

Rift Valley Fever Virus Structural and Nonstructural Proteins: Recombinant Protein Expression and Immunoreactivity Against Antisera from Sheep

Bonto Faburay,¹ William Wilson,² D. Scott McVey,² Barbara S. Drolet,² Hana Weingartl,^{3,4} Daniel Madden,¹ Alan Young,⁵ Wenjun Ma,¹ and Juergen A. Richt¹

Abstract

The Rift Valley fever virus (RVFV) encodes the structural proteins nucleoprotein (N), aminoterminal glycoprotein (Gn), carboxyterminal glycoprotein (Gc), and L protein, 78-kD, and the nonstructural proteins NSm and NSs. Using the baculovirus system, we expressed the full-length coding sequence of N, NSs, NSm, Gc, and the ectodomain of the coding sequence of the Gn glycoprotein derived from the virulent strain of RVFV ZH548. Western blot analysis using anti-His antibodies and monoclonal antibodies against Gn and N confirmed expression of the recombinant proteins, and *in vitro* biochemical analysis showed that the two glycoproteins, Gn and Gc, were expressed in glycosylated form. Immunoreactivity profiles of the recombinant proteins in western blot and in indirect enzyme-linked immunosorbent assay against a panel of antisera obtained from vaccinated or wild type (RVFV)-challenged sheep confirmed the results obtained with anti-His antibodies and demonstrated the suitability of the baculo-expressed antigens for diagnostic assays. In addition, these recombinant proteins could be valuable for the development of diagnostic methods that differentiate infected from vaccinated animals (DIVA).

Key Words: Rift Valley fever virus—Structural proteins—Nonstructural proteins—Immunoreactivity profiles—Sheep.

Introduction

RIFT VALLEY FEVER (RVF), CAUSED BY Rift Valley fever virus (RVFV), represents a potential threat to global human and animal health. The virus belongs to the genus *Phlebovirus* of the family Bunyaviridae and causes severe epidemics among ruminants and humans in Africa and the Arabian Peninsula (Jupp and Cornel 1988, Shoemaker et al. 2002). Although the RVFV natural infection cycle has been linked to the *Aedes* and *Culex* spp. mosquitos, other mosquito species, including those found in North America, can be infected and subsequently transmit the virus (McIntosh and Jupp 1981, Gad et al. 1987, Gargan et al. 1988, Jupp and Cornel 1988, Turell et al. 2010). The fact that a variety of mosquito species can act as vectors increases the likelihood of RVFV becoming

endemic in areas outside of its traditional geographic range (Turell et al. 1988). The potential risk of spread of RVFV to temperate regions is exacerbated by global climate change, the almost ubiquitous presence of potential mosquito vectors, and international trade and travel. Currently, there are multiple challenges associated with efforts to combat RVF disease. Most important amongst them are the development and availability of an effective vaccine and pen-side diagnostic assays. In Africa, a live attenuated vaccine based on the Smithburn strain is available and provides lasting protective immunity, but it is abortigenic in pregnant livestock (Smithburn 1949, Hunter, et al. 2002, Botros et al. 2006), thus limiting its use. Killed or inactivated vaccines require multiple inoculations to be efficacious, which poses logistical challenges in developing countries where Rift Valley fever is endemic. The

¹Department of Diagnostic Medicine/Pathobiology, College of Veterinary Medicine, Kansas State University, Manhattan, Kansas.

²United States Department of Agriculture, Agricultural Research Service, Arthropod Borne Animal Disease Research Unit, Manhattan, Kansas.

³National Center for Foreign Animal Disease, Canadian Food Inspection Service, Winnipeg, Manitoba, Canada.

⁴Department of Medical Microbiology, University of Manitoba, Winnipeg, Canada.

⁵Department of Veterinary and Biomedical Science, South Dakota State University, Brookings, South Dakota.

use of live-attenuated vaccines in nonendemic countries is less desirable due to concerns of introduction of live RVFV into a nonendemic region and the potential risk of reversion to virulence. Importantly, development of sensitive diagnostics, as well as effective vaccines, will require the identification and expression of sufficiently immunoreactive and potentially immunoprotective proteins that could be used in diagnostic assays or as candidates for vaccine development.

Like all members of the Bunyaviridae family, the RVFV genome comprises three single negative-stranded RNA strands, composed of large (L), medium (M), and small (S) segments (Elliott 1996, Giorgi 1996, Schmaljohn 1996). The L segment encodes the viral RNA-dependent RNA polymerase. The M segment encodes the structural glycoproteins, Gn and Gc, which are expressed as a polyprotein precursor that is processed by cellular proteases during maturation (Collett 1986, Suzich et al. 1990, Gerrard and Nichol 2007). It has been shown that these structural glycoproteins elicit production of virus-neutralizing antibodies important for protective immunity (Niklasson et al. 1985, Besselaar and Blackburn 1994, Bird et al. 2011, Papin et al. 2011, Piper et al. 2011). The M segment also encodes a nonstructural protein, NSm, which was shown to have an antiapoptotic function and a 78-kD protein whose function is not known yet (Anderson and Peters 1988, Won et al. 2007). The S segment encodes the nucleocapsid protein, N, and a nonstructural protein, NSs, which is recognized as a major virulence factor of the virus by counteracting host innate immunity (Vialat et al. 2000, Bouloy et al. 2001, Billecocq et al. 2004, Le May et al. 2004). In this study, we describe the expression of various RVFV proteins using the baculovirus system and profile their reaction pattern with antisera derived from RVFV vaccinated or challenged sheep.

Materials and Methods

Cloning and construction of recombinant bacmid

The full-length coding sequences of the RVFV nucleoprotein (N) and the nonstructural protein (NSs) were amplified by PCR using a proofreading DNA polymerase, *Pfx50* (Life Technologies/Invitrogen, Carlsbad, CA), and primers (Table

1) designed from published sequences of RVFV strain ZH548 (accession no. DQ380151). Plasmid pET30 Ek/LIC containing the entire coding region of the S segment of ZH548 was used as template for the PCR. The ectodomain and full-length coding sequences of the virus structural glycoproteins Gne and Gc, respectively, as well as the nonstructural M protein NSm were synthesized according to the published sequences of the ZH548 strain (Genewiz, San Diego, CA). Amplicons were purified using a Qiagen gel or PCR purification kit (Qiagen, Valencia, CA). The purified products were cloned into a pFastBac/CT-TOPO vector (Life Technologies/Invitrogen, Carlsbad, CA). The TOPO cloning reactions were transformed into One Shot Mach1 T1 chemically competent *Escherichia coli* to produce the respective donor plasmids pRF-N, pRF-NSs, pRF-Gne, pRF-Gc, and pRF-NSm. The sizes and sequences of the inserts in the donor plasmids were confirmed by PCR using gene-specific primers (Table 1) that were confirmed by restriction enzyme analysis and DNA sequencing. Donor plasmids containing the gene of interest were purified from the *E. coli* transformants using a Qiagen Miniprep kit (Qiagen, Valencia, CA). The constructs were transformed into MAX Efficiency DH10Bac-competent *E. coli* to generate a recombinant bacmid by site-specific transpositioning. Recombinant bacmids were purified using HiPure Plasmid Miniprep kit. Transpositioning of the gene of interest into a recombinant bacmid was confirmed by PCR using M13F and M13R primers (Heidecker et al. 1980).

