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## *Acrocomia crisper* fruits lipid extract prevents LPS-induced acute lung injury in mice

[Extracto lipídico de los frutos de *Acrocomia crisper* previene el daño pulmonar agudo inducido por LPS en ratones]

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**Abstract:** The aim of this study was to evaluate the effects of single oral doses of D-005 (a lipid extract obtained from the fruit oil of *Acrocomia crisper*) on LPS-induced acute lung injury (ALI) in mice. D-005 batch composition was: lauric (35.8%), oleic (28.4%), myristic (14.2%), palmitic (8.9%), stearic (3.3%), capric (1.9%), caprylic (1.2%), and palmitoleic (0.05%) acids, for a total content of fatty acids of 93.7%. D-005 (200 mg/kg) significantly reduced lung edema (LE) ( $\approx$  28% inhibition) and Lung Weight/Body Weight ratio (LW/BW) (75.8% inhibition). D-005 (25, 50, 100 and 200 mg/kg) produced a significant reduction of Histological score (59.9, 56.1, 53.5 and 73.3% inhibition, respectively). Dexamethasone, as the reference drug, was effective in this experimental model. In conclusion, pretreatment with single oral doses of D-005 significantly prevented the LPS-induced ALI in mice.

**Keywords:** *Acrocomia crisper*; D-005; Fatty acids; Acute lung injury; Lipopolysaccharide; Mice

**Resumen:** El objetivo de este estudio fue evaluar los efectos de dosis orales únicas de D-005 (extracto lipídico obtenido del aceite de frutos de *Acrocomia crisper*) sobre el daño pulmonar agudo (DPA) inducido por LPS en ratones. La composición del lote de D-005 fue: ácido láurico (35.8%), oleico (28.4%), mirístico (14.2%), palmítico (8.9%), esteárico (3.3%), cáprico (1.9%), caprílico (1.2%) y palmitoleico (0.05%), con un contenido total de ácidos grasos de 93.7%. D-005 (200 mg/kg) redujo significativamente el edema pulmonar (EP) ( $\approx$  28% de inhibición) y la relación peso pulmón/peso corporal (PP/PC) (75.8% de inhibición). D-005 (25, 50, 100 y 200 mg/kg) produjo una reducción significativa de la puntuación histológica (59.9, 56.1, 53.5 y 73.3% de inhibición, respectivamente). La dexametasona, fármaco de referencia, fue efectiva en este modelo experimental. En conclusión, el pretratamiento con dosis orales únicas de D-005 previno significativamente el DPA inducido por LPS en ratones.

**Palabras clave:** *Acrocomia crisper*; Ácidos grasos; Daño pulmonar agudo; Lipopolisacárido; Ratones.

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## INTRODUCTION

The acute respiratory distress syndrome (ARDS) was firstly described in 1967 (Ashbaugh *et al.*, 1967) in patients with acute attacks of tachypnea and hypoxia, being considered as a severe particular case of acute lung injury (ALI) (Bernard *et al.*, 1994; Ranieri *et al.*, 2012).

ARDS/ALI causes could be from a multifactorial origin and generally are associated to diffuse alveolar damage characterized by clinical symptoms of dyspnea, arterial hypoxaemia and bilateral pulmonary infiltrates in absence of clinical evidence of left atrial hypertension. Both of them are responsible for thousands of adult and pediatric deaths annually worldwide, showing a significant mortality rate (30 - 60% of all reported cases) (Singh *et al.*, 2014), while the most common initiating event of ALI/ARDS involved: gastric acid aspiration, pneumonia, pulmonary contusion, sepsis, shock, trauma, multiple transfusions, pancreatitis, etc (Matute *et al.*, 2008; Travieso & Blanco, 2009).

Inflammation is an important etiological factor in the development and progression of this pathology. An acute inflammatory response in the airways causes damage to the epithelium and pulmonary endothelium, with the subsequent release of oxidizing substances and proteases. On the other hand, arachidonic acid derivatives such as prostaglandins, thromboxanes and leukotrienes, are implicated as pro-inflammatory mediators of ALI. Significant modifications in the levels of thromboxanes in tissues, as well as circulating prostacyclines promote the formation of edema and changes in pulmonary perfusion due to the increase in the synthesis of mediators of COX-2 (Min *et al.* 2012; Howden *et al.* 2012; Bulmuş *et al.* 2013).

