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# Repellent activity of *Glycosmis* plant extracts against two stored product insects

[Actividad repelente de extractos de plantas *Glycosmis* contra dos insectos de productos almacenados]

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**Abstract:** In the present study, the repellent activities of the leaf and/or stem crude extracts of *Glycosmis lucida* Wall. ex Huang, *G. craibii* var. *glabra*, *G. craibii* Tanaka, *G. oligantha* Huang, *G. pentaphylla* (Retz) Correa. and *G. esquirolii* (Levl.) Tanaka were analyzed by using assays on petri dishes against *Tribolium castaneum* and *Liposcelis bostrychophila*. The leaf and stem extracts of *G. lucida*, *G. craibii* var. *glabra*, *G. craibii* Tanaka, *G. oligantha* and *G. esquirolii* possessed significant repellent activities against *T. castaneum*, the same level repellent with the positive control, DEET. However, the extracts of *G. pentaphylla*, no repellency but some insect attractant was observed. Moreover, they also showed repellent activities against *L. bostrychophila*. These results indicate that extracts from *G. lucida* and *G. oligantha* leaf could be a source of novel repellent against insects.

Keywords: extraction, Glycosmis, repellency, Tribolium castaneum, Liposcelis bostrychophila

**Resumen:** En el presente estudio, las actividades repelentes de la hoja y/o tronco de los extractos crudos de *Glycosmis lucida* Wall. ex Huang, *G. craibii* var. *glabra*, *G. craibii* Tanaka, *G. oligantha* Huang, *G. pentaphylla* (Retz) Correa y *G. esquirolii* (Levl.) Tanaka se analizaron mediante el uso de ensayos en placas de Petri contra *Tribolium castaneum* y *Liposcelis bostrychophila*. Los extractos de las hojas y tallo de *G. lucida*, *G. craibii* var. *glabra*, *G. craibii* Tanaka, *G. oligantha* y *G. esquirolii* poseían actividades repelentes significativas contra *T. castaneum*, el mismo nivel repelente del control positivo, el DEET. Sin embargo, los extractos de *G. pentaphylla*, no se observó la repelencia pero sí actividad atrayente de insectos. Por otra parte, también se mostraron las actividades repelentes contra *L. bostrychophila*. Estos resultados indican que los extractos de hojas de *G. lucida* y *G. oligantha* podrían ser una fuente de repelente contra los insectos.

Palabras clave: Extracción, Glycosmis, repelencia, Tribolium castaneum, Liposcelis bostrychophila.

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### **INTRODUCTION**

Antagonistic storage has been used as one of traditional Chinese medicinal materials conservation methods. It mainly utilizes some traditional Chinese medicinal materials having special volatile odor to store with medicinal materials vulnerable to insects, so as to prevent the insects. According to our survey of literatures, many cases which use this method to prevent the insects were reported (Xia et al., 2000; Wang, 2003; Han et al., 2004; Luo et al., 2012). In order to inherit and develop the traditional method of prevention and control of stored grain insects, we took five Glycosmis species as research object and two traditional Chinese medicinal materials storage (Tribolium castaneum and insects Liposcelis bostrychophila) as the target insects. It was expected that this research work would provide some theoretical basis for the conception of antagonistic storage.

The red flour beetle (Tribolium castaneum Herbst) (Coleoptera: Tenebrionidae) is world-widely distributed and among the most economically important stored product pests (Garcia et al., 2005). Infestations can also result in elevated temperature and moisture conditions that lead to an accelerated growth of molds, including toxigenic species (Magan 2003). The booklouse (Liposcelis et al., *bostrychophila* Badonnel) (Psocoptera) is an important emerging pest of stored products. It feed on a wide variety of foods, but they are particularly associated with amylaceous products, such as grain and flour (Nayak, 2006; Throne et al., 2006; Zhou et al., 2012). It is widely distributed in several tropical and sub-tropical countries in Asia, Europe, North America, South America, Africa and Australia (Turner, 1999; Li et al., 2011). This insect's parthenogenic mode of reproduction coupled with its short life cycle in favourable conditions make it particularly troublesome, as it can rapidly infest susceptible commodities (Fisher, 1985; Mills et al., 1992; Phillips & Throne, 2010). Infestations of stored product insects could be controlled by insecticidal treatment of commodities and surfaces (Zettler & Arthur, 2000). However, increasing attention is being given to the development of more ecologically and economically sustainable control methods for storedproduct pests, mainly because of the problems already mentioned, for example, the management of the habitat and the use of semiochemical-baited traps, repellents or of biological control (Phillips & Throne, 2010).

