

LONDON  
SCHOOL of  
HYGIENE  
& TROPICAL  
MEDICINE



LSHTM Research Online

Kollmann, Tobias R; Kampmann, Beate; Mazmanian, Sarkis K; Marchant, Arnaud; Levy, Ofer; (2017) Protecting the Newborn and Young Infant from Infectious Diseases: Lessons from Immune Ontogeny. IMMUNITY, 46 (3). pp. 350-363. ISSN 1074-7613 DOI: <https://doi.org/10.1016/j.immuni.2017.03.009>

Downloaded from: <http://researchonline.lshtm.ac.uk/4650734/>

DOI: <https://doi.org/10.1016/j.immuni.2017.03.009>

**Usage Guidelines:**

Please refer to usage guidelines at <https://researchonline.lshtm.ac.uk/policies.html> or alternatively contact [researchonline@lshtm.ac.uk](mailto:researchonline@lshtm.ac.uk).

Available under license: <http://creativecommons.org/licenses/by-nc-nd/2.5/>

<https://researchonline.lshtm.ac.uk>

## **Protecting the Newborn & Young Infant from Infectious Diseases: Lessons from Immune Ontogeny**

Tobias R. Kollmann\*, Arnaud Marchant, Beate Kampmann, Sarkis K. Mazmanian, Ofer Levy

**Tobias R. Kollmann** MD, PhD (\* communicating author)

Division of Infectious Disease, Department of Pediatrics,  
University of British Columbia, Vancouver, Canada

BC Children's Hospital

938 W 28<sup>th</sup> Ave, A5-175, Vancouver, BC V5Z4H4

Tel: 604-875-2466; Fax: 604-875-2226

email: [tkollm@mac.com](mailto:tkollm@mac.com)

**Arnaud Marchant** MD, PhD

Institute for Medical Immunology

Université Libre de Bruxelles, Belgium

Tel: +32 2 650 9588, Fax: +32 2 650 9562

e-mail: [arnaud.marchant@ulb.ac.be](mailto:arnaud.marchant@ulb.ac.be)

**Beate Kampmann** MD, FRCPCH, DTM&H, PhD

Centre of International Child Health

Department of Paediatrics, Imperial College London, UK  
and MRC Unit The Gambia, West Africa

Tel: +44 207 594 2758, Fax: +44 207 594 3894

e-mail: [b.kampmann@imperial.ac.uk](mailto:b.kampmann@imperial.ac.uk) or [bkampmann@mrc.gm](mailto:bkampmann@mrc.gm)

**Sarkis K. Mazmanian**, PhD

Division of Biology and Biological Engineering

California Institute of Technology

1200 E California Bl. (Mailcode: 156-29), Pasadena CA 91125

Tel: 626-395-2356

Email: [sarkis@caltech.edu](mailto:sarkis@caltech.edu)

**Ofer Levy** MD, PhD

Department of Medicine, Division of Infectious Diseases

Director, *Precision Vaccines Program*

Boston Children's Hospital & Harvard Medical School

Boston, MA, USA

Tel: (617) 919-2904; Fax: (617) 730-0255

e-mail: [ofer.levy@childrens.harvard.edu](mailto:ofer.levy@childrens.harvard.edu)

**Abstract** Infections in the first year of life are common and often severe. The newborn host demonstrates both quantitative and qualitative differences to the adult in nearly all aspects of host immunity, which at least partially explain the increased susceptibility to infection. However, these differences are not solely the result of a state of immaturity, but rather reflect an evolutionary conserved, highly effective adaptation to the particular demands placed on the immune system around the time of birth. This plasticity observed in early life immunity also provides the potential to leverage protection of the young against infection and disease through a number of powerful interventions. This review focuses on the mechanisms that underlie the increased susceptibility as well as on immune-based approaches to broadly increase protection from infectious disease in early life.

**I. Introduction** | Infectious diseases are a predominant causes of childhood death (Bhutta and Black; Hostetter; Liu et al.). Neonatal infection in particular remains a common tragedy, with ~ 7 million cases and ~700,000 deaths per year, currently accounting for 40% of mortality in those under five years of age (Bhutta and Black; Blencowe et al.; Darmstadt et al.; Ghazal et al.; Hostetter; Lawn et al.; Liu et al.; Murray et al.; Seale et al.). Although neonatal morbidity and mortality due to infection also represents a significant hurdle in resource-rich countries (Heron; Oestergaard et al.; Wang et al.), newborns in resource-poor areas are most severely affected. (Agarwal; Bhutta and Black; Chan and Lake; Sepulveda and Murray) Given the magnitude of this problem, even modestly effective interventions would save millions of lives and billions of dollars. Broadly enhancing protection from infection and disease through immune modulation offers a feasible approach. However, optimal design and implementation of immunomodulatory interventions requires a deeper understanding of the developmental changes occurring in the neonatal immune system at the cellular and molecular levels. Herein we review how the immune system contributes to distinct host defense in early life and how specific developmental events increase the risk of particular infectious diseases. We place these insights into context of interventions that have the capacity to broadly enhance immune-mediated protection from a wide range of infectious diseases in the newborn period and early infancy.

## **II. The developing immune system has to satisfy opposing demands.**

Human immunity develops from a single-cell state in the early stages of embryonic life during which *cell autonomous immunity* (CAI) provides protection, to one of biochemical communication amongst collections of cells at which point *nutritional immunity* plays an increasing role. After several weeks of gestation, specialized cells within the developing fetus provide *barrier* and *innate immune protection*. Only after these stages does T- and B-cell-based *adaptive immunity* in the fetus become effective, as these require highly specialized tissues (Turvey and Broide). All of these components of immunity remain active throughout postnatal life, with substantial overlap and cross-regulation. Furthermore, each has been shaped throughout evolution to ensure species survival (Figure 1).

In addition to these genetically 'hard-wired' programs, the immune system also remains responsive to the rapidly changing demands of each individual's specific environment. This requires straddling of sometimes opposing demands such as preservation of semiallogeneic existence *in utero* while protecting against a multitude of potentially infectious microbes. The ability of the immune system in early life to handle both genetically-encoded and environmental-driven programming emphasizes its enormous dynamic capacity. As a result, well-placed immunomodulatory interventions can leverage this powerful early life plasticity and direct the trajectory of immune ontogeny to enhance resistance to infectious disease while maintaining immune homeostasis.

### *1. Cell Autonomous Immunity: the most ancient form of self-defence.*

Cell autonomous immunity (CAI) guards cells against intracellular infection via a system of compartmentalization, responsive to the threat of pathogens able to cross cell membranes (Randow et al.). Host cells express sensory machinery at these boundaries such as pattern recognition receptors (PRRs) that detect pathogen-associated molecular patterns (PAMPs)

and receptors that respond to danger-associated molecules (DAMPs) (Oh and Lee). For example, viral nucleic acids in the cytosol induce a type 1 IFN-dependent CAI response via RIG-I-like receptors (RLRs) that activate autophagy (Richetta and Faure; Wu and Chen). Newborns demonstrate age-dependent differences in regulation of CAI such as autophagy, likely because the mechanisms involved in CAI are also crucially important during normal development, i.e. beyond host defense against infection (Di Bartolomeo et al.). Not surprisingly then, activation of CAI following viral infection can have devastating consequences especially during early life. For example, Zika virus infection activates autophagy in infected human fetal neural stem cells via Akt-mTOR, which in turn inhibits normal neurogenesis during development (Liang et al.). On the other hand, Herpes Simplex virus (HSV) encephalitis is particularly severe in young children in part because of lower neonatal HSV-induced type 1 IFN production relative to the adult, which results in impaired autophagy and decreased viral control resulting in permanent central nervous system damage (Gantt and Muller; Wilcox et al.). The molecular mechanisms of lower IFN production in infected cells of the nervous system of newborns vs. adult have not been delineated. But newborn plasmacytoid DCs (pDCs), key to anti-viral defenses, demonstrate impaired production of Type 1 IFN as compared to adults due to reduced interaction of IFN-regulatory factor 3 (IRF3) with cAMP-responsive-element-binding protein (CREB)-binding protein (CBP) and their target DNA sequence (Kollmann et al.). The relevance of this pathway for host defense has been confirmed for humans, as genetic mutations in the PRR-IRF3-IFN signalling cascade predispose to severe HSV encephalitis in childhood (Abel et al.; Casanova et al.). Of note, environmental influences such as nutrient availability alter CAI, indicating that targeted interventions could optimize CAI in early life (Cuervo and Macian).

## *2. Nutritional Immunity provides broadly effective protection.*

Nutritional immunity provides host protection via regulation of metabolic pathways and availability of essential nutrients (Beisel). One example is iron (Fe), an element essential for the survival of all living organisms (Hood and Skaar). The human body is a rich reservoir of Fe and to prevent infection, it restricts access of microbes to Fe. The master-switch for free Fe levels is hepcidin, a peptide produced in the liver, where its expression is increased in response to inflammation or pathogen recognition. Specifically, interleukin (IL)-6 and other cytokines activate the signal transducer and activator of transcription 3 (Stat3) pathway thereby upregulating hepcidin expression and reducing free Fe (Recalcati et al.; Ruchala and Nemeth). This hepcidin pathway is likely an important component of newborn host defence, as human neonatal plasma Fe levels directly correlate with susceptibility to sepsis (Bullen et al.; Johnson and Wessling-Resnick; Nairz et al.; Oppenheimer; Wander et al.). Specifically, a dramatic physiological drop in serum Fe within hours of birth reduces the risk for neonatal sepsis (Bullen et al.; Hay et al.; Recalcati et al.; Sturgeon; Szabo et al.). Conversely, supplemental Fe given to Fe-replete infants increases the risk for sepsis and death (Oppenheimer; Oppenheimer; Sazawal et al.). Neonatal and even prenatal (i.e. maternal) nutritional immunity is thus a highly effective means of host protection that is tightly regulated yet can be readily manipulated, providing a distinct avenue to enhance early life protection from infection (Rochette et al.).

## *3. Physical barrier functions are enhanced by antimicrobial effector molecules.*

Protective barrier functions include physical and chemical components of placenta, skin and mucous membranes. The outermost layer of the skin acts as a physical barrier; however, its toughest layer, the *stratum corneum*, only fully develops during the first two weeks of life (Figure 2, *Skin*) (Marchant et al.). As some measure of counterbalance, the skin of full-term infants displays high production of antimicrobial proteins and peptides (APPs); APPs are in fact expressed in an age-dependent pattern by nearly all human tissues and cells exposed to microbes (Wiesner and Vilcinskis). Examples of APPs include defensins such as human  $\beta$ -defensins, bactericidal/permeability-increasing protein (BPI), whey acidic protein motif-containing proteins, secretory leukocyte protease inhibitor, elafin (antiproteinase 3; skin derived antileuko-proteinase), lactoferrin, and lysozyme. (King et al.; Wiesner and Vilcinskis) The skin in particular produces  $\beta$ -defensins and cathelicidins (Dorschner et al.). Of note, a waxy coating (*vernix caseosa*) produced by fetal sebaceous glands *in utero* covers the term newborn as a microbicidal shield for the first few days of life. The vernix contains multiple APPs such as lysozyme,  $\beta$ -defensins, ubiquitin and psoriasin, as well as antimicrobial free fatty acids (Tollin et al.). As the *vernix caseosa* is mainly formed during the last trimester of gestation, the preterm newborn is left relatively more exposed and vulnerable. (Marchant et al.). Clinical trials of prophylactic and therapeutic application of APPs to newborns and infants have shown promising results (Battersby et al.). For example, intravenous administration of the antibacterial and endotoxin-neutralizing recombinant N-terminal fragment of BPI (rBPI<sub>21</sub>) to a pediatric cohort with meningococcal sepsis, including newborns as young as 2 weeks of age, was associated with clinical benefit (Levin et al.). And oral administration of bovine lactoferrin, an 80kDa cationic multi-functional protein with iron-binding, immunomodulatory and direct membrane-perturbing microbicidal activity, to preterm, very low birthweight newborns (<1500 g) was associated with reduced incidence of late-onset sepsis, necrotizing enterocolitis, and fungal infection (Legrand; Manzoni et al.; Manzoni et al.; Manzoni et al.)