Up to the present there is no evidence of effective pharmacological therapies to treat ALI/ARDS, which justifies the search for new pharmacotherapeutic strategies as a current issue. Emerging therapies currently under investigation to prevent or treat ALI will provide further strategies to reduce these mortality rates, making a noteworthy emphasis on the natural products. In these sense, medicinal plants play an essential role as an important source of pharmacological active metabolites.

The palm family Arecaceae is a wide group comprising approximately 217 genera and 2,500 arboreal species to be found throughout equatorial, tropical, and subtropical areas of the world (Sosnowska and Henrik, 2009). Palm fruits are an important oleaginous resource for the development of the pharmaceutical and cosmetic industries. The species *Elaeis guineensis* Jacq, *Cocos nucifera* L.

and *Serenoa repens* L. are widely used for these purposes (Rodríguez *et al.* 2007; González. and Bravo, 2017; Ghorbanpour *et al.*, 2017). In Cuba this family is represented by a great diversity of species (Muñiz & Borhide, 1982), *Roystonea regia* (Cuban royal palm) being a specie widely studied for medicinal uses as antioxidant (López *et al.*, 2009; Rodríguez *et al.*, 2010; Oyarzábal *et al.*, 2011; González *et al.*, 2013), antiinflammatory (Menéndez *et al.*, 2006; Ravelo *et al.*, 2011, González *et al.*, 2013) and in benign prostatic hyperplasia (Pérez *et al.*, 2010; Oyarzábal *et al.*, 2013; Guzmán *et al.*, 2013; Oyarzábal *et al.*, 2015), while other species such as *Acrocomia crisper* (Cuban belly palm), another Cuban endemic palm, with medicinal potentialities has not been sufficiently studied nor employed industrially before. Up to the present there is no documentation of its use in folk medicine.

D-005, a lipid extract obtained from the fruit oil of *A. crisper*, contains a reproducible mixture of free fatty acids, mainly lauric, oleic, myristic, and palmitic acids; while stearic, capric, caprylic and palmitoleic acids are present in lower concentrations. (Sierra *et al.*, 2015) Previous studies showed that some of these fatty acids, such as myristic, stearic, palmitic, oleic, and several oily extracts containing these fatty acids inhibited cyclooxygenase (COX) and lipooxygenase (LOX) activities *in vitro* (Chan *et al.*, 1996; Zhang *et al.*, 2002; Henry *et al.*, 2002; Menéndez *et al.*, 2007).

In consonance with this data, a more recent study demonstrated that D-005 inhibited COX-2 and 5-LOX enzyme activities, with highest affinity for COX-2. The dual inhibition of COX-2 and 5-LOX suggested that D-005 could produce anti-inflammatory effects (Pérez *et al.*, 2017).

Taking into account the primary role played by the COX-2 and 5-LOX enzymes in the pulmonary inflammatory process involved in the development and progression of ALI (Min *et al.* 2012; Howden *et al.* 2012; Bulmuş *et al.* 2013), it is logical to assume that D-005 could provide a protective effect on ALI due to its anti-inflammatory profile dual (Pérez *et al.*, 2017). In addition, ALI is one of the most serious health problems due to its high mortality rate (Singh *et al.*, 2014), and until now is an orphan disease of effective pharmacological therapies, that is, without a cure. So, it becomes justifies characterize the lipid extract of fruits of *A. crisper* and evaluate its relevance in the context of acute lung injury.

Several experimental models and different species of animals in the preclinical evaluation of some drugs have been used extensively to reproduce the essential manifestation and the most

important events of ALI. However, ALI induced by lipopolysaccharide (LPS) in mice is a classical model being widely developed by the researchers worldwide (Matute *et al.*, 2008; Mena *et al.*, 2016).

In light of these issues, this study was carried out to evaluate the effects of D-005 on LPS-induced ALI in mice.

## MATERIALS AND METHODS

### *Extract preparation*

Ripe fruits of *A. crisper* were collected and authenticated (No. 1982-1031) in the National Botanic Garden (Havana, Cuba). Fruits were dried under shade in a well-ventilated area, powdered with a hammer mill at 2.36 mm, and extracted with n-hexane at room temperature (three extractions of 24 h). Solvent was removed at 60°C under vacuum, in a Büchi rotary evaporator, and the obtained oil was saponified at reflux with a hydro alcoholic potassium hydroxide solution (0.5 mol/L). Afterwards, the fatty acids were released with HCl (10%) until pH 1-2, and the organic phase was washed with water and dried under vacuum at 110°C. The product obtained was the D-005 batch used in this study.

