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RESEARCH ARTICLE

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Associations of AMP and adenosine induced dyspnea sensation to large and small airways dysfunction in asthma

Claire A. Cox^{1,2*}, Ilse M. Boudewijn^{1,2}, Sebastiaan J. Vroegop³, Siebrig Schokker³, Anne J. Lexmond⁴, Henderik W. Frijlink⁴, Paul Hagedoorn⁴, Judith M. Vonk^{2,5}, Martijn P. Farenhorst¹, Nick H. T. ten Hacken^{1,2}, Huib A. M. Kerstjens^{1,2} and Maarten van den Berge^{1,2}

Abstract

Background: Bronchial provocation is often used to confirm asthma. Dyspnea sensation, however, associates poorly with the evoked drop in FEV₁. Provocation tests only use the large airways parameter FEV₁, although dyspnea is associated with both large- and small airways dysfunction. Aim of this study was to explore if adenosine 5'- monophosphate (AMP) and adenosine evoke an equal dyspnea sensation and if dyspnea associates better with large or small airways dysfunction.

Methods: We targeted large airways with AMP and small airways with dry powder adenosine in 59 asthmatic (ex)smokers with \geq 5 packyears, 14 ± 7 days apart. All subjects performed spirometry, impulse oscillometry (IOS), and Borg dyspnea score. In 36 subjects multiple breath nitrogen washout (MBNW) was additionally performed. We analyzed the association of the change (Δ) in Borg score with the change in large and small airways parameters, using univariate and multivariate linear regression analyses. MBNW was analyzed separately.

Results: Provocation with AMP and adenosine evoked similar levels of dyspnea. Δ FEV₁ was not significantly associated with Δ Borg after either AMP or adenosine provocation, in both univariate and multivariate analyses. In multivariate linear regression, a decrease in FEF₂₅₋₇₅ during adenosine provocation was independently associated with an increase in Borg. In the multivariate analyses for AMP provocation, no significant associations were found between Δ Borg and any large or small airways parameters.

Conclusion: AMP and adenosine induce equally severe dyspnea sensations. Our results suggest that dyspnea induced with dry powder adenosine is related to small airways involvement, while neither large nor small airways dysfunction was associated with AMP-induced dyspnea.

Trail registration: NCT01741285 at www.clinicaltrials.gov, first registered Dec 4th, 2012.

Keywords: Borg score, Dry powder adenosine, AMP, Provocation, Dyspnea

²Groningen Research Institute for Asthma and COPD, University of Groningen, University Medical Center Groningen, PO box 30.0001, 9700, RB, Groningen, The Netherlands

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^{*} Correspondence: c.a.cox@umcg.nl

¹Department of Pulmonary Diseases, University of Groningen, University Medical Center Groningen, PO box 30.0001, 9700, RB, Groningen, The Netherlands

Background

Airway hyperresponsiveness (AHR) is a distinct asthma characteristic. Bronchial provocation tests can be used to assess AHR, can help to diagnose asthma and monitor asthma control [1]. However, the patient's dyspnea perception associates poorly with the provocation test [2]. In clinical practice, patients often experience dyspnea before the provocative agent causes the forced expiratory volume in the first second (FEV₁) to drop 20% [3]. On the other hand, others experience no dyspnea even when the FEV₁ has dropped more than 20% [4]. A provocation test is based on the FEV₁, which is believed to be a marker for the larger airway [5]. However, dyspnea sensation is associated with both large- and small airways dysfunction [6-8]. To evaluate the small airways, for example, the forced expiratory flow between 25 and 75% of the expiration (FEF₂₅₋₇₅) or the difference in resistance between 5 Hz and 20 Hz (R_5-R_{20}) measured with impulse oscillometry (IOS) can be used [5]. Provocation tests with subsequent IOS measurements have suggested that dyspnea induced with a provocative agent corresponds better to small- than to large airways dysfunction [3, 9, 10].

