

OPTIMIZATION OF ULTRASOUND ASSISTED EXTRACTION AND  
ANTIOXIDANT ACTIVITY OF *PHALERIA MACROCARPA*

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Especially dedicated to M, thank you for the hard work.

AS, thank you for willingly went through hard ship especially the last 10 days.

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## ABSTRACT

In the past decades, markets had been flooded with herbal based products including *Phaleria macrocarpa* (*P. macrocarpa*) or known as Mahkota Dewa. Previous studies reported that this plant contains high amount of active compound that are responsible for various bioactivities. However, these active compounds are sensitive towards many factors; temperature, pH, light and others in the production line. Thus, this study was conducted to optimize the extraction condition using ultrasound assisted extraction (UAE), determine the primary metabolites of the optimized extract and investigate the antioxidant activities using 2, 2-diphenyl-1-picrylhydrazine radical scavenging activity and ferric reducing antioxidant power assays. The result showed that concentration of solvent, solid to solvent ratio and sonication time affected percentage yield, total phenolic content (TPC), total flavonoid content (TFC) and antioxidant activities. The optimized parameters for the extraction are 75% methanol, 1: 31 solid to solvent ratio and 4.06 minutes of sonication time. Under these conditions, *P. macrocarpa* extract yielded 18.54% of crude extract, 81.59 mg GAE/g (milligram gallic acid/gram) of TPC and 28.17 mg QUE/g (milligram quercetin/gram) of TFC. Primary metabolite assays showed that *P. macrocarpa* optimized extract contained  $40.427 \pm 1.127$  mg BSA/g (milligram bovine serum albumin/gram) of protein,  $23.981 \pm 0.2306$  mg aescin/g of saponin and  $0.853 \pm 0.0452$  mg glucose/g of polysaccharides. Meanwhile, quantification of phenolic compound (gallic acid) and flavonoid compounds (quercetin, rutin and kaempferol) were done using high performance liquid chromatography and it was found that all compounds were present in the optimized extract with amount of 23.908 mg/g, 6.262 mg/g, 0.393 mg/g and 0.077 mg/g for rutin, gallic acid, kaempferol and quercetin, respectively. This extract was then tested for antioxidant activities and it exhibited potent antioxidant activities in both assays due to the relatively high TPC and TFC. This study suggested that extraction using UAE under controlled parameters contributed to the high antioxidant activities in *P. macrocarpa* and therefore can improve the quality of herbal products in the market.

## ABSTRAK

Sejak beberapa dekad yang lalu, pasaran telah dibanjiri dengan pelbagai produk berasaskan herba termasuk *Phaleria macrocarpa* (*P. macrocarpa*) atau dikenali sebagai Mahkota Dewa. Kajian sebelum ini melaporkan bahawa tumbuhan ini mengandungi sebatian aktif tersebut yang tinggi yang bertanggungjawab terhadap pelbagai bioaktiviti. Walau bagaimanapun, sebatian aktif adalah sensitif terhadap pelbagai faktor; suhu, pH, cahaya dan lain-lain dalam barisan pengeluaran. Oleh itu, kajian ini dijalankan untuk mengoptimumkan keadaan proses pengekstrakan menggunakan pengekstrakan dengan bantuan ultrasonik (UAE), menentukan metabolit utama ekstrak yang telah dioptimumkan dan menyiasat aktiviti antioksidan ekstrak tersebut menggunakan aktiviti cerakin memerangkap radikal 2, 2-difenil-1-pikrilhidrazin dan cerakin kuasa antioksidan penurunan ferik. Hasilnya menunjukkan bahawa kepekatan pelarut, nisbah pepejal kepada pelarut dan tempoh sonikasi mempengaruhi peratusan hasil, jumlah kandungan fenolik (TPC), jumlah kandungan flavonoid (TFC) serta aktiviti antioksidan. Parameter yang optimum untuk proses pengekstrakan adalah 75% kepekatan pelarut, 1: 31 nisbah pepejal kepada pelarut dan 4.06 minit tempoh sonikasi. Di bawah keadaan ini, ekstrak *P. macrocarpa* menghasilkan 18.54% peratusan hasil, 81.59 mg GAE/g (miligram asid galik/gram) TPC dan 28.17 mg QUE/g (miligram kuersetin/gram) TFC. Cerakin metabolit utama menunjukkan bahawa ekstrak *P. macrocarpa* yang optimum mengandungi  $40.427 \pm 1.127$  mg BSA/g (miligram albumin serum lembu/gram) protein,  $23.981 \pm 0.2306$  mg aasin/g saponin dan  $0.853 \pm 0.0452$  mg glukosa/g polisakarida. Sementara itu, pengkuantitian sebatian fenolik (asid galik) dan sebatian flavonoid (kuersetin, rutin dan kaempferol) telah dilakukan menggunakan kromatografi cecair berprestasi tinggi dan didapati bahawa semua sebatian wujud dalam ekstrak optimum dengan jumlah 23.908 mg/g, 6.262 mg/g, 0.393 mg/g dan 0.0767 mg/g masing-masing untuk rutin, asid galik, kaempferol dan kuersetin. Kemudian, ekstrak ini diuji untuk aktiviti antioksidan dan ekstrak tersebut menunjukkan aktiviti antioksidan yang kuat pada kedua-dua cerakin selari dengan kandungan TPC dan TFC yang tinggi di dalamnya. Kajian ini mencadangkan bahawa pengekstrakan menggunakan UAE di bawah parameter yang dikawal menyumbang kepada aktiviti antioksidan yang tinggi bagi *P. macrocarpa* dan dengan itu boleh meningkatkan kualiti produk herba di pasaran.