### *Sample preparation for chromatographic analysis*

D-005 batch composition was assessed by a validated specific gas chromatographic method. Fatty acids were prepared for analysis as fatty acid methyl esters using a validated one-step extraction-transesterification method (Rodríguez *et al.*, 2009). Approximately 100 mg of D-005 were placed into a 15 mL tube with Teflon-lined screw cap. One millilitre of methanol containing 1 mg of the internal standard tridecanoic acid (Sigma-Aldrich, St. Louis, MO, USA), 3 mL of freshly prepared acetyl chloride (10%) in methanol and 5 mL hexane were added. The tube was capped, shaken, and incubated at 85 °C for 30 min in a dried thermostat, with occasional shaking. After allowing the tube to cool to room temperature, 3 mL of sodium hydroxide (2 mol/L) were added, the tube was shaken in a horizontal shaker for 15 min, and then it was let to stand around two minutes until phase separation. One mL of the supernatant was carefully transferred to a vial, 1 mL hexane was added, the vial was capped, shaken for homogenizing, and 1 µL was taken for chromatographic analysis.

### *Fatty Acid Methyl Esters Gas Chromatography-Flame Ionization Detector Analysis*

Prepared samples of the batch (n=3) were analysed for fatty acid methyl esters using an Agilent equipment model 7890A (Agilent Technologies, SA) with a BPX-5wide-bore capillary column (30 m × 0.53 mm × *df* 1.5 µm, SGE, Australia). Injection port and detector temperatures were maintained at 320 °C. Hydrogen was used as the carrier gas at a flow rate of 8 mL/min. A temperature gradient was used with an initial oven temperature of 120 °C (2 min) that increased to 320 °C at 10 °C/min. Fatty acid methyl esters were identified by their retention times using the FAME standard mix (Supelco, USA) and quantified using Agilent Chem Station software.

### *Animals and Study protocol*

BALB/c male mice (18-20 g), from the Center for Laboratory Animals Production (CENPALAB, Habana, Cuba) were adapted for 7 days to controlled laboratory conditions: temperature (25 ± 2 °C, 60 ± 5 % relative humidity and 12 hours' light/dark cycles). Food (rodent pellets supplied by CENPALAB) and water were provided *ad libitum*. Animal handle was conducted according to the Cuban Code for the Use of Laboratory Animals and ethical principles for animal management No.39/04. The independent ethical board of the Center of Natural Products approved the use of animals and the study protocol.

### *Substances Preparation, administration and dosage*

D-005, immediately before use was suspended in a vehicle (Tween 65/water, 2%) and Dexamethasone injectable (Dex) (Carlos J. Finlay laboratory, Havana, Cuba) was used as reference drug. LPS, inductor agent of ALI, (*Escherichia coli* serotype 055: B5; Sigma Chemical Co., St. Louis, MO), was dissolved in 20 ml of saline solution (0.9%).

After quarantine period, mice were randomly divided in eight groups (n=10). Experimental groups were as follows: a negative control group (Tween 65/water oral vehicle administration and 0.2 ml i.p saline solution) without damage induction and seven groups injected with LPS (20 mg/kg, i.p): Positive control (Tween 65/water vehicle), D-005 (5, 25, 50, 100, 200 mg/kg) and Dex (20 mg/kg) group. Oral administration by gastric gavage of D-005 and Dex were started 60 min before the intra-peritoneal injection of saline or LPS (20 mg/kg).

### Experimental Procedure

One hour before D-005 or Dex administration, except the negative control group, animals were injected with LPS (20mg/kg, ip). Twenty-four hours later, anesthesia state was induced by thiopental sodium injection (40 mg/kg, ip) and animals were sacrificed by exsanguinations. Afterwards, lungs and tissue samples were collected to quantify the lung edema (LE) and lung weight/body weight ratio (LW/BW ratio) and to determine the histopathological changes, respectively.

### Determination of LW/BW ratio and lung edema

Mice Body weight (BW) was determined in an analytical balance before the sacrifice, while, lung weight (LW) was immediately determined after to extraction. The LW/BW ratio was calculated taking into account the values of LW and BW.