Signal peptide prediction and modification of Gn and Gc

The RVFV M segment, which encodes the envelope glycoproteins Gn and Gc, has at least four translation initiation sites within the single mRNA transcribed from the M segment. We hypothesized that sequences starting from one of the initiation codons (ATG) to the start of Gn may serve as a signal peptide and guide the translocation of the polyprotein from the cytoplasm to the endoplasmic reticulum (ER). Prediction of signal peptides and signal peptidase cleavage sites was performed using hidden Markov model and neural network model prediction programs at SignalP 3.0. A 54-nucleotide sequence beginning from the

TABLE 1. PRIMERS USED FOR THE AMPLIFICATION AND CLONING OF RIFT VALLEY FEVER VIRUS PROTEIN-CODING SEQUENCES

Primer	Orientation	Sequences	Target/description	Reference
JAR979	Forward	CACCATGGACAACACTATCAAGACCTTGCGATCC	N cds	Designed by this study
JAR980	Reverse	GGCTGCTGTCTTGTAAGCCTGAGCG	N cds	Designed by this study
JAR981	Forward	CACCATGGATTACTTTCCTGTGATATCTGTTGATTTG	NSs cds	Designed by this study
JAR982	Reverse	ATCAACCTCAACAAATCCATCATCATCACTCTCC	NSs cds	Designed by this study
JAR975	Forward	CACCATGATTGAAGGAGCTTGGGATTC	NSm cds	Designed by this study
JAR976	Reverse	AGCAAAAACAACAGGTGCCAAAGC	NSm cds	Designed by this study
JAR996 ^a	Forward	CACCATGACAGTCCTTCCAGCCTTAG	Gne or Gc cds	Designed by this study
JAR111	Reverse	GGCACTGAGAGCAGTGTGACACTG	Gne cds	Designed by this study
JAR987	Reverse	TGAGGCCTTCTTAGTGGCAGCAAG	Gc cds	Designed by this study
JAR990	Forward	CCCAGTCACGACGTTGTA AAAACG	M13 sequencing primer	Heidecker et al. 1980
JAR991	Reverse	AGCGGATAACAATTTACACACAGG	M13 sequencing primer	Heidecker et al. 1980

^aA signal peptide specific primer common for structural glycoproteins, Gne and Gc.

N, nucleoprotein; cds, coding sequence; NSs, nonstructural protein S; NSm, nonstructural protein m; Gne, ectodomain of Gn glycoprotein; Gc, Gc glycoprotein.

fifth ATG of the RVFV M segment, upstream of Gn coding sequence, was identified as a strong signal peptide with a single signal peptidase cleavage site. To ensure expression of glycosylated forms of the proteins, this signal sequence was fused to the amino terminus of Gne and Gc and amplified by PCR using primers shown in Table 1. The sequences were cloned into recombinant bacmids and expressed in a baculovirus expression system (Life Technologies/Invitrogen, Carlsbad, CA).

Expression and purification of recombinant RVFV proteins

To express recombinant RVFV proteins, highly purified recombinant bacmids were transfected, using Cellfectin II reagent (Life Technologies/Invitrogen Carlsbad, CA), into *Spodoptera frugiperda*, Sf9, cells (Life Technologies/Invitrogen) grown in Sf-900 II SFM medium (Life Technologies/Invitrogen, Carlsbad, CA) supplemented with 10% fetal bovine serum (FBS) and 100 U/mL–100 µg/mL penicillin–streptomycin. Protein expression was carried out using P2 or higher passage recombinant baculovirus stock (>10⁷ pfu/mL) in T75 or T175 Biolite flasks (ThermoFisher Scientific, Dubuque, IA). To visualize expression of recombinant proteins, samples were subjected to western blot analysis by resolving on 12% Bis-Tris polyacrylamide gel and detected using anti-His-horseradish peroxidase (HRP) antibody. Purification of recombinant proteins was carried out using Ni-NTA superflow resin (Novagen, Rockland, MA) according to manufacturer's instructions. Recombinant proteins were eluted with an elution buffer containing 300 mM NaCl, 50 mM Na₂PO₄ (pH 8.0), and 250 mM imidazole and dialyzed overnight against storage buffer, phosphate-buffered saline (PBS; pH 7.4), and 5% glycerol. The purified proteins were stained with Coomassie Blue, and protein concentrations were determined using the bicinchoninic acid (BCA) assay (Thermo Scientific, Rockford, IL) at an absorbance of 562 nm, using bovine serum albumin (BSA; Sigma-Aldrich, St. Louis, Mo) as the protein standard. Aliquots were stored at –80°C until used.

Immunofluorescence antibody assay

To confirm expression of recombinant RVFV proteins, we carried out an immunofluorescence antibody assay. Briefly, Sf9 cells were infected with recombinant baculovirus carrying RVFV N- or Gne-coding sequences. About 36 h later, cells were harvested and adhered to glass slides using a cytocentrifuge (Cytopro, Wescor, Logan, UT) according to manufacturer's instruction and fixed in acetone for 5 min at –20°C. Slides were blocked in PBS containing 1% BSA at 37°C for 45 min and then incubated for 30 min at 37°C with mouse monoclonal antibodies ID8 and 4D4 (provided by Dr. Connie W. Schmaljohn, the United States Army Medical Research Institute for Infectious Diseases, USAMRIID), against N and Gn proteins, respectively. Slides were rinsed for 10 min in 1× fluorescence antibody (FA) rinse buffer (2.85 g Na₂CO₃, 8.4 g NaHCO₃, 2.125 g NaCl, distilled water to 1000 mL; pH 9 to 9.5) and the probed with anti-mouse fluorescein isothiocyanate (FITC)-conjugated secondary antibody for 30 min at 37°C (Sigma-Aldrich, St. Louis, MO). Slides were rinsed for 10 min in 1× FA rinse buffer and counterstained with 4',6-diamidino-2-phenylindole (DAPI; Invitrogen, Molecular

Probes[®]) containing mounting medium. Slides were examined under a fluorescent microscope (Nikon, Eclipse 90i[®]) at 100× magnification.