Provocation tests can be performed with either direct or indirect acting agents. Direct stimuli, such as histamine and methacholine, stimulate the airway smooth muscle, resulting in airway contraction [11]. Indirect stimuli, on the other hand, induce the release of mediators from inflammatory cells, such as histamine, leukotrienes, and prostaglandins causing airway contraction [12]. Examples of indirect stimuli are mannitol, nebulized adenosine 5'monophosphate (AMP), and dry powder adenosine. The well-established AMP is dose restricted (as AMP becomes insoluble above 320–400 mg/mL) [13], whereas mannitol and the newly available dry powder adenosine are not [14]. AMP and dry powder adenosine are well tolerated by patients [15], but mannitol evokes discomforting cough [16, 17]. AMP and dry powder adenosine appear to act via the same indirect pathways, but can consist of differently sized particles. Nebulized AMP commonly has a mass median aerodynamic diameter (MMAD) between 5.1- $8.5 \,\mu m$ [18], depending on the nebulizer settings and AMP concentration [18, 19]. Dry powder adenosine, on the other hand, can be produced with an MMAD as small as $2.6-2.9 \,\mu\text{m}$ [20], with a much smaller distribution in particle size which is independent of the dose [20]. Therefore, dry powder adenosine was postulated to reach the small peripheral airways to a larger extent compared to nebulized AMP, especially when inhaled at a low flow [21]. Thus, to target the small airways specifically, without a dose restriction and cough, adenosine may be valuable.

In this study we evaluated whether there is a difference between the perception of dyspnea induced with the assumed small airways trigger dry powder adenosine or the

Table 1 Baseline characteristics

Gender (M/F)	24/35
Age (years)	47.0 (37.0–55.0)
BMI (kg/m²)	26.8 (23.1–31.4)
Smoking status (Current/Ex)	30/29
Number of packyears (years)	16.8 (11.0–26.0)
Adenosine provocation (pos/neg)	45/14
Positive Adenosine (mg)	3.11 (0.87–6.38)
AMP provocation (pos/neg)	40/19
Positive AMP (mg/mL)	14.67 (4.7–44.88)
Borg score (points)	0.0 (0.0-2.0)
FEV ₁ (L)	2.93 (2.36–3.44)
FEV ₁ percentage of predicted (%)	85 (74–96)
FVC (L)	4.14 (3.52–4.94)
FVC percentage of predicted (%)	105 (94–116)
FEV ₁ /FVC (%)	70 (62–77)
FEF ₂₅	4.86 (3.46–6.42)
FEF ₂₅ percentage of predicted (%)	72 (48–96)
FEF ₅₀	2.35 (1.70–3.27)
FEF_{50} percentage of predicted (%)	51 (36–65)
FEF ₇₅	0.67 (0.46–1.14)
FEF ₇₅ percentage of predicted (%)	36 (25–56)
FEF ₂₅₋₇₅	1.79 (1.30–2.74)
FEF_{25-75} percentage of predicted (%)	49 (35–65)
R_5 (kPa sL ⁻¹)	0.53 (0.42–0.67)
$R_{20} (kPa sL^{-1})$	0.42 (0.35–0.47)
$R_{5}-R_{20} (kPa sL^{-1})$	0.08 (0.04–0.22)
AX ($kPa L^{-1}$)	0.64 (0.24–1.82)
X ₅ (kPa sL ⁻¹)	-0.13 (- 0.220.09)
F_{res} (s ⁻¹)	16.78 (12.33–21.83)
LCI _{2.5%} ^a	9.27 (8.60–11.28)
LCI _{5%} ^a	6.22 (5.76–7.37)
S _{cond} ^a	0.04 (0.02–0.06)
S _{acin} a	0.14 (0.10-0.19)

Data is presented as count or median (inter quartile range (IQR)). *pos* positive response, ≤ 20 mg for adenosine and ≤ 160 mg/ml for AMP, *neg* negative response, > 20 mg for adenosine and > 160 mg/ml for AMP, *FEV*₁ forced expiratory volume in the first second, *FVC* forced vital capacity, *FEF*₂₅ forced expiratory flow at 25% of FVC, *FEF*₅₀ forced expiratory flow at 50% of FVC, *FEF*₂₅₋₇₅ forced expiratory flow at 25% of FVC, *FEF*₂₅₋₇₅ forced expiratory flow at 25% of FVC, *FEF*₂₅₋₇₅ forced expiratory flow at 25 to 75% of FVC, *Rs* resistance to 5 Hz, *R*₂₀ resistance to 20 Hz, *Rs*-*R*₂₀ difference in resistance to 5 Hz and 20 Hz, *AX* reactance area, *X*₅ reactance to 5 Hz, *Fres* resonance frequency, *LCI* lung clearance index, *S*_{cons} ventilation heterogeneity of the conducting airways, *S*_{acin} ventilation heterogeneity of the acinar airways. ^a = multiple breath nitrogen washout (MBNW) was measured in 36 subjects

