed using Cell Quest software. For each sample, 100 to 1000 protoplasts were gated on forward light scatter and the GFP fluorescence per population of cells corresponds to the average fluorescence intensity of the population of cells after subtraction of autofluorescence determined with non transformed BY-2 protoplasts. The data are normalised using an experiment, in presence of 50 µM 2,4 D, with protoplasts transformed with the reporter vector in combination with the vector used as the effector plasmid but lacking *Sl-ARF* coding region.

RNA isolation and Quantitative RT-PCR

Total RNA from fruit was extracted according to the method of Hamilton *et al.* [57]. Total RNA from leaves and seedlings was extracted using a Plant RNeasy Mini kit (Qiagen) according to the manufacturer's instruction. Total RNA was treated by DNase I to remove any genomic DNA contamination. First strand cDNA was

reverse transcribed from 2 µg of total RNA using Omniscript kit (Qiagen) according to the manufacturer's instruction. The qRT-PCR analysis was performed as previously described [3]. The sequences of primers are listed in Table S3 in File S1. Relative fold differences were calculated based on the comparative Ct method using the Sl-Actin as an internal standard. To determine relative fold differences for each sample in each experiment, the Ct value of genes was normalized to the Ct value for Sl-Actin-51 (accession number Q96483/Solyc11g005330) and was calculated relative to a calibrator using the formula $2^{-\Delta\Delta C_t}$. At least two to three independent RNA isolations were used for cDNA synthesis and each cDNA sample was subjected to real-time PCR analysis in triplicate. Heat map representation was performed using centring and normalized ΔC_t value, with Cluster 3.0 software and Java Tree view to visualize dendrogram.

Hormone treatment

For auxin treatment on light grown seedlings, 12-day-old tomato seedlings (30 seedlings) were soaked in liquid MS medium with or without (mock treatment) 10 µM IAA for 2 hours. The efficiency of the treatment was checked by measuring the induction of the tomato early auxin-responsive SAUR gene. For ethylene treatment on dark grown seedlings, 5-days-old Micro-Tom seedlings (100 seedlings) were treated with air or ethylene gas (50 µL/L) for 5 hours. The efficiency of the treatment was checked by measuring the induction of the tomato ethylene-responsive E4 gene. Experiment was repeated for 3 biological times.

RNA-Sequencing and RT-PCR

Total RNA was extracted from bud, flower and post-flower (4DPA) for three biological repeats using a TRIZOL Reagent (Invitrogen) according to the manufacturer's instruction. Total RNA was treated by DNase I to remove any genomic DNA contamination and checked by RNA gel and Agilent RNA 6000 Nano Assay, which the RIN value above 7 was determined to be qualified. After that, the best RNA were sent out for deep RNA sequencing using Illumina HiSeq2000 and the reads generated were mapped to the tomato genome sequence SL2.40. The data are deposited at NCBI SRA database under the accession number SRP029978. The gene expression was calculated for each annotated tomato gene (iTAG2.30). For continuous validation, first strand cDNA was synthesized as previously described and PCR was performed using primers designed from the intron and exon of 7 *Sl-ARF* genes. The primer sequences are listed in Table S4 in File S1. An aliquot of 1 µl of the product was used as a template. The PCR amplification cycle was as follows: 95°C for 30 s, 56–60°C for 40 s, 72°C for 30 s–2.5 min. Samples were

References

- Quint M, Gray WM (2006) Auxin signaling. *Curr Opin Plant Biol* 9: 448–453.
- Teale WD, Ditegou FA, Dovzhenko AD, Li X, Molendijk AM, et al. (2008) Auxin as a model for the integration of hormonal signal processing and transduction. *Mol Plant* 1: 229–237.
- Wang H, Schauer N, Usadel B, Frasse P, Zouine M, et al. (2009) Regulatory features underlying pollination-dependent and -independent tomato fruit set revealed by transcript and primary metabolite profiling. *Plant Cell* 21: 1428–1452.
- Sagar M, Chervin C, Mila I, Hao Y, Roustan JP, et al. (2013) SIARF4, an auxin response factor involved in the control of sugar metabolism during tomato fruit development. *Plant Physiol* 161: 1362–1374.
- Sagar M, Chervin C, Bouzayen M, Zouine M (2013) Under-expression of the Auxin Response Factor Sl-ARF4 improves post-harvest behavior of tomato fruits. *Plant Signal Behav* 8.
- Theologis A, Huynh TV, Davis RW (1985) Rapid induction of specific mRNAs by auxin in pea epicotyl tissue. *J Mol Biol* 183: 53–68.

taken after 25, 30 or 35 cycles and 10 µl of the PCR product was visualized on a 2–2.5% agarose gel. All PCRs were carried out in a Mastercycler (Eppendorf, Hamburg, Germany). DNA was stained with ethidium bromide in the gel. Sl-Ubi3 expression was used as an internal control.