Tunicamycin assay

A baculovirus expression system was used to produce glycosylated recombinant proteins, a posttranslational modification shown to enhance antigenicity/immunogenicity (Gavrilov et al. 2011). To confirm that the modified Gne and Gc proteins were glycosylated, a tunicamycin glycosylation inhibition assay was performed. For this, six-well plates were seeded with 2×10⁶ Sf9 cells per well and infected with recombinant baculoviruses expressing RVFV Gne or Gc at a multiplicity of infection (MOI) of 1. Immediately following infection, tunicamycin was added to each well at varying concentrations of 0.5 µg/mL, 1 µg/mL, 3 µg/mL, 6 µg/mL, 8 µg/mL, and 10 µg/mL. Cells were harvested 5 days post-infection (pi) and resuspended in PBS (pH 7.4) containing 1× Complete protease inhibitor (Roche Diagnostics[®], Indianapolis, IN). The samples were separated by sodium dodecyl sulfate polyacrylamide gel electrophoresis (SDS-PAGE) in NuPAGE 12% Bis-Tris gels (Life Technologies/Invitrogen) and transferred on to polyvinylidene difluoride (PVDF) membranes (GE Healthcare, Amersham Hybond-P). The membranes were probed as described above using anti-His (carboxy-terminal)-HRP monoclonal antibody (Life Technologies/Invitrogen, Carlsbad, CA). Reactivity was detected using the AEC (3-amino-9-ethyl-carbazole) peroxidase substrate system (Abcam, Cambridge, MA).

Rift Valley fever virus anti-sera

Sheep (Rambouillet) were vaccinated subcutaneously with 10⁶ plaque-forming units (pfu) of RVFV vaccine strain MP12. These studies were performed at Kansas State University (Biosecurity Research Institute [BRI] BSL-3Ag facility) or the Arthropod-Borne Animal Diseases Research Unit (ABADRU) large animal isolation building (LAIB) in Laramie, WY (day 28 sheep sera, P1–P6). Blood samples were collected from individual animals at specific time points (days) postvaccination (pv) for sera. Prevacination blood samples were collected from each individual sheep for use as preimmune control sera. Day-28 pi antisera were obtained from previous sheep challenged at the CFIA BSL-3Ag facility in Winnipeg, Canada, with the wild-type RVFV ZH501 strain (sheep P7–P10). All sera were stored at –80°C, and sera from the CFIA facility were heat-inactivated with 0.5% Tween (56°C for 30 min) prior to removal from the BSL-3Ag laboratory. Institutional Animal Care and Use Committee (IACUC) protocols were approved by Kansas State University and University of Wyoming (ABADRU-LAIB).

Western blot analysis

Briefly, approximately 5 µg of each of recombinant RVFV protein was subjected to electrophoresis in 12% Bis-Tris polyacrylamide gel in 1×3-(*N*-morpholino)propanesulfonic acid (MOPS) running buffer (Life Technologies/Invitrogen). The proteins were transferred by electroblotting onto PVDF membranes according to standard protocols. The membrane was blocked in 0.1% Tween-20 in PBS (pH 7.4) containing 3% BSA at room temperature for 1 h. The blot was washed three

times for 5 min each in 0.1% Tween-20 in PBS. All subsequent washing steps were carried out as described above. For analysis of recombinant protein expression, the membrane was incubated with anti-His(carboxyterminal)-HRP (Life Technologies/Invitrogen) diluted 1:5000 in blocking solution. Expression of recombinant proteins N and Gne was further confirmed using a primary antibody, mouse anti-N (R3-ID8) and mouse anti-Gn monoclonal antibody (4D4), respectively, at a dilution of 1:2000. For analysis of reactivity against sheep sera, the membrane was incubated with a 1:100 dilution of each test serum for 1 h at room temperature. After washing, the membrane was incubated for 1 h at room temperature with Protein G-HRP (Abcam, Cambridge, MA) diluted 1:25,000. After the final washing steps, specific reactivity was detected using AEC peroxidase substrate (Sigma-Aldrich, St. Louis, MO).

Indirect enzyme-linked immunosorbent assay

A 96-well plate (Nunc, Maxisorp[®]) was used for the indirect enzyme-linked immunosorbent assay (ELISA). Each well was coated overnight at 4°C with 100 ng of recombinant protein in 100 μ L of Dulbecco's coating buffer (pH 7.4) (Life Technologies/Invitrogen). Plates were blocked for 15 min at 37°C with PBS (pH 7.4) containing 0.1% Tween and 1% skim milk. After washing three times with 0.1% Tween-20 in PBS, a volume of 200 μ L of test serum, diluted 1:200 in the blocking solution, was added and incubated at 37°C for 1 h. All subsequent washing steps were carried out three times as indicated above. Each serum sample was tested in duplicate. Each test included a positive control, obtained from sheep challenged with a virulent strain of RVFV (ZH501), and negative controls obtained from the respective sheep prior to vaccination.

After washing, plates were incubated with Protein G-HRP (Abcam, Cambridge, MA), diluted 1:50,000 in blocking solution, at 37°C for 1 h. Protein G has high binding affinity to immunoglobulin (IgG) from sheep, goats, horses, and rabbits, with little or no binding affinity to IgM. After washing, 100 μ L of substrate buffer containing 0.1 mg/mL 3,3',5,5'-tetramethylbenzidine (TMB) (Thermo Scientific, Rockford, IL) and hydrogen peroxide (H₂O₂) was added and the plates were incubated in the dark for 25 min. The reaction was stopped with 2 M H₂SO₄, and optical densities (OD) were measured at 450 nm using a microplate reader (Fluostar Omega, BMG Labtech, Cary, NC). For each ELISA, the cutoff OD value was determined by addition of 2 standard deviations (SD) to the mean OD value of serum obtained from prevaccinated/non-infected sheep.

Statistical analysis

Data were analyzed using a *t*-test of independent samples with equal or unequal variances. To reduce the effect of variable individual host immune response on reactivity, we calculated geometric means of OD values and analyzed the differences for statistical significance.

