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## Salmonella-induced thrombi in mice develop asynchronously in the spleen and liver and are not effective bacterial traps

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## **Supplemental Materials and Methods**

### **Mice and infection with STm**

Wild-type (WT), C57BL/6 mice (6-8 weeks old; Harlan OLAC Ltd.) were used in accordance with local and national ethical approval HO licence numbers 3028/50 and P2E63AE7B. Mice were infected intraperitoneally (i.p.) or intravenously (i.v.) with  $1-5 \times 10^5$  attenuated STm SL3261 or virulent SL1344<sup>12</sup>. Non-infected (Day 0), vehicle-immunized mice served as controls. Viable bacterial numbers in tissues were calculated by plating out tissue homogenates and counting the numbers of colonies<sup>13</sup>.

### **Immunohistology, fluorescent microscopy and quantification of thrombi and bacteria**

Tissues were frozen in liquid nitrogen. Five-six  $\mu\text{m}$  acetone-fixed cryosections were stained either for immunohistochemistry (IHC) or immunofluorescence (IF) as described elsewhere<sup>13,14</sup>. Briefly, primary antibodies were incubated for 45min at RT before adding HRP-conjugated or biotin-conjugated secondary antibodies. ABCComplex alkaline phosphatase (Vector) was used. Signal was detected using diaminobenzidine for HRP activity and naphthol AS-MX phosphate with Fast Blue salt and levamisole for alkaline phosphatase activity. Antibodies were used to detect CD41, CD31, fibrin/fibrinogen, Ly6G, Ly6C, F4/80, *Salmonella* and nuclei (DAPI; supplemental Table 1)<sup>9</sup>. Images were obtained at x20 using a Zeiss (Jena, Germany) AxioScan.Z1 Scanner. Quantification of frequency and proportion of thrombus per total tissue section and bacteria per thrombus was performed using Zen 2012 blue edition (Jena, Germany) software.

### **Clodronate treatment**

Mice were treated i.p. with either 200  $\mu\text{l}$  (5 mg/ml) of clodronate or PBS liposomes (Liposoma, B.V., Amsterdam, The Netherlands) 24 h before STm infection as described elsewhere<sup>15,16</sup>

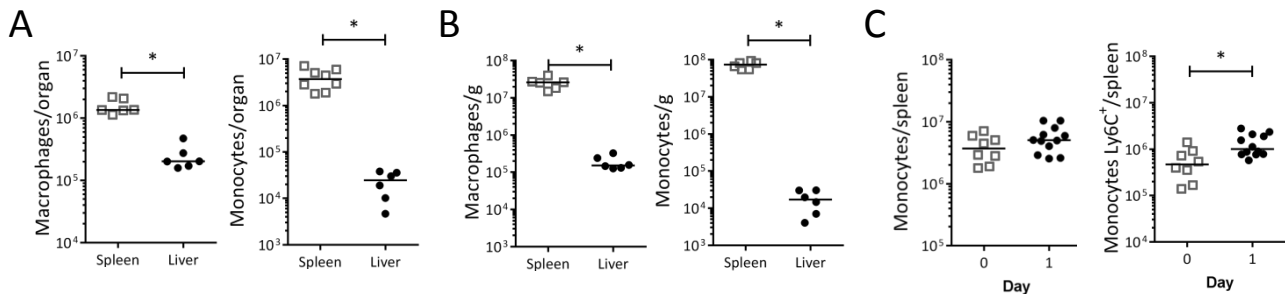
### **Statistical and data analysis**

Statistical significance was determined using the 2-tailed Mann-Whitney non-parametric sum of ranks test, 1-wayANOVA with the Kruskal-Wallis test. P values were calculated using GraphPad Prism software (GraphPad, La Jolla, CA) and were considered statistically significant when  $p < 0.05$ . Data presented are medians unless stated