It is well known that Rutaceae are used in many countries for their repellent properties against several insects (Venkatachalam & Jebanesan, 2001; Ngassoum et al., 2007; Fogang et al., 2012; Mehmood et al., 2012: Conti et al., 2013: Ali et al., 2013). The Glycosmis genus is a species of shrub or dungarunga of the Rutaceae family and distributes in south and southest of Aisa, northest of Australia and mainly located in south of nanling, south of Yunnan and southeast of Tibet in China (Huang, 1997). A survey of literature has shown that the chemical composition of genus Glycosmis possessed the insecticide activities against several insects (Greger et al., 1996; Latha & Joseph, 1999). Among the Glycosmis genus, the repellency of G. pentaphylla essential oil against the insect was evaluated (Khan et al., 1983; Pandey et al., 2013). However, little is known about the effectiveness of other Glycosmis species extract against stored product insects.

To our knowledge, nothing was reported on the repellent activity of *G lucida*, *G craibii* var. glabra, *G craibii* Tanaka, *G oligantha*, *G pentaphylla* and *G esquirolii* extracts against *T. castaneum* and *L. bostrychophila*. In this study, their repellent activity against the red flour beetle and the booklouse were investigated for the first time.

# MATERIAL AND METHODS

#### Plant materials

The leaf and stem samples of G. lucida, G. craibii var. glabra, G. craibii Tanaka, G. oligantha, G. pentaphylla and G. esquirolii plants were collected in Xishuangbanna, Yunnan province, China (northern latitude: 21°08'~22°36'; longitude: east 99°56'~101°50'), between 2011 June and 2013 June. The specimens were identified by Dr. LX Zhang, Yunnan branch of the Institute of Medicinal Plant Development (IMPLAD), Chinese Academy of Medical Sciences, Yunnan, China, and the voucher specimens were deposited at the Herbarium (BNU) of the College of Resources Science and Technology, Beijing Normal University.

#### Extraction

The plant samples were dried naturally for a week. The dried plant samples (100 g) were ground to powders using a grinding mill (High speed grinder 6202, Beijing huanya tianyuan machinery technology co. LTD, Beijing, China) and extracted thrice with methanol (800 mL, 30 min each) at room temperature by using ultrasonic irradiation. All extracts were filtered through Whatman N° 1 filter paper,

#### Insects

The red flour beetle, *T. castaneum* was obtained from laboratory cultures maintained for the last 2 years in the dark, in incubators at 28-30° and 70-80% relative humidity. The beetles were reared on wheat flour mixed with yeast (10:1, w/w) at 12-13% moisture content. Unsexed adult beetles used in the experiments were about 2 weeks old.

The booklouse, *L. bostrychophila* was obtained from laboratory cultures maintained for the last 2 years in the dark, in incubators at  $29-30^{\circ}$  and 70-80% relative humidity and was reared on a 1:1:1 mixture, by mass, of milk powder, active yeast and flour. All containers housing insects and the Petri dishes used in experiments were made escape proof with a coating of polytetrafluoroethylene (Fluon, *Sino-Rich*, Beijing, China). Insects used in the experiments were 1 week old.

#### Repellent activity bioassay

The repellent effects of the essential oil against *T. castaneum* and *L. bostrychophila* were assessed by