Results

Expression of RVFV recombinant proteins

Using recombinant baculovirus, RVFV structural (Gc, Gn, and N) and nonstructural (NSs and NSm) proteins, each

containing a hexahistidine tag at their carboxyl terminus, were expressed in a eukaryotic expression system using Sf9 cells. To ensure translocation in the ER and glycosylation of the structural glycoproteins Gne and Gc, a signal peptide was fused upstream of the amino terminus of both proteins. Infection of Sf9 cells with recombinant baculovirus carrying the full-length Gn coding sequence resulted in no or a low amount of Gn protein. To minimize interactions of the full-length Gn with cellular membranes, only the ectodomain of the protein Gne, without the transmembrane and cytosolic regions, was expressed. The proteins were purified and detected by western blot using anti-His-HRP monoclonal antibody (Fig. 1A). Recombinant proteins of the expected molecular weights were expressed (Gc, 60 kD; Gne, 54 kD; N, 30 kD; NSs, 33 kD; NSm, 17 kD). Expression of N and Gn proteins was further confirmed by immunoreaction with anti-N or anti-Gn mouse monoclonal antibodies (Fig. 1B). Coomassie-stained gels of the purified proteins are shown in Figure 1C. Immunofluorescent antibody assay using the mouse monoclonal antibodies against the Gn and N proteins further confirmed the expression of the proteins in Sf9 cells (Fig. 2). A specific green immunofluorescent signal was observed around the periphery of the nucleus of Gne-expressing cells, whereas N appeared to show a more diffuse staining (Fig. 2).

Baculovirus-expressed Gn and Gc proteins are N-glycosylated

It is known that the RVFV Gn and Gc glycoproteins carry one and four putative *N*-glycosylation sites, respectively (Gerrard and Nichol 2007). To characterize glycosylation patterns of Gne and Gc, biochemical inhibition of *N*-glycosylation by treatment of cells with tunicamycin, a potent inhibitor of bacterial and eukaryote *N*-acetylglucosamine transferases, was carried out. Treatment of Sf9 cells infected with either recombinant Gc or Gne baculoviruses with varying concentrations of tunicamycin (0.5–10 μ g) resulted in inhibition of glycosylation of the proteins demonstrated by a lower molecular weight of the reactive proteins when compared to nontreated controls. The shift was more obvious for Gc (Fig. 3A), known to have four putative glycosylation sites when compared to Gne (showing marginal shift), known to have only one glycosylation site (Fig. 3B). As a control, baculovirus-expressed sheep prion protein (PrP) expressed in Sf9 cells was used. Treatment with tunicamycin resulted in significant inhibition of *N*-glycosylation (Fig. 3C).

Western blot analysis

We examined antibody reactivity against the recombinant proteins N, NSs, NSm, Gne, and Gc using sera collected at different time points *pv* (days 3, 10, and 28) from MP12 vaccinated sheep. Recombinant N was reactive with day-3 *pv* sera, and both N and NSs proteins showed antibody reactivity with day-10 *pv* and day-28 *pv* sera, with N showing consistently stronger reactivity than NSs (Fig. 4). Reactivity of the structural glycoproteins Gne and Gc showed that Gne was reactive with 1 of 2 serum samples on day 3 *pv*. Thereafter, it remained consistently reactive with all sera obtained on day 10 and 28 *pv* (Table 2). The Gc protein showed no reactivity at day 3 *pv* and weak reactivity with day 10 *pv* sera, where only 1 of the 4 serum samples tested was positive; it was 90%

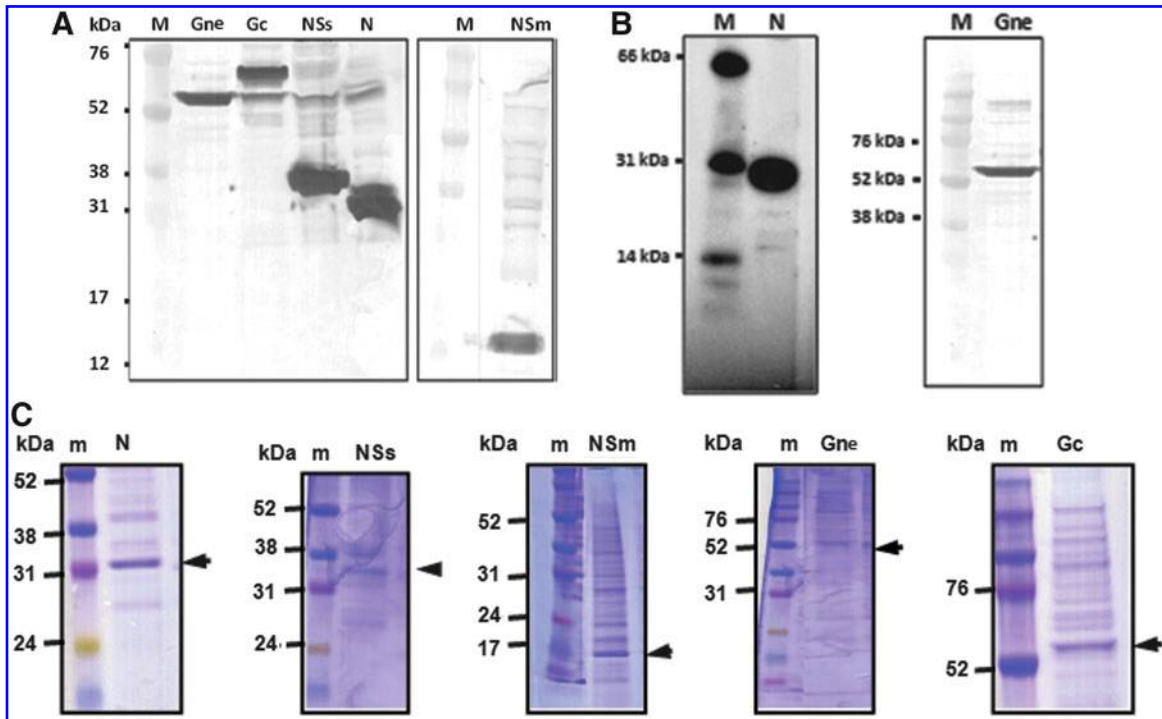


FIG. 1. Baculovirus-expressed Rift Valley fever virus (RVFV) proteins. Proteins were purified and detected using anti-His(C-term)HRP monoclonal antibody. Gne, 54 kD; Gc, 60 kD; NSs, 34 kD; N, 31 kD; NSm, 14 kD; M, marker (A). Monoclonal antibodies against N and Gn, IDE8 and 4D4, respectively, were used to confirm expression of the respective proteins (B). (C) Coomassie Blue staining of the purified proteins. N, nucleoprotein; NSs, nonstructural protein S segment; NSm, nonstructural protein m segment; Gne, ectodomain of amino-terminal glycoprotein; Gc, carboxy-terminal glycoprotein; M, molecular weight marker.