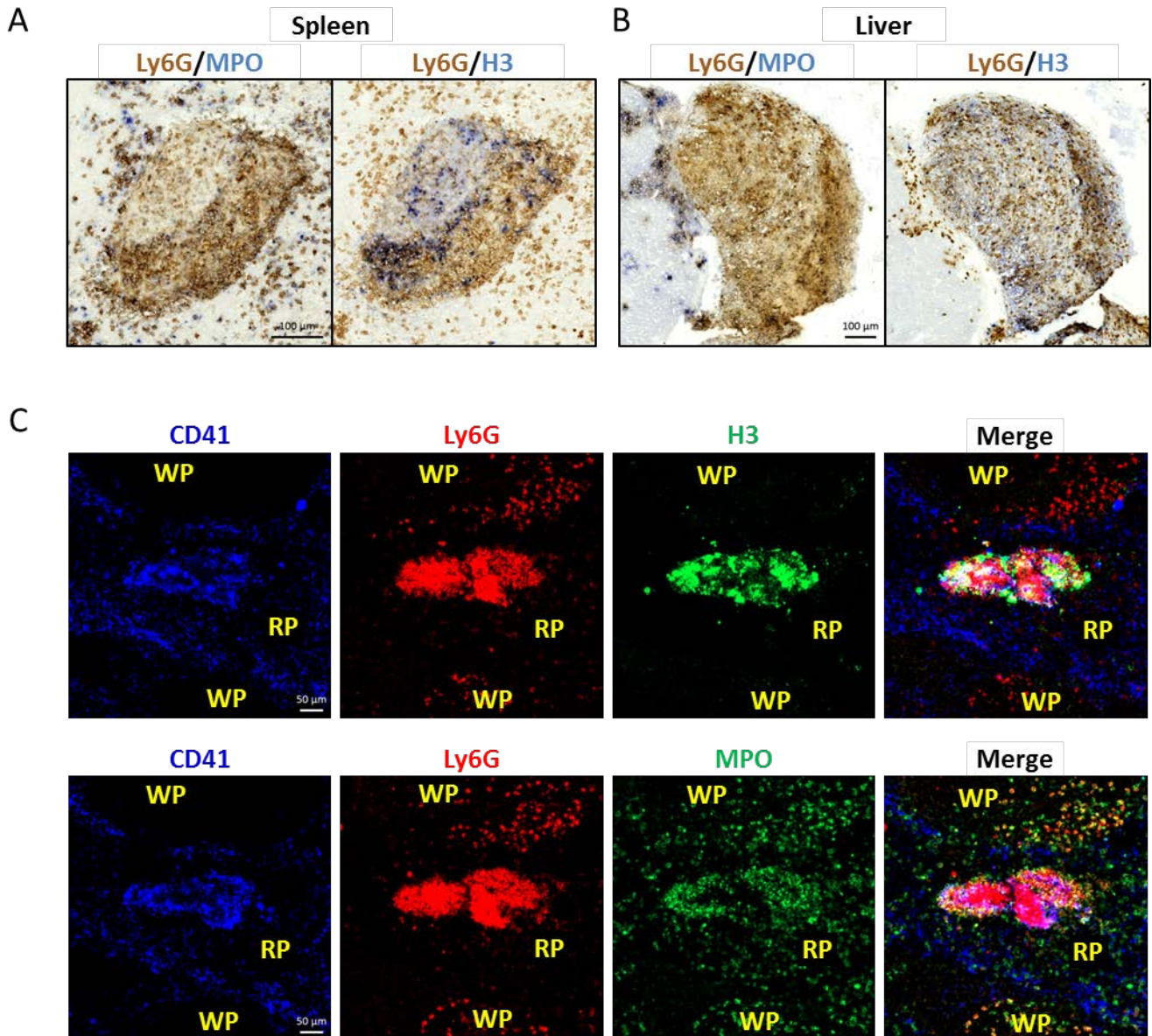
Supplemental Information

Table S1. Antibodies used for immunohistology, immunofluorescence and flow cytometry staining.