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**LIST OF SYMBOLS**

%	-	Percentage
°C	-	Degree Celsius
μl	-	Microliter
μg	-	Microgram
g	-	Gram
Ha	-	Alternative hypothesis
Ho	-	Null hypothesis
mg	-	Milligram
min	-	Minute
ml	-	Milliliter
nm	-	Nanometer
ppm	-	Part per million
R <sup>2</sup>	-	Coefficient of multiple determination



## LIST OF ABBREVIATIONS

ANOVA	-	Analysis of variance
BSA	-	Bovine serum albumin
C	-	Concentration
CCD	-	Central composite design
Df	-	Dilution factor
DPPH	-	2, 2-diphenyl-1-picrylhydrazine
FRAP	-	Ferric reducing antioxidant power activity
GAE	-	Gallic acid
GC-MS	-	Gas Chromatography-Mass Spectrophotometer
LC-MS	-	Liquid Chromatography- Mass Spectrophotometer
HPLC	-	High Performance Liquid Chromatography
P	-	Probability
<i>P. macrocarpa</i>	-	<i>Phaleria macrocarpa</i>
QUE	-	Quercetin
RSM	-	Response surface methodology
TFC	-	Total flavonoid content
TPC	-	Total phenolic content
Uv-vis	-	Ultraviolet-visible

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## CHAPTER 1

### INTRODUCTION

#### 1.1 General Background

*Phaleria macrocarpa* (*P. macrocarpa*) is the scientific name for Mahkota Dewa and can be found throughout the year in tropical areas. It is traditionally used to treat cancer, impotency, hemorrhoids, diabetes mellitus, allergies, liver and heart diseases, kidney disorders, blood related diseases, acne, stroke, migraine, and various skin ailments (Zhang *et al.*, 2006; Hendra *et al.*, 2011a and Hendra *et al.*, 2011b). *P. macrocarpa* attracts scientists around the globe to do extensive research and revealed the bioactivity potential of this plants.

*P. macrocarpa* based product had been penetrating herbal market few years back and the demand towards the product kept increasing. Most products available in market nowadays claim for high antioxidant activity, that includes *P. macrocarpa* based product. However, compounds responsible for antioxidant may degrade in the processing line due to the sensitivity towards extreme conditions. Therefore, it is important to choose the best method of extraction and control the conditions of extraction so that the active compounds can be retained till the end product. There are several advance methods of extraction that has numerous advantages in many ways. Previous study suggested that ultrasound assisted extraction (UAE) method

can easily be employed in the existing extraction machine for manufacturing plant. UAE had been proven extracting in a short time with higher percentage yield of active compounds (Vilkhu *et al.*, 2008; Gil-Chávez *et al.*, 2013).

There are a growing number of studies regarding *P. macrocarpa* in the past few years. However, there have been no reports on optimizing extraction of *P. macrocarpa* using ultrasonic-assisted extraction method as well as comparing antioxidant activity between 2, 2-diphenyl-1-picrylhydrazil (DPPH) Radical Scavenging Activity and ferric reducing antioxidant power activity (FRAP). By employing UAE method of extraction, percentage yield of active compounds can be increased and thus, contributed to the high antioxidant activity of that particular extract. In accordance with a tendency of increasing demand to consume products rich in antioxidants, optimization extraction of the *P. macrocarpa* extract and antioxidant activity is studied in this research.

## 1.2 Problem Statement

Malaysian's herbal market is saturated with a lot of local herbs mainly in nutraceutical, pharmaceutical and cosmeceutical fields. Many products claimed to have lots of biological benefits, for example supplement high in antioxidants as the result of increasing awareness among consumers. However, the increasing awareness on consuming natural products leads to high demand on natural and herbal market. There is no doubt that the increasing awareness among consumers provides positive economic growth in Malaysia however, for manufacturers, high demand leads to the massive production. Production, especially the one involving extraction of active compounds needs large volume of raw material. There are a lot of challenges need to be tackled in order to meet the growing market demand. One of the most vital phases in manufacturing is the processing.

Herbal plants naturally contained high antioxidant activity due to the presence of various active compounds for example phenolic and flavonoid compounds. However, this activity is reduced with the reduction of total active compounds remain after extraction process. Some active compounds are heat sensitive, light sensitive, pH sensitive and others. These make them unstable and most likely will denature due to the unfavorable conditions during drying and extraction process. In extraction process, methods and parameters play significant role in contributing the quantity and quality of the end product; the extract. Many scholars agreed that conventional methods of extraction are no longer suitable in term of yield, environmental friendly and even production cost to meet market demand. Therefore, in the past decade, a lot of studies proposed the improvised conventional method and others provide a totally new and advance method like supercritical fluid extraction.