Supporting Information

File S1 Supporting tables and figures. Table S1. uORF prediction in the 5'UTR leader sequences of Sl-ARFs. **Table S2.** *In silico* analysis of *Sl-ARF* gene promoters. **Table S3.** Quantitative RT-PCR primers of *Sl-ARF* genes. **Table S4.** PCR primers for identifying the alternative splicing expressed forms in *Sl-ARF* genes. **Figure S1. *Sl-ARF* genes genomic distribution on the tomato chromosomes.** The arrows next to gene names show the direction of transcription. The number near to each *Sl-ARF* designates the position megabases (Mb) of the first ATG in the tomato chromosome pseudomolecules (tomato genome version SL2.40). The chromosome numbers and their corresponding size are indicated at the top and bottom of each bar. **Figure S2. Phylogenetic relationship between tomato Sl-ARF genes.** The unrooted tree was generated using MEGA4 program by neighbor-joining method. Bootstrap values (above 50%) from 1000 replicates are indicated at each branch. Sl-ARFs with a star (*) are deprived of domain III and IV necessary for interaction with Aux/IAAs. **Figure S3.1–6. Predicted alternative splicing in six Sl-ARFs (Figure S3.1 to Figure S3.6).** RNA-seq reads generated during the fruit-set and mapped on the corresponding *Sl-ARF* gene sequence (*Sl-ARF2B*, 3, 4, 8A, 8B, and 2A) showing predicted alternative splicing events. RNA-seq reads are represented by red and blue rod arrows. (PDF)

Acknowledgments

This work benefited from the networking activities within the European funded COST ACTION FA1106 “Qualityfruit”. The authors thank C. Pecher and A. Zakaroff-Girard for their technical assistance and expertise in flow cytometry (Cytometry and cell sorting platform, INSERM UPS UMR 1048, Toulouse RIO imaging platform). The authors also thank O. Boucher and N. Marsaud for their technical assistance and expertise in RNA-seq data production (plateforme genotoul, GeT).

Author Contributions

Conceived and designed the experiments: MZ MB. Performed the experiments: MZ YF ALCB IM PF HW CA. Analyzed the data: MZ YF MB JPR. Contributed reagents/materials/analysis tools: MZ ALCB IM JPR. Wrote the paper: MZ MB JPR.

- Guilfoyle T, Hagen G, Ulmasov T, Murfett J (1998) How does auxin turn on genes? *Plant Physiol* 118: 341–347.
- Tiwari SB, Hagen G, Guilfoyle T (2003) The roles of auxin response factor domains in auxin-responsive transcription. *Plant Cell* 15: 533–543.
- Hayashi K (2012) The interaction and integration of auxin signaling components. *Plant Cell Physiol* 53: 965–975.
- Szemenyei H, Hannon M, Long JA (2008) TOPLESS mediates auxin-dependent transcriptional repression during Arabidopsis embryogenesis. *Science* 319: 1384–1386.
- Causier B, Lloyd J, Stevens L, Davies B (2012) TOPLESS co-repressor interactions and their evolutionary conservation in plants. *Plant Signal Behav* 7: 325–328.
- Ulmasov T, Murfett J, Hagen G, Guilfoyle TJ (1997) Aux/IAA proteins repress expression of reporter genes containing natural and highly active synthetic auxin response elements. *Plant Cell* 9: 1963–1971.