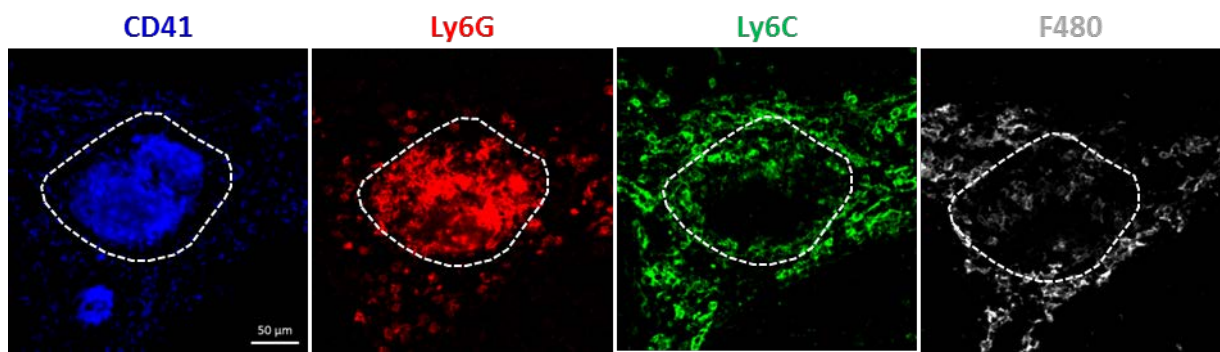
Reactivity	Isotype	Clone	Conjugate	Supplier
CD11b	Rat IgG2b, k	M1/70	eFluor 450	eBioscience
CD31	Rat IgG2a, k	390	Biotin	eBioscience
CD41	Rat IgG1, k	eBioMWRReg30	Purified	eBioscience
CD41	Rat IgG1, k	eBioMWRReg30	PE	eBioscience
F4/80	Rat IgG2a, k	BM8	APC	eBioscience
Fibrin/Fibrinogen	Goat, IgG	Polyclonal	Purified	Accurate Chemical & Scientific corporation
Goat	Donkey, IgG	Polyclonal	Alexa Fluor 647	Jackson Immunoresearch
Ly6C	Rat IgG2c, k	HK1.4	Biotin	eBioscience
Ly6G	Rat IgG2a, k	1A8	Biotin	BioLegend
Rabbit	Donkey, IgG	Polyclonal	Alexa Fluor 488	Jackson Immunoresearch
<i>Salmonella</i>	Rabbit, IgG	Polyclonal	Purified	Abcam
Myeloperoxidase	Rabbit, IgG	Polyclonal	Purified	Dako
Citrullinated histone H3	Rabbit, IgG	Polyclonal	Purified	Abcam
Streptavidin			Alexa Fluor 555	Invitrogen
Streptavidin			PE-Texas Red	BD Pharmingen



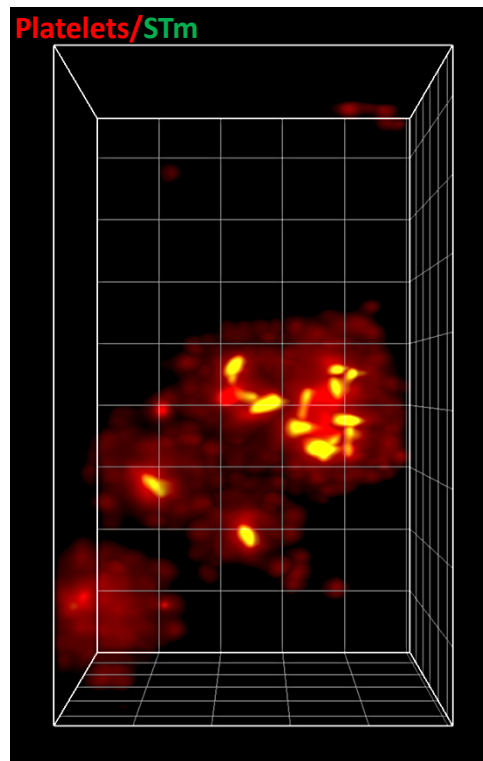
**Figure S1. Numbers of monocytic cells before and after infection.** (A) Graphs showing the numbers of macrophages (left, F4/80<sup>hi</sup>CD11b<sup>lo</sup>Ly6G<sup>-</sup> cells) and total monocytes (right, F4/80<sup>lo</sup>CD11b<sup>hi</sup>Ly6G<sup>-</sup> cells) (gating strategy from Rose, Misharin et al. 2012, Cytom; Tam et. al. 2014, Infect. & Immun.) present in the spleen and liver in non-infected mice. (B) Shows equivalent cell numbers per 1 g of tissue to adjust for organ size. (C) Graphs display number of total monocytes (left, F4/80<sup>lo</sup>CD11b<sup>hi</sup>Ly6G<sup>-</sup> cells) and inflammatory monocytes (right, F4/80<sup>lo</sup>CD11b<sup>hi</sup>Ly6G<sup>-</sup>Ly6C<sup>+</sup> cells) present in the spleen before and 1 day after infection with  $5 \times 10^5$  STm SL3261.