In order to meet consumer demand, manufacturers choose to scaling up the production. Scaling up in industrial production often involve huge amount of investment and effort, however, it does not ensuring promised returns. However, optimization processing method and conditions need to be completed prior to scaling up. Manufacturer should not only consider most suitable method. They should also consider the parameters of that particular method. The optimization parameters during extraction may help in increasing extracted active compound. Based on previous study of other plants, UAE showed a great supremacy of extracting active compounds in a short period of time. Since the extraction of active compounds increased, the antioxidant activity which was contributed greatly by the presence of active compounds will most like to increase as well. Thus, this research studied on ultrasonic-assisted extraction (UAE) method by controlling the condition of extraction. Subsequently, the percentage yield, total phenolic content (TPC) and total flavonoid content (TFC) are increased. Thereby, applying correct method and conditions of extraction is crucial to optimize percentage yield, TPC as well as TFC of *P. macrocarpa*.

### 1.3 Significant of the Study

With the increase of awareness among consumers regarding healthy living, the development of high antioxidant content based product had been increased rapidly for the past few years. This plant especially can be capitalized as source of antioxidant agent and can be commercialized as value-added ingredient into nutraceutical and cosmeceutical based product. At the end of this study, the optimized parameter of UAE can guide herbal related manufacturers to boost their production. The quantity and quality of extract can be increased with the employment of best extraction method. These will leads to the decrement of natural resources wastage

### 1.4 Objective of the Study

The objectives of this study are:

- i. To optimize the yield of *P. macrocarpa* fruit extract using ultrasonic-assisted extraction (UAE) method and determine the primary metabolite of the optimized extract.
- ii. To investigate antioxidant activity of *P. macrocarpa* fruit extract using antioxidant activity assay of DPPH scavenging activity and Ferric Reducing Antioxidant Power (FRAP) assay.

## 1.5 Scope of the Study

- i. Screening of parameter conditions (percentage of solvent, %, solid to solvent ratio, g/g, sonication time, min) of extraction process using ultrasonic-assisted extraction (UAE) method and screening of selected phytochemical compounds onto the extracts.
- ii. Optimization extraction of phytochemical compounds from *P. macrocarpa* using UAE method with methanol as solvent.
- iii. Quantification of selected phenolic and flavonoid compounds using high performance liquid chromatography (HPLC) method.
- iv. Determination of primary metabolites (total polysachharide, total glycosaponin and total protein) on the optimized extract.
- v. Investigation of antioxidant activity assays of *P. macrocarpa* using 2, 2-diphenyl-1-picrylhydrazine scavenging activity (DPPH) and ferric reducing antioxidant power (FRAP) assay.

## REFERENCES

- Agung, B. S., Faried, A., Arifin, M. Z., Wiriadisastra, K., and Ohta, T. (2008). Herbal Medicine Isolation, *Phaleria macrocarpa* for Primary Glioblastoma Multiforme Cells. *Ann Epidemiol.* 18,708–741.
- Ahmadi, M., Vahabzadeh, F., Bonakdarpour, B., Mofarrah, E. and Mehranian, M. (2005). Application of the Central Composite Design and Response Surface Methodology to the Advanced Treatment of Olive Oil Processing Wastewater Using Fenton's Peroxidation. *Journal of Hazardous Materials.* B123, 187–195.
- Ahmed, N. R. (2016). Optimization and Characterization of Extraction and Spray Drying Parameters of *Ficus deltoidea* Extract. Master of Engineering, Universiti Teknologi Malaysia, Johor Bahru.
- Alim, N. A. S. M. A., Sulaiman, A. Z., and Ajit, A. (2016). Application of Ultrasound on the Extraction of Vitexin from *Ficus deltoidea* Leaves. *ARPN Journal of Engineering and Applied Sciences.* 11, 2199-2204.
- Altaf, R., Asmawi, M. Z., Dewa, A., Sadikun, A., and Muhammad Ihtisham Umar, M. I. (2013). Phytochemistry and Medicinal Properties of *Phaleria macrocarpa* (Scheff.) Boerl. Extracts. *Pharmacogn Rev.* 7, 73–80.
- Andrean, D., Prasetyo, S., Kristijarti, A. P., and Hudaya, T. (2014). The Extraction and Activity Test of Bioactive Compounds in *Phaleria macrocarpa* as Antioxidants. *Procedia Chemistry.* 9, 94–101.
- Ali, R. B., Atangwho, I. J., Kaur, N., Abraika, O. S., Ahmad, M., Mahmud, R., and Asmawi, M. Z. (2012). Bioassay-Guided Antidiabetic Study of *Phaleria macrocarpa* Fruit Extract. *Molecules.* 17 4986–5002.
- Altemimi, A., Choudhry, R., Watson, D. G. and Lightfoot, D. A. (2015). Effect of Ultrasonic Treatments on the Polyphenol and Antioxidant Content of Spinach Extract. *Ultrasonic Sonochemistry.* 24, 247-255.