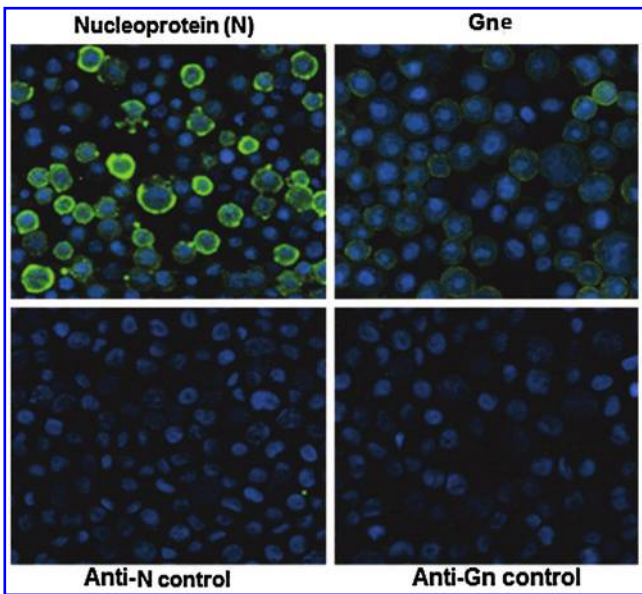


FIG. 2. Immunofluorescence antibody assay confirms expression of the recombinant proteins Gne and N in Sf9 cells. Monoclonal antibodies 4D4 and ID8 were used to detect expression of Gne and N, respectively. A specific green fluorescent signal around the nucleus of Gne-expressing cells is seen indicating that the recombinant protein is within the cell. Anti-Gn control, noninfected Sf9 cells stained with Gn monoclonal antibody (4D4); anti-N control, noninfected Sf9 cells stained with N monoclonal antibody (ID8) show negative staining. The cell nucleus is stained in blue color.

reactive with day 28 pv sera. To examine reactivity of RVFV proteins further, we performed immunoblot analysis using day-28 pi sera obtained from sheep infected with the wild-type strain ZH501 of RVFV. Recombinant N and Gne proteins showed consistent and strong reactivity against all sera (4/4); NSs and Gc were also reactive against all sera (4/4) but with relative lower signal intensities. We did not detect specific reactivity with the NSm protein, either with the MP12-vaccinated or ZH501-infected sheep sera.

Indirect ELISA

An indirect ELISA was developed to assess antibody reactivity of the recombinant proteins against sera obtained from sheep vaccinated with RVFV MP12 vaccine strain or challenged with wild-type ZH501 RVFV strain. For MP12-vaccinated sheep sera, the recombinant proteins N, NSs, Gc, and Gne showed time-dependent increase in reactivity, shown by an increase in OD values (Fig. 5A–D). A consistent strong reactivity for all proteins was observed with day-28 sera. There was particularly strong antibody reactivity with N and to a lesser extent with Gne on day 3 pv (Fig. 5A, C). The level of reactivity with NSs was also strong but had comparatively lower OD values (Fig. 5B). Reactivity with Gc was high on day 10 pv and was strongest on day 28 (Fig. 5D). For the wild type exposure-derived sheep sera, the recombinant proteins, N, NSs, Gne, and Gc were reactive with the day 28 pi (data not shown). In contrast, NSm showed low reactivity manifested by relatively low OD values with sera from both MP12 and wild-type infected sheep (Fig. 5E). Sera from three sheep (infected with the wild-type ZH501, P7 and P10, and

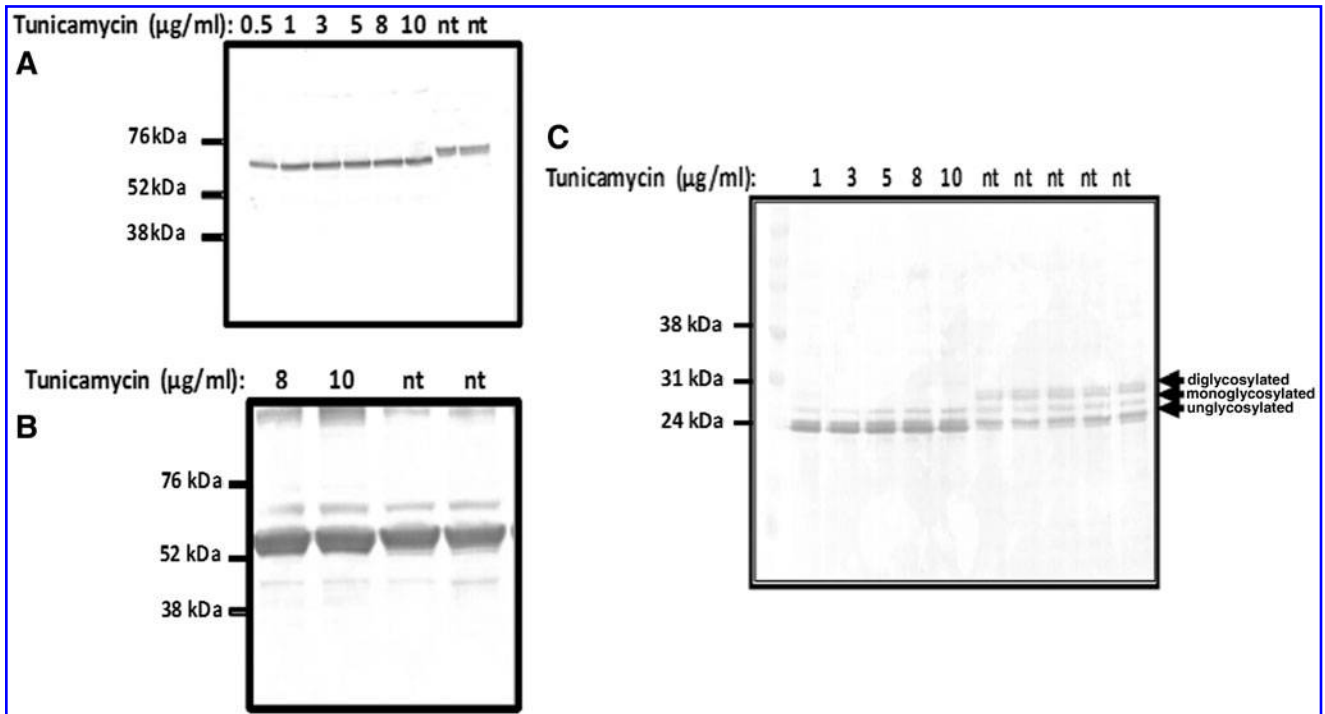


FIG. 3. *In vitro* glycosylation assay of Rift Valley fever glycoproteins, Gc and Gne. (A) Treatment of Gc-recombinant baculovirus-infected Sf9 cells with varying concentrations of tunicamycin (0.5 $\mu\text{g}/\text{mL}$ to 10 $\mu\text{g}/\text{mL}$) resulted in inhibition of glycosylation shown by a shift in electrophoretic migration. (B) Similar treatment of Gne (8 $\mu\text{g}/\text{mL}$ and 10 $\mu\text{g}/\text{mL}$) resulted in marginal molecular weight shift (compare with nontreated, nt), since the protein has one putative *N*-glycosylation site; Gc has four putative *N*-glycosylation sites. Treatment of baculovirus expressed sheep prion protein (PrP) with varying concentrations of tunicamycin (1 $\mu\text{g}/\text{mL}$ to 10 $\mu\text{g}/\text{mL}$) resulted in inhibition of glycosylation of the protein (C). nt, nontreated controls; m, molecular weight marker.

