METABOLISM OF SODIUM NITROPRUSSIDE (SNP) IN RAT RED BLOOD CELLS

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Sodium nitroprusside (SNP) is an established drug, clinically used in the treatment of hypertensive emergencies. The aim of this study is further elucidation of the molecular mechanism of the SNP action, particularly its metabolism in rat erythrocytes and reticulocytes. Rats erythrocyte and reticulocyte-rich suspensions were aerobically incubated without (control) or in the presence of SNP (0.1, 0.25, 0.5, 1.0 i 1.5 mM). The concentrations of reactive nitrogen species (RNS) and reactive oxygen species (ROS) were determined after incubation. In rat erythrocytes, SNP did not alter nitrite level (NO+ ion indicator), while significantly increased concentrations of hydroxylamine (NO- ion indicator), S-nitrosothiols (SNO) and 3-nitrotyrosine (peroxynitrite indicator). Concentration of superoxide anion (O2-) decreased in the

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presence of low doses of SNP only, while level of hydrogen peroxide (H₂O₂) increased in dose-dependent manner in rat erythrocytes. On the other hand, SNP significantly increased nitrite, hydroxylamine and 3-nitrotyrosine concentrations in rat reticulocytes. In addition, low doses of SNP induced decrease of O₂- level. Concentration of H₂O₂ did not alter in rat reticulocytes. On the basis of these data, we can conclude: SNP spontaneously liberated nitric oxide as NO- ion in rat erythrocytes and reticulocytes. In addition, applied experimental doses of SNP induced strong nitrosative and oxidative stress in these cells.

Key words: erythrocytes, nitroxyl anion, reticulocytes, sodium nitroprusside.

INTRODUCTION

Corresponding to their intermediate position in the differentiation program, the reticulocytes do not possess full range of metabolic pathways of proliferating cells, but they are still equipped with a set of metabolic pathways (corresponding to presence of mitochondria and ribosomes), most of which are lost during their transition to the mature erythrocytes (RAPOPORT, 1986). Reactive oxygen species (ROS) and their derivatives are present in living tissues at low but measurable concentrations, which are determined by the balance between the rates of radical production and their corresponding rates of clearance (DROGE, 2002). In conditions of disturbed prooxidant-antioxidant balance in favour of the prooxidants, leading to potential damage of cells, the oxidative stress occurs (SIES, 1991). Nitric oxide (NO) is a small hydrophobic molecule with chemical properties which make it uniquely suitable for both intra- and intercellular messenger. In the reactions with O2-/O2, NO generated reactive nitrogen species (RNS) which affected almost every molecule in cells (WINK and MITCHELL, 1998). Diverse and important physiological roles of NO implicate that exogenous donation of NO may be useful in the treatment of some disease states. Transition metal NO complex, sodium nitroprusside (SNP) spontaneously releases NO, mainly as nitrosonium ion - NO+ (Hou et al., 1999).

The aim of this study is further elucidation of the molecular mechanism of the SNP action, particularly its metabolism in rat erythrocytes and reticulocytes.

MATERIAL AND METHODS

Erythrocyte and reticulocyte-rich red blood cell suspensions of rats (Wistar albino rats of 250-350 g body mass) were used in this study. Reticulocytosis was induced by phenylhydrazine hydrochloride treatment (35 mg/kg body mass during three days) (Kostić *et al.*, 1990). After 7-8 days, rats were anaesthetized by ether, and blood was taken by exanguination. Reticulocytes amounted to 86.57±1.28 %. Three times washed red blood cells were resuspended in incubation buffer containing: 50 mM Hepes, 100 mM NaCl, 1 mM MgCl₂, 1 mM NaH₂PO₄, 5

mM glucose and 2 mM CaCl₂, pH 7.4 at 37°C (Kostić *et al.*, 1990). Cell suspensions (final hematocrit value about 0.20) were aerobically incubated for 2 hours, without (control), or in the presence of different concentrations of SNP: 0.1, 0.25, 0.5, 1.0 and 1.5 mM. The SNP was added at the beginning of incubation (0 min). Samples extractions were carried out after incubation.

Concentrations of RNS and ROS were determined in L-Arg extracts: 1/2 vol 3 M perchloacetic acid and 2 vol 20 mM EDTA were added to 1 vol cell suspension. After extraction on ice (15 min) and centrifugation 4 min/15 000 rpm, extracts were neutralized by 2 M K₂CO₃. Spectrophotometric determination of nitrites (NO+ indicator) (Green et al., 1982), hydroxylamine (NO+ indicator) (Arnelle and Stamler, 1996), S-nitrosothiols (SNO) (Green et al., 1982), 3-nitrotyrosine - (peroxynitrite indicator) (RIORDAN and VALLEE, 1972), superoxide anion (O₂-) (Auclair and Voisin, 1985) and hydrogen peroxide (H₂O₂) concentrations (Pick and Keisari, 1980) were performed in L-Arg extracts.