- Antolovich, M., Prenzler, P., Patsalides, E., McDonald, S. and Robards, K. (2002). Methods for Testing Antioxidant Activity. *The Analyst*. 127, 183-198.
- Astuti, E., Raharjo, T. J., and Eviane, D. (2007). Cytotoxicity of *Phaleria macrocarpa* (Scheff) boerl fruit Flesh and Seed Extract of Ethanol and Its Effect against P53 and Bcl-2 Genes Expression of Normal Cell. *Proceedings of the International Conference on Chemical Sciences*. Yogyakarta Indonesia, 1–4.
- Azmir, J., Zaidul, I. S. M., Rahman, M. M., Sharif, K. M., Mohamed, A., Sahena, F., Jahurul, M. H. A., Ghafoor, K., Norulaini, N. A. N. and Omar, A. K. M. (2013). Techniques for extraction of bioactive compounds from plant materials: A review. *Journal of Food Engineering*. 117, 426–436.
- Azmir, J., Zaidul, I. S. M., Sharif, K. M., Uddin, M. S., Jahurul, M. H. A., Jinap, S., Hajeb, P., and Mohamed, A. (2014). Supercritical Carbon Dioxide Extraction of Highly Unsaturated Oil from *Phaleria macrocarpa* Seed. *Food Res Int*. 65, 394–400.
- Badarinath, A. V., RAO, K. M., Chetty, C. M. S., Ramkanth, S., Rajan, T. V. S., and Gnanaprakash, K. (2010). A Review on In-vitro Antioxidant Methods: Comparisons, Correlations and Considerations. *International Journal of PharmTech Research*. 2, 1276-1285.
- Bag, G.C., Grihanjali, D. P., and Bhaigyabati, T. (2015). Assessment of Total Flavonoid Content and Antioxidant Activity of Methanolic Rhizome Extract of Three Hedychium Species of Manipur Valley. *Int. J. Pharm. Sci. Rev. Res*. 30, 154-159.
- Baldi, A., Pandit, M. K., and Ranka, P. (2012). Amelioration of *in-vivo* Antioxidant Activity by Banana Extracts. *International Journal of Pharmaceutical & Biological Archives*. 3, 157-161.
- Benzie, I. F. F. and Strain, J. J. (1996). The Ferric Reducing Ability of Plasma (FRAP) as a Measure of “Antioxidant Power”. *Analytical Biochemistry*. 239, 70-76.
- Biesaga, M. (2011). Influence of extraction methods on stability of flavonoids. *Journal of Chromatography A*. 1218, 2505–2512.
- Brahim, M., Gambier, F. and Brosse, N. (2014). Optimization of polyphenols extraction from grape residues in water medium. *Industrial Crops and Products*. 52, 18-22.

- Cacace, J. E. and Mazza, G. (2003a). Mass Transfer Process during Extraction of Phenolic Compounds from Milled Berries. *Journal of Food Engineering*. 59, 379–389.
- Cacace, J. E. and Mazza, G. (2003b). Optimization of Extraction of Anthocyanins from Black Currants with Aqueous Ethanol. *Journal of Food Science*. 68, 240-248.
- Carocho, M., and Ferreira, I. C. F. R. (2013). A Review on Antioxidants, Prooxidants and Related Controversy: Natural and Synthetic Compounds, Screening and Analysis Methodologies and Future Perspectives. *Food and Chemical Toxicology*. 51, 15–25.
- Chemat, F., Zill-e-Huma and Khan, M. K. (2011). Applications of ultrasound in food technology: Processing, preservation and extraction. *Ultrasonics Sonochemistry*. 18, 813–835.
- Chen, M., Zhao, Y. and Yu, S. (2015). Optimisation of ultrasonic-assisted extraction of phenolic compounds, antioxidants, and anthocyanins from sugar beet molasses. *Food Chemistry*. 172, 543-550.
- Chen, R., Liu, Z., Zhao, J., Chen, R., Meng, F., Zhang, M. and Ge, W. (2011). Antioxidant and Immunobiological Activity of Water-Soluble Polysaccharide Fractions Purified from *Acanthopanax senticosu*. *Food Chemistry*. 127, 434–440.
- Cheok, C. Y., Chin, N. L., Yusof, Y. A., Talib, R. A. and Law, C. L. (2013). Optimization of Total Monomeric Anthocyanin (TMA) and Total Phenolic Content (TPC) Extractions from Mangosteen (*Garcinia mangostana* Linn.) Hull Using Ultrasonic Treatments. *Industrial Crops and Products*. 50, 1-7.
- Chong, S. C., Dollah, M. A., Chong, P. P., and Maha, A. (2011). *Phaleria macrocarpa* (Scheff.) Boerl fruit Aqueous Extract Enhances LDL Receptor and PCSK9 Expression *in vivo* and *in vitro*. *J Ethnopharmacol*. 137, 817–827.
- Chua, L. S., Amin, N. A. M, Neoc, J. C. H., Lee, T. H., Lee, C. T., Sarmidi, M. R. and Aziz, R. A. (2011). LC–MS/MS-based metabolites of *Eurycoma longifolia* (Tongkat Ali) in Malaysia (Perak and Pahang). *Journal of Chromatography B*. 879, 3909– 3919.
- Dias, D. A., Hill, C. B., Jayasinghe, N. S., Atieno, J., Sutton, T. and Roessner, U. 2015. Quantitative profiling of polar primary metabolites of two chickpea