LE was determined according to the method described by He et al (He *et al.*, 2014). We evaluated LPS-induced edema by measuring the content of lung water as follow: the left lung was weighed out in analytical balance to obtain the wet weight (WW) and then it was exposed to a temperature of 85°C for 48 h to determine dry weight (DW).

The LE was expressed by the mathematical expression:  $LE = (WW - DW)/WW \times 100\%$ .

### Histological analysis

The lungs were extracted and fixed in buffered formaldehyde solution (10%). After fixation, the lung tissue was sectioned sagittally and embedded in paraffin. The sections were stained with hematoxylin – eosin (HE). Histological analysis of lung damage was evaluated according to the following variables: neutrophil infiltration, haemorrhages, oedema, exudation, necrosis, congestion, atelectasis and alveolar thickening. The severity of lung injury was evaluated establishing a histological score according to the following criteria: No damage, 0; damage in 25% of the fields, 1; damage in 50% of the fields, 2; damage in 75% of the fields, 3; diffuse damage, 4 (He *et al.*, 2014).

### Statistical Analysis

Comparisons among groups were done by using the Kruskal-Wallis test and paired comparisons between treated and control groups were done by using Mann-Whitney U test. The level of statistical significance was set at  $\alpha=0.05$ . Data were processed with the Statistics Software for Windows (Release 6.1 Stat Soft Inc, Tulsa OK, USA).

## RESULTS

### Chemical analysis of D-005 batch

D-005 fatty acid composition, analyzed by Gas Chromatography (Figure No. 1), was as follows (w/w, %): lauric (35.8%), oleic (28.4%), myristic (14.2%), palmitic (8.9%), stearic (3.3%), capric (1.9%), caprylic (1.2%), and palmitoleic (0.05%), for a total content of fatty acids of 93.7%. Other fatty acids were analyzed and quantified, but only the eight major fatty acids, are reported individually. The mean percent coefficient of variance between duplicates for total fatty acids was 0.8%.

### Biological effects of D-005 on LPS-injection induced ALI.

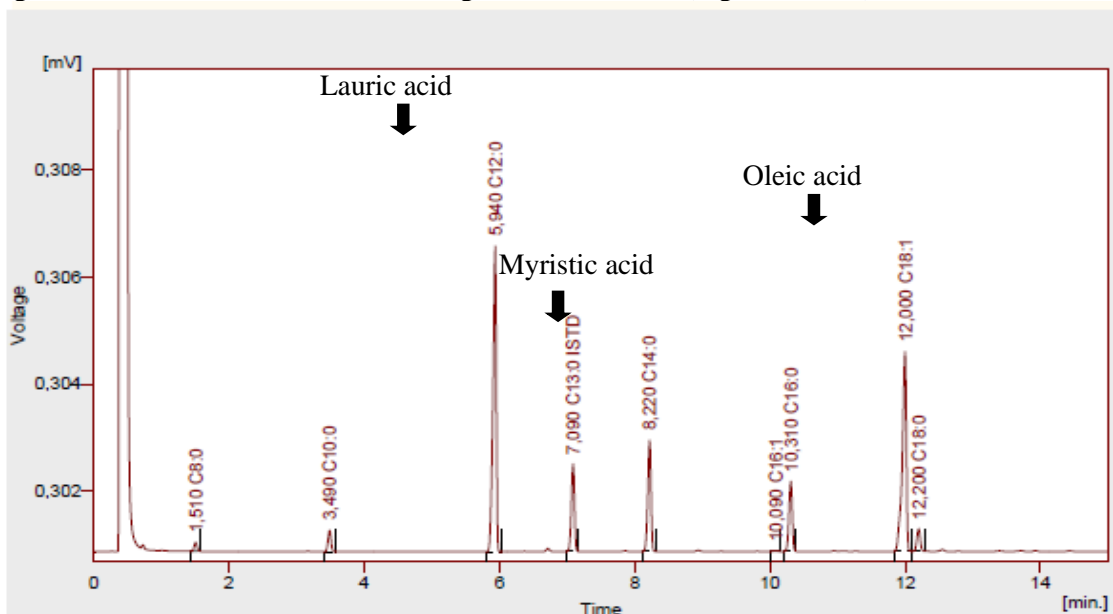
Table No. 1 shows the effects of single oral administration of D-005 on LE and LW/BW ratio in mice with ALI induced by LPS. The i.p. injection with LPS produced a significant ( $p<0.01$ ) increase of LE and LW/BW ratio in animals of positive control group compared to animals of negative control group.