All values are expressed as mean \pm SEM. Statistical evaluation was calculated by Student's t-test for paired observations. For all comparisons p < 0.05 was considered as significant.

RESULTS AND DISCUSSION

Results presented in Figure 1 showed that SNP did not alter nitrite level, while significantly (p < 0.05) increased concentrations of hydroxylamine, S-nitrosothiols and 3-nitrotyrosine in rat erythrocytes. Literature data (Hou et al., 1999) showed that SNP as transition metal NO complex, spontaneously releases NO, mainly as NO⁺ ion. However, according to results of our study, there is dominant generation of nitroxyl anion (NO⁻) due to SNP metabolism in rat erythrocytes. These data have new implications, because NO⁻ ion has discrete chemistry versus the other RNS and accomplish strong cytotoxic effects (MIRANDA et al., 2003).

Concentration of O_2 decreased (p < 0.05) in the presence of low doses of SNP only (Fig. 2). These data is probably consequence of peroxynitrite generation (Fig. 1). Namely, NO in the reaction with O_2 , as well as NO ion in reaction with O_2 generated peroxynitrite, which damaged almost every molecule in cells (WINK and MITCHELL, 1998). Level of H_2O_2 increased in dose-dependent manner in the presence of SNP (Fig. 2), indicating oxidative stress (SIES, 1991) that occurs in rat erythrocytes.

In rat reticulocytes SNP significantly increased nitrite, hydroxylamine and 3-nitrotyrosine concentrations (Fig. 3). Apparently, SNP is metabolized to NO ion in reticulocytes, too. Generation of RNS, indicate SNP-induced nitrosative stress in these cells. In addition, low doses of SNP induced decrease of O2-level, while high doses of SNP even increased O2- concentration (Fig. 4). Concentration of H2O2 did not alter in rat reticulocytes (Fig. 4). These alterations of ROS is consequence of SNP-induced inhibition of Cu, Zn-containing superoxid dismutase and Mn-containing superoxid dismutase (MALETIĆ *et al.*, unpublished data).

Our previous study has been showed SNP-induced inhibition of the process of oxidative phosphorylation in rat reticulocyte mitochondria (MALETIĆ *et al.*, 2004) and stimulation of the process of glycolysis in rat reticulocytes (MALETIĆ *et al.*, 2004) and erythrocytes (MALETIĆ *et al.*, 2000). Data presented now (current study) indicate that SNP-induced alteration of energy production in rat red blood cells are mediated dominantly by NO ion, as well as RNS and H₂O₂ (Fig. 1, 2, 3, 4).

On the basis of obtained data, we can conclude: SNP spontaneously liberated nitric oxide as NO ion in rat erythrocytes and reticulocytes. In addition, applied experimental doses of SNP induced strong nitrosative and oxidative stress in these cells.

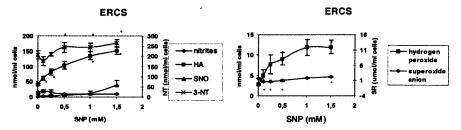


Fig. 1. - Alterations of nitrite, hydroxylamine, Snitrosothiols and 3-nitrotyrosine concentrations in the presence of SNP in rat erythrocytes Values represent mean ± SEM for 4 experiments *p < 0.05, control (0 mM SNP) versus SNP (other concentrations)

Fig. 2. - Alterations of superoxide anion radical and hydrogen peroxide concentrations in the presence of SNP in rat erythrocytes

Values represent mean ± SEM for 4 experiments

*p < 0.05, control (0 mM SNP) versus SNP (other concentrations)

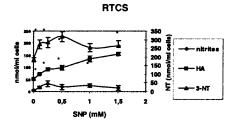
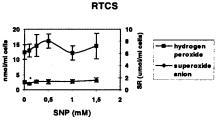


Fig. 3. - Alterations of nitrite, hydroxylamine and 3-nitrotyrosine concentrations in the presence of SNP in rat reticulocytes

Values represent mean ± SEM for 4 experiments

*p < 0.05, control (0 mM SNP) versus SNP (other concentrations)



and hydrogen peroxide concentrations in the presence of SNP in rat reticulocytes

Values represent mean ± SEM for 4 experiments

*p < 0.05, control (0 mM SNP) versus SNP (other concentrations)