MP-12-immunized sheep, P5) showed significant reactivity ($p < 0.05$) in comparison with the negative control (day 0 serum). The positive sera were obtained from sheep on day 28 pi with the wild-type RVFV ZH501.

Discussion

There is increasing demand for sensitive and safe diagnostic tests and efficacious vaccines for zoonotic pathogens,

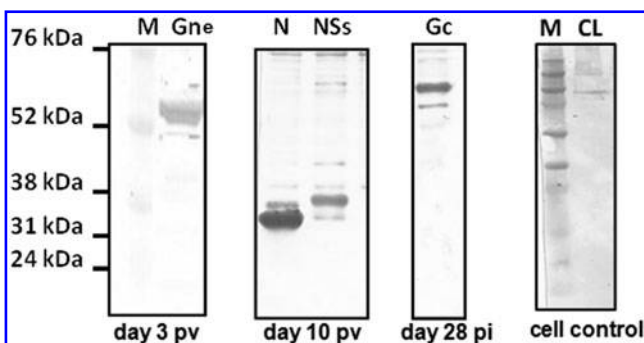


FIG. 4. Western blot showing a sample of immunoreactivity of purified baculovirus-expressed proteins with RVFV antisera from sheep. Reactivity shows that the proteins were expressed in the correct conformation. Gne, ectodomain of amino-terminal glycoprotein; N, nucleoprotein; NSs, non-structural protein S segment; Gc, carboxy-terminal glycoprotein; CL, noninfected cell lysate; m, molecular weight marker; pv, postvaccination; pi, postinfection.

including RVFV, to protect human and animal health. The recent spread of RVFV beyond its traditional endemic boundaries in Africa to the Arabian Peninsula (Jupp and Cornel 1988, Ikegami and Makino 2009, Abdo-Salem et al. 2011) has resulted in increased interest for RVFV vaccines, rapid diagnostics, and associated immunoreagents. At present, diagnosis of RVFV infection is achieved using various techniques including virus isolation (Shope and Sather 1979, Anderson et al. 1989), antigen detection (Niklasson et al. 1983, Meegan et al. 1987, Drolet et al. 2012), nucleic acid amplification techniques (Ibrahim et al. 1997, Garcia et al. 2001, Drosten et al. 2002, Drolet et al. 2012), and detection of RVFV-specific antibodies (Niklasson et al. 1984, Swanepoel et al. 1986, Meegan et al. 1987, Paweska et al. 2003).

The use of virus isolation is not user-friendly, takes an extended period of time, and requires suitable biosafety protocols or measures; antigen or nucleic acid detection in the serum or blood of animals or humans only works in cases of host viremia, which in the case of RVFV infection is a narrow viremic window, lasting on average about 3 days in serum (Morrill et al. 1990, Nfon et al. 2012, Smith et al. 2012) or longer in whole blood (Grolla et al. 2012). The classical methods for detection of antibodies to RVFV include various forms of virus neutralization and hemagglutination-inhibition tests. Disadvantages of these techniques include health risk to laboratory personnel (Smithburn 1949), as well as restrictions to high biocontainment laboratories for their use outside RVF endemic areas (Barnard 1979).

TABLE 2. REACTIVITY OF BACULOVIRUS-EXPRESSED RIFT VALLEY FEVER VIRUS PROTEINS WITH SHEEP ANTISERA DETERMINED BY WESTERN BLOT

Recombinant proteins	MP12 vaccinated sheep sera: days pv			Wild type (ZH501) challenged sheep sera: day 28 pi (nsp/total sample tested)
	Day 3	Day 10 (nsp/total sample tested)	Day 28	
Gne	1/2	4/4	10/10	4/4
Gc	0/2	1/4	9/10	4/4
N	2/2	4/4	10/10	4/4
NSs	0/2	4/4	9/10	4/4
NSm	0/2	0/4	0/10	0/4

pv, postvaccination; pi, postinfection; nsp, number of samples tested positive; Gne, ectodomain of Gn glycoprotein; Gc, Gc glycoprotein; N, nucleoprotein; NSs, nonstructural protein S; NSm, nonstructural protein m.

On the other hand, application of ELISA to detect IgG antibody to RVFV relied largely on the use of inactivated whole virus lysate, which is also associated with potential health risks (Swanepoel et al. 1986a, Swanepoel et al. 1986b, Meegan et al. 1987, Paweska et al. 1995). The nucleocapsid (N) protein is the most abundant and highly immunogenic component of the RVF virion and has been used for development of diagnostic assays for detection of RVFV-specific antibodies in

human and animal sera (Fafetine et al. 2007, Jansen van Vuren et al. 2007, Paweska et al. 2007, Paweska et al. 2008). Although the N protein is shown to be highly conserved among members of the Bunyaviridae family (Schwarz, et al. 1996, Magurano and Nicoletti 1999), a previous indirect ELISA based on the recombinant protein did not show cross-reactivity with other African phleboviruses that could hamper the reliability of using this protein in assays for serodiagnosis of RVFV