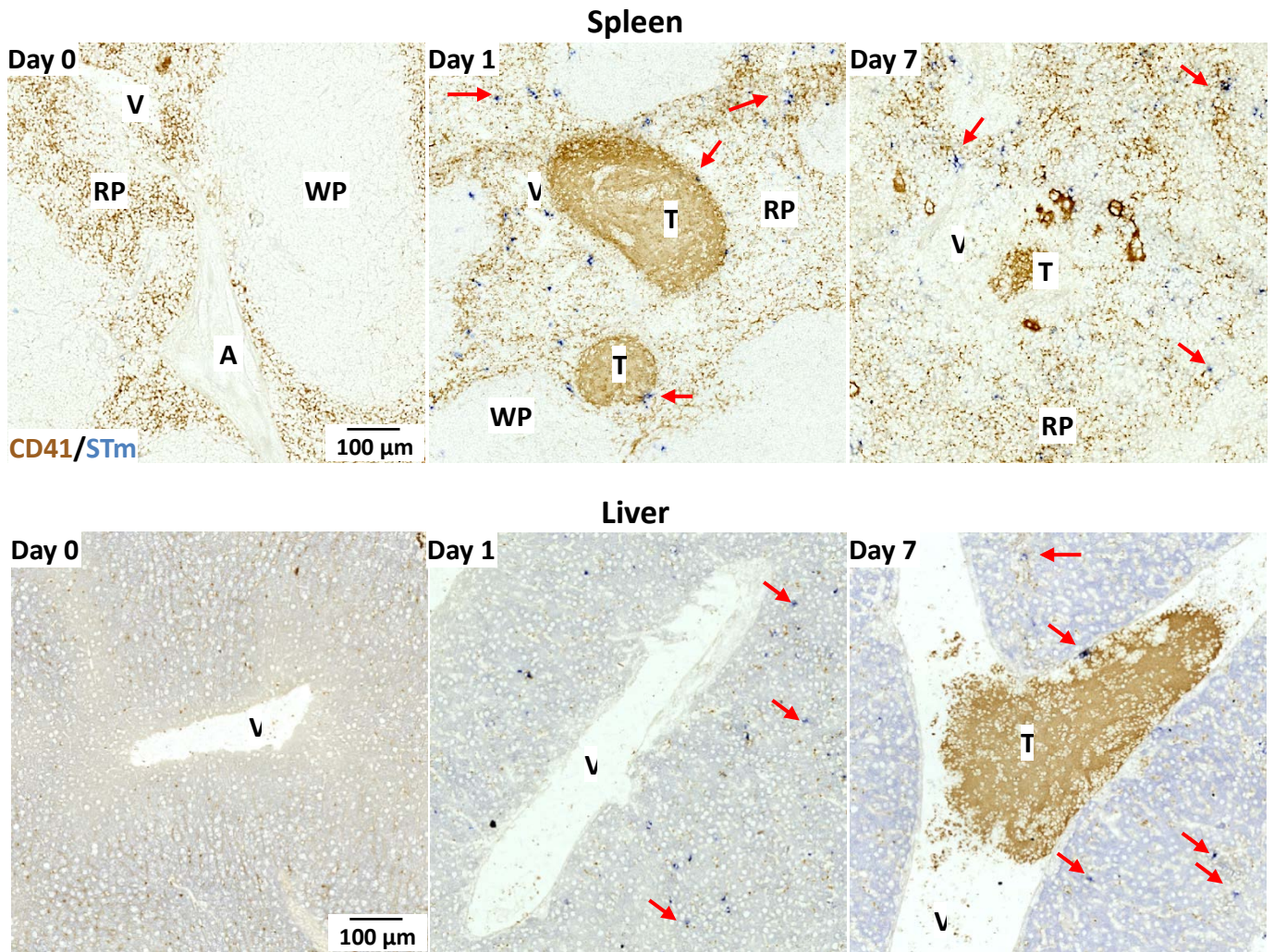
**Figure S2. MPO and citrullinated histone H3 are detected within thrombi after STm infection.** (A) Representative photomicrographs of a thrombus from the spleen of a mouse infected with  $5 \times 10^5$  STm SL3261 for 24h. (B) Representative photomicrographs of a thrombus from the liver of a mouse infected with  $5 \times 10^5$  STm SL3261 for 7 days. For A and B serial sections were stained to identify neutrophils (Ly6G; brown) together with either Myeloperoxidase (MPO; blue) or citrullinated histone H3 (H3; blue). (C) Immunofluorescence of a thrombus in the spleen from a 1 day-infected ( $5 \times 10^5$  STm SL3261) mouse co-incidentally stained for H3, Ly6G and CD41 (top row) or MPO, Ly6G and CD41 (bottom row).