- cultivars with contrasting responses to salinity. *Journal of Chromatography B*. 1000, 1-13.
- Diouf, P. N., Stevanovic, T. and Cloutier, A. (2009). Study on Chemical Composition, Antioxidant and Anti-Inflammatory Activities of Hot Water Extract From *Piceam mariana* Bark and Its Proanthocyanidin-Rich Fractions. *Food Chemistry*. 113, 897-902.
- Dudonnè, S., Vitrac, X., Coutière, P., Woillez, M., and Mèrillon, J. M. (2009). Comparative Study of Antioxidant Properties and Total Phenolic Content of 30 Plant Extracts of Industrial Interest Using DPPH, ABTS, FRAP, SOD, and ORAC Assays. *J. Agric. Food Chem.* 57, 1768–1774.
- Easmin, M. T. S., Sarker, M. Z. I., Ferdosh, S., Shamsudin, S. H., Yunus, K., Uddin, M. S., Sarker, M. M. R., Akanda, M.J. H., Hossain, M. S., and Khalil, H. P. S. A. (2014). Bioactive Compounds and Advanced Processing Technology: *Phaleria macrocarpa* (sheff.) Boerl, a Review. *J Chem Technol Biotechnol*. 2014, 1-11.
- El-Olemyl, M. M., Al-Muhtadi, F. J., and Afifi, A. A. (1994). Experimental Phytochemistry. *AQ Laboratory Manual, College of Pharmacy, Kingsaud University*. 1-134. King Saud University Press.
- Faried, A., Kurnia, D., Faried, L. S., Usman, N., Miyazaki, T., and Kato H. (2007). Anticancer Effects of Gallic Acid Isolated from Indonesian Herbal Medicine, *Phaleria macrocarpa* (Scheff.) Boerl, on Human Cancer Cell Lines. *Int J Oncol*. 30, 605–613.
- Fariza, N., Fadzureena, J., Zunoliza, A., Chuah, A. L., Pin, K. Y., and Adawiah, I. (2012). Anti-inflammatory Activity of the Major Compound from Methanol Extract of *Phaleria macrocarpa* Leaves. *Journal of Applied Science*. 12, 1195–1198.
- Fariza, N., Chuah, L., Pin, K. Y., Dayang Radiah, A. B., Umi Kalsom, Y. and Adawiah, I (2014). Optimisation of Extraction of *Phaleria macrocarpa* Leaves. *Med Aromat Plants*. 3, 1-3.
- Ghasemzadeh, A., Jaafar, H. Z. E. and Rahmat, A. (2011). Effects of Solvent Type on Phenolics and Flavonoids Content and Antioxidant Activities In Two Varieties of Young Ginger (*Zingiber officinale* Roscoe ) Extracts, *Journal of Medicinal Plants Research*. 5, 1147–1154.

- Gil-Chávez, G.J., Villa, J.A., Ayala-Zavala, J. F., Heredia, J. B., Sepulveda, D., Yahia E Mand González-Aguilar G. A. Y. E. (2013). Technologies for Extraction and Production of Bioactive Compounds to be used as Nutraceuticals and Food Ingredients: An Overview. *Comp Rev Food Sci F.* 12, 5–23.
- Giovana, B. C., Ghanem, A., Brooks, M. S. (2015). Optimization of Ultrasound-Assisted Extraction of Anthocyanins from Haskap Berries (*Lonicera caerulea L.*) using Response Surface Methodology. *Ultrasonic Sonochemistry.* 27, 449-455.
- Halliwell, B. (1991) Reactive Oxygen Species in Living Systems: Source, Biochemistry, and Role in Human Disease. *American Journal of Medicine.* 91, 14-22.
- Harborne, J. B. (1973). *Phytochemical Methods- A Guide to Modern Techniques of Plant Analysis.* Third edition, Chapman and Hall publishers, London, 7-279.
- Hasmida, M. N., Nur Syukriah, A. R., Liza, M. S. and Mohd Azizi, C. Y. (2014). Effect of Different Extraction Techniques on Total Phenolic Content and Antioxidant Activity of *Quercus infectoria* Galls. *International Food Research Journal.* 21, 1075-1079.
- Hendig W. E. K. W. (2009). Benzophenone Glucoside Isolated Fromethyl Acetate Extract of the Bark of Mahkota Dewa [*Phaleria macrocarpa* (Scheff.) Boerl.] and Its Inhibitory Activity on Leukemia L1210 Cell Line. *Indo J Chem.* 9, 142–145.
- Hendra, R., Ahmad, S., Oskoueian, E., Sukari, A. and Shukor, M. Y. (2011a). Antioxidant, Anti-inflammatory and Cytotoxicity of *Phaleria macrocarpa* (Boerl.) Scheff Fruit. *BMC Complementary and Alternative Medicine.* 2011, 110.
- Hendra, R., Ahmad, S., Sukari, A., Shukor, M. Y. and Oskoueian, E. (2011b). Flavonoid Analyses and Antimicrobial Activity of Various Parts of *Phaleria macrocarpa* (Scheff.) Boerl Fruit. *International Journal of Molecular Sciences.* 12, 3422–3431.
- Hartati, M. S., Mubarika, S., Gandjar, I. G., Hamann, M. T., Rao, K. V. and Wahyuono, S. (2005). Phalerin, A New Benzophenoic Glucoside Isolated