Similar to the skin, the host defense of the intestinal mucosa via expression of APPs is also developmentally regulated and equally dependent on colonization with beneficial microbes (Figure 2, *Mucous membranes*). (Hornef and Fulde) The small intestinal epithelium of neonatal mice expresses the cathelicidin cathelin-related antimicrobial peptide (CRAMP) that exerts antibacterial activity against commensal and pathogenic bacteria. Production of Paneth cell-derived APPs like cryptidins and cryptin-related sequence (CRS)-peptides on the other hand begins only after birth, due to the delayed appearance of small intestinal Paneth cells during the postnatal period. Intestinal epithelial CRAMP expression wanes after the postnatal period, reflecting a switch in the APP repertoire and production site from epithelial CRAMP expression to Paneth cell-secreted cryptidin and CRS peptides after weaning. The mucosa of the respiratory tract also produces a variety of APPs, such as BPI, lysozyme, lactoferrin, and defensins (Diamond et al.; Travis et al.). And as in the intestine, APPs in the respiratory tract are mainly produced in response to PRR stimulation following microbial encounters, a response that increases with gestational age and correlates with decreased susceptibility to infection with e.g. *Bordetella pertussis* (Elahi et al.; Starner et al.). Overall, the development of barrier function appears tightly linked to, and apparently driven by, microbial exposure. In this context, deliberate exposure to specific non-pathogenic microbes could provide a means of accelerating barrier-mediated host protection.

#### 4. Innate immunity provides immediate effector function and integrates diverse environmental signals.

Coordination of host immune responses is especially important for host protection in more complex, multicellular organisms as well as in the progressively complex fetus (Fig. 1). Via recently appreciated immunometabolic pathways, innate immunity coordinates CAI and nutritional immunity (O'Neill et al.). For example, innate immune activation following treatment of human monocytes with the Dectin-1 agonist  $\beta$ -glucan induces a shift to cellular aerobic glycolysis via an Akt/HIF1 $\alpha$ -mediated activation of the mammalian target of rapamycin (mTOR) pathway resulting in increased subsequent TNF responses to PRR agonists such as LPS or heat killed bacteria (Cheng et al.). This immunometabolic pathway directs the memory-like function of innate immunity, i.e. epigenetic changes that lead to long-lasting alteration of innate immune memory, also known as “trained immunity” (Cheng et al.; Levy and Netea; Netea et al.; O'Neill et al.). The activities of innate immunity are both rapid (preventing microbial proliferation/spread) and broad (enabling protection against multiple diverse pathogens) (Buchmann).

The innate immune system exerts its effector functions through soluble (e.g. complement and APPs) as well as cellular components (Figure 2). (Pettengill et al.) With respect to soluble factors, levels of most individual complement proteins are lower in neonates compared to adults, resulting in lower complement activity in early life. Similarly, most APPs (e.g. Lactoferrin, BPI, and cathelicidin anti-microbial peptide 18 (also called LL-37)) have lower constitutive plasma concentrations in early vs. adult life, especially in preterm and low birth weight newborns (Singh et al.; Strunk et al.). Early life APP deficiency impacts host defense, as lower serum levels of cathelicidin, for example, are associated with increased severity of acute bacterial respiratory infection in children aged 0–24 month. (Battersby et al.; Mansbach et al.)

Specialized innate immune cells include the myeloid lineages, namely granulocytes (e.g. neutrophils), monocytes, macrophages, and dendritic cells (DCs), as well as innate lymphocytes. While neutrophils are present in human fetal liver parenchyma by as early as 5 weeks gestation (De Kleer et al.), neutrophils in early life demonstrate quantitatively and qualitatively different responses under stress conditions as compared to adult neutrophils, including reduced chemotaxis, respiratory burst and formation of extracellular traps, scaffolds for APPs that serve to capture and kill extracellular bacteria. (Carr). Limitations in neonatal neutrophil function may in part reflect higher expression levels of inhibitory receptors (De Kleer et al.).

Monocytes appear in the fetal circulation as soon as self-renewing hematopoietic stem cells (HSPC) have seeded the fetal liver (De Kleer et al.). During early development, monocyte progenitors also colonize various organs and differentiate into tissue-resident macrophages that self-maintain throughout life (De Kleer et al.). Following PRR stimulation, human newborn monocytes/macrophages and DCs produce a cytokine profile that differs substantially from those of their adult counterparts (Figure 2, *Blood*) (De Kleer et al.; Kollmann et al.). Specifically, upon PRR stimulation *in vitro*, neonatal APCs produce less proinflammatory (IL-1 $\beta$ , TNF $\alpha$ ) and Th-1 promoting cytokines (IL-12p70, type 1 IFN), but

equal or greater amounts of Th-17 promoting cytokines (IL-23, IL-6) compared with adult cells. Robust neonatal production of IL-6 (*a*) induces a physiological hepatic acute phase response at birth, including induction of mannose binding lectin (MBL), C-reactive protein (CRP) and LPS-binding protein (LBP) that rise in the first week of life, possibly broadly enhancing resistance to infection and (*b*) contributes to healing of tissues injured during birth (Jones; Levy). Newborn monocytes and conventional DC (cDC) also produce more IL-10 compared to adults, likely reflecting the importance of anti-inflammatory responses in early life. Mechanisms that lead to this early life pattern of innate cytokine response include (*a*) high mononuclear cell levels of intracellular cyclic adenosine monophosphate (cAMP), a secondary messenger that suppresses Th1 but enhances Th2 and anti-inflammatory cytokine production (Levy et al.) and (*b*) altered DNA binding capacity of transcription factors such as IRF3 to the promoter regions of cytokine genes secondary to age-specific chromatin remodelling (Lissner et al.).

Innate lymphocytes develop early during human gestation (Figure 2, *Blood*). Fetal  $\gamma\delta$  T cells are dominated by cells expressing a canonical V $\gamma$ 9V $\delta$ 2 TCR and are programmed to express type 1 effector molecules (Dimova et al., 2015; Vermijlen and Prinz, 2014). Non-V $\gamma$ 9V $\delta$ 2 effector T cells are known to be induced in utero by congenital CMV infection (Vermijlen et al.). Innate lymphocytes expressing rearranged TCR, including iNKT cells and mucosal-associated invariant T (MAIT) cells, as well as non-TCR-expressing innate lymphoid cells (ILCs), also develop early during fetal life and are programmed to express effector functions, although possibly at lower levels than adult cells (Chan et al., 2013; Hong DK; Nakazawa et al., 1997).

Collectively, these observations indicate that many cellular and molecular components of innate immune responses develop early during fetal life but are regulated distinctly from their adult counterparts.

### *5. Adaptive Immunity – diverse yet targeted effectors.*

Development of adaptive immunity requires close cooperation between multiple elements of the innate immune system, as well as nutritional innate immunity via metabolic pathways that shape the function of effector and memory lymphocytes (Buck et al.; Iwasaki and Medzhitov).

Mature fetal  $\alpha\beta$  T lymphocytes can be detected from ~14 weeks of gestation onward, i.e. several months later than a number of innate lymphocyte subsets (Hong DK). The repertoire of fetal T cell receptors diversifies during the second and third trimesters of gestation (Rechavi et al.). In the fetus and the newborn, the majority of  $\alpha\beta$  T lymphocytes are recent thymic emigrants (RTEs) (Haines et al.). In comparison to mature naïve T lymphocytes, RTEs have a distinct functional program involving epigenetic modifications at key cytokine loci (Fink). Newborns often display limited Th1-type responses to some vaccines and pathogens, correlating with a lower capacity of CD4 T cells to produce IFN- $\gamma$  and of APCs to produce Th1-polarizing cytokines (Debock and Flamand; White et al.). However, this reduced Th1 capacity is not absolute as newborns and young infants develop adult-type Th1 responses to e.g. BCG or whole cell pertussis vaccines, and fetuses can develop



Th1 responses to CMV infection (Huygens et al.; Huygens et al.; Marchant et al.; Mascart et al.). This indicates that the quality and magnitude of signals present at the time of naïve CD4 T cell priming determine the development of immune responses in early life. Specifically, the profile of cytokines produced by newborn APCs, including IL-6 and IL-23 suggests a robust ability to mount Th17 and follicular helper T (TFH) cells at similar or even higher levels than adults (Debock and Flamand). Of note, fetal  $\alpha\beta$  T lymphocytes may already acquire a specific phenotype of memory cells *in utero*, programmed to effectively produce Th1-, Th2- or Th17-cytokines (Zhang et al., 2014). Furthermore, multiple pathogens, including CMV, HIV and *T. cruzi* can induce effector CD8 T lymphocytes in the fetus, clearly indicating that cell-mediated immune responses are not intrinsically deficient in early human life (Hermann et al.; Muenchhoff et al.). However, the anti-microbial properties of fetal and newborn effector T lymphocytes may be limited by a more rapid onset of functional exhaustion (Huygens et al.).

Mature B lymphocytes can be detected in the fetal liver from 8 weeks of gestation (Hong DK). Somatic hypermutation of peripheral and marginal zone (MZ) B cells develops and isotype switching begins *in utero* already and leads to fetal diversification of the B cell receptor repertoire (Hong DK; Rechavi et al.). The capacity of the newborn to develop antibody responses depends on the nature of the immune stimulus as reflected in the immunogenicity of standard childhood vaccines. For example, hepatitis B vaccine (HBV) immunization induces at least equivalent antibody responses in newborns and adults; in contrast, antibody response to oral polio, measles and rubella vaccines increase with age at immunization (Ota et al.; Siegrist and Aspinall). Importantly, irrespective of their primary response, neonatal immunization can induce potent memory B cell responses that promote immunogenicity of subsequent vaccine booster doses (Halsey and Galazka). The mechanisms underlying the early life maturation of effector B lymphocyte responses are currently unclear but could involve the upregulation of complement receptors, of the ecto-enzyme CD73, of T cell co-stimulatory molecules expressed by neonatal B lymphocytes, or the gradual age-dependent enhancement of interactions between infant B and TFH cells, all of which are likely influenced by maternal antibodies (Debock and Flamand; Pettengill and Levy; Siegrist and Aspinall).

#### *6. Microbial colonization provides key signals for immune development and protection.*

Within hours after birth the neonate is colonized by bacteria (Figure 2). (Arrieta et al.) While the composition of this microbiota rapidly evolves during the first 2-3 years of life, microbial communities are largely unique to each individual, as both host genetic and environmental factors influence the composition of the intestinal microbiota via cross-talk between microbes and their hosts (Arrieta et al.; Chu and Mazmanian; Dorrestein et al.; Landwehr-Kenzel and Henneke). Bacterial colonization is in fact essential for optimal host immune development, illustrated by the finding that germ-free mice are at increased risk for infectious as well as autoimmune diseases (Arrieta et al.; Chu and Mazmanian; Khosravi et al.; Renz et al.). Germ-free mice also display reduced hematopoiesis of macrophages from both the bone marrow and yolk sac early in life, leading to impaired clearance of systemic *Listeria monocytogenes* infections<sup>56</sup>. And in mice, LPS derived from Gram-negative bacteria induces microRNA-146a, that down-regulates IL-1 receptor associated kinase 1 (IRAK1)

thereby changing TLR4 signaling towards a state of tolerance following bacterial colonization (Lotz et al.). The human commensal *Bacteroides fragilis* produces polysaccharide A (PSA) that induces TLR2-mediated development of regulatory T ( $T_{reg}$ ) responses (Chu and Mazmanian), an activity also found in other microbes such as the probiotic *Bifidobacterium breve*. Beyond their impact on immune tolerance, commensal microbiota also prepare the host to rapidly mount immune responses upon pathogen encounter, as germ-free or antibiotic-treated mice demonstrate impaired clearance of systemic bacterial infection. As mentioned above, interactions of the infant gut with the microbiota are likely important for maturation of mucosal APP expression (Fig. 2B), whose dysregulation has been associated with necrotizing enterocolitis (Maynard et al.; Salzman et al.). This physiological process is readily altered by administration of antibiotics in early life. For example, treatment of mice with broad-spectrum antibiotics to deplete the microbiota reduces circulating peptidoglycan and leads to impaired neutrophil-mediated killing of *Streptococcus pneumoniae* and *Staphylococcus aureus*. (Chu and Mazmanian) This likely relates to the ability of commensal microbes to promote maintenance of neutrophils and other myeloid cells (Arrieta et al.; Renz et al.). Furthermore, antibiotic treatment reduces: (a) expression of IFN-responsive genes in peripheral blood monocytes and DCs of young mice due to epigenetic changes including reduced H<sub>3</sub>K<sub>4</sub>me<sub>3</sub> deposits at specific promoter regions (De Kleer et al.); (b) adaptive immune responses to intranasal infection with influenza virus, reducing virus-specific antibody titers and frequency of virus-specific CD4 and CD8 T cells (Chu and Mazmanian); (c) flagellin production by commensal flora important for driving TLR5-mediated lymph node plasma cell differentiation to enhance antibody responses to inactivated influenza and polio vaccines (Oh et al.). These data also highlight the still largely underappreciated role for microbiota in influencing immunity to immunization (Huda et al.).