assumed larger airways trigger nebulized AMP. In addition, we evaluated for both triggers if the perception of dyspnea during a provocation test is more closely associated to changes in large- or small airways function.

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Methods

Study design and patients

This study was performed with baseline data from the previously published OLiVIA study (clinical trial number: NCT01741285, www.clinicaltrials.gov) [22]. Included subjects were asthmatics (doctor's diagnosis), current or ex-smokers (>5 pack years), aged between 18 and 65 vears, and all had a preserved lung function (FEV₁ > 50% predicted and > 1.2 L). Excluded were subjects with a recent (< 6 weeks) exacerbation or upper airway infection, females who were pregnant or lactating, and subjects with clinically unstable concomitant diseases. The screenings phase of the OLiVIA study incorporated two provocation tests. First an AMP provocation and 14 ± 7 days later a dry powder adenosine provocation, performed after a washout period of four to six weeks for asthma maintenance therapy and eight hours for short acting β 2-antagonists (SABAs). In the Olivia study only subjects with hyperresponsiveness to adenosine ($\geq 20\%$ drop in FEV₁ on < 20mg adenosine) were included. In the current study, all subjects who performed both provocation tests were accepted, on condition that they experienced dyspnea (increase in Borg > 1) evoked by the challenge.

Measurements

Provocation tests

Wet nebulized AMP (MMAD $5.1-8.5 \mu m$) [18] was administered in doubling concentrations ranging from 0.04 to 320 mg/mL. The AMP solutions were inhaled during two minutes of tidal breathing, without a breath-holding period, using the APS Pro System (CareFusion) with the

SideStream nebulizer (Philips Respironics) at an output rate of 0.13 mL/min. Consecutive concentrations were inhaled at five-minute intervals until the concentration caused the FEV₁ to drop \geq 20% (PC₂₀) or the highest concentration was administered.

Dry powder adenosine (MMAD 2.6–2.9 μ m) [20] was administered in doubling doses of 0.04 to 80 mg. The powder was inhaled from functional residual capacity (FRC) to total lung capacity (TLC) at a low flow rate of 20 30 L/min guided by an inspiratory flow meter, as described previously [23]. After each inhalation subjects held their breath for 10 s at TLC to allow for optimal airway deposition [24]. The procedure was repeated at three-minute intervals until the administered dose evoked a \geq 20% drop in FEV₁ (PD₂₀) or the highest dose was administered.

Pulmonary function tests

Before and after each provocation test pulmonary function tests were performed. In all subjects spirometry and IOS measurements were performed to obtain parameters for large (i.e. FEV_1 , R_{20}) and small airways (i.e. FEF_{25-75} , R_5 - R_{20}), using the classification from the review by Van der Wiel et al. [5]. Due to availability of the measurement device, multiple breath nitrogen washout (MBNW) was only measured in a subset of subjects in one of the centers. MBNW provided the index for the ventilation heterogeneity of the acinar (S_{acin}) and conductive airways (S_{cond}), and the lung clearance index (LCI).







Dyspnea score

Before and after the provocation test dyspnea was assessed with the Borg dyspnea score [25], scoring dyspnea sensation from 0 = 'no dyspnea at all' to 10 = 'maximal dyspnea'.