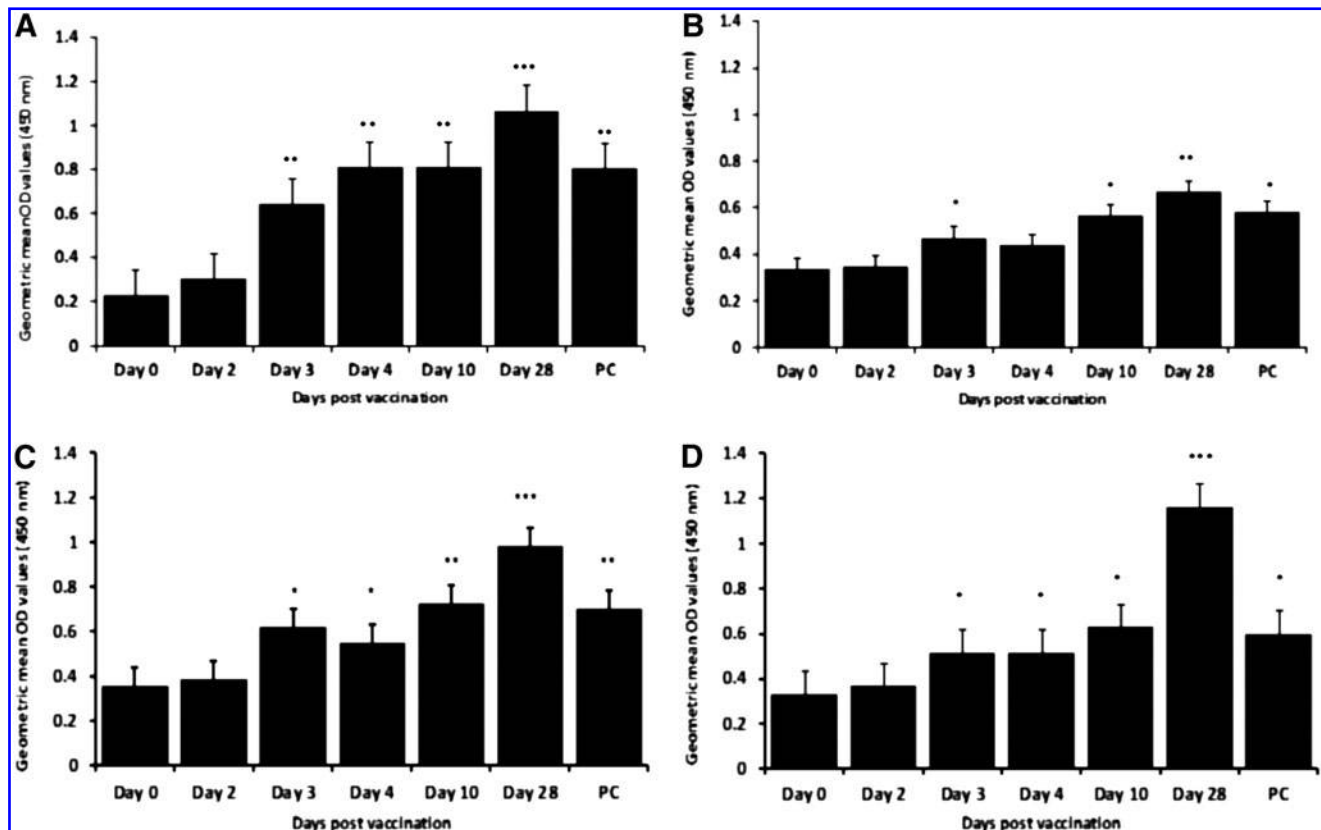


FIG. 5. Reactivity of recombinant RVFV proteins, nucleoprotein, N (A), nonstructural protein, NSs (B), glycoprotein, Gne (C), glycoprotein, Gc (D), and nonstructural protein NSm (E) with antisera from MP12 vaccinated sheep. Day 0, pre-vaccination sera; PC, positive control sera derived from sheep challenged with RVFV wild-type, ZH501; P1–P6, day-28 sera from sheep vaccinated with MP12 RVFV strain (Laramie, WY); P7–P10, day-28 sera from sheep infected with the wild-type virus (ZH501). Asterisks (*) denote level of statistical significance and show that differences in optical density (OD) values of sera tested for each of the time-points was significantly different ($p < 0.05$) from day 0 (prevaccination) sera. Cutoff OD value for each enzyme-linked immunosorbent assay (ELISA) was determined by addition of 2 standard deviations to the mean OD value of serum obtained from prevaccinated/noninfected sheep. N, 0.320; NSs, 0.358; Gn, 0.395; Gc, 0.387; NSm, 0.208.

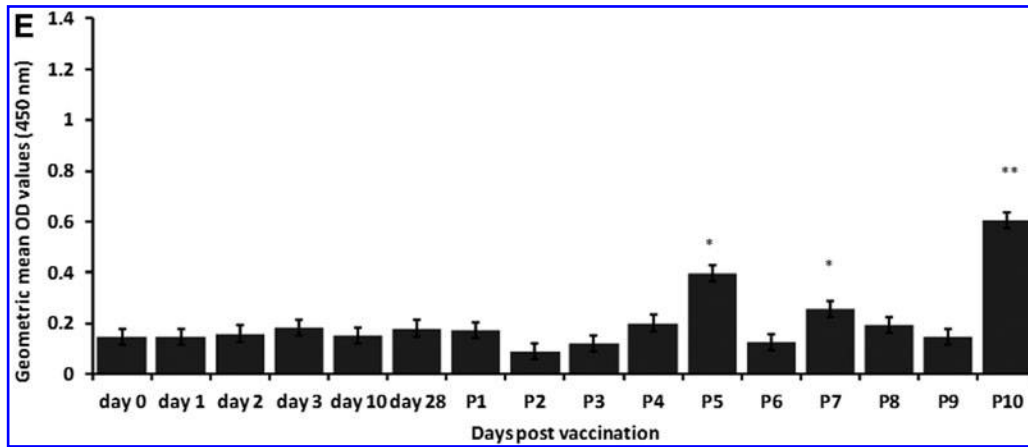


FIG. 5. (Continued).

infection (Paweska et al. 2007). However, the N protein did show serological cross-reactivity with an unidentified agent among some sera from US and Canadian sheep (Wilson et al., personal communication).

Recombinant proteins provide a safe platform for development of improved diagnostics and, in some cases, for subunit vaccines. In this study, we have expressed a panel of RVFV proteins (N, NSs, NSm, Gne, and Gc) using the baculovirus expression system and evaluated their potential as diagnostic targets. One of the advantages of using the baculovirus expression system is the ability to scale up expression; it also allows posttranslational protein modification, which in the case of glycoproteins, enhances antigenicity and immunogenicity. Sequence modifications made to Gn and Gc resulted in enhanced expression, and analysis of Gne expression by immunofluorescence showed that the protein was present within the cell, manifested by detection of green fluorescence around the cell nucleus, in the periphery of the plasma membrane of infected cells (Fig. 2).