Importantly, there appears to be an age-restricted 'window of opportunity', as colonization of germ-free or antibiotic-treated mice early in life with commensal microbes promotes normal immune function while colonization of adult germ-free or antibiotic-treated mice fails to do so (Chu and Mazmanian; Khosravi et al.). For example, invariant NKT (iNKT) cells appear to be activated and home to the gut and promote inflammation in germ-free mice; colonization with a complex microbiota in early life restores homeostasis, but not when gut bacteria are introduced later in adulthood. (Olszak et al.) A critical early life period of beneficial microbial-host immune interaction has also been identified in studying the impact of feeding- or birth-mode (Ardeshtir et al.; Cox et al.). For example, the decreased diversity of the dominant *Bacteroidetes* phylum in infants born by Caesarean section is associated with lower plasma concentrations of CXCL10 and CXCL11, two IFN-dependent chemokines important for leukocyte migration to sites of infection (De Kleer et al.). These effects likely relate to the impact of the maternal microbiome on newborn immune function; interestingly, this impact is dependent on maternal antibody transfer, suggesting complex cross-regulatory interactions between host and microbiota even across generations (Gomez de Agüero et al.). Finally, although controversial, a fetal microbiome has been described, suggesting this host-microbe interaction may already be prior to birth (Aagaard et al.). Overall, these observations suggest a critical perinatal (and possibly even prenatal) window wherein the interaction of the host with the microbiome drives optimal immune development and exposure to specific microbes, including probiotics, may

enhance immune mediated homeostasis (Arrieta et al.; Dominguez-Bello and Blaser).

### 7. Regulation of immunity in early life.

The transition around birth from a largely shielded environment *in utero* to postnatal life as an '*animal in a microbial world*' (McFall-Ngai et al.) represents the most dramatic life event for our mammalian immune system. Specifically, the semi-allogeneic state of the mother/fetus requires suppression of rejection, in part accomplished via a 'default' tolerogenic immune response involving adaptive T<sub>reg</sub> cells as well as pronounced PRR-mediated IL-10 production by neonatal APCs (Kollmann et al.). The normal birth process on the other hand is the result of targeted inflammation necessary to separate the maternal-fetal layers, requiring immune suppressive counter-regulation to prevent systemic inflammation (Gomez-Lopez et al.). In addition, hypoxia suffered during labour can cause tissue damage that in turn enhances inflammation causing further damage (Sharma et al.). Such potential for perinatal inflammatory immunopathology is counterbalanced by a strong immune bias towards resolution of inflammation and healing, i.e. the well-described Th2 dominated response in early life (Iwasaki and Medzhitov). The postnatal period is characterized by rapidly changing environmental and microbial exposures that relentlessly stimulate immune development, requiring immune regulation to preserve homeostasis (MacGillivray and Kollmann);

Managing such rapidly shifting, diverse functional demands is in part achieved in part through compartmentalization where immune responses appear strictly confined to specific tissue compartments (Iwasaki and Medzhitov; Kollmann et al.; Thome et al.). Such 'spatial' regulation is complemented through regulatory mechanisms depending on 'time'. Specifically via the impact of layered immune maturation, whereby fetal immune cells, functionally distinct from their adult counterparts, arise from discrete HSPC at different stages of development. (Krow-Lucal and McCune) Such layered immunity appears to provide for the abundance of active immune suppressive cells and functions in fetal and early postnatal life, including T<sub>reg</sub> cells, myeloid-derived suppressor cells (MDSCs), and erythroid (nucleated RBC) suppressor cells (Elahi; Gantt et al.; Gervassi et al.; Pandiyan et al.; Power Coombs et al.). For example, natural Tregs develop in the thymus in parallel with naïve T cells during fetal life (Hong DK). Fetal naïve CD4 T cells preferentially differentiate to Treg cells in peripheral tissues in response to non-inherited maternal HLA antigens and could thereby play a central role in fetal tolerance to maternal cells (Boer et al.; Mold et al.; Mold et al.). Furthermore, Treg cells continue to comprise a larger proportion of CD4 T cells in peripheral tissues in young children as compared to adults, supporting their role in the maintenance of immune homeostasis in early life. (Thome et al.) MDSCs are a heterogeneous population of granulocytic or monocytic cells that suppress innate as well as adaptive immune function (Gantt et al.; Gervassi et al.). MDSCs express suppressive factors such as arginase-1, reactive oxygen species, and inducible nitric oxide synthase, which inhibit T cell proliferation and cytotoxicity, induce the expansion of Treg, and block NK cell activation. Granulocytic MDSCs are present in large numbers in pregnant women and in cord blood, yet wane rapidly during infancy. Cord blood MDSCs suppress CD4 and CD8 T cell and NK cell responses, suggesting a significant role in immune homeostasis in early human life. Similarly, physiologically abundant CD71<sup>+</sup> erythroid cells in neonatal mice and

human newborns exhibit immunosuppressive properties (Elahi). Specifically, neonatal CD71<sup>+</sup> cells express the immunosuppressive enzyme arginase-2 and ablation of CD71<sup>+</sup> cells in neonatal mice increases resistance to the perinatal pathogens *L. monocytogenes* and *E. coli*, yet not to polymicrobial sepsis (Elahi et al.; Wynn et al.). Lastly, compared to adults, neonatal cord blood plasma has higher amounts of adenosine-generating enzymes (soluble CD73 and alkaline phosphatase) and lower levels of adenosine deaminase (ADA), the enzyme that metabolizes and inactivates adenosine, resulting in high plasma concentrations of adenosine, an endogenous purine metabolite that inhibits TLR-mediated Th1-polarizing cytokine induction (Pettengill et al.; Power Coombs et al.), and also reduces neutrophil activation (Hasko and Cronstein).

Behind the complexity of regulating immune ontogeny stands a surprisingly simple basic principle: balancing cost to the species vs. the individual of an immune response directed at the microbial world while maintaining homeostasis (Iwasaki and Medzhitov). All immune effector responses can be viewed on a spectrum defined by the costs, including potential immunopathology, associated with their maximal deployment. Evolution has presumably selected for immune responses that minimize cost while at the same time providing sufficient protection for survival of the human species. In this light, the development of multiple highly effective immune regulatory strategies in parallel with immune effector functions suggests that our immune system in early life is not simply 'immature' (meaning 'not ripe', 'not perfect'), but shaped to satisfy the complex demands of early life ready to adapt to new challenges.

## **II. Age-dependent susceptibility to infection in early life.**

The intensity and rapid kinetics of the opposing immunologic demands of host protection in early life versus immunoregulation to avoid tissue damage suggests that alterations in a given individual along these physiological trajectories must be highly regulated. In this context, the clinically observed increased risk for infection in early life can be viewed as an imbalance of the phylogenetically selected beneficial survival programs vs. specific environmental demands exerted on the individual during ontogeny. This concept appears useful in correlating the particular aspects of immune regulation during ontogeny with the most pertinent infections during the same age period. For example, the decreased CAI and barrier function of the skin and mucous membranes of pre-term infants predicts the known risk for invasion with skin- and mucosa-colonizing microbes (Marchant et al.). Furthermore, the relative deficiencies in the complement system and APPs along with reduced phagocyte migration indicate a potential increased vulnerability of the newborn infant, especially those born preterm, to systemic spread and infection with extracellular microbes. Indeed, newborns display heightened susceptibility to pyogenic infection with Gram-positive extracellular bacteria such as *Staphylococcus spp.* (Power Coombs et al.) and *Streptococcus agalactiae* (Group B Streptococcus) (Landwehr-Kenzel and Henneke), Gram-negative infections such as *E. coli*, as well as certain fungal infections (Hsieh et al.; Rao and Ali; Vergnano et al.). The distinct aspects of early life innate and adaptive immune ontogeny summarized above would further predict an increase of infection in early life with pathogens controlled by Th1 type immune responses. This is also the case, as an increased risk for severe infection with intracellular pathogens requiring Th1 protective responses for effective host defense, including bacteria (e.g., *L. monocytogenes*, *Salmonella spp.*),

mycobacteria and viruses (egg. HSV, HIV) is observed in newborns and young infants (Chirico et al.; Garcia-Vidal et al.; Sherrid and Kollmann; Speer et al.; Vanden Driessche et al.; Weiner and Kaufmann). Indeed, viral infections including respiratory syncytial virus and influenza virus are often more severe and/or prolonged in early life as compared to adult life (Bertoletti and Hong; Clark and Lynch; Gantt and Muller; Huygens et al.; Muenchhoff et al.). However, given the current limited insights into cause-effect relationships, linking susceptibility of particular infections in early life to specific immune parameters during immune ontogeny remains an area that is still incompletely understood.

### **III) Opportunities for Intervention**

As summarized above, the early life immune system is not in a fixed state of 'immaturity', but rather rapidly adapts to environmental cues. It is thus possible, and indeed likely, that protection from infection can be enhanced by providing broadly active immune modulatory stimuli during early life. We already mentioned some specific examples in the respective sections on immune ontogeny above; we here focus on a range of interventions that can provide broad protection against a wide range of pathogens.

The traditional pathogen-centric approach has led to the successful development of childhood vaccines that prevent ~ 2.5 million deaths each year worldwide (Barnighausen et al.; Clemens et al., 2010; Levine, 2011; UNICEF). However, vaccine-mediated prevention of infections occurring at birth or soon after birth is limited by reduced or slower immune responses to a number of vaccines administered in early life. In the future, it may be possible to enhance responses to early life vaccines by inclusion of novel adjuvants that demonstrate age-specific immune-enhancing activity (Oh et al.; van Haren et al.). In the meantime, maternal immunization offers an attractive complement to infant immunization as it allows the transfer of high quality pathogen-specific IgG across the placenta during the second half of pregnancy, effectively protecting the newborn and young infant. Maternal immunization has already proven effective in reducing neonatal and infant disease due to tetanus, pertussis and influenza, and may become a central component of the control of group B streptococcus (GBS) and RSV infections in young infants (Amirthalingam et al.; Beigi et al.; Dauby et al.; Lindsey et al.; Niewiesk; Vidarsson et al., 2014).

Further progress on this pathogen-specific focused avenue has however been limited by our current lack of understanding of the mechanistic requirements for protective immunity (Iwasaki and Medzhitov). While immunogenicity is the key parameter measured, in the form of B cell-derived antibody or T lymphocyte cell-mediated immunity, which are often used as a surrogate for protective immunity, immunogenicity only indicates the ability to induce an immune response while protective immunity denotes the ability to eliminate the pathogen without hurting the host (Iwasaki and Medzhitov). Although counterintuitive to many, increased protection from infectious diseases in early life could in fact require a decrease rather than increase and a more balanced immune response, resulting in decreased immunopathology and thus cost. The potential benefit in certain contexts of attenuated immune responses has already been shown for viral infections such as hepatitis B virus (HBV) or human immune deficiency virus (HIV), where the immune regulatory mechanisms dominant during early life prevent immune-mediated harm to the host (Bertoletti and Hong; Huygens et al.; Muenchhoff et al.).

It is increasingly appreciated that young infants can be protected from infectious pathogens through non pathogen-specific mechanisms. For example, vaccines have effects beyond inducing classic antigen-specific T and B cell-mediated adaptive immune responses targeting a specific pathogen (Aaby et al.). Protective heterologous (“non-specific”) effects of vaccines have been demonstrated for live attenuated vaccines such as Bacillus Calmette–Guérin (BCG), oral polio and measles vaccines, all of which reduce morbidity and mortality far beyond that attributable to prevention of the target disease. (Aaby et al.; de Castro et al.; Goodridge et al.; Lund et al.; Sorup et al.). Of note, BCG activates autophagy as part of CAI (Buffen et al.), iron sequestering as part of nutritional immunity (Kochan et al.), epigenetic changes of innate cells as part of trained innate immunity (Netea and van Crevel), and promotes T- and B cell responses to unrelated antigens (Kleinnijenhuis et al.; Libraty et al.). Such wide-ranging immune-modulatory activity of BCG may contribute to its broadly protective, heterologous effects.

And as mentioned above, multiple arms of the immune system are enhanced following microbial colonization, from CAI to barrier and innate immune function (Arrieta et al.; Chu and Mazmanian; Khosravi et al.; Renz et al.). For example, in neonatal mice, microbiota regulate neutrophil homeostasis and host resistance to sepsis in a TLR4- and myeloid differentiation factor 88 (MyD88)-dependent pathway via IL-17 production in Group 3 innate lymphoid cells (ILCs) (Deshmukh et al.). And in humans, certain enteral probiotics reduce not only the risk of necrotising enterocolitis in prematurely born infants but also infection-related mortality (Alfaleh and Anabrees, 2014; Denkel L.A.; Oncel et al., 2014; Panigrahi; Roy et al., 2014; Samanta et al., 2009; Strunk et al.). Similarly, enhancing APP-mediated host innate immune defenses directly via supplementation with oral bovine lactoferrin reduced late onset sepsis in human preterm newborns (Pammi and Abrams). Taken together these data suggest that, in addition to interventions targeting antigen-specific immunity, non-pathogen-specific CAI, nutritional and/or leukocyte-based innate immune functions can be harnessed to prevent infectious diseases in early life.