**Figure S3. Association of different myeloid cells with thrombi.** Immunofluorescence photomicrographs of the same thrombus from a spleen taken from a mouse infected for 24h with  $5 \times 10^5$  STm SL3261. Sections were stained to identify platelets (CD41, blue), neutrophils (Ly6G, red), Ly6C<sup>+</sup> cells (green) and F4/80<sup>+</sup> cells (grey). Dotted line outlines the thrombus.

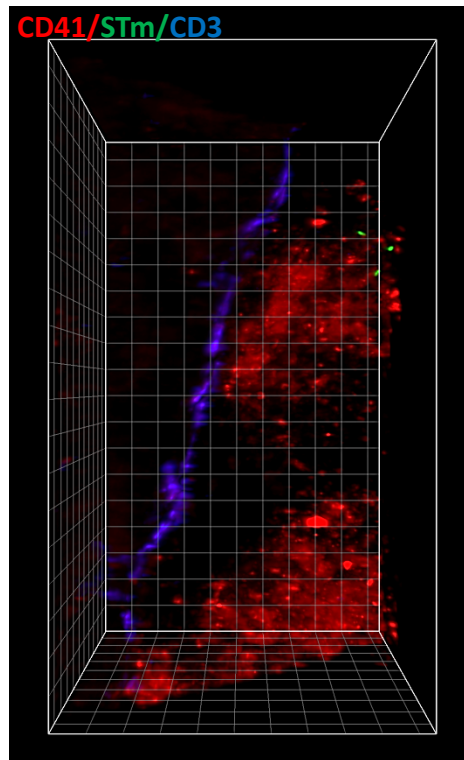


**Figure S4. *Salmonella* are found throughout platelet aggregates formed *in vitro*.** Platelet-Rich Plasma (PRP) was obtained from healthy donors and mixed with DiOC6 (3,3'-dihexyloxycarbocyanine iodide) to stain platelets. The PRP was mixed, with agitation, for 30 min with mCherry-expressing *Salmonella* Typhimurium (SL1344) at a ratio 5 platelets to 3 bacteria. At the end of the incubation, aggregates were fixed in 4% (w/v) paraformaldehyde. Whole aggregates were placed on a coverslip and imaged using Dual Inverted Selective Plane Illumination Microscopy (diSPIM). The image shows a representative z-stack, the grid size is 10  $\mu\text{m}$ . A 3D reconstruction can be observed in supplemental video 1.