- from The Methanolic Extract of Mahkota Dewa (*Phaleria macrocarpa*) Leaves. *Majalah Farmasi Indones.* 16, 51–7.
- Hussein, M. A., 2011. A Convenient Mechanism for the Free Radical Scavenging Activity of Resveratrol. *International Journal of Phytomedicine.* 3, 459-469.
- Idolo, I., Marshall, L. J., Ho, P. and Williamson, G. (2016). *Hibiscus sabdariffa* (Roselle) Extracts and Wine: Phytochemical Profile, Physicochemical Properties, and Carbohydrase Inhibition. *Journal of Agriculture of Food Chemistry.* 64, 4921–4931.
- Ilaiyarajaa, N., Likhithb, K. R., Babua, G. R. S. and Khanuma, F. (2015). Optimisation of extraction of bioactive compounds from *Feronia limonia* (wood apple) fruit using response surface methodology (RSM). *Food Chemistry.* 173, 348–354.
- Innovative Ultrasonic. *Ultrasonic Innovations in the Food Industry.* 3rd Innovative Food Centre Conference, Melbourne, Australia. 16-17 October 2007. Keynote Lecture. Available at:  
<http://www.innovativeultrasonics.com/publications/Ultrasonic-Innovations-in-the-Food-Industry-From-the-Laboratory-to-Commercial-Production/>
- Ismail, H. I., Chan, K. W., Mariod, A. A. and Ismail M. (2010). Phenolic Content and Antioxidant Activity of Cantaloupe (*Cucumis melo*) Methanolic Extracts. *Food Chemistry.* 119, 643-647.
- Karimi, E., Oskoueian, E., Hendra, R. and Jaafar, H. Z. E. (2010). Evaluation of *Crocus sativus L. stigma* Phenolic and Flavonoid Compounds and Its Antioxidant Activity. *Molecules.* 15, 6244-6256.
- Kaur, S. and Mondal, P. (2014). Study of Total Phenolic and Flavonoid Content, Antioxidant Activity and Antimicrobial Properties of Medicinal Plants. *Journal of Microbiology & Experimentation.* 1, 1-6.
- Khaled, A. S., Nahla S. A. A., Ibrahim, A. S., Hegazy, M. E. F., El-Missiry, M. M. and Hammouda, F. M. (2015). Green Technology: Economically and Environmentally Innovative Methods for Extraction of Medicinal & Aromatic Plants (MAP) in Egypt. *Journal of Chemical and Pharmaceutical Research.* 7, 1050-1074.
- Khoddami, A., Wilkes, M. A. and Roberts, T. H. (2013). Techniques for Analysis of Plant Phenolic Compounds. *Molecules.* 18, 2328–75.

- Khoo, H. E. and Azlan, A. and Ismail, A. and Abas, F. (2013). Response Surface Methodology Optimization for Extraction of Phenolics and Antioxidant Capacity in Defatted *Dabai* Parts. *Sains Malaysiana*. 42, 949–954.
- Kim, W. J., Veriansyah, B., Lee, Y. W., Kim, J. and Kim, J. D. (2010). Extraction of Mangiferin from Mahkota Dewa (*Phaleria macrocarpa*) Using Subcritical Water. *J Ind Eng Chem*. 16, 425–30.
- Krishnaiah, D., Sarbatly, R., and Nithyanandam, R. (2011). A Review of the Antioxidant Potential of Medicinal Plant Species. *Food and Bioproducts Processing*. 89, 217–233.
- Kutchan, T.M. (2001). Ecological Arsenal and Developmental Dispatcher. The Paradigm of Secondary Metabolism. *Plant Physiol*. 125, 58-60.
- Lattanzio, V., Lattanzio, V. M. T. and Cardinali, A. (2006). Role of phenolics in the resistance mechanisms of plants against fungal pathogens and insects. *Phytochemistry: Advances in Research*. 2006, 23-67.)
- Lay, M. M., Karsani, S. A., Banisalam, B., Mohajer, S. and Abd Malek, S. N. (2014a). Antioxidants, Phytochemicals, and Cytotoxicity Studies on *Phaleria macrocarpa* (Scheff.) Boerl Seeds. *BioMed Research International*. 2014, 1-13.
- Lay, M. M., Karsani, S. A., Mohajer, S. and Abd Malek, S. N. (2014b). Phytochemical constituents, nutritional values, phenolics, flavonols, flavonoids, antioxidant and cytotoxicity studies on *Phaleria macrocarpa* (Scheff.) Boerl fruits. *BMC Complementary and Alternative Medicine* 14, 1-12.
- Li, H. Z., Zhang, Z. J., Hou, T. Y., Li, X. J. and Chen, T. (2015). Optimization of ultrasound-assisted hexane extraction of perilla oil using response surface methodology. *Industrial Crops and Products*. 76, 18-24.
- Li, W. K. and Fitzloff J. F. (2002). Determination of Andrographolide in Commercial Andrographis (*Andrographis paniculata*) Products Using HPLC with Evaporative Light Scattering Detection. *Journal of Liquid Chromatography and Related Technologies*. 25, 1335-1343.
- Luque-Garcia, J. L., and Luque de Castro, M. D. (2003). Where Is Microwave Based Analytical Treatment for Solid Sample Pre-Treatment Going? *Trends Anal. Chem*. 22, 90–99.