## **V. CONCLUSION:**

The imbalance of environmental cues and demands in the context of the genetic constraints of the particular host are what lead to the readily observed increase in clinical disease following infection in early life. However, the plasticity of the early life immune system makes it amenable to therapeutic interventions to combat infection. It is thus possible, and indeed likely that protection from infection can safely yet effectively be enhanced by providing broadly active immune modulatory stimuli during ontogeny. Specifically, the human immune system develops early during fetal life and appears to be functionally programmed to promote tolerance to the maternal environment *in utero* and to commensal microbes, while tightly regulating perinatal inflammatory reactions. The efficacy of microbial stimuli- whether acquired within a commensal microbiome or as live attenuated vaccines- in safely enhancing host protection from infection or disease suggests these interventions do not bypass immune regulatory mechanisms but instead enhance them in the context of healthy immune homeostasis in early life. Boosting of CAI, nutritional and other innate defense mechanisms may reduce the risk for infectious disease for a wide range of different pathogens that threaten the newborn and young infant. Such interventions may benefit the young host by promoting immune regulation and

homeostasis rather than simply increasing effector functions. Elucidating the underlying molecular mechanisms is a key area for future research that will shed new light into immune ontogeny and inform development of age-specific immunomodulatory interventions. Together with the established pathogen-targeting approaches such as antigen-specific immunization of mother, newborn and infant, additional efforts aimed at enhancing heterologous host resistance through optimized development and delivery of vaccines or probiotics will provide additional effective, practical and affordable approaches for protecting newborns and young infants from the heavy burden of infectious diseases.

## Figure legends

### **Figure 1. Key elements of immunity share a common phylogeny and ontogeny.**

Parallels are drawn between the phylogeny and ontogeny of immunity. Host protection from infection has developed in evolution (phylogeny) and develops in a given human across an individual's lifespan (ontogeny) from: (a) a unicellular state wherein cell-autonomous immunity (CAI) is key to survival as noted in single cell organism such as an amoeba and early after conception; (b) multicellular organisms such as fungi that expresses biochemical communication via nutritional immunity (NI) amongst collections of cells as noted in ontogeny during the morula blastocyst stage; (c) multicellular animals of lower complexity such as *C. elegans* that express leukocyte-based innate immunity as is noted in the first month embryo; and (d) animals of higher complexity such as mice that express classic adaptive immunity with specialized cells (e.g., T and B cells) and tissues (e.g., lymph nodes) as is noted in the human fetus of 2 months or greater gestational age. All of these aspects of immunity contribute to perinatal host defense.

### **Fig. 2. Ontogeny of fetal, neonatal & infant host defense.**

Host-protective barrier functions include physical, chemical and functional components of the epithelial of skin and mucous membranes. These have to be understood in the context of age-specific developmental challenges as outlined near the top of the figure. (A) *Skin*: While physical and chemical barriers are reduced early in life, especially in the preterm, the *vernix caseosa* and skin epithelia of full-term newborns robustly expresses APPs. (B) *Mucous membranes*: In parallel with and induced by an increasingly complex microbiota, the newborn intestinal mucosal epithelium rapidly changes structurally with increase in crypts, and crypt-based Paneth cells, as well as functionally with increasing APP expression (Maynard et al.). (C) *Blood*: The composition of neonatal blood is distinct, with relatively low concentrations of complement components and APPs (Dorschner et al.; Hackam et al.; Hong DK; Hornef and Fulde; Marchini et al.; Tourneur and Chassin; Underwood et al.; Visscher and Narendran; Yoshio et al.) and high concentrations of the immunosuppressive purine metabolite adenosine. Plasma also contains maternal antibodies transferred beginning mid-gestation, and supplemented by postnatal factors derived from breastmilk (Hanson and Korotkova). Innate immunity is detectable from the end of the first month of gestation, with changes driven largely by the increasing exposure to environmental microbes (De Kleer et al.; Dowling and Levy; Kollmann et al.; Pettengill et al.; Vermijlen and Prinz). Neonatal APCs such as blood monocytes express PRRs (e.g., TLRs) with distinct functional responses including limited Th1-polarizing cytokine production to most stimuli. Age-dependent differences in activity of interferon response factor (IRF) transcription factors as well as epigenetic changes contribute to this cytokine ontogeny (Buffen et al.; Danis et al.; Kleinnijenhuis et al.). Adaptive immunity develops from 4 weeks of gestation onwards, with changes driven by an evolving chimerism reflecting fetal (liver-derived, shaded cells) Treg-rich lymphocytes and more adult-like (bone marrow-derived, not shaded cells) lymphocytes (Krow-Lucal and McCune) with distinct epigenetically encoded functional programs (Hanson and Korotkova; White et al.)



**Fig. 3. Interventions that broadly enhance host defense against infectious disease in early life.** There are key windows of opportunity during prenatal life and early postnatal life to enhance host resistance to specific infections via homologous – i.e., pathogen and thus classic antigen-specific responses (top panels) as well as broadly protective heterologous (“non-specific”) responses (bottom panels). (A) *Maternal immunization* leverages passive transfer of maternal IgG antibodies across the placenta that can protect the fetus and newborn. The specificity of the maternal IgG reflects past maternal exposures thereby targeting specific pathogens. (B-top panel) *Breastfeeding* provides secretory IgA, with specificities reflecting maternal microbiota, transferred across the gut along with maternal IgG bound to antigen; (B-bottom) breastmilk also contains soluble factors, including cytokines, lipids and fatty acids, that broadly enhance mucosal resistance to infection. (C) Early life *immunization* of the newborn or young infant reduces risk for infection with (C-top) specifically-targeted pathogens (Clemens et al., 2010; Levine, 2011); (C-bottom) live attenuated vaccines such as Bacille Calmette Guérin (BCG) provide broader heterologous (“non-specific”) protection, possibly via “trained immunity” mediated by epigenetic reprogramming of monocytes (Aaby et al.). (D) *Probiotics* reduce infection (Alfaleh and Anabrees, 2014; Oncel et al., 2014; Panigrahi; Roy et al., 2014; Samanta et al., 2009). Mechanisms underlying probiotic effects remain under study and may include, for example, (D-top panel) enhancement of colonization resistance (Buffie and Pamer; Sassone-Corsi and Raffatellu) wherein bacteriocin production by probiotic bacteria targets specific pathogens without affecting commensal flora, and (D-bottom) mucosal PRR signalling-mediated enhancement of immune development, including intestinal epithelial cell expression of antimicrobial protein and peptide (APPs) as well as innate lymphoid cells and mucosal Th17 and T-reg development.

## REFERENCES

- Aaby, P., Kollmann, T.R., and Benn, C.S. (2014). Nonspecific effects of neonatal and infant vaccination: public-health, immunological and conceptual challenges. *Nat Immunol* 15, 895-899.
- Aagaard, K., Ma, J., Antony, K.M., Ganu, R., Petrosino, J., and Versalovic, J. (2014). The placenta harbors a unique microbiome. *Science translational medicine* 6, 237ra265.
- Abel, L., Plancoulaine, S., Jouanguy, E., Zhang, S.Y., Mahfoufi, N., Nicolas, N., Sancho-Shimizu, V., Alcais, A., Guo, Y., Cardon, A., *et al.* (2010). Age-dependent Mendelian predisposition to herpes simplex virus type 1 encephalitis in childhood. *The Journal of pediatrics* 2010 Oct;157, 623-629.
- Agarwal (2016). Characterisation and antimicrobial resistance of sepsis pathogens in neonates born in tertiary care centres in Delhi, India: a cohort study. *The Lancet. Global health* 4, e752-760.
- Alfaleh, K., and Anabrees, J. (2014). Probiotics for prevention of necrotizing enterocolitis in preterm infants. *Cochrane Database Syst Rev* 4, CD005496.
- Amirthalingam, G., Andrews, N., Campbell, H., Ribeiro, S., Kara, E., Donegan, K., Fry, N.K., Miller, E., and Ramsay, M. (2014). Effectiveness of maternal pertussis vaccination in England: an observational study. *Lancet* 384, 1521-1528.
- Ardeشير, A., Narayan, N.R., Mendez-Lagares, G., Lu, D., Rauch, M., Huang, Y., Van Rompay, K.K., Lynch, S.V., and Hartigan-O'Connor, D.J. (2014). Breast-fed and bottle-fed infant rhesus macaques develop distinct gut microbiotas and immune systems. *Sci Transl Med.* 6, 252ra120. doi: 210.1126/scitranslmed.3008791.
- Arrieta, M.C., Stiemsma, L.T., Amenyogbe, N., Brown, E.M., and Finlay, B. (2014). The intestinal microbiome in early life: health and disease. *Frontiers in immunology* 5, 427.
- Arrieta, M.C., Stiemsma, L.T., Dimitriu, P.A., Thorson, L., Russell, S., Yurist-Doutsch, S., Kuzeljevic, B., Gold, M.J., Britton, H.M., Lefebvre, D.L., *et al.* (2015). Early infancy microbial and metabolic alterations affect risk of childhood asthma. *Sci Transl Med.* 7, 307ra152. doi: 310.1126/scitranslmed.aab2271.
- Barnighausen, T., Bloom, D.E., Cafiero-Fonseca, E.T., and O'Brien, J.C. (2014). Valuing vaccination. *Proc Natl Acad Sci U S A* 111, 12313-12319.
- Battersby, A.J., Khara, J., Wright, V.J., Levy, O., and Kampmann, B. (2016). Antimicrobial Proteins and Peptides in Early Life: Ontogeny and Translational Opportunities. *Frontiers in immunology* 7, 309.
- Beigi, R.H., Fortner, K.B., Munoz, F.M., Roberts, J., Gordon, J.L., Han, H.H., Glenn, G., Dormitzer, P.R., Gu, X.X., Read, J.S., *et al.* (2014). Maternal immunization: opportunities for scientific advancement. *Clin Infect Dis* 59 Suppl 7, S408-414.
- Beisel, W.R. (1992). History of nutritional immunology: introduction and overview. *The Journal of nutrition* 122, 591-596.
- Bertoletti, A., and Hong, M. (2014). Age-Dependent Immune Events during HBV Infection from Birth to Adulthood: An Alternative Interpretation. *Frontiers in immunology* 5, 441.
- Bhutta, Z.A., and Black, R.E. (2013). Global maternal, newborn, and child health--so near and yet so far. *N Engl J Med* 369, 2226-2235.
- Blencowe, H., Vos, T., Lee, A.C., Philips, R., Lozano, R., Alvarado, M.R., Cousens, S., and Lawn, J.E. (2013). Estimates of neonatal morbidities and disabilities at regional and global levels for 2010: introduction, methods overview, and relevant findings from the Global Burden of Disease study. *Pediatr Res* 74 Suppl 1, 4-16.

Boer, M.C., Joosten, S.A., and Ottenhoff, T.H. (2015). Regulatory T-Cells at the Interface between Human Host and Pathogens in Infectious Diseases and Vaccination. *Front Immunol.* 6:217, 10.3389/fimmu.2015.00217. eCollection 02015.

Buchmann, K. (2014). Evolution of Innate Immunity: Clues from Invertebrates via Fish to Mammals. *Frontiers in immunology* 5, 459.

Buck, M.D., O'Sullivan, D., and Pearce, E.L. (2015). T cell metabolism drives immunity. *J Exp Med.* 212, 1345-1360. doi: 1310.1084/jem.20151159. Epub 20152015 Aug 20151110.

Buffen, K., Oosting, M., Quintin, J., Ng, A., Kleinnijenhuis, J., Kumar, V., van de Vosse, E., Wijmenga, C., van Crevel, R., Oosterwijk, E., *et al.* (2014). Autophagy controls BCG-induced trained immunity and the response to intravesical BCG therapy for bladder cancer. *PLoS pathogens* 10, e1004485.

Buffie, C.G., and Pamer, E.G. (2013). Microbiota-mediated colonization resistance against intestinal pathogens. *Nat Rev Immunol.* 13, 790-801. doi: 710.1038/nri3535. Epub 2013 Oct 1037.

Bullen, J., Griffiths, E., Rogers, H., and Ward, G. (2000). Sepsis: the critical role of iron. *Microbes and infection / Institut Pasteur* 2, 409-415.

Carr, R. (2000). Neutrophil production and function in newborn infants. *British journal of haematology* 110, 18-28.