D-005 (5-200 mg/kg) reduced moderately ( $\approx 28\%$  inhibition) the levels of LE, but only the doses of 200mg/kg, produced a significant reduction ( $p<0.05$ ). Furthermore, D-005 decreased the LW/BW ratio, although only the effect of the highest dose (200 mg/kg) was marked (75.86% inhibition) and significant ( $p<0.05$ ). Dex (20 mg/kg), reference drug, reduced marked (65.60% inhibition) and significantly ( $p<0.05$ ) not only the LE but also the LW/BW ratio (58.62% inhibition).

ALI induction by LPS caused a significant increase of the HS in the positive control group ( $p<0.001$ ) compared to the negative control group. The Table No. 2 shows the effects of D-005 oral administration on histological score (HS) in mice with ALI induced by LPS. The lowest assayed dose (5 mg/kg) moderately, but not significantly, reduced the HS while doses of D-005 (25, 50, 100 and 200 mg/kg) produced a significant ( $p<0.05$ ) and marked inhibition (59.9, 56.1, 53.5 and 73.2%, respectively) of HS. Dex (20 mg/kg), reference drug, significantly ( $p<0.05$ ) and markedly inhibited HS by 59.9%.

Normal structure of lung tissues was observed in all negative control mice. The Figure No. 2A shows the lung tissue from an animal of the negative control group with a normal distribution of alveolar spaces, also a normal appearance of the squamous epithelium and capillaries without signs of inflammation. By the opposite, lung tissue of all positive controls displayed marked signs of ALI such as severe neutrophil infiltration and alveolar

thickening almost in 50-75% of the histological fields (Figure No. 2B).



**Figure No. 1**  
**Gas Chromatogram of D-005 showing majorly compounds from the total fatty acid composition by black arrows**

**Table No. 1**  
**Effects of D-005 on LE and LW/BW ratio on LPS induced ALI in mice**

Groups	LE [%]	I [%]	LW/BW	I [%]
Negative control (Vehicle)	78,22 ± 1,08 **	-	0,60±0.08**	-
Positive control (Vehicle + LPS)	83,86±1,08	-	0,89±0,05	-
D-005 5 mg/kg + LPS	81,81±0,92	36,3	0,78±0,42	37,9
D-005 25 mg/kg + LPS	82,24±0,77	28,7	0,78±0,43	37,9
D-005 50 mg/kg + LPS	82,63±0,91	21,8	0,76±0,03	44,8
D-005 100 mg/kg + LPS	82,56±0,92	23,0	0,76±0,04	44,8
D-005 200 mg/kg + LPS	82,25±0,46*	28,5	0,67±0,01*	75,8
Dex 20 mg/kg + LPS	80,16±0,64*	65,6	0,72±0,04*	58,6

I: inhibition, LPS: lipopolysaccharide, LE: lung edema,

LW: lung weight, BW: body weight, Dex: dexamethasone.

\* p<0.01; \*\* p <0.001. Comparisons with the positive LPS control (Mann Whitney U test).

**Table No. 2**  
**Effects of D-005 on histological score on LPS induced ALI in mice.**

GROUPS	HS	I [%]
Negative control (Vehicle)	0,75±0,16***	-
Positive control (Vehicle + LPS)	2,62±0,74	-
D-005 5 mg/kg + LPS	1,87±0,29	40,1
D-005 25 mg/kg + LPS	1,50±0,18*	59,9
D-005 50 mg/kg + LPS	1,57±0,20*	56,1
D-005 100 mg/kg + LPS	1,62±0,18*	53,5
D-005 200 mg/kg + LPS	1,25±0,25**	73,2
Dex 20 mg/kg + LPS	1,50±0,32*	59,9

I: inhibition, HS: histological score, Dex: dexamethasone.  
 \* p<0.01; \*\* p<0.001, \*\*\*p<0.001. Comparisons with the positive LPS control (Mann Whitney U test).