Statistical analysis

All analyses were performed on the change (Δ) in a parameter induced by the provocation test; calculated by subtracting the pre-provocation value from the post-provocation value. To check if adenosine and AMP induced similar responses, we compared changes in parameters between the two tests with a two sided Student's paired t-test or a two-sided Wilcoxon test, in accordance with the normality of distribution. With Spearman's correlation the change in Borg score (Δ Borg) was univariately correlated to the change in each parameter of spirometry, IOS and MBNW, for both AMP and adenosine. Subsequently, multivariate linear regression models were constructed, to investigate the origin of dyspnea. A large- and a small airways parameter from both spirometry and IOS, was selected for the model. The parameter had to have the lowest p-value in the univariate correlation analysis and were corrected for co-linearity (correlation < 0.7). Because of assumed clinical relevance, gender and smoking status were added to the model. Models were ran once without reducing or increasing the amount of parameters. As MBNW was measured in fewer subjects, a separate model was constructed expanding the models with the MBNW parameter with the lowest p-value.

Results

Study population

For this study 77 subjects were screened. However, 18 subjects were excluded as they were unable to perform



flow at 75% of FVC (Δ FEF₇₅), and **d**. the change in forced expiratory flow at 25 to 75% of the FVC (Δ FEF₂₅₋₇₅)

spirometry or a provocation test adequately (n = 5), complete the medication washout period (n = 8), had chronic non-asthmatic respiratory diseases (n = 2), or had other unstable non-respiratory diseases (n = 3) [22]. A total of 59 subjects underwent both provocation tests of which 36 performed a MBNW test. Baseline characteristics are shown in Table 1.

Comparison of adenosine and AMP provocation

Provocation with adenosine and AMP evoked a decreases in FEV₁ of $23.4 \pm 8\%$ and $21.1 \pm 8\%$, respectively. The severity of dyspnea evoked with adenosine and AMP was not significantly different, with an increase in Borg of 3.95 ± 2.1 and 3.77 ± 2.1 points, respectively (p = 0.65). Spearman's correlation between Δ Borg after adenosine and Δ Borg after AMP was moderate (rho 0.56, p < 0.001) (Fig. 1). AMP provocation evoked a greater increase in R₂₀ (p = 0.04) compared to a denosine, while a denosine evoked a greater increase in LCI_{2.5%} (p = 0.03) and S_{acin} (p = 0.01) (Fig. 2). An overview of all comparisons is shown in (see Additional file 1: Table S1).

Univariate associations with $\Delta Borg$

In the univariate analyses, $\Delta Borg$ for provocation with adenosine was significantly correlated with ΔFEF_{25} (Ls^{-1}) , ΔFEF_{75} (Ls^{-1}) , and ΔFEF_{25-75} (Ls^{-1}) and showed a trend toward an association with the ΔFEF_{50} (Ls^{-1}) (Fig. 3). The $\Delta Borg$ for provocation with AMP was significantly associated with ΔAX (kPa L^{-1}) and ΔX_5 (kPa sL⁻¹) and there was a trend towards a correlation with ΔFEV_1 (L) and ΔR_5 - R_{20} (kPa sL⁻¹) (Fig. 4). Results of all correlation analyses are shown in (see Additional file 1: Table S2).



Multivariate associations with ΔBorg Spirometry-IOS models

The multivariate model for adenosine included ΔFEV_1 (L), $\Delta \text{FEF}_{25-75}$ (Ls⁻¹), ΔR_{20} (kPa sL⁻¹) and $\Delta \text{R}_5\text{-R}_{20}$ (kPa sL⁻¹) (Table 2). This model showed an independent significant, negative association of $\Delta \text{FEF}_{25-75}$ (Ls⁻¹) with ΔBorg (R² = 20.9%). The model for AMP included ΔFEV_1 (L), ΔFEF_{50} (Ls⁻¹), ΔR_{20} (kPa sL⁻¹), and ΔX_5 (kPa sL⁻¹) (Table 2) and showed no independent associations to ΔBorg (R² = 4.3%).

Spirometry-IOS-MBNW models

In the subgroup analysis incorporating MBNW data, ΔS_{cond} was added to the adenosine model (Table 2). The result shows that $\Delta Borg$ had the best association with ΔFEF_{25-75} (kPa sL⁻¹), yet not significant (p = 0.09). The model incorporating ΔS_{cond} had an improved R² (R² = 26.5%). The AMP model with MBNW incorporated $\Delta LCI_{5\%}$ (Table 2), which shows no independent association to $\Delta Borg$ (R² = 5.4%).