Casanova, J.L., Abel, L., and Quintana-Murci, L. (2011). Human TLRs and IL-1Rs in host defense: natural insights from evolutionary, epidemiological, and clinical genetics. *Annu Rev Immunol* 29:447-91, 10.1146/annurev-immunol-030409-101335.

Chan, A.C., Leeansyah, E., Cochrane, A., d'Udekem d'Acoz, Y., Mittag, D., Harrison, L.C., Godfrey, D.I., and Berzins, S.P. (2013). Ex-vivo analysis of human natural killer T cells demonstrates heterogeneity between tissues and within established CD4(+) and CD4(-) subsets. *Clin Exp Immunol.* 172, 129-137. doi: 110.1111/cei.12045.

Chan, M., and Lake, A. (2012). Towards ending preventable child deaths. *Lancet* 379, 2119-2120.

Cheng, S.C., Joosten, L.A., and Netea, M.G. (2014). The interplay between central metabolism and innate immune responses. *Cytokine & growth factor reviews* 25, 707-713.

Chirico, G., Marconi, M., De Amici, M., Gasparoni, A., Mingrat, G., Chiara, A., Rondini, G., and Ugazio, A.G. (1985). Deficiency of neutrophil bactericidal activity in term and preterm infants. A longitudinal study. *Biology of the neonate* 47, 125-129.

Chu, H., and Mazmanian, S.K. (2013). Innate immune recognition of the microbiota promotes host-microbial symbiosis. *Nat Immunol* 14, 668-675.

Clark, N.M., and Lynch, J.P., 3rd (2011). Influenza: epidemiology, clinical features, therapy, and prevention. *Seminars in respiratory and critical care medicine* 32, 373-392.

Clemens, J., Holmgren, J., Kaufmann, S.H., and Mantovani, A. (2010). Ten years of the Global Alliance for Vaccines and Immunization: challenges and progress. *Nat Immunol* 11, 1069-1072.

Cox, L.M., Yamanishi, S., Sohn, J., Alekseyenko, A.V., Leung, J.M., Cho, I., Kim, S.G., Li, H., Gao, Z., Mahana, D., *et al.* (2014). Altering the intestinal microbiota during a critical developmental window has lasting metabolic consequences. *Cell* 158, 705-721.

Cuervo, A.M., and Macian, F. (2012). Autophagy, nutrition and immunology. *Mol Aspects Med.* 33, 2-13. doi: 10.1016/j.mam.2011.1009.1001. Epub 2011 Oct 1011.

Danis, B., George, T.C., Goriely, S., Dutta, B., Renneson, J., Gatto, L., Fitzgerald-Bocarsly, P., Marchant, A., Goldman, M., Willems, F., and De Wit, D. (2008). Interferon regulatory

factor 7-mediated responses are defective in cord blood plasmacytoid dendritic cells. *Eur. J. Immunol.* *38*, 507-517.

Darmstadt, G.L., Kinney, M.V., Chopra, M., Cousens, S., Kak, L., Paul, V.K., Martines, J., Bhutta, Z.A., and Lawn, J.E. (2014). Who has been caring for the baby? *Lancet* *384*, 174-188.

Dauby, N., Goetghebuer, T., Kollmann, T.R., Levy, J., and Marchant, A. (2012). Uninfected but not unaffected: chronic maternal infections during pregnancy, fetal immunity, and susceptibility to postnatal infections. *Lancet Infect Dis* *12*, 330-340.

de Castro, M.J., Pardo-Seco, J., and Martinon-Torres, F. (2015). Nonspecific (Heterologous) Protection of Neonatal BCG Vaccination Against Hospitalization Due to Respiratory Infection and Sepsis. *Clin Infect Dis.* *60*, 1611-1619. doi: 1610.1093/cid/civ1144. Epub 2015 Feb 1627.

De Kleer, I., Willems, F., Lambrecht, B., and Goriely, S. (2014). Ontogeny of myeloid cells. *Frontiers in immunology* *5*, 423.

Debock, I., and Flamand, V. (2014). Unbalanced Neonatal CD4(+) T-Cell Immunity. *Frontiers in immunology* *5*, 393.

Denkel L.A., S.F., Geffers C., Gastmeier P., Garten L., Piening B. (2015). Probiotics prevent necrotizing enterocolitis, sepsis and mortality in preterm infants: a multicenter analysis of more than 10,000 VLBW infants in German NICUs. *Antimicrobial Resistance and Infection Control* *4*, 039.

Deshmukh, H.S., Liu, Y., Menkiti, O.R., Mei, J., Dai, N., O'Leary, C.E., Oliver, P.M., Kolls, J.K., Weiser, J.N., and Worthen, G.S. (2014). The microbiota regulates neutrophil homeostasis and host resistance to *Escherichia coli* K1 sepsis in neonatal mice. *Nat Med.* *20*, 524-530. doi: 510.1038/nm.3542. Epub 2014 Apr 1020.

Di Bartolomeo, S., Nazio, F., and Cecconi, F. (2010). The role of autophagy during development in higher eukaryotes. *Traffic* *11*, 1280-1289.

Diamond, G., Legarda, D., and Ryan, L.K. (2000). The innate immune response of the respiratory epithelium. *Immunol Rev.* *173*, 27-38.

Dimova, T., Brouwer, M., Gosselin, F., Tassignon, J., Leo, O., Donner, C., Marchant, A., and Vermijlen, D. (2015). Effector Vgamma9Vdelta2 T cells dominate the human fetal gammadelta T-cell repertoire. *Proc Natl Acad Sci U S A.* *112*, E556-565. doi: 510.1073/pnas.1412058112. Epub 1412052015 Jan 1412058123.

Dominguez-Bello, M.G., and Blaser, M.J. (2015). Asthma: Undoing millions of years of coevolution in early life? *Sci Transl Med.* *7*, 307fs339. doi: 310.1126/scitranslmed.aad2741.

Dorrestein, P.C., Mazmanian, S.K., and Knight, R. (2014). Finding the missing links among metabolites, microbes, and the host. *Immunity.* *40*, 824-832. doi: 810.1016/j.immuni.2014.1005.1015.

Dorschner, R.A., Lin, K.H., Murakami, M., and Gallo, R.L. (2003). Neonatal skin in mice and humans expresses increased levels of antimicrobial peptides: innate immunity during development of the adaptive response. *Pediatr Res* *53*, 566-572.

Dowling, D.J., and Levy, O. (2014). Ontogeny of early life immunity. *Trends Immunol* *35*, 299-310.

Elahi, S. (2014). New insight into an old concept: role of immature erythroid cells in immune pathogenesis of neonatal infection. *Front Immunol.* *5*:376., 10.3389/fimmu.2014.00376. eCollection 02014.

Elahi, S., Buchanan, R.M., Attah-Poku, S., Townsend, H.G., Babiuk, L.A., and Gerdtts, V. (2006). The host defense peptide beta-defensin 1 confers protection against *Bordetella pertussis* in newborn piglets. *Infect Immun.* 74, 2338-2352.

Elahi, S., Ertelt, J.M., Kinder, J.M., Jiang, T.T., Zhang, X., Xin, L., Chaturvedi, V., Strong, B.S., Qualls, J.E., Steinbrecher, K.A., *et al.* (2013). Immunosuppressive CD71+ erythroid cells compromise neonatal host defence against infection. *Nature* 504, 158-162.

Fink, P.J. (2013). The biology of recent thymic emigrants. *Annu Rev Immunol* 31:31-50., 10.1146/annurev-immunol-032712-100010. Epub 032012 Nov 032711.

Gantt, S., Gervassi, A., Jaspan, H., and Horton, H. (2014). The role of myeloid-derived suppressor cells in immune ontogeny. *Frontiers in immunology* 5, 387.

Gantt, S., and Muller, W.J. (2013). The immunologic basis for severe neonatal herpes disease and potential strategies for therapeutic intervention. *Clinical & developmental immunology* 2013, 369172.

Garcia-Vidal, C., Viasus, D., and Carratala, J. (2013). Pathogenesis of invasive fungal infections. *Current opinion in infectious diseases* 26, 270-276.

Gervassi, A., Lejarcegui, N., Dross, S., Jacobson, A., Itaya, G., Kidzeru, E., Gantt, S., Jaspan, H., and Horton, H. (2014). Myeloid derived suppressor cells are present at high frequency in neonates and suppress in vitro T cell responses. *PLoS One.* 9, e107816. doi: 107810.101371/journal.pone.0107816. eCollection 0102014.

Ghazal, P., Dickinson, P., and Smith, C.L. (2013). Early life response to infection. *Current opinion in infectious diseases* 26, 213-218.

Gomez de Agüero, M., Ganál-Vonarburg, S.C., Fuhrer, T., Rupp, S., Uchimura, Y., Li, H., Steinert, A., Heikenwalder, M., Hapfelmeier, S., Sauer, U., *et al.* (2016). The maternal microbiota drives early postnatal innate immune development. *Science* 351, 1296-1302.

Gomez-Lopez, N., StLouis, D., Lehr, M.A., Sanchez-Rodriguez, E.N., and Arenas-Hernandez, M. (2014). Immune cells in term and preterm labor. *Cellular & molecular immunology* 11, 571-581.

Goodridge, H.S., Ahmed, S.S., Curtis, N., Kollmann, T.R., Levy, O., Netea, M.G., Pollard, A.J., van Crevel, R., and Wilson, C.B. (2016). Harnessing the beneficial heterologous effects of vaccination. *Nature reviews.*

Hackam, D.J., Afrazi, A., Good, M., and Sodhi, C.P. (2013). Innate immune signaling in the pathogenesis of necrotizing enterocolitis. *Clinical & developmental immunology* 2013, 475415.

Haines, C.J., Giffon, T.D., Lu, L.S., Lu, X., Tessier-Lavigne, M., Ross, D.T., and Lewis, D.B. (2009). Human CD4+ T cell recent thymic emigrants are identified by protein tyrosine kinase 7 and have reduced immune function. *J Exp Med.* 206, 275-285. doi: 210.1084/jem.20080996. Epub 20082009 Jan 20080926.

Halsey, N., and Galazka, A. (1985). The efficacy of DPT and oral poliomyelitis immunization schedules initiated from birth to 12 weeks of age. *Bulletin of the World Health Organization* 63, 1151-1169.

Hanson, L.A., and Korotkova, M. (2002). The role of breastfeeding in prevention of neonatal infection. *Seminars in neonatology : SN* 7, 275-281.

Hasko, G., and Cronstein, B. (2013). Regulation of inflammation by adenosine. *Frontiers in immunology* 4, 85.

Hay, G., Refsum, H., Whitelaw, A., Melbye, E.L., Haug, E., and Borch-Johnsen, B. (2007). Predictors of serum ferritin and serum soluble transferrin receptor in newborns and their

associations with iron status during the first 2 y of life. *The American journal of clinical nutrition* *86*, 64-73.

Hermann, E., Truyens, C., Alonso-Vega, C., Even, J., Rodriguez, P., Berthe, A., Gonzalez-Merino, E., Torrico, F., and Carlier, Y. (2002). Human fetuses are able to mount an adultlike CD8 T-cell response. *Blood* *100*, 2153-2158.

Heron, M. (2013). Deaths: leading causes for 2010. *Natl Vital Stat Rep.* *62*, 1-96.

Hong DK, L.D. (2015). Developmental Immunology and Role of Host Defenses in Fetal and Neonatal Susceptibility to Infection. In *Infectious Diseases of the Fetus and Newborn Infant*, V.N. Christopher B Wilson, Yvonne Maldonado, Jack S Remington, Jerome O Klein, ed. (Philadelphia: Elsevier Saunders), pp. 81-188.

Hood, M.I., and Skaar, E.P. (2012). Nutritional immunity: transition metals at the pathogen-host interface. *Nature reviews. Microbiology* *10*, 525-537.

Hornef, M.W., and Fulde, M. (2014). Ontogeny of intestinal epithelial innate immune responses. *Frontiers in immunology* *5*, 474.

Hostetter, M.K. (2012). What we don't see. *N Engl J Med* *366*, 1328-1334.

Hsieh, E., Smith, P.B., Jacqz-Aigrain, E., Kaguelidou, F., Cohen-Wolkowicz, M., Manzoni, P., and Benjamin, D.K., Jr. (2012). Neonatal fungal infections: when to treat? *Early Hum Dev* *88 Suppl 2*, S6-s10.

Huda, M.N., Lewis, Z., Kalanetra, K.M., Rashid, M., Ahmad, S.M., Raqib, R., Qadri, F., Underwood, M.A., Mills, D.A., and Stephensen, C.B. (2014). Stool microbiota and vaccine responses of infants. *Pediatrics* *134*, e362-372.