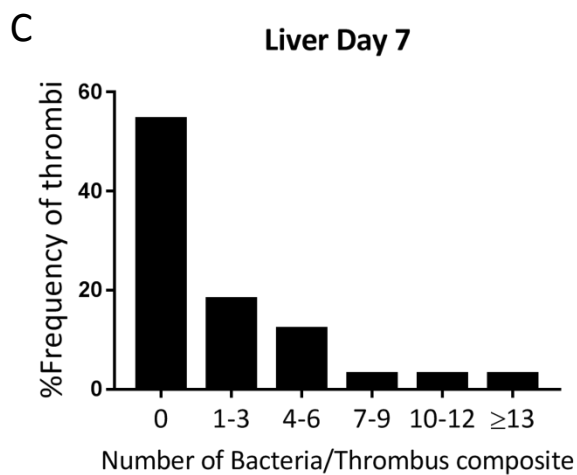
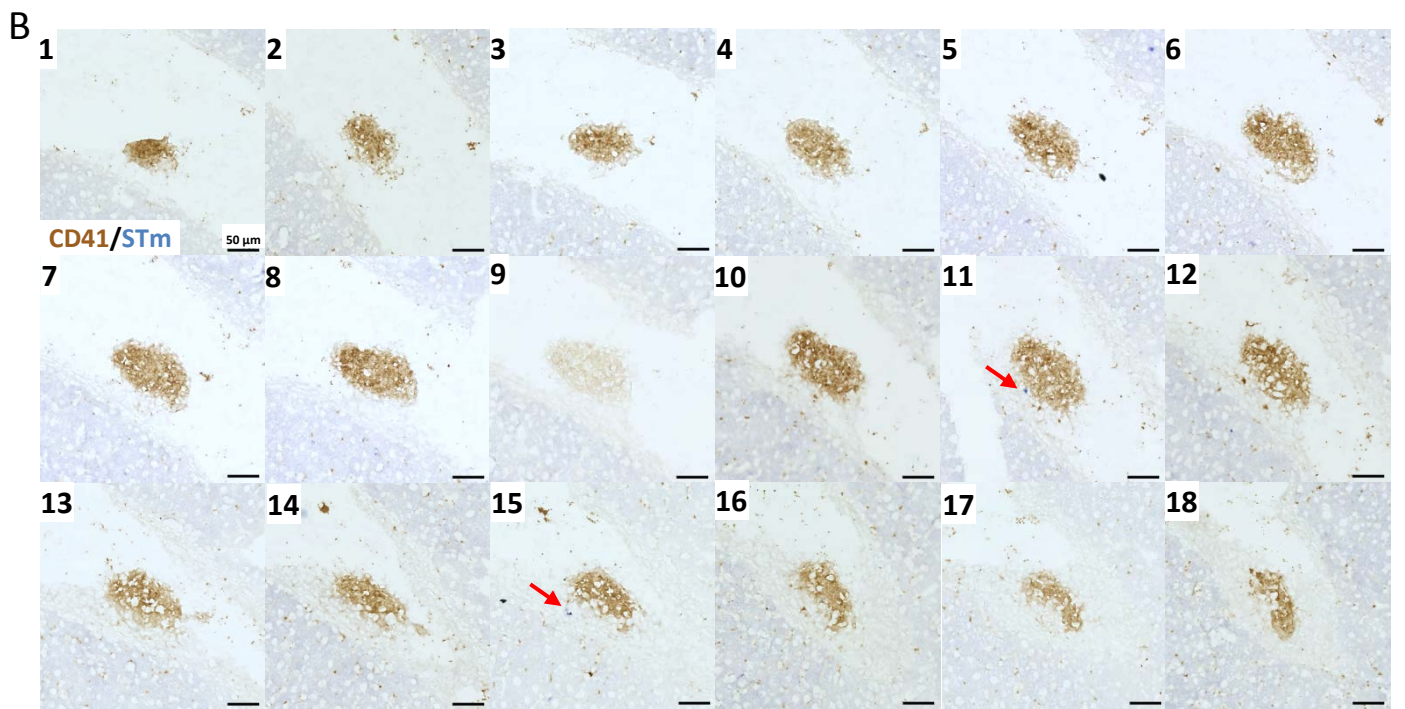
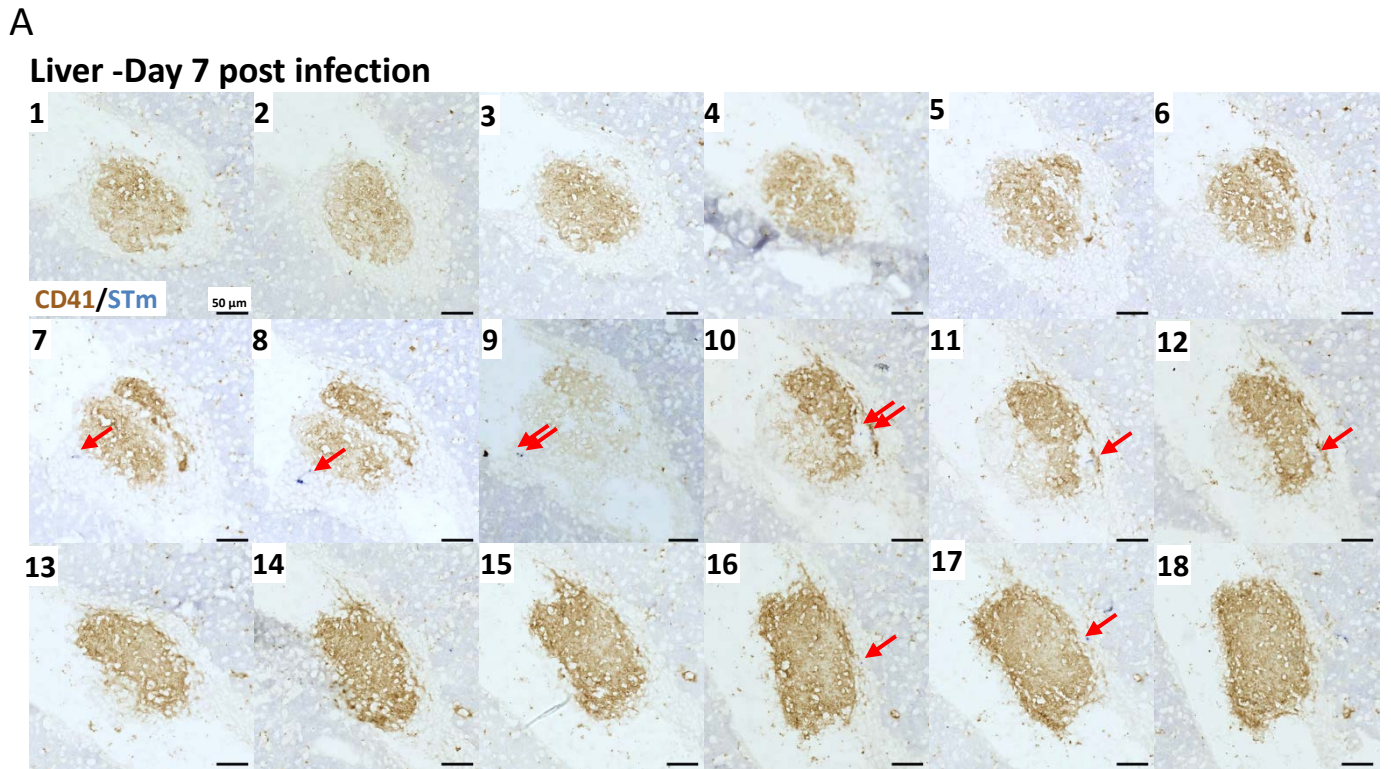