Table 2 Multivariate models predicting Δ Borg in AMP and adenosine provocation

	AMP	
	Α.	В.
	B (p-value)	B (p-value)
Gender	0.25 (0.78)	0.55 (0.62)
Smoking status	-0.11 (0.90)	- 0.23 (0.81)
Δ FEV $_1$ (L)	-0.97 (0.62)	- 0.74 (0.72)
$\Delta \; \text{FEF}_{50} \; (\text{Ls}^{-1})$	0.50 (0.54)	0.60 (0.48)
Δ R ₂₀ (kPa sL ⁻¹)	-2.00 (0.76)	- 1.01 (0.88)
Δ X ₅ (kPa sL ⁻¹)	-0.64 (0.67)	- 0.69 (0.65)
Δ LCI _{5%}		0.22 (0.65)
	Adenosine	
	Α.	В.
	B (p-value)	B (p-value)
Gender	-0.83 (0.37)	-0.81 (0.37)
Smoking status	0.32 (0.65)	0.45 (0.51)
Δ FEV $_1$ (L)	1.50 (0.43)	0.98 (0.60)
$\Delta \; \text{FEF}_{\text{25-75}} \; (\text{Ls}^{-1})$	-2.18 (0.04)	- 1.82 (0.09)
$\Delta~\text{R}_{20}~(\text{kPa sL}^{-1})$	-8.28 (0.11)	-6.53 (0.20)
Δ R_5-R_{20} (kPa sL^{-1})	2.61 (0.27)	3.20 (0.18)
Δ S _{cond}		14.56 (0.16)

A. The models based on all subjects and B. the models incorporating multiple breath nitrogen washout (MBNW). Δ = change (post-pre); FEV₁ = forced expiratory volume in the first second; FEF₅₀ = forced expiratory flow at 50% of FVC; FEF₂₅₋₇₅ = forced expiratory flow at 25 to 75% of FVC; R₂₀ = resistance to 20 Hz; X₅ = reactance to 5 Hz; R₅-R₂₀ = difference in resistance to 5 Hz and 20 Hz; LCI_{5%} = lung clearance index at 5%; S_{cons} = ventilation heterogeneity of the conducting airways

Discussion

We found that dry powder adenosine and AMP evoke equal increases in dyspnea sensation, with a similar decrease in FEV₁. However, during adenosine and AMP provocation, the increase in dyspnea sensation was differentially associated with large- and small airways dysfunction. The only independently association with dyspnea induced by dry powder adenosine was the decrease in FEF_{25–75}, whereas dyspnea induced by AMP was not associated with changes in either large- or small airways dysfunction.

Our aim was to selectively target the small airways with dry powder adenosine. Therefore, we expected that dyspnea induced by dry powder adenosine would associate primarily with small airways parameters. Our findings were partly in line with this as we found the increase in Borg dyspnea score after inhalation of adenosine to associate with the decrease in FEF_{25-75} , both in the univariate and multivariate analysis. However, the adenosine-induced change in other small airways parameters, such as $R_{5}\mathchar`-R_{20}$, $S_{\rm cond}$ and $S_{\rm acin}$, did not associate with Δ Borg. This was in contrast to our expectations, as these parameters are considered to be measures of the more peripheral small airways. A possible explanation could be that the measurements provide different information, yet there is no gold standard to determine which parameter is most accurate. Another possible explanation could be that the adenosine did not reach the more peripheral small airways even though it was designed to reach the small airways, consist of relatively small particles (MMAD of 2.6-2.9 µm) [20], and was inhaled with a low flow of 30 L/min [21]. Unfortunately we lack information on the exact deposition as radiolabeling for adenosine was not performed and our conclusions are thus based on assumed differential deposition.