Huygens, A., Dauby, N., Vermijlen, D., and Marchant, A. (2014). Immunity to cytomegalovirus in early life. *Front Immunol.* *5*:552, 10.3389/fimmu.2014.00552. eCollection 02014.

Huygens, A., Lecomte, S., Tackoen, M., Orlislagers, V., Delmarcelle, Y., Burny, W., Van Rysselberge, M., Liesnard, C., Larsen, M., Appay, V., *et al.* (2015). Functional Exhaustion Limits CD4+ and CD8+ T-Cell Responses to Congenital Cytomegalovirus Infection. *J Infect Dis.* *212*, 484-494. doi: 410.1093/infdis/jiv1071. Epub 2015 Feb 1095.

Iwasaki, A., and Medzhitov, R. (2015). Control of adaptive immunity by the innate immune system. *Nat Immunol.* *16*, 343-353. doi: 310.1038/ni.3123.

Johnson, E.E., and Wessling-Resnick, M. (2012). Iron metabolism and the innate immune response to infection. *Microbes and infection / Institut Pasteur* *14*, 207-216.

Jones, S.A. (2005). Directing transition from innate to acquired immunity: defining a role for IL-6. *J Immunol.* *175*, 3463-3468.

Khosravi, A., Yanez, A., Price, J.G., Chow, A., Merad, M., Goodridge, H.S., and Mazmanian, S.K. (2014). Gut microbiota promote hematopoiesis to control bacterial infection. *Cell Host Microbe.* *15*, 374-381. doi: 310.1016/j.chom.2014.1002.1006.

King, A.E., Kelly, R.W., Sallenave, J.M., Bocking, A.D., and Challis, J.R. (2007). Innate immune defences in the human uterus during pregnancy. *Placenta.* *28*, 1099-1106. Epub 2007 Jul 1030.

Kleinnijenhuis, J., Quintin, J., Preijers, F., Benn, C.S., Joosten, L.A., Jacobs, C., van Loenhout, J., Xavier, R.J., Aaby, P., van der Meer, J.W., *et al.* (2014). Long-lasting effects of BCG vaccination on both heterologous Th1/Th17 responses and innate trained immunity. *Journal of innate immunity* *6*, 152-158.

Kleinnijenhuis, J., van Crevel, R., and Netea, M.G. (2015). Trained immunity: consequences for the heterologous effects of BCG vaccination. *Transactions of the Royal Society of Tropical Medicine and Hygiene* 109, 29-35.

Kochan, I., Golden, C.A., and Bukovic, J.A. (1969). Mechanism of tuberculostasis in mammalian serum. II. Induction of serum tuberculostasis in guinea pigs. *Journal of bacteriology* 100, 64-70.

Kollmann, T.R., Levy, O., Montgomery, R.R., and Goriely, S. (2012). Innate Immune Function by Toll-like Receptors: Distinct Responses in Newborns and the Elderly. *Immunity* 37, 771-783.

Krow-Lucal, E.R., and McCune, J.M. (2014). Distinct functional programs in fetal T and myeloid lineages. *Frontiers in immunology* 5, 314.

Landwehr-Kenzel, S., and Henneke, P. (2014). Interaction of *Streptococcus agalactiae* and Cellular Innate Immunity in Colonization and Disease. *Frontiers in immunology* 5, 519.

Lawn, J.E., Blencowe, H., Oza, S., You, D., Lee, A.C., Waiswa, P., Lalli, M., Bhutta, Z., Barros, A.J., Christian, P., *et al.* (2014). Every Newborn: progress, priorities, and potential beyond survival. *Lancet* 384, 189-205.

Legrand, D. (2016). Overview of Lactoferrin as a Natural Immune Modulator. *The Journal of pediatrics* 173 Suppl, S10-15.

Levin, M., Quint, P.A., Goldstein, B., Barton, P., Bradley, J.S., Shemie, S.D., Yeh, T., Kim, S.S., Cafaro, D.P., Scannon, P.J., and Giroir, B.P. (2000). Recombinant bactericidal/permeability-increasing protein (rBPI21) as adjunctive treatment for children with severe meningococcal sepsis: a randomised trial. rBPI21 Meningococcal Sepsis Study Group. *Lancet* 356, 961-967.

Levine, M.M. (2011). "IDEAL" vaccines for resource poor settings. *Vaccine* 29 Suppl 4, D116-125.

Levy, O. (2007). Innate immunity of the newborn: basic mechanisms and clinical correlates. *Nature reviews* 7, 379-390.

Levy, O., Coughlin, M., Cronstein, B.N., Roy, R.M., Desai, A., and Wessels, M.R. (2006). The adenosine system selectively inhibits TLR-mediated TNF-alpha production in the human newborn. *J Immunol* 177, 1956-1966.

Levy, O., and Netea, M.G. (2014). Innate immune memory: implications for development of pediatric immunomodulatory agents and adjuvanted vaccines. *Pediatr Res* 75, 184-188.

Liang, Q., Luo, Z., Zeng, J., Chen, W., Foo, S.S., Lee, S.A., Ge, J., Wang, S., Goldman, S.A., Zlokovic, B.V., *et al.* (2016). Zika Virus NS4A and NS4B Proteins Deregulate Akt-mTOR Signaling in Human Fetal Neural Stem Cells to Inhibit Neurogenesis and Induce Autophagy. *Cell stem cell*.

Libraty, D.H., Zhang, L., Woda, M., Acosta, L.P., Obcena, A., Brion, J.D., and Capeding, R.Z. (2014). Neonatal BCG vaccination is associated with enhanced T-helper 1 immune responses to heterologous infant vaccines. *Trials in vaccinology* 3, 1-5.

Lindsey, B., Jones, C., and Kampmann, B. (2012). Bridging the gap: maternal immunisation as a means to reduce neonatal deaths from infectious diseases. *Pathogens and global health* 106, 137-138.

Lissner, M.M., Thomas, B.J., Wee, K., Tong, A.J., Kollmann, T.R., and Smale, S.T. (2015). Age-Related Gene Expression Differences in Monocytes from Human Neonates, Young Adults, and Older Adults. *PLoS One*. 10, e0132061. doi: 10.1371/journal.pone.0132061. eCollection 0132015.

Liu, L., Johnson, H.L., Cousens, S., Perin, J., Scott, S., Lawn, J.E., Rudan, I., Campbell, H., Cibulskis, R., Li, M., *et al.* (2012). Global, regional, and national causes of child mortality: an updated systematic analysis for 2010 with time trends since 2000. *Lancet* 379, 2151-2161.

Lotz, M., Gutle, D., Walther, S., Menard, S., Bogdan, C., and Hornef, M.W. (2006). Postnatal acquisition of endotoxin tolerance in intestinal epithelial cells. *J Exp Med* 203, 973-984.

Lund, N., Andersen, A., Hansen, A.S., Jepsen, F.S., Barbosa, A., Biering-Sorensen, S., Rodrigues, A., Ravn, H., Aaby, P., and Benn, C.S. (2015). The Effect of Oral Polio Vaccine at Birth on Infant Mortality: A Randomized Trial. *Clin Infect Dis*.

MacGillivray, D.M., and Kollmann, T.R. (2014). The role of environmental factors in modulating immune responses in early life. *Frontiers in immunology* 5, 434.

Mansbach, J.M., Piedra, P.A., Borregaard, N., Martineau, A.R., Neuman, M.I., Espinola, J.A., and Camargo, C.A., Jr. (2012). Serum cathelicidin level is associated with viral etiology and severity of bronchiolitis. *J Allergy Clin Immunol.* 130, 1007-1008.e1001. doi: 10.1016/j.jaci.2012.1007.1044. Epub 2012 Sep 1001.

Manzoni, P., Meyer, M., Stolfi, I., Rinaldi, M., Cattani, S., Pagni, L., Romeo, M.G., Messner, H., Decembrino, L., Laforgia, N., *et al.* (2014). Bovine lactoferrin supplementation for prevention of necrotizing enterocolitis in very-low-birth-weight neonates: a randomized clinical trial. *Early Hum Dev* 90 Suppl 1, S60-65.

Manzoni, P., Rinaldi, M., Cattani, S., Pagni, L., Romeo, M.G., Messner, H., Stolfi, I., Decembrino, L., Laforgia, N., Vagnarelli, F., *et al.* (2009). Bovine lactoferrin supplementation for prevention of late-onset sepsis in very low-birth-weight neonates: a randomized trial. *Jama* 302, 1421-1428.

Manzoni, P., Stolfi, I., Messner, H., Cattani, S., Laforgia, N., Romeo, M.G., Bollani, L., Rinaldi, M., Gallo, E., Quercia, M., *et al.* (2012). Bovine lactoferrin prevents invasive fungal infections in very low birth weight infants: a randomized controlled trial. *Pediatrics* 129, 116-123.

Marchant, A., Goetghebuer, T., Ota, M.O., Wolfe, I., Ceesay, S.J., De Groote, D., Corrah, T., Bennett, S., Wheeler, J., Huygen, K., *et al.* (1999). Newborns develop a Th1-type immune response to *Mycobacterium bovis* bacillus Calmette-Guerin vaccination. *J Immunol.* 163, 2249-2255.

Marchant, E.A., Boyce, G.K., Sadarangani, M., and Lavoie, P.M. (2013). Neonatal sepsis due to coagulase-negative staphylococci. *Clinical & developmental immunology* 2013, 586076.

Marchant, E.A., Kan, B., Sharma, A.A., van Zanten, A., Kollmann, T.R., Brant, R., and Lavoie, P.M. (2015). Attenuated innate immune defenses in very premature neonates during the neonatal period. *Pediatr Res*.

Marchini, G., Lindow, S., Brismar, H., Stabi, B., Berggren, V., Ulfgren, A.K., Lonne-Rahm, S., Agerberth, B., and Gudmundsson, G.H. (2002). The newborn infant is protected by an innate antimicrobial barrier: peptide antibiotics are present in the skin and vernix caseosa. *The British journal of dermatology* 147, 1127-1134.

Mascart, F., Verscheure, V., Malfroot, A., Hainaut, M., Pierard, D., Temerman, S., Peltier, A., Debie, A.S., Levy, J., Del Giudice, G., and Loch, C. (2003). Bordetella pertussis infection in 2-month-old infants promotes type 1 T cell responses. *J Immunol.* 170, 1504-1509.

Maynard, C.L., Elson, C.O., Hatton, R.D., and Weaver, C.T. (2012). Reciprocal interactions of the intestinal microbiota and immune system. *Nature* 489, 231-241.

McFall-Ngai, M., Hadfield, M.G., Bosch, T.C., Carey, H.V., Domazet-Lozo, T., Douglas, A.E., Dubilier, N., Eberl, G., Fukami, T., Gilbert, S.F., *et al.* (2013). Animals in a bacterial world, a



new imperative for the life sciences. *Proc Natl Acad Sci U S A.* 110, 3229-3236. doi: 3210.1073/pnas.1218525110. Epub 1218522013 Feb 1218525117.

Mold, J.E., Michaelsson, J., Burt, T.D., Muench, M.O., Beckerman, K.P., Busch, M.P., Lee, T.H., Nixon, D.F., and McCune, J.M. (2008). Maternal alloantigens promote the development of tolerogenic fetal regulatory T cells in utero. *Science.* 322, 1562-1565. doi: 1510.1126/science.1164511.

Mold, J.E., Venkatasubrahmanyam, S., Burt, T.D., Michaelsson, J., Rivera, J.M., Galkina, S.A., Weinberg, K., Stoddart, C.A., and McCune, J.M. (2010). Fetal and adult hematopoietic stem cells give rise to distinct T cell lineages in humans. *Science* 330, 1695-1699.

Muenchhoff, M., Adland, E., Karimanzira, O., Crowther, C., Pace, M., Csala, A., Leitman, E., Moonsamy, A., McGregor, C., Hurst, J., *et al.* (2016). Nonprogressing HIV-infected children share fundamental immunological features of nonpathogenic SIV infection. *Science translational medicine* 8, 358ra125.

Muenchhoff, M., Prendergast, A.J., and Goulder, P.J. (2014). Immunity to HIV in Early Life. *Frontiers in immunology* 5, 391.

Murray, C.J., Vos, T., Lozano, R., Naghavi, M., Flaxman, A.D., Michaud, C., Ezzati, M., Shibuya, K., Salomon, J.A., Abdalla, S., *et al.* (2012). Disability-adjusted life years (DALYs) for 291 diseases and injuries in 21 regions, 1990-2010: a systematic analysis for the Global Burden of Disease Study 2010. *Lancet* 380, 2197-2223.

Nairz, M., Haschka, D., Demetz, E., and Weiss, G. (2014). Iron at the interface of immunity and infection. *Frontiers in pharmacology* 5, 152.