- Ma, Y. Q., Chen, J. C., Liu, D. H. and Ye, X. Q. (2009). Simultaneous extraction of phenolic compounds of citrus peel extracts: Effect of ultrasound. *Ultrasonics Sonochemistry*. 16, 57–62.
- Matsuda, Kuroyanagi, M., Sugiyama, S., Umehara, K., Ueno, A., and Nishi, K. (1994). Cell Differentiation-inducing Diterpenes from *Andrographis paniculata* Nees. *Chemical and Pharmaceutical Bulletin*. 42, 1216-1225.
- Majd, M. H., Rajaeic, A., Bashi, D. S., Mortazavif, S. A. and Bolouriang, S. (2014). Optimization of Ultrasonic-Assisted Extraction of Phenolic Compounds from Bovine Pennyroyal (*Phlomidosema parviflorum*) Leaves Using Response Surface Methodology. *Industrial Crops and Products*. 57, 195–202.
- Nazzaro, M., Mottola, M. V., Cara, F. L., Monaco, G. D., Aquino, R. P., Volpe, M. G. (2012). Extraction and Characterization of Biomolecules from Agricultural Wastes. *Chemical Engineering Transaction*. 27, 331-336.
- Opalić, M., Domitran, Z., Kom, D., Belščak, A., Horžić, D. and Karlović, D. (2009). The effect of Ultrasound Pre-Treatment and Air-Drying on the Quality of Dried Apples. *Czech Journal of Food Science*. 27, 297-300.
- Oshimi, S., Zaima, K., Matsuno, Y., Hirasawa, Y., Iizuka, T., and Studiawan, H. (2008). Studies on the Constituents from the Fruits of *Phaleria macrocarpa*. *J Nat Med*. 62, 207–10.
- Petigny, L., Périno-Issartier, S., Wajsman, J., and Chemat, F. (2013). Batch and Continuous Ultrasound Assisted Extraction of Boldo Leaves (*Peumus boldus* Mol.). *International Journal of Molecular Sciences*. 14, 5750–64.
- Pouillot, A., Polla, L. L., Tacchini, P., Neequaye, A., Polla, A., and Polla, B. (2011). *Formulating, Packaging, and Marketing of Natural Cosmetic Products, First Edition*. John Wiley and Sons Publishers, 239-257.
- Prior, R. L., Wu, X., And Schaich, K. (2005). Standardized Methods for the Determination of Antioxidant Capacity and Phenolics in Foods and Dietary Supplements. *Journal of Agriculture Food Chemistry*. 53, 4290-4302.
- Rastaon, N. and Tuah, P. M. (2016). Quantitative Analysis of Quercetin in Various Parts of *Phaleria macrocarpa* (Scheff.) Boerl Extracts. *Transactions on Science and Technology*. 3, 203 – 208.
- Riwanto, I., Budijitno, S., Dharmana, E., Handojo, D., Prasetyo, S. A., and Eko, A. (2011). Effect of *Phaleriamacrocarpa* Supplementation on Apoptosis and

- Tumor Growth of C3H Mice with Breast Cancer under Treatment with Adriamycin-Cyclophosphamide. *Int Surg.* 96, 164–170 (2011).
- Rodrigues, S., Gustavo, A. S. P. and Fabiano, A. N. F. (2008). Optimization of ultrasound extraction of phenolic compounds from coconut (*Cocos nucifera*) shell powder by response surface methodology. *Ultrasonics Sonochemistry.* 15, 95–100.
- Rosli, N. A. (2012). Preparation of Activated Carbon from Pineapple Peel and Pomelo Peel for Dyes Removal: Equilibrium, Kinetic and Thermodynamic Studies. Master of Science, Universiti Sains Malaysia, Penang.
- Rostagno, M. A., Miguel, P. and Barroso, C. G. (2003). Ultrasound-assisted extraction of soy isoflavones. *Journal of Chromatography A.* 1012, 119–128.
- Saikia, S., Mahnot, N. K. and Mahanta, C. L. (2015). Optimisation of phenolic extraction from *Averrhoa carambola* pomace by response surface methodology and its microencapsulation by spray and freeze drying. *Food Chemistry.* 171, 141-152.
- Sarkar, F. H., Padhye, S., and Ahmad, A. (2012) Role of Novel Nutraceuticals Garcinol, Plumbagin and Magniferin in the Prevention and Therapy of Human Malignancies: Mechanism of Anticancer Activity. *Nutraceuticals Cancer.* 2012, 179–199.
- Saufi, A., Heimendah, C. B., Alfermann, A. W., and Fuss E. (2008). Stereochemistry of Lignans in *Phaleria macrocarpa* (Scheff.) Boerl. *Z Naturforsch C.* 63, 13–16.
- Siddiqui, M. J., Hafizoh, S. N., Ismail, Z., Sahib, H. B., Helal, M. S. H. and Abdul Majid, A. M. S. (2009). Analysis of Total Proteins, Polysaccharides and Glycosaponins Contents of *Orthosiphon stamineus* Benth. In Spray and Freeze Dried Methanol: Water(1:1) extract and its Contribution to Cytotoxic and Antiangiogenic Activities. *Phcog Res.* 1, 320-326.
- Singleton, V. L., and Rossi, J. A. J. (1965). Colorimetry of Total Phenolics with Phosphomolybdic-Phosphotungstic Acid Reagents. *Am. J. Enol. Vitic.* 16, 144-158.
- Slater, T. F. (1984). Free-radical Mechanisms in Tissue Injury. *Journal of Biochemistry.* 222, 1–15.