13. Wang L, Kim J, Somers DE (2013) Transcriptional corepressor TOPLESS complexes with pseudoresponse regulator proteins and histone deacetylases to regulate circadian transcription. *Proc Natl Acad Sci U S A* 110: 761–766.
14. Audran-Delalande C, Bassa C, Mila I, Regad F, Zouine M, et al. (2012) Genome-wide identification, functional analysis and expression profiling of the Aux/IAA gene family in tomato. *Plant Cell Physiol* 53: 659–672.
15. Rademacher EH, Lokere AS, Schlereth A, Llavata-Peris CI, Bayer M, et al. (2012) Different auxin response machineries control distinct cell fates in the early plant embryo. *Dev Cell* 22: 211–222.
16. Cole M, Chandler J, Weijers D, Jacobs B, Comelli P, et al. (2009) DORNROSCHEN is a direct target of the auxin response factor MONOPTEROS in the Arabidopsis embryo. *Development* 136: 1643–1651.
17. Lim PO, Lee IC, Kim J, Kim HJ, Ryu JS, et al. (2010) Auxin response factor 2 (ARF2) plays a major role in regulating auxin-mediated leaf longevity. *J Exp Bot* 61: 1419–1430.
18. Wilmoth JC, Wang S, Tiwari SB, Joshi AD, Hagen G, et al. (2005) NPH4/ARF7 and ARF19 promote leaf expansion and auxin-induced lateral root formation. *Plant J* 43: 118–130.
19. Ellis CM, Nagpal P, Young JC, Hagen G, Guilfoyle TJ, et al. (2005) AUXIN RESPONSE FACTOR1 and AUXIN RESPONSE FACTOR2 regulate senescence and floral organ abscission in Arabidopsis thaliana. *Development* 132: 4563–4574.
20. Marin E, Jouannet V, Herz A, Lokere AS, Weijers D, et al. (2010) miR390, Arabidopsis TAS3 tasiRNAs, and their AUXIN RESPONSE FACTOR targets define an autoregulatory network quantitatively regulating lateral root growth. *Plant Cell* 22: 1104–1117.
21. Yoon EK, Yang JH, Lee WS (2010) Auxin and Abscisic Acid Responses of Auxin Response Factor 3 in Arabidopsis Lateral Root Development. *Journal of Plant Biology* 53: 150–154.
22. Varaud E, Brioude F, Szecei J, Leroux J, Brown S, et al. (2011) AUXIN RESPONSE FACTOR8 regulates Arabidopsis petal growth by interacting with the bHLH transcription factor BIGPETALp. *Plant Cell* 23: 973–983.
23. Goetz M, Vivian-Smith A, Johnson SD, Koltunow AM (2006) AUXIN RESPONSE FACTOR8 is a negative regulator of fruit initiation in Arabidopsis. *Plant Cell* 18: 1873–1886.
24. Goetz M, Hooper LC, Johnson SD, Rodrigues JC, Vivian-Smith A, et al. (2007) Expression of aberrant forms of AUXIN RESPONSE FACTOR8 stimulates parthenocarpic in Arabidopsis and tomato. *Plant Physiol* 145: 351–366.
25. Guillon F, Philippe S, Bouchet B, Devaux MF, Frasse P, et al. (2008) Down-regulation of an Auxin Response Factor in the tomato induces modification of fine pectin structure and tissue architecture. *J Exp Bot* 59: 273–288.
26. de Jong M, Wolters-Arts M, Feron R, Mariani C, Vriezen WH (2009) The Solanum lycopersicum auxin response factor 7 (SlARF7) regulates auxin signaling during tomato fruit set and development. *Plant J* 57: 160–170.
27. Li H, Johnson P, Stepanova A, Alonso JM, Ecker JR (2004) Convergence of signaling pathways in the control of differential cell growth in Arabidopsis. *Dev Cell* 7: 193–204.
28. Jain M, Khurana JP (2009) Transcript profiling reveals diverse roles of auxin-responsive genes during reproductive development and abiotic stress in rice. *Febs Journal* 276: 3148–3162.
29. de Jong M, Wolters-Arts M, Garcia-Martinez JL, Mariani C, Vriezen WH (2011) The Solanum lycopersicum AUXIN RESPONSE FACTOR 7 (SlARF7) mediates cross-talk between auxin and gibberellin signalling during tomato fruit set and development. *J Exp Bot* 62: 617–626.
30. Li J, Dai X, Zhao Y (2006) A role for auxin response factor 19 in auxin and ethylene signaling in Arabidopsis. *Plant Physiol* 140: 899–908.
31. Liu ZH, Yu YC, Xiang FN (2011) [Auxin response factors and plant growth and development]. *Yi Chuan* 33: 1335–1346.
32. Vert G, Walcher CL, Chory J, Nemhauser JL (2008) Integration of auxin and brassinosteroid pathways by Auxin Response Factor 2. *Proc Natl Acad Sci U S A* 105: 9829–9834.
33. Guilfoyle TJ, Hagen G (2007) Auxin response factors. *Curr Opin Plant Biol* 10: 453–460.
34. Guilfoyle TJ, Hagen G (2001) Auxin response factors. *Journal of Plant Growth Regulation* 20: 281–291.
35. Ulmasov T, Hagen G, Guilfoyle TJ (1999) Activation and repression of transcription by auxin-response factors. *Proceedings of the National Academy of Sciences of the United States of America* 96: 5844–5849.