Nakazawa, T., Agematsu, K., and Yabuhara, A. (1997). Later development of Fas ligand-mediated cytotoxicity as compared with granule-mediated cytotoxicity during the maturation of natural killer cells. *Immunology.* 92, 180-187.

Netea, M.G., Latz, E., Mills, K.H., and O'Neill, L.A. (2015). Innate immune memory: a paradigm shift in understanding host defense. *Nat Immunol.* 16, 675-679. doi: 610.1038/ni.3178.

Netea, M.G., and van Crevel, R. (2014). BCG-induced protection: Effects on innate immune memory. *Seminars in immunology* 26, 512-517.

Niewiesk, S. (2014). Maternal antibodies: clinical significance, mechanism of interference with immune responses, and possible vaccination strategies. *Frontiers in immunology* 5, 446.

O'Neill, L.A., Kishton, R.J., and Rathmell, J. (2016). A guide to immunometabolism for immunologists. *Nature reviews* 16, 553-565.

Oestergaard, M.Z., Inoue, M., Yoshida, S., Mahanani, W.R., Gore, F.M., Cousens, S., Lawn, J.E., and Mathers, C.D. (2011). Neonatal mortality levels for 193 countries in 2009 with trends since 1990: a systematic analysis of progress, projections, and priorities. *PLoS Med.* 8, e1001080. doi: 1001010.1001371/journal.pmed.1001080. Epub 1002011 Aug 1001030.

Oh, D.Y., Dowling, D.J., Ahmed, S., Choi, H., Brightman, S., Bergelson, I., Berger, S.T., Sauld, J.F., Pettengill, M., Kho, A.T., *et al.* (2016). Adjuvant-induced Human Monocyte Secretome Profiles Reveal Adjuvant- and Age-specific Protein Signatures. *Molecular & cellular proteomics : MCP* 15, 1877-1894.

Oh, J.E., and Lee, H.K. (2014). Pattern recognition receptors and autophagy. *Front Immunol.* 5:300., 10.3389/fimmu.2014.00300. eCollection 02014.

Oh, J.Z., Ravindran, R., Chassaing, B., Carvalho, F.A., Maddur, M.S., Bower, M., Hakimpour, P., Gill, K.P., Nakaya, H.I., Yarovinsky, F., *et al.* (2014). TLR5-mediated sensing of gut microbiota

is necessary for antibody responses to seasonal influenza vaccination. *Immunity* 41, 478-492.

Olszak, T., An, D., Zeissig, S., Vera, M.P., Richter, J., Franke, A., Glickman, J.N., Siebert, R., Baron, R.M., Kasper, D.L., and Blumberg, R.S. (2012). Microbial exposure during early life has persistent effects on natural killer T cell function. *Science* 336, 489-493.

Oncel, M.Y., Sari, F.N., Arayici, S., Guzoglu, N., Erdeve, O., Uras, N., Oguz, S.S., and Dilmen, U. (2014). Lactobacillus Reuteri for the prevention of necrotising enterocolitis in very low birthweight infants: a randomised controlled trial. *Arch Dis Child Fetal Neonatal Ed* 99, F110-115.

Oppenheimer, S. (2012). Iron and infection: narrative review of a major iron supplementation study in Papua New Guinea undertaken by the Department of Tropical Paediatrics, Liverpool School of Tropical Medicine, 1979-1983, its aftermath and the continuing relevance of its results. *Paediatrics and international child health* 32 Suppl 2, S21-29.

Oppenheimer, S.J. (2001). Iron and its relation to immunity and infectious disease. *The Journal of nutrition* 131, 616S-633S; discussion 633S-635S.

Ota, M.O., Vekemans, J., Schlegel-Haueter, S.E., Fielding, K., Whittle, H., Lambert, P.H., McAdam, K.P., Siegrist, C.A., and Marchant, A. (2004). Hepatitis B immunisation induces higher antibody and memory Th2 responses in new-borns than in adults. *Vaccine* 22, 511-519.

Pammi, M., and Abrams, S.A. (2015). Oral lactoferrin for the prevention of sepsis and necrotizing enterocolitis in preterm infants. *Cochrane Database Syst Rev.* 2:CD007137, 10.1002/14651858.CD14007137.pub14651854.

Pandiyan, P., Zheng, L., and Lenardo, M.J. (2011). The molecular mechanisms of regulatory T cell immunosuppression. *Front Immunol.* 2:60, 10.3389/fimmu.2011.00060. eCollection 02011.

Panigrahi, P. (2013). Effectiveness of a Lactbacillus plantarum synbiotic therapy for preventing late onset sepsis in neonates. In PAS Annual Meeting (Washington D.C., Pediatric Academic Society), p. Abstract 3618.3612.

Pettengill, M., Robson, S., Tresenriter, M., Millan, J.L., Usheva, A., Bingham, T., Belderbos, M., Bergelson, I., Burl, S., Kampmann, B., *et al.* (2013). Soluble ecto-5'-nucleotidase (5'-NT), alkaline phosphatase, and adenosine deaminase (ADA1) activities in neonatal blood favor elevated extracellular adenosine. *J Biol Chem* 288, 27315-27326.

Pettengill, M.A., and Levy, O. (2016). Circulating Human Neonatal Naive B Cells are Deficient in CD73 Impairing Purine Salvage. *Frontiers in immunology* 7, 121.

Pettengill, M.A., van Haren, S.D., and Levy, O. (2014). Soluble mediators regulating immunity in early life. *Frontiers in immunology* 5, 457.

Power Coombs, M.R., Kronforst, K., and Levy, O. (2013). Neonatal host defense against Staphylococcal infections. *Clinical & developmental immunology* 2013, 826303.

Randow, F., MacMicking, J.D., and James, L.C. (2013). Cellular self-defense: how cell-autonomous immunity protects against pathogens. *Science* 340, 701-706.

Rao, S., and Ali, U. (2005). Systemic fungal infections in neonates. *Journal of postgraduate medicine* 51 Suppl 1, S27-29.

Recalcati, S., Locati, M., and Cairo, G. (2012). Systemic and cellular consequences of macrophage control of iron metabolism. *Seminars in immunology* 24, 393-398.

Rechavi, E., Lev, A., Lee, Y.N., Simon, A.J., Yinon, Y., Lipitz, S., Amariglio, N., Weisz, B., Notarangelo, L.D., and Somech, R. (2015). Timely and spatially regulated maturation of B and T cell repertoire during human fetal development. *Science translational medicine* 7, 276ra225.

Renz, H., Brandtzaeg, P., and Hornef, M. (2012). The impact of perinatal immune development on mucosal homeostasis and chronic inflammation. *Nature reviews* 12, 9-23.

Richetta, C., and Faure, M. (2013). Autophagy in antiviral innate immunity. *Cell Microbiol.* 15, 368-376. doi: 310.1111/cmi.12043. Epub 12012 Nov 12047.

Rochette, L., Gudjoncik, A., Guenancia, C., Zeller, M., Cottin, Y., and Vergely, C. (2015). The iron-regulatory hormone hepcidin: A possible therapeutic target? *Pharmacology & therapeutics* 146c, 35-52.

Roy, A., Chaudhuri, J., Sarkar, D., Ghosh, P., and Chakraborty, S. (2014). Role of Enteric Supplementation of Probiotics on Late-onset Sepsis by *Candida* species in Preterm Low Birth Weight Neonates: A Randomized, Double Blind, Placebo-controlled Trial. *North American journal of medical sciences* 6, 50-57.

Ruchala, P., and Nemeth, E. (2014). The pathophysiology and pharmacology of hepcidin. *Trends in pharmacological sciences* 35, 155-161.

Salzman, N.H., Underwood, M.A., and Bevins, C.L. (2007). Paneth cells, defensins, and the commensal microbiota: a hypothesis on intimate interplay at the intestinal mucosa. *Seminars in immunology* 19, 70-83.

Samanta, M., Sarkar, M., Ghosh, P., Ghosh, J., Sinha, M., and Chatterjee, S. (2009). Prophylactic probiotics for prevention of necrotizing enterocolitis in very low birth weight newborns. *Journal of tropical pediatrics* 55, 128-131.

Sassone-Corsi, M., and Raffatellu, M. (2015). No vacancy: how beneficial microbes cooperate with immunity to provide colonization resistance to pathogens. *J Immunol.* 194, 4081-4087. doi: 4010.4049/jimmunol.1403169.

Sazawal, S., Black, R.E., Ramsan, M., Chwaya, H.M., Stoltzfus, R.J., Dutta, A., Dhingra, U., Kabole, I., Deb, S., Othman, M.K., and Kabole, F.M. (2006). Effects of routine prophylactic supplementation with iron and folic acid on admission to hospital and mortality in preschool children in a high malaria transmission setting: community-based, randomised, placebo-controlled trial. *Lancet* 367, 133-143.

Seale, A.C., Blencowe, H., Manu, A.A., Nair, H., Bahl, R., Qazi, S.A., Zaidi, A.K., Berkley, J.A., Cousens, S.N., and Lawn, J.E. (2014). Estimates of possible severe bacterial infection in neonates in sub-Saharan Africa, south Asia, and Latin America for 2012: a systematic review and meta-analysis. *Lancet Infect Dis* 14, 731-741.

Sepulveda, J., and Murray, C. (2014). The state of global health in 2014. *Science* 345, 1275-1278.

Sharma, A.A., Jen, R., Butler, A., and Lavoie, P.M. (2012). The developing human preterm neonatal immune system: a case for more research in this area. *Clin Immunol.* 145, 61-68. doi: 10.1016/j.clim.2012.1008.1006. Epub 2012 Aug 1017.

Sherrid, A.M., and Kollmann, T.R. (2013). Age-Dependent Differences in Systemic and Cell-Autonomous Immunity to *L. monocytogenes*. *Clinical & developmental immunology* 2013, 917198.

Siegrist, C.A., and Aspinall, R. (2009). B-cell responses to vaccination at the extremes of age. *Nature reviews* 9, 185-194.

Singh, V.V., Chauhan, S.K., Rai, R., Kumar, A., Singh, S.M., and Rai, G. (2013). Decreased pattern recognition receptor signaling, interferon-signature, and bactericidal/permeability-increasing protein gene expression in cord blood of term low birth weight human newborns. *PloS one* 8, e62845.

Sorup, S., Benn, C.S., Poulsen, A., Krause, T.G., Aaby, P., and Ravn, H. (2014). Live vaccine against measles, mumps, and rubella and the risk of hospital admissions for nontargeted infections. *Jama* 311, 826-835.

Speer, C.P., Gahr, M., Wieland, M., and Eber, S. (1988). Phagocytosis-associated functions in neonatal monocyte-derived macrophages. *Pediatr Res* 24, 213-216.

Starner, T.D., Agerberth, B., Gudmundsson, G.H., and McCray, P.B., Jr. (2005). Expression and activity of beta-defensins and LL-37 in the developing human lung. *J Immunol* 174, 1608-1615.

Strunk, T., Doherty, D., Richmond, P., Simmer, K., Charles, A., Levy, O., Liyanage, K., Smith, T., Currie, A., and Burgner, D. (2009). Reduced levels of antimicrobial proteins and peptides in human cord blood plasma. *Archives of disease in childhood. Fetal and neonatal edition* 94, F230-231.

Strunk, T., Kollmann, T., and Patole, S. (2015). Probiotics to prevent early-life infection. *Lancet Infect Dis.* 15, 378-379. doi: 310.1016/S1473-3099(1015)70088-70085.

Sturgeon, P. (1954). Studies of iron requirements in infante and children. I. Normal values for serum iron, copper and free erythrocyte protoporphyry. *Pediatrics* 13, 107-125.

Szabo, M., Vasarhelyi, B., Balla, G., Szabo, T., Machay, T., and Tulassay, T. (2001). Acute postnatal increase of extracellular antioxidant defence of neonates: the role of iron metabolism. *Acta Paediatr* 90, 1167-1170.

Thome, J.J., Bickham, K.L., Ohmura, Y., Kubota, M., Matsuoka, N., Gordon, C., Granot, T., Griesemer, A., Lerner, H., Kato, T., and Farber, D.L. (2016). Early-life compartmentalization of human T cell differentiation and regulatory function in mucosal and lymphoid tissues. *Nature medicine* 22, 72-77.

Tollin, M., Bergsson, G., Kai-Larsen, Y., Lengqvist, J., Sjovall, J., Griffiths, W., Skuladottir, G.V., Haraldsson, A., Jornvall, H., Gudmundsson, G.H., and Agerberth, B. (2005). Vernix caseosa as a multi-component defence system based on polypeptides, lipids and their interactions. *Cell Mol Life Sci.* 62, 2390-2399.

Tourneur, E., and Chassin, C. (2013). Neonatal immune adaptation of the gut and its role during infections. *Clinical & developmental immunology* 2013, 270301.