Pretreatments with oral doses of D-005 (25-200 mg/kg) ameliorated the pulmonary lesions, while the lowest dose assayed (5 mg/kg) achieved a slight but not significant inhibition of damage. The Figure No. 2C shows the principal findings observed in lung tissue of animals pretreated with this dose, being neutrophil infiltration and alveolar thickening, evident signs of ALI, while the doses of 25 (Figure No. 2D) and 50 mg/kg (Figure No. 2E) only showed slight alterations characterized by a poorly neutrophil infiltration.

In contrast, minor damages were found in lung tissues of animals pretreated with D-005 (100 and 200 mg/kg). The Figure No. 2F and Figure No. 2G illustrate a prominent attenuation of LPS-induced pulmonary histopathologic findings observed in these D-005 treated groups.

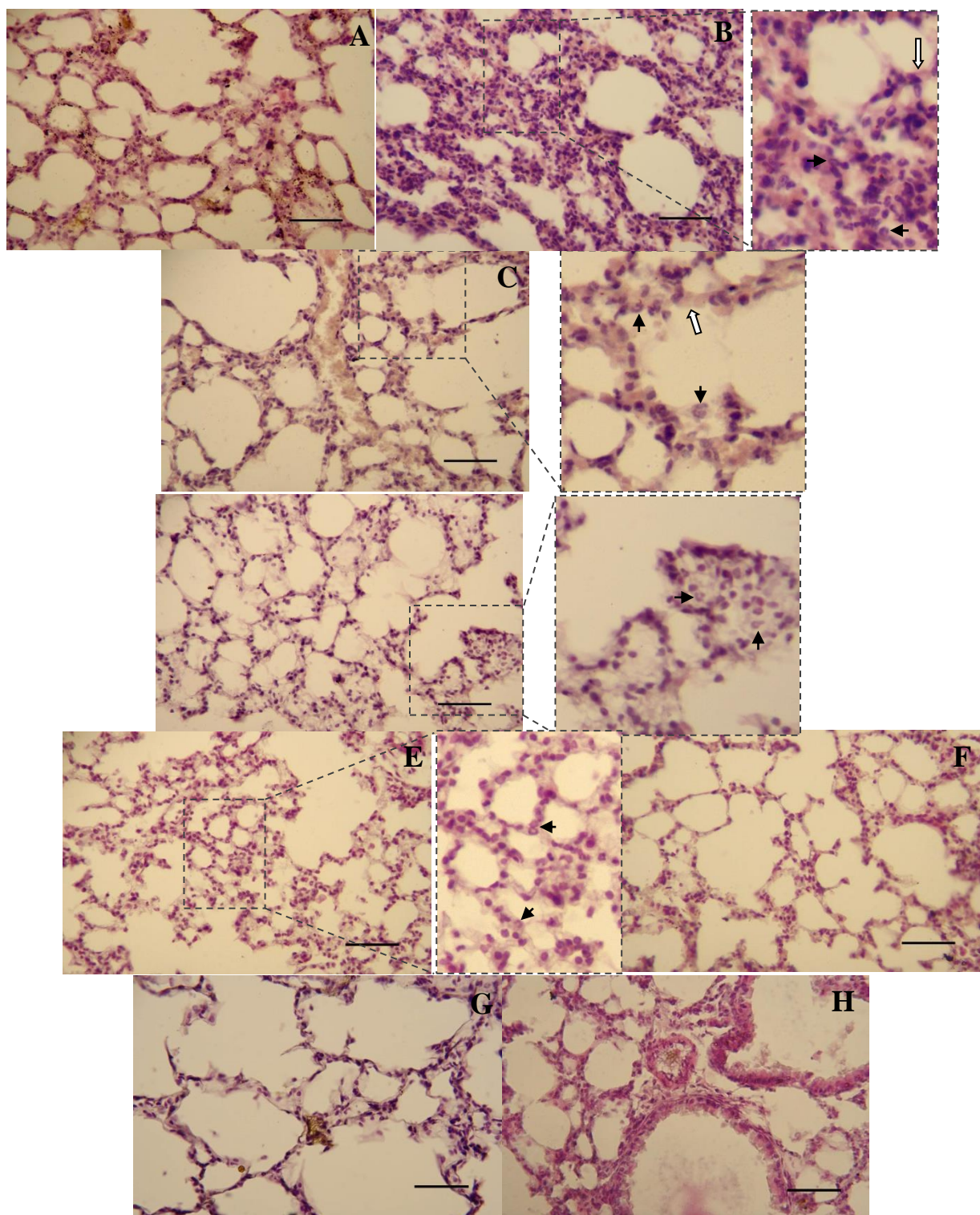
Dex was effective as reference drug achieving a marked and significant inhibition of ALI (Figure No. 2H).