With respect to AMP-induced dyspnea, multivariate analysis showed no large or small airways parameters that independently associated with Δ Borg. This may suggest that other factors than airway caliber or resistance play a role in the sensation of induced dyspnea. AMP acts on adenosine receptors which are located on various inflammatory cells including mast cells, eosinophils, and neutrophils, and their activation induces a cascade resulting in airway contraction [12]. Adenosine receptors are also found on afferent nerve endings [26]. It could be speculated that activation of afferent nerve endings plays a role in the dyspnea sensation after inhalation of AMP, independent of the presence of airway contraction. This activation may be direct or indirect through bronchial interstitial edema. In the context of direct activation, the findings of Burki et al. [27] are of interest. They administered intravenous adenosine to six asthmatic and six healthy subjects. Both groups reported a significant increase in dyspnea, with a higher intensity of the dyspnea

in asthmatics. The FEV_1 , however, remained unchanged, indicating the absence of airway constriction. Based on these observations, they concluded that afferent nerve endings may be involved in the adenosine-induced sensation of dyspnea, which in asthmatics might be sensitized due to inflammation. In the context of indirect activation, interstitial edema may arise when the adenosine-induced inflammatory response induces alveolar-capillary leakage [28], which triggers the J-receptors to induce dyspnea sensation [29]. This, combined with the knowledge that afferent nerve endings are mainly seen in the upper and central airways [30], where we assume AMP primarily deposits, supports our speculation.

Although dry powder adenosine and AMP provocation may induce dyspnea through different processes, the degree of dyspnea after the final dose was not different. In addition, both tests were well tolerated and, apart from dyspnea, only led to minor cough in some subjects. This confirms previous findings in a small proof of concept study, that the relatively new adenosine provocation test is well tolerated [23].

We only included current and ex-smokers with asthma. It is therefore unclear whether these findings can be extrapolated to never-smoking asthmatics, as previous studies have shown a decreased dyspnea perception attributed to smoking, in asthmatics [31]. Never-smoking asthmatics may have had greater increases in dyspnea as a result of the provocations, but what this would have done to the association of dyspnea to large- and small airways parameters cannot be speculated.

Conclusion

Our study shows that provocation with dry powder adenosine and AMP evoke similar levels of dyspnea. Dyspnea sensation evoked with dry powder adenosine shows small airways involvement independent of large airways involvement, while AMP evoked dyspnea associated with neither large- nor small airways dysfunction. This may indicate that dry powder adenosine and AMP evoke dyspnea via different processes.

Additional file

Additional file 1: This document contains supplementary Tables S1 and S2, as referred to in the text. Table S1 is The change (post - pre) in all pulmonary function parameters evoked by the provocation. Table S2 shows Spearman's univariate correlation of the change in Borg dyspnea score with gender, smoking status, and all pulmonary function parameters. (DOCX 27 kb)

Abbreviations

AHR: Airway hyperresponsiveness; AMP: Adenosine 5' monophosphate; AX: Reactance area; FEF₂₅: Forced expiratory flow at 25% of FVC; FEF₂₅₋₇₅: Forced expiratory flow between 25 and 75% of the expiration; FEF₅₀: Forced expiratory flow at 50% of FVC; FEF₇₅: Forced expiratory flow at 75% of FVC; FEV₁: Forced expiratory volume in the first second; FRC: Functional residual capacity; FVC: Forced vital capacity; IOS: Impulse oscillometry; LCI: Lung clearance index; MMAD: Mass median aerodynamic diameter; PC₂₀: Provocative concentration causing the FEV₁ to drop $\geq 20\%$; PD₂₀: Provocative dose causing the FEV₁ to drop $\geq 20\%$; R₂₀: Airway resistance to 20 Hz; R₅: Airway resistance to 5 Hz; R₅-R₂₀: Difference between airway resistance to 5 Hz and 20 Hz; SABA: Short acting B₂-antagonists; S_{acin}: Ventilation heterogeneity of the acinar airways; S_{cond}: Ventilation heterogeneity of the conductive airways; TLC: Total lung capacity; X₅: Ractance to 5 Hz; **A**: Change; pre-provocation minus post-provocation

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Availability of data and materials

The datasets used and/or analyzed during the current study are available from the corresponding author on reasonable request.

Authors' contributions

CC, IB, SV, SS, AL, HF, PH, JV, MF, NtH, HK, and MvdB: full access to study data and accountable for all aspects of the work, data interpretation, critical revision of the manuscript for important intellectual content, and approval of the final version of the manuscript for submission. IB, NtH, and MvdB were involved in the study design. IB and SS performed data acquisition and patient enrolment. AL, IB and CC performed data entry and analysis. CC performed the statistical analysis. JM, HK, and MvdB advised on the statistical analysis. CC, HK and MvdB wrote the manuscript.