Travis, S.M., Conway, B.A., Zabner, J., Smith, J.J., Anderson, N.N., Singh, P.K., Greenberg, E.P., and Welsh, M.J. (1999). Activity of abundant antimicrobials of the human airway. *Am J Respir Cell Mol Biol.* 20, 872-879.

Turvey, S.E., and Broide, D.H. (2010). Innate immunity. *The Journal of allergy and clinical immunology* 125, S24-32.

Underwood, M.A., Gilbert, W.M., and Sherman, M.P. (2005). Amniotic fluid: not just fetal urine anymore. *Journal of perinatology : official journal of the California Perinatal Association* 25, 341-348.

UNICEF (2014). IMMUNIZATION SUMMARY - A statistical reference containing data through 2013. U.N.C.s.F. (UNICEF), ed. (New York, NY, UNICEF).

van Haren, S.D., Ganapathi, L., Bergelson, I., Dowling, D.J., Banks, M., Samuels, R.C., Reed, S.G., Marshall, J.D., and Levy, O. (2016). In vitro cytokine induction by TLR-activating vaccine adjuvants in human blood varies by age and adjuvant. *Cytokine* 83, 99-109.

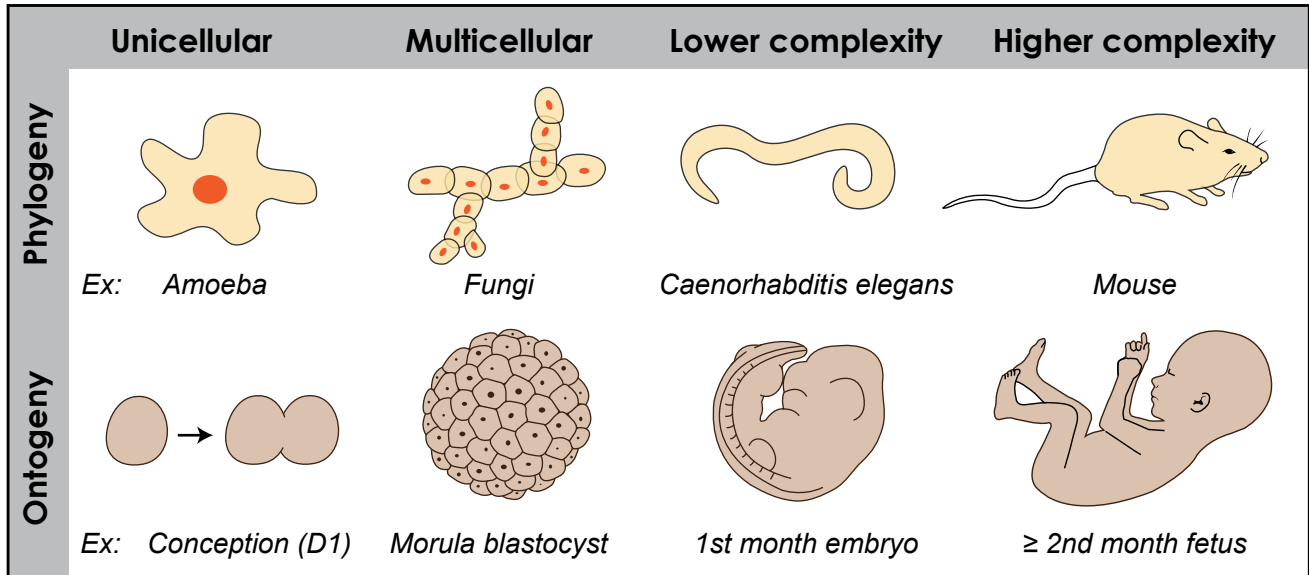
- Vanden Driessche, K., Persson, A., Marais, B.J., Fink, P.J., and Urdahl, K.B. (2013). Immune vulnerability of infants to tuberculosis. *Clinical & developmental immunology* 2013, 781320.
- Vergnano, S., Sharland, M., Kazembe, P., Mwansambo, C., and Heath, P.T. (2005). Neonatal sepsis: an international perspective. *Archives of disease in childhood. Fetal and neonatal edition* 90, F220-224.
- Vermijlen, D., Brouwer, M., Donner, C., Liesnard, C., Tackoen, M., Van Rysselberge, M., Twite, N., Goldman, M., Marchant, A., and Willems, F. Human cytomegalovirus elicits fetal gammadelta T cell responses in utero. *J Exp Med* 207, 807-821.
- Vermijlen, D., and Prinz, I. (2014). Ontogeny of Innate T Lymphocytes - Some Innate Lymphocytes are More Innate than Others. *Frontiers in immunology* 5, 486.
- Vidarsson, G., Dekkers, G., and Rispens, T. (2014). IgG subclasses and allotypes: from structure to effector functions. *Front Immunol.* 5:520., 10.3389/fimmu.2014.00520. eCollection 02014.
- Visscher, M., and Narendran, V. (2014). The Ontogeny of Skin. *Advances in wound care* 3, 291-303.
- Wander, K., Shell-Duncan, B., and McDade, T.W. (2009). Evaluation of iron deficiency as a nutritional adaptation to infectious disease: an evolutionary medicine perspective. *American journal of human biology : the official journal of the Human Biology Council* 21, 172-179.
- Wang, H., Liddell, C.A., Coates, M.M., Mooney, M.D., Levitz, C.E., Schumacher, A.E., Apfel, H., Iannarone, M., Phillips, B., Lofgren, K.T., *et al.* (2014). Global, regional, and national levels of neonatal, infant, and under-5 mortality during 1990-2013: a systematic analysis for the Global Burden of Disease Study 2013. *Lancet.* 384, 957-979. doi: 910.1016/S0140-6736(1014)60497-60499. Epub 62014 May 60492.
- Weiner, J., 3rd, and Kaufmann, S.H. (2014). Recent advances towards tuberculosis control: vaccines and biomarkers. *Journal of internal medicine* 275, 467-480.
- White, G.P., Hollams, E.M., Yerkovich, S.T., Bosco, A., Holt, B.J., Bassami, M.R., Kusel, M., Sly, P.D., and Holt, P.G. (2006). CpG methylation patterns in the IFNgamma promoter in naive T cells: variations during Th1 and Th2 differentiation and between atopics and non-atopics. *Pediatr Allergy Immunol* 17, 557-564.
- White, G.P., Watt, P.M., Holt, B.J., and Holt, P.G. (2002). Differential patterns of methylation of the IFN-gamma promoter at CpG and non-CpG sites underlie differences in IFN-gamma gene expression between human neonatal and adult CD45RO- T cells. *J Immunol* 168, 2820-2827.
- Wiesner, J., and Vilcinskas, A. (2010). Antimicrobial peptides: the ancient arm of the human immune system. *Virulence.* 1, 440-464. doi: 410.4161/viru.4161.4165.12983.
- Wilcox, D.R., Wadhvani, N.R., Longnecker, R., and Muller, W.J. (2015). Differential reliance on autophagy for protection from HSV encephalitis between newborns and adults. *PLoS Pathog.* 11, e1004580. doi: 1004510.1001371/journal.ppat.1004580. eCollection 1002015 Jan.
- Wu, J., and Chen, Z.J. (2014). Innate immune sensing and signaling of cytosolic nucleic acids. *Annu Rev Immunol* 32:461-88., 10.1146/annurev-immunol-032713-120156.
- Wynn, J.L., Scumpia, P.O., Stocks, B.T., Romano-Keeler, J., Alrifai, M.W., Liu, J.H., Kim, A.S., Alford, C.E., Matta, P., Weitkamp, J.H., and Moore, D.J. (2015). Neonatal CD71+ Erythroid

Cells Do Not Modify Murine Sepsis Mortality. *J Immunol.* 195, 1064-1070. doi: 1010.4049/jimmunol.1500771. Epub 1502015 Jun 1500722.

Yoshio, H., Tollin, M., Gudmundsson, G.H., Lagercrantz, H., Jornvall, H., Marchini, G., and Agerberth, B. (2003). Antimicrobial polypeptides of human vernix caseosa and amniotic fluid: implications for newborn innate defense. *Pediatr Res* 53, 211-216.

Zhang, X., Mozeleski, B., Lemoine, S., Deriaud, E., Lim, A., Zhivaki, D., Azria, E., Le Ray, C., Roguet, G., Launay, O., *et al.* (2014). CD4 T cells with effector memory phenotype and function develop in the sterile environment of the fetus. *Sci Transl Med.* 6, 238ra272. doi: 210.1126/scitranslmed.3008748.

Figure 1



**Cell Autonomous Innate Immunity**

Independent of other cells; Compartmentalization within cell

**Nutritional Innate Immunity**

Communication amongst cells; No cells specialized in host defense

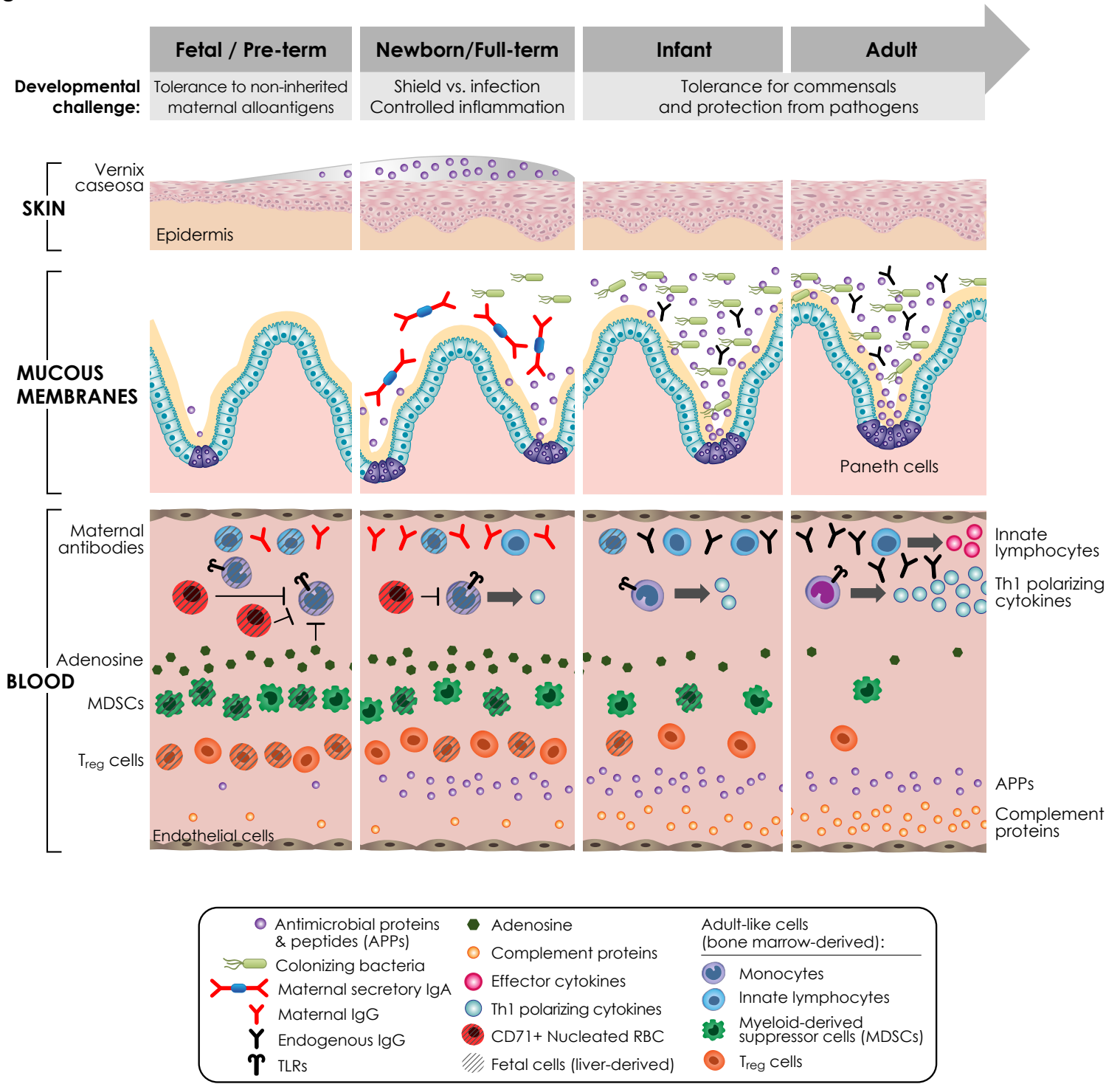
**Leukocyte-based Innate Immunity**

Cells specialized in host defense

**Antigen-specific Adaptive Immunity**

Specialized cells & tissues

**Figure 2**





**Figure 3**