Indeed, other studies reported specific targeting of recombinant RVFV glycoproteins to the plasma membrane of insect cells using the baculovirus expression system (Takehara et al. 1990, Liu et al. 2008) or localization of the proteins on the cell surface when expressed using alphavirus replicons (Filone et al. 2006). To ensure that Gne and Gc are translocated into the ER and glycosylated, we added a signal peptide sequence carrying a signal peptidase cleavage site upstream of the 5' end of the coding sequence. This ensured correct protein trafficking through the lumen of the ER, where glycosylation enzymes and signal peptidase are located (Gerrard and Nichol 2002, Gerrard and Nichol 2007). Biochemical characterization of glycosylation of Gne and Gc showed that the glycoproteins were *N*-glycosylated. Treatment with tunicamycin, a potent inhibitor of bacterial and eukaryote *N*-acetylglucosamine transferases, resulted in inhibition of glycosylation as detected by electrophoretic migration of the proteins (Fig. 3A, B), strongly indicating that Gne and Gc were subjected to posttranslational *N*-glycosylation.

Analysis of antibody reactivity of the recombinant proteins with RVFV antisera by western blot and indirect ELISA showed the proteins were immunoreactive, a strong indication of correct structural conformation. In western blot analyses (Fig. 4) and ELISA (Fig. 5) using MP12 vaccinated and

wild-type-challenged sera, the RVFV nucleocapsid protein was the most reactive. As the most abundant and highly immunogenic structural component of RVFV virion, the N protein has been considered the best choice for the development of immunoreagents for antigen detection assays (Pepin et al. 2010).

The early immune response and strong antibody reactivity against N protein with MP12-vaccinated sera might suggest a possible role of N protein in protective immunity to RVFV infection. Indeed, vaccination with N antigen has been shown to protect mice from wild-type RVFV challenge (Wallace et al. 2006, Liu et al. 2008, Lorenzo et al. 2008, Lagerqvist et al. 2009, Jansen van Vuren et al. 2010, Lorenzo et al. 2010). Remarkably, the glycoprotein Gne also showed an early-response reactivity (day 3 pv) in western blot and ELISA with MP12-vaccinated sera and was detected by day-28 pi sera from all ZH501-infected sheep. This would qualify Gne, in addition to the N-protein, as a good choice as a diagnostic antigen. Moreover, the early and specific antibody response to Gne found with MP12-vaccinated sheep makes this protein a good candidate for vaccine development. Importantly, the RVFV glycoproteins Gne and Gc have been shown to contain epitopes that induce production of neutralizing antibodies, a correlate of protective immunity against RVFV infection (Niklasson et al. 1985, Morrill et al. 1987, Hubbard et al. 1991, Morrill et al. 1997).

Therefore, future studies should aim at establishing a possible correlation between Gne reactivity and the presence of neutralizing antibodies. Also, baculovirus-expressed recombinant Gne alone or in combination with recombinant N could improve the sensitivity and specificity of herd screening for RVFV infections in areas where cross-reactivity with other bunyaviruses/phleboviruses may be an issue. Recombinant Gc also showed strong reactivity with sheep sera, but seemed to be less sensitive when compared with Gne (Fig. 5). However, the induction of a strong antibody response to Gc appears later in MP12-vaccinated animals compared to N and Gn. For improved RVFV detection, molecular tools for detection of RVFV nucleic acids should be used in parallel with antibody detection to identify early-infected ruminants.

The recombinant NSs protein showed early-onset reactivity with MP12 sera, but had an overall lower reactivity than the

N protein. This may be attributed to the typically low antibody titers raised against the NSs protein after RVFV infections (Pepin et al. 2010). The RVFV NSs protein is responsible for general suppression of the host transcriptional machinery, including type I interferons (Le May et al. 2004, Le May et al. 2008). In contrast to NSs, the other nonstructural protein, NSm, showed poor reactivity with sera from MP12 vaccinated and wild-type-challenged animals in western blot. Further assessment of its reactivity in an ELISA test showed little or no reactivity with MP12-vaccinated sera, demonstrated by extremely low OD values (Fig. 5E). Sera from 3 sheep (2 wild type-infected, P7 and P10, and 1 MP12-vaccinated, P5), at day 28 pi showed relatively high reactivity when compared to day-0 sera (Fig. 5E). Interestingly, in a virulent RVFV challenge experiment using ELISA, Bird et al. (2011) reported seropositivity for NSs antibodies in all 3 control sheep, whereas only 2 of the 3 animals were seropositive for NSm; this was the first time immunoreactivity against NSm had been detected in a natural host species (Bird et al. 2011). This might indicate that NSm is less immunogenic when compared to NSs or is expressed at lower levels resulting in a lower antibody response in mammalian hosts.

The development of a diagnostic tool that differentiates infected from vaccinated animals (DIVA) is very much needed in regions nonendemic for RVFV. The apparent poor immunogenicity of RVFV NSm in ruminant hosts reported in this paper and also shown previously (Bird et al. 2011) makes the use of NSm as a target for developing a DIVA diagnostic test questionable. In this study, NSs-specific antibodies were consistently detected in RVFV-infected/vaccinated sheep, which suggests that the NSs protein may serve as a candidate for the development of a DIVA diagnostic assay. Previous investigators have demonstrated variability of detection of antibodies to NSs (Fernandez et al. 2012). Therefore, further optimization and standardization of the use of this protein in diagnostic assays may be required for this antigen to be a reliable DIVA marker.

Research efforts aimed at the development and validation of a new generation of safe and accurate diagnostic immunoreagents and assays based on RVFV recombinant antigens are critical (Pepin et al. 2010). This is the first study to carry out expression of several RVFV proteins and assess their reactivity against a panel of immune sera from natural hosts. Analysis of antibody reactivity revealed that the proteins were expressed in the correct conformation in a baculovirus expression system. Addition of a fusion signal peptide sequence to the structural glycoproteins Gne and Gc ensured processing of the proteins through the cellular glycosylation pathway. We have shown that baculovirus-expressed N, Gne, Gc, and NSs could be used as potential serodiagnostic targets for monitoring host humoral immune response to infection and/or vaccination. The immunoreactivity toward NSm was rather weak making this RVFV antigen a less desirable diagnostic target. The general immunoreactivity profile suggests that N and Gne would represent desirable targets for development of highly sensitive serodiagnostic assays. In addition, future vaccine formulations will require DIVA compatibility with companion diagnostic DIVA tests. We have shown that baculovirus-expressed N and NSs assays, depending on the vaccine composition, can be used for such companion assays.

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Author Disclosure Statement

No competing financial interests exist.

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Address correspondence to:

Juergen A. Richt

Department of Diagnostic Medicine/Pathobiology

College of Veterinary Medicine

Kansas State University

1800 Denison Avenue

Mosier Hall K-224

Manhattan, KS 66506

E-mail: jricht@vet.k-state.edu