## DISCUSSION

Chromatographic analysis of this D-005 batch confirmed the presence of fatty acids between 8 and 18 carbon atoms, which were identified by comparison of their relative retentions with commercial standards. The free fatty acids composition shows the lauric acid as the major component (35.8%), followed by oleic acid (28.4%), and myristic acid (14.2%). This result is similar to that obtained in a previous study (Sierra

et al., 2015). Composition of D-005 differs from those obtained for the oily extracts of the hole fruits of *S. repens* (C18:1 = 43.29%, C12:0 = 26.60%, C14:0 = 11.49%) (Rodríguez et al., 2012) and *R. regia* (C18:1 = 48.50%, C12:0 = 23.30%, C14:0 = 11.00%) (Rodríguez et al., 2007), which present higher proportions of oleic acid and lower proportions of lauric acid, although the saturated fatty acids predominate in the three extracts. In addition, it is important to highlight that total free fatty acids content in D-005 (> 90 %) is higher than those present in the oily extract of other Arecaceae as *Serenoa repens* (40.7- 80.7 %) (Habib, 2009).

Arecaceae family is broadly distributed in Cuba and interestingly around 90% of palms species are endemic (Muñiz & Borhede, 1982). *R. regia* and *A. crisper* fruit extracts have a similar fatty acid composition (Rodríguez et al., 2007) and both species have been described as anti-inflammatory (Ravelo et al. 2011, González et al., 2013). The compounds mainly differ between species and are found in a variable percentage in oil extracts (Sierra et al., 2014). In accordance, it has been described that some oily extracts containing myristic, steraic, palmitic and oleic acid have an in vitro inhibitory effect on cyclooxygenase (COX) and lipooxygenase (LOX) activities, blockading an important pathway of pro-inflammatory precursors release. (Chan et al., 1996; Henry et al., 2002; Zhang et al., 2002; Menéndez et al., 2007; Pérez et al., 2017)



**Figure No. 2**

Lung sections, H&E, 40 X. A) Negative control. Normal appearance of alveolar walls without signs of inflammation; B) Positive control showing severe neutrophil infiltration and alveolar thickening; C) D-005 (5 mg/kg) showing neutrophil infiltration and alveolar thickening; D) D-005 (25 mg/kg) and E) D-005 (50 mg/kg), showing poorly neutrophil infiltration. F) D-005 (100 mg/kg), G) D-005 (200 mg/kg), H) Dexamethasone 200 mg/kg. In F, G and H a significant attenuation of LPS-induced pulmonary histopathologic changes are observed. Arrowheads: neutrophils, arrows: alveolar wall. The rectangles with dashed lines in B, C, D and E enclose regions that have been magnified to the right of each figure. Bars: 50  $\mu$ m.

Intraperitoneal injection with LPS (20 mg/kg) induced ALI in mice. ALI was evidenced by a significant increase of LE, LW/BW ratio and the histological changes observed in positive control animals compared to negative control animals. In such regard, it is well known that LPS mechanism to induce ALI involves the LPS binding to specific proteins (P) in pulmonary endothelial cells to form a complex (LPS:P) which activates CD14/TLR4 receptors on monocytes and macrophages promoting the inflammatory mediators release (Yang *et al.*, 1998; Matute *et al.*, 2008) mediated by the Nuclear factor- $\kappa$ B (NF- $\kappa$ B) pathway activation. NF- $\kappa$ B, a nuclear transcriptional factor, mediates the ALI progression as regulator of different genes encoding cytokines, chemokines, adhesion molecules and growth factors (Blackwell & Christman, 1997; Lawrence *et al.*, 2001; Kalemci *et al.*, 2013).

Moreover, it has been shown that COX-2 overexpression could be induced by *in vitro* inoculation of LPS on alveolar macrophages (Hempel *et al.*, 1994; Uhlig *et al.*, 1996), in pulmonary epithelial cells (Mitchell *et al.*, 1994) and rat lung tissue (Chida *et al.*, 1994) promoting the subsequently acute inflammatory response.