**Figure S5. Detection of bacteria within the spleen and liver after infection with STm.** Frozen tissue sections (spleen and liver) from WT mice infected with  $5 \times 10^5$  STm SL3261 were stained by IHC with anti-*Salmonella* antibody (blue) to identify bacteria and anti-CD41 (brown) to identify platelets, megakaryocytes and thrombi. Representative photomicrographs of spleen and liver sections from WT mice at day 0, 1, and 7 post-infection. Red arrows highlight some STm bacteria, V=Vein, RP=Red Pulp, WP=White Pulp, T=Thrombus.





**Figure S6. Detection of STm proximal to a thrombus.** 30- $\mu\text{m}$  liver sections from mice infected with  $5 \times 10^5$  STm SL3261 for 7 days were stained with anti-*Salmonella* antibody to detect bacteria (green), anti-CD41 (red) to detect platelets and anti-CD31 (blue) to delineate the blood vessel. Imaging was performed using Dual Inverted Selective Plane Illumination Microscopy (diSPIM). The image shows a representative z-stack, the grid size is 10  $\mu\text{m}$ . A 3D reconstruction can be observed in supplemental video 2.



**Figure S7. Serial sections of thrombi show that most contain no or few bacteria.** A) and B) 6- $\mu\text{m}$  serial sections of two individual thrombi in livers from two different mice infected with  $5 \times 10^5$  STm SL3261 for 7 days were stained with anti-*Salmonella* antibody to detect bacteria (blue) and anti-CD41 (brown) to detect platelet-rich thrombi. Red arrows=STm. C) Graph showing numbers of bacteria counted in the serial sections from up to 19 sections from 34 thrombi from two different mice. Data are. Scale bar= 50 $\mu\text{m}$