Ethics approval and consent to participate

All subjects provided written informed consent. The OLiVIA study was approved by the ethics committee of the University Medical Center Groningen on March 19th, 2013 and is known under reference number M13.133950.

Consent for publication

Not applicable.

Competing interests

CC, IB, SV, SS, AL, JV, MF and NtH have nothing to disclose. HF reports other from MEDA, other from AstraZeneca, outside the submitted work. PH has a patent on the Novolizer with royalties paid, a patent on the Genuair with royalties paid, and a patent on the Twincer with royalties paid. HK reports that his institution has received grants from TEVA in relation to the submitted work, as well as consultancy fees from Novartis, GlaxoSmithKline, Fluidda, AstraZeneca, and Boehringer Ingelheim outside the submitted work. MvdB reports grants paid to the University from Astra Zeneca, TEVA, GSK, Chiesi, outside the submitted work.

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Author details

¹Department of Pulmonary Diseases, University of Groningen, University Medical Center Groningen, PO box 30.0001, 9700, RB, Groningen, The Netherlands. ²Groningen Research Institute for Asthma and COPD, University of Groningen, University Medical Center Groningen, PO box 30.0001, 9700, RB, Groningen, The Netherlands. ³Department of Pulmonary Diseases, Martini Hospital Groningen, PO box 30, 033 9700, RM, Groningen, The Netherlands. ⁴Department of Pharmaceutical Technology and Biopharmacy, University of Groningen, Antonius Deusinglaan 1, 9713, AV, Groningen, The Netherlands. ⁵Department of Epidemiology, University of Groningen, University Medical Center Groningen, PO box 30, 001 9700, RB, Groningen, The Netherlands.

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- References
- Cockcroft DW, Davis B. Direct and indirect challenges in the clinical assessment of asthma. Ann Allergy Asthma Immunol. 2009;103:363–9 quiz 369–372, 400.
- Ottanelli R, Rosi E, Romagnoli I, Ronchi MC, Lanini B, Grazzini M, et al. Perception of bronchoconstriction and bronchial hyper-responsiveness in asthma. Clin Sci (Lond). 2000;98:681–7.
- Segal LN, Goldring RM, Oppenheimer BW, Stabile A, Reibman J, Rom WN, et al. Disparity between proximal and distal airway reactivity during methacholine challenge. COPD. 2011;8:145–52.
- Brand PL, Rijcken B, Schouten JP, Koëter GH, Weiss ST, Postma DS. Perception of airway obstruction in a random population sample. Relationship to airway hyperresponsiveness in the absence of respiratory symptoms. Am Rev Respir Dis. 1992;146:396–401.
- Van Der Wiel E, ten Hacken NHT, Postma DS, Van Den Berge M. Small-airways dysfunction associates with respiratory symptoms and clinical features of asthma: a systematic review. J Allergy Clin Immunol. 2013;131:646–57.
- Perez T, Chanez P, Dusser D, Devillier P. Small airway impairment in moderate to severe asthmatics without significant proximal airway obstruction. Respir Med. 2013;107:1667–74.
- Takeda T, Oga T, Niimi A, Matsumoto H, Ito I, Yamaguchi M, et al. Relationship between small airway function and health status, dyspnea and disease control in asthma. Respiration. 2010;80:120–6.
- Van Der Wiel E, Postma DS, Van Der Molen T, Schiphof-Godart L, ten Hacken NHT, Van Den Berge M. Effects of small airway dysfunction on the clinical expression of asthma: a focus on asthma symptoms and bronchial hyper-responsiveness. Allergy Eur J Allergy Clin Immunol. 2014;69:1681–8.
- Mansur AH, Manney S, Ayres JG. Methacholine-induced asthma symptoms correlate with impulse oscillometry but not spirometry. Respir Med. 2008;102:42–9.
- Boudewijn IM, Telenga ED, Van Der Wiel E, Van Der Molen T, Schiphof L, ten Hacken NHT, et al. Less small airway dysfunction in asymptomatic bronchial hyperresponsiveness than in asthma. Allergy Eur J Allergy Clin Immunol. 2013;68:1419–26.
- 11. Cockcroft DW, Davis BE. Mechanisms of airway hyperresponsiveness. J Allergy Clin Immunol. 2006;118:551–9 quiz 560-1.
- 12. Joos GF, O'Connor B, Anderson SD, Chung F, Cockcroft DW, Dahlén B, et al. Indirect airway challenges. Eur Respir J. 2003;21:1050–68.
- Chan W, Cushley MJ, Holgate ST. The effect of inhaled adenosine 5'monophosphate (AMP) on airway calibre in normal and asthmatic subjects. Clin Sci. 1986;70:65p–6p.
- Cockcroft DW. Direct challenge tests: airway hyperresponsiveness in asthma: its measurement and clinical significance. Chest. 2010;138:185–245.
- Lexmond AJ, Boudewijn IM, Hagedoorn P, Schokker S, Cox CA, Vonk JM, et al. Bronchial provocation testing can be improved by using dry powder adenosine instead of nebulized AMP. Am J Respir Crit Care Med. 2017;197: 391-4.
- Sverrild A, Porsbjerg C, Backer V. The use of inhaled mannitol in the diagnosis and management of asthma. Expert Opin Pharmacother. 2012;13: 115–23.
- Brannan JD, Anderson SD, Perry CP, Freed-Martens R, Lassig AR, Charlton B, et al. The safety and efficacy of inhaled dry powder mannitol as a bronchial provocation test for airway hyperresponsiveness: a phase 3 comparison study with hypertonic (4.5%) saline. Respir Res. 2005;6:1–12.
- Lexmond AJ, Hagedoorn P, Frijlink HW, de Boer AH. Challenging the twominute tidal breathing challenge test. J Aerosol Med Pulm Drug Deliv. 2013;26:380–6.
- Cohen J, Postma DS, Douma WR, Vonk JM, De B a H, ten Hacken NHT. Particle size matters: diagnostics and treatment of small airways involvement in asthma. Eur Respir J. 2011;37:532–40.
- Lexmond AJ, Hagedoorn P, van der Wiel E, ten Hacken NHT, Frijlink HW, de Boer AH. Adenosine dry powder inhalation for bronchial challenge testing, part 1: inhaler and formulation development and in vitro performance testing. Eur J Pharm Biopharm. 2014;86:105–14.
- 21. Usmani OS, Biddiscombe MF, Barnes PJ. Regional lung deposition and bronchodilator response as a function of β 2 -agonist particle size. Am J Respir Crit Care Med. 2005;172:1497–504.