Fig. 4. - Alterations of superoxide anion radical

REFERENCES

- ARNELLE, L. and STAMLER, J. (1996): Detection of hydroxylamine. *In*: Feelisch, M. and Stamler, J.S. (eds)., Methods in Nitric Oxide Research. Wiley, London, p. 541-552.
- AUCLAIR, C. and VOISIN, E. (1985): Nitroblue tetrazolium reduction. *In*: Greenwald, R.A. ed., Handbook of Methods for Oxygen Radical Research. CRC Press, Inc, Boka Raton, p. 123-132.
- DROGE, W. (2002): Free radicals in the physiological control of cell function. Physiol. Rev. 82: 47-95.
- Green, L. C., Wagner, D. A., Glogowski, J., et al. (1982): Analysis of nitrate, nitrite and [15N]nitrate in biological fluids. Anal. Biochem. 126: 131-138.
- HOU, Y. C., JANEZUK, A., and WANG, P. G. (1999): Current trends in the development of nitric oxide donors. Curr. Pharm. Des. 15: 417-441.
- Kostić, M. M., ŽIVKOVIĆ, R. V., and RAPOPORT, S. M. (1989): Maturation-dependent changes of the rat reticulocyte energy metabolism and hormonal reponsiveness. Biomed. Biochim. Acta. 49: 178-182.
- MALETIĆ, S. D., DRAGIĆEVIĆ-ĐOKOVIĆ, LJ. M., ŽIKIĆ, R. V., et al. (2000): Effects of nitric oxide donors on energy metabolism of rat erythrocytes. J Environ Pathol. Toxicol. Oncol. 19: 383-390.
- MALETIĆ, S. D., DRAGIĆEVIĆ-ĐOKOVIĆ, LI. M., OGNJANOVIĆ, B.I., et al. (2004): Effects of exogenous donor of nitric oxide sodium nitroprusside on energy production of rat reticulocytes. Physiol. Res. 53: (In Press).
- MIRANDA, K. M., NIMS, R.W., THOMAS, D. D., et al. (2003): Comparasion of the reactivity of nitric oxide and nitroxyl with heme proteins. A chemical discussion of the differential biological effects of these redox related products of NOS. J. Inorg. Biochem. 93: 52-60.
- PICK, E. and KEISARI, Y. (1980): A simple colorimetric method for the measurement of hydrogen peroxide produced by cells in culture. J. Immunol. Meth. 38: 161-70.
- RAPOPORT, S. M. (1986): The reticulocyte. CRC Press Inc., Bocca Raton, Florida.
- RIORDAN, J. F. and VALLEE, B. L. (1972): Nitration with tetranitromethane. *In*: Hirs, C. H. W. and Timasheff, S.N. (eds)., Methods in Enzymology. Academic Press, New York, Vol. 25. p. 515-521.
- SIES, H. (1991): Oxidative stress: oxidants and antioxidants. Academic Press, London.
- WINK, D. A. and MITCHELL, J. B. (1998): Chemical biology of nitric oxide: insights into regulatory, cytotoxic, and cytoprotective mechanisms of nitric oxide. Free Radic. Biol. Med. 25: 434-456.

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METABOLIZAM NATRIJUM NITROPRUSIDA (SNP) U CRVENIM KRVNIM ĆELIJAMA PACOVA

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Natrijum nitroprusid (SNP) je lek koji se klinički koristi u tretmanu hipertenzija. Cili ovog rada je da se objasne molekularni mehanizmi delovanja SNPa, sa posebnim akcentom na metabolizam ovog leka u eritrocitima i retikulocitima pacova. Suspenzije eritrocita i crvenih krvnih ćelija bogate retikulocitima su aerobno inkubirane bez (kontrola) ili u prisustvu različitih koncentracija SNPa (0.1, 0.25, 0.5, 1.0 i 1.5 mM). Koncentracije reaktivnih vrsta azota (RNS) i kiseonika (ROS) su određivane nakon inkubacije. U eritrocitima pacova. SNP ne menja koncentraciju nitrita (indikator NO+ jona), dok značajno povećava koncentracije hidroksilamina (indicator NO jona), S-nitrozotiola (SNO) and 3-nitrotirozina (indikator peroksinitrita). Koncentracija superoksid anjon radikala (O₂·) je smanjena samo u prisustvu niskih doza SNP, dok je nivo vodonik peroksida (H2O2) povećan na dozno-zavisan način u eritrocitima pacova. S druge strane, SNP značajno povećava koncentracije nitrita, hidroksilamina i 3nitrotirozina u retikulocitima pacova. Niske doze SNPa indukuju smanjenje O₂nivoa. Koncentracije H₂O₂ nisu promenjenje u retikulocitima pacova. Na osnovu iznetih podataka, možemo zaključiti: SNP spontano oslobađa azot monoksid kao NO jon u eritrocitima i retikulocitima pacova. Primenjene eksperimentalne doze SNPa indukuju snažan nitrozacioni i oksidacioni stress u ovim ćelijama.

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