36. Nagpal P, Ellis CM, Weber H, Ploense SE, Barkawi LS, et al. (2005) Auxin response factors ARF6 and ARF8 promote jasmonic acid production and flower maturation. *Development* 132: 4107–4118.
37. Jones B, Frasse P, Olmos E, Zegzouti H, Li ZG, et al. (2002) Down-regulation of DR12, an auxin-response-factor homolog, in the tomato results in a pleiotropic phenotype including dark green and blotchy ripening fruit. *Plant Journal* 32: 603–613.
38. Wang DK, Pei KM, Fu YP, Sun ZX, Li SJ, et al. (2007) Genome-wide analysis of the auxin response factors (ARF) gene family in rice (*Oryza sativa*). *Gene* 394: 13–24.
39. Kalluri UC, Difazio SP, Brunner AM, Tuskan GA (2007) Genome-wide analysis of Aux/IAA and ARF gene families in *Populus trichocarpa*. *BMC Plant Biol* 7: 59.
40. Paterson AH, Bowers JE, Bruggmann R, Dubchak I, Grimwood J, et al. (2009) The Sorghum bicolor genome and the diversification of grasses. *Nature* 457: 551–556.
41. Xing H, Pudake RN, Guo G, Xing G, Hu Z, et al. (2011) Genome-wide identification and expression profiling of auxin response factor (ARF) gene family in maize. *BMC Genomics* 12: 178.
42. Kumar R, Tyagi AK, Sharma AK (2011) Genome-wide analysis of auxin response factor (ARF) gene family from tomato and analysis of their role in flower and fruit development. *Molecular Genetics and Genomics* 285: 245–260.
43. Wu J, Wang F, Cheng L, Kong F, Peng Z, et al. (2011) Identification, isolation and expression analysis of auxin response factor (ARF) genes in *Solanum lycopersicum*. *Plant Cell Reports* 30: 2059–2073.
44. Ottenschlager I, Wolff P, Wolverton C, Bhalerao RP, Sandberg G, et al. (2003) Gravity-regulated differential auxin transport from columella to lateral root cap cells. *Proc Natl Acad Sci U S A* 100: 2987–2991.
45. Maza E, Frasse P, Senin P, Bouzayen M, Zouine M (2013) Comparison of normalization methods for differential gene expression analysis in RNA-Seq experiments: A matter of relative size of studied transcriptomes. *Communicative & Integrative Biology* 6.
46. (2012) The tomato genome sequence provides insights into fleshy fruit evolution. *Nature* 485: 635–641.
47. Serrani JC, Ruiz-Rivero O, Fos M, Garcia-Martinez JL (2008) Auxin-induced fruit-set in tomato is mediated in part by gibberellins. *Plant Journal* 56: 922–934.
48. de Jong M, Mariani C, Vriezen WH (2009) The role of auxin and gibberellin in tomato fruit set. *Journal of Experimental Botany* 60: 1523–1532.
49. Wang H, Jones B, Li Z, Frasse P, Delalande C, et al. (2005) The tomato Aux/IAA transcription factor IAA9 is involved in fruit development and leaf morphogenesis. *Plant Cell* 17: 2676–2692.
50. Devghalaere F, Doucen T, Guitton B, Keeling J, Payne W, et al. (2012) A genomics approach to understanding the role of auxin in apple (*Malus x domestica*) fruit size control. *BMC Plant Biol* 12: 7.
51. Wang JW, Wang LJ, Mao YB, Cai WJ, Xue HW, et al. (2005) Control of root cap formation by MicroRNA-targeted auxin response factors in Arabidopsis. *Plant Cell* 17: 2204–2216.
52. Williams L, Carles CC, Osmont KS, Fletcher JC (2005) A database analysis method identifies an endogenous trans-acting short-interfering RNA that targets the Arabidopsis ARF2, ARF3, and ARF4 genes. *Proc Natl Acad Sci U S A* 102: 9703–9708.
53. Wu MF, Tian Q, Reed JW (2006) Arabidopsis microRNA167 controls patterns of ARF6 and ARF8 expression, and regulates both female and male reproduction. *Development* 133: 4211–4218.
54. Zhou F, Roy B, von Arnim AG (2010) Translation reinitiation and development are compromised in similar ways by mutations in translation initiation factor eIF3h and the ribosomal protein RPL24. *BMC Plant Biol* 10: 193.
55. Rosado A, Li R, van de Ven W, Hsu E, Raikhel NV (2012) Arabidopsis ribosomal proteins control developmental programs through translational regulation of auxin response factors. *Proc Natl Acad Sci U S A* 109: 19537–19544.
56. Abel S, Theologis A (1994) Transient transformation of Arabidopsis leaf protoplasts: a versatile experimental system to study gene expression. *Plant J* 5: 421–427.
57. Hamilton AJ, Lycett GW, Grierson D (1990) Antisense Gene That Inhibits Synthesis of the Hormone Ethylene in Transgenic Plants. *Nature* 346: 284–287.