- Smith, M. R., Penner, M. H., Bennett, S. E. and Bakalinsky, A. T. (2011). Quantitative Colorimetric Assay for Total Protein Applied to the Red Wine Pinot Noir. *Journal of Agriculture Food Chemistry*. 59, 6871–6876.
- Sofowora, A. (1985). *Medicinal Plants and Traditional Medicine in Africa*. John Wiley and Sons Publishers, Chichester, 190-201.
- Sufi A. (2007). Lignans in *Phaleria macrocarpa* (Scheff.) Boerl and in *Linum flavum var compactum* L. Faculty of Mathematics and Natural Sciences (Mataram: Heinrich-Heine-University Dusseldorf), p. 104.
- Sugiwati, S., and Setiasih, S. (2010). Antidiabetic Activity of Mahkota Dewa [*Phaleria macrocarpa* (Scheff.) Boerl.] Stem Extracts as an Inhibitor of Alpha-Glucosidase. *Journal of Indonesian Medicinal Plant*. 3, 94–100.
- Sun, Y., Liu, J. and Kennedy, J. F. (2010). Application of response surface methodology for optimization of polysaccharides production parameters from the roots of *Codonopsis pilosula* by a central composite design. *Carbohydrate Polymers*. 80, 949-953.
- Susilawati, Matsjeh, S., Pranowo, H. D., and Anwar, C. (2012). Macrone, A Novel Diepoxy lignan from Bark of Mahkota Dewa (*Phaleria macrocarpa* (Scheff.) Boerl.) and Its Antioxidant Activity, *Indo. Journal of Chemistry*. 12, 62–69.
- Tahir, M. M., Ibrahim, N. and Yaacob, W. A. (2014). Cytotoxicity and Antiviral Activities of *Asplenium nidus*, *Phaleria macrocarpa* and *Eleusine indica*. *Proceeding of AIP Conference*. Bangi, Selangor. 1614, 549-552.
- Thaipong, K., Boonprakob, U., Crosby, K., Cisneros-Zevallos, L., and Byrne, D. H. (2006). Comparison of ABTS, DPPH, FRAP, and ORAC Assays for Estimating Antioxidant Activity from Guava Fruit Extracts. *Journal of Food Composition and Analysis*. 19, 669–675.
- Tjandrawinata, R. R., Nofiarny, D., Susanto, L. W., Hendri P., and Clarissa, A. (2011). Symptomatic Treatment of Premenstrual Syndrome and/or Primary Dysmenorrhea with DLBS1442, A Bioactive Extract of *Phaleria macrocarpa*. *International Journal of General Medicine*. 4, 465–76.
- Trease, G. E. and Evans, W. C. (1978). *A Textbook of Pharmacognocny*. 11<sup>th</sup> Edition, Bailliere Tindall, London, 530.
- Tri, W. A., Eko, S., Ismail, M. A., and Shafiur, M. R. (2012). Effect of Aloe Vera (*Alloe Vera*) and Crown of God Fruit (*Phaleria macrocarpa*) On Sensory,

- Chemical, and Microbiological Attributes of Indian Mackerel (*Restrelliger Neglectus*) During Ice Sto. *Int Food Res J.* 1, 119–125.
- Triastuti, A., Paltiel, H. J., and Choi, J. W. (2009). *Phaleria macrocarpa* Suppress Nephropathy by Increasing Renal Antioxidant Enzyme Activity in Alloxan-Induced Diabetic Rats. *Nat Prod Sci.* 15, 167–172.
- Tuba, A., and Gülçin, I. (2008). Antioxidant and Radical Scavenging Properties of Curcumin. *Chemico-Biological Interactions.* 174, 27–37.
- Vilkhu, K., Mawson, R., Simons, L. and Bates D. (2008). Applications and Opportunities for Ultrasound Assisted Extraction in the Food Industry — A Review. *Innovative Food Science and Emerging Technologies.* 9, 161–169.
- Wang, J., Sun, B., Cao, Y., Tian, Y. and Li, X. (2008). Optimisation of ultrasound-assisted extraction of phenolic compounds from wheat bran. *Food Chemistry.* 106, 804–810.
- Wang, Z., Pan, Z., Ma, H. and Atungulu, G. G. (2011). Extract of Phenolics from Pomegranate Peels The Open Food Science Journal. 5, 17-25.
- Wu, J., Lidong, L. and Chau F. (2001). Ultrasound-assisted extraction of ginseng saponin roots and cultured ginseng cells. *Ultrasonic Sonochemistry.* 8, 347-352.
- Wu, J., Gamage, T. V., Vilkhu, K. S., Simons, L. K. and Mawson R. (2008). Effect of Thermosonication on Quality Improvement of Tomato Juice. *Innovative Food Science and Emerging Technologies.* 9, 186–195.
- Yosie, A., Effendy, M. A. W., Sifzizul, T. M. T., and Habsah M. (2011). Antibacterial, Radical-Scavenging Activities and Cytotoxicity Properties of *Phaleria macrocarpa* (Scheff.) Boerl leaves in HEPG2 cell lines. *Int J Pharm Sci Res.* 2, 1700–1706.
- Zhang, Y. B., Xu, X. J., and Liu, H. M. (2006). Chemical Constituents from Mahkota Dewa. *Journal of Asian Natural Products Research.* 8, 119–123.
- Zohra, M., and Atik, F. (2011). Impact of Solvent Extraction Type on Total Polyphenols Content and Biological Activity from *Tamarix aphylla* (L.) Karst. *International Journal of Pharma and Bio Science.* 2, 609-615.
- Zou, T. B., Xia, E. Q., He, T. P., Huang, M. Y., Jia, Q. and Li, H. W. (2014). Ultrasound-Assisted Extraction of Mangiferin from Mango (*Mangifera indica* L.) Leaves Using Response Surface Methodology. *Molecules.* 19, 1411-1421.