- 22. Cox CA, Boudewijn IM, Vroegop SJ, Schokker S, Lexmond AJ, Frijlink HW, et al. Extrafine compared to non-extrafine particle inhaled corticosteroids in smokers and ex-smokers with asthma. Respir Med. 2017;130:35–42.
- Lexmond AJ, van der Wiel E, Hagedoorn P, Bult W, Frijlink HW, ten Hacken NHT, et al. Adenosine dry powder inhalation for bronchial challenge testing, part 2: proof of concept in asthmatic subjects. Eur J Pharm Biopharm. 2014; 88:148–52.
- 24. Usmani OS. Treating the small airways. Respiration. 2012;84:441-53.
- Kendrick KR, Baxi SC, Smith RM. Usefulness of the modified 0-10 Borg scale in assessing the degree of dyspnea in patients with COPD and asthma. J Emerg Nurs. 2000;26:216–22.
- Caruso M, Holgate ST, Polosa R. Adenosine signalling in airways. Curr Opin Pharmacol Elsevier. 2006;6:251–6.
- 27. Burki NK, Alam M, Lee L-Y. The pulmonary effects of intravenous adenosine in asthmatic subjects. Respir Res. 2006;7:139.
- Eckle T, Koeppen M, Eltzschig HK. Role of extracellular adenosine in acute lung injury. Physiology. 2009;24:298–306.
- 29. Paintal AS. Sensations from J receptors. Am J Phys. 1995;10:238-43.
- Widdicombe J. Reflexes from the lungs and airways: historical perspective. J Appl Physiol. 2006;101:628–34.
- 31. Kleis S, Chanez P, Delvaux M, Louis R. Perception of dyspnea in mild smoking asthmatics. Respir Med. 2007;101:1426–30.

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