In the context of these findings, substances acting on the NF- $\kappa$ B pathway and on the COX-2 overexpression could provide an important pharmacological benefit, by mitigating the inflammatory process involved in ALI.

As can be observed a significant increase of histological markers of lung damage was evidenced in our experimental conditions, being an expected result according to the reports of other authors (Everthart *et al.*, 2006; Kalemci *et al.*, 2013; He *et al.*, 2014). Additionally, Dex (20 mg/kg), as reference substance, was effective on the analysed variables, which confirms the experimental model validity.

Oral administration of D-005 showed beneficial effects in mice with LPS-induced ALI. D-005 significantly attenuated the presence of neutrophil infiltration and also the alveolar thickening, which are considered very relevant histological evidences of lung tissue injury in ALI (Matute-Bello *et al.*, 2011). The minimum effective dose was 25 mg/kg since the lower dose tested (5 mg/kg) produced a slight reduction in HS but it was not significant.

On the other side, a maximum effective dose could not be characterized by considering that 200 mg/kg achieved the highest inhibition on HS, but higher doses were not evaluated in this experimental protocol. Therefore, further studies

should elucidate the efficacy of high doses in this ALI model.

Inflammation has been described as an important etiologic factor in ALI initiation (Lee *et al.*, 2001b; Matute *et al.*, 2008). It is mainly associated to a substantial increase of the mediators of arachidonic acid synthesis as thromboxane and prostaglandins that promote the formation of edema and changes in pulmonary perfusion. Indeed, the increased synthesis of mediators from COX-2 causes activation and mast cell degranulation releasing proteases and chemokines responsible for the increased vascular permeability (PGE<sub>2</sub>) and produces vasodilatation (PGL<sub>2</sub>), promoting the edema formation. Meanwhile, 5-LOX mediators promotes chemotaxis and the recruitment of macrophages and mast cells to alveolar space (LTB<sub>4</sub>) promoting the inflammatory infiltration (Guo *et al.*, 2012).

ALI/ARDS are characterized by lung edema due to increased permeability of the alveolar-capillary barrier and subsequent impairment of arterial oxygenation. Lung edema, endothelial and epithelial injury are accompanied by an influx of neutrophils into the interstitium and bronchoalveolar space. Hence, activation and recruitment of neutrophils are regarded to play a key role in progression of ALI/ARDS (Grommes & Soehnlein, 2011). For these reasons, the inhibition of lung edema and of inflammatory infiltration to the lungs constitutes a possible target for those drugs with potential benefit for the treatment of ALI.

On this regards, previously *in vitro* studies showed the non-competitive inhibitory effect of D-005 on COX-2 and 5-LOX activities (Pérez *et al.*, 2017). Thus, the efficacy of D-005 to reduce LE, LW/BW ratio and attenuate the signs of LPS-induced ALI in mice (neutrophil infiltration and the alveolar thickening) could be associated, at least partially to both enzymes inhibition.

On the other hand, it has been demonstrated that esterified fatty acids as Methyl palmitate (MP), methylester of palmitic acid, attenuate the pulmonary histopathological changes in a murine model of LPS-mediated ALI and such effects involved the inhibition of NF- $\kappa$ B activation (Kalemci *et al.*, 2013). Controversially it was demonstrated by Lee *et al.* that saturated fatty acids could induce the activation of NF- $\kappa$ B and COX-2 expression in a murine monocytic cell line (RAW 264.7). However, a premise discussed in these work was the fact that unsaturated fatty acids could not only inhibit the saturated fatty acid effect on LPS-induced NF- $\kappa$ B activation but also the expression of



COX-2 and other inflammatory markers (Lee *et al.*, 2001a).

Presumably, these findings could clarify why D-005 inhibits the activities of COX-2 and 5-LOX and reduces lung damage in the ALI induced by LPS, although the pharmacological mechanism to elucidate its effectiveness has not been fully elucidated.

This study constitutes the first report of the beneficial effects of D-005 on ALI and establishes a fundamental stage to develop new studies and provide new evidences about its effects on this pathological entity.

## CONCLUSIONS

Pretreatment with single oral doses of D-005 markedly and significantly prevented the LPS-induced ALI in mice by reducing not only the histological score but also LE and LW/BW ratio.

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