using assays on petri dishes (Chaubey, 2007). Petri dishes 9 cm in diameter were used to confine beetles during the experiment. The extracts of G. lucida, G. craibii var. glabra, G. craibii Tanaka, G. oligantha, G. pentaphylla and G. esauirolii were prepared in ethanol (157.26, 31.45, 6.29 nL/cm<sup>2</sup>), and absolute ethanol was used as the control. Filter paper 9 cm in diameter was cut in half and 500 µL of each concentration was applied separately to half of the filter paper as uniformly as possible with a micropipette. The other half (control) was treated with 500 µL of absolute ethanol. Both the treated half and the control half were then air-dried to evaporate the solvent completely (2 min). A full disk was carefully remade by attaching the tested half to the negative control half with tape. Each reassembled filter paper after treatment with solid glue was placed in a petri dish with the seam oriented in one of four randomly selected different directions to avoid any insecticidal stimuli affecting the distribution of insects. Twenty insects were released in the center of each filter paper disk, and a cover was placed over the petri dish. Five replicates were used, and the experiment was repeated three times. Counts of the insects present on each strip were made after 2 and 4 h. The percent repellency (PR) of each volatile oil was then calculated using the formula:

## **PR** (%) = $[(N_c - N_t) / (N_c + N_t)] \times 100$

Where  $N_c$  is the number of insects present in the negative control half and  $N_t$  is the number of insects present in the treated half. The averages were then assigned to different classes (0 to V) using the following scale (percentage repellency) (Liu & Ho, 1999). Class, % repellency: 0, >0.01 to <0.1; I, 0.1-20.0; II, 20.1-40.0; III, 40.1-60.0; IV, 60.1-80.0; and V, 80.1-100.

As for the booklouse, petri dishes and filter papers were changed to 6 cm in diameter and the concentration of the oil used in the experiments were 126.32, 25.26, 5.05 nL/cm<sup>2</sup>. The half filter paper was treated with 150  $\mu$ L of the solution. As a positive control, a commercial repellent DEET (*N*, *N*-diethyl-3-methylbenzamide), was used under the conditions as the oil. Analysis of variance (ANOVA) and LSD test were conducted by using SPSS statistics 20 for Windows 2007. Percentage was subjected to an arcsine square-root transformation before ANOVA and LSD tests.

#### **RESULTS AND DISCUSSION**

Methanol extracts were obtained before carrying out each specific biological assay. The plant extracts yields of six Glycosmis species were 1.84% to 7.80% (Table 1). The repellent activity of the leaf and/or stem extracts of medicinal plants against *T. castaneum* and *L. bostrychophila* was determined by using assays on petri dishes (Table 2 and Table 3).

According to our results, except for *G* pentaphylla, other five *Glycosmis* species extracts possessed strong repellency against *T. castaneum*, especially, the *G lucida* leaf extract displayed the highest repellency against *T. castaneum* (Table 2). When at the lowest tested concentrations of 6.29  $nL/cm^2$ , the extract of *G lucida* leaf still showed stronger repellency (94% and 84%, class V) than the positive control, DEET (78% and 68%, class IV) at 2

h and 4 h after exposure (Table 2). The extracts of *G* oligantha leaf, *G* esquirolii leaf displayed the same level repellency against *T*. castaneum compared with the positive control. When at the concentration of  $31.45 \text{ nL/cm}^2$ , the extracts of *G* craibii var. glabra leaf showed strong repellency (88%, class V) at 2 h after exposure and the *G* craibii Tanaka leaf extracts also exhibited strong repellency (90% and 88%, class

V) against *T. castaneum* at 2 h and 4 h after exposure (Table 2). With the concentration decreasing, the repellency of the *Glycosmis* species extracts also weakened gradually. However, the extracts of *G. pentaphylla* leaf and stem especially the *G. pentaphylla* stem, no repellency but some insect attractant properties against the red flour beetle were observed at 2 and 4 h after exposure (Table 2).

Table 1						
Methanol extracts of the 6 Glycosmis species						
Species	Source	Yield (%)	Family			
G. lucida	Leaf	5.48	Rutaceae			
G craibii var. glabra	Leaf	7.80	Rutaceae			
G craibii Tanaka	Leaf	1.84	Rutaceae			
	Leaf	2.66				
G oligantha	Stem	2.86	Rutaceae			
	Leaf	2.79				
G pentaphylla	Stem	4.34	Rutaceae			
~	Leaf	6.22				
G. esquirolii	Stem	5.08	Kutaceae			

At the same time, the extracts of the six Glycosmis species also possessed repellency against L. bostrychophila. When at the highest concentration of 126.32 nL/cm<sup>2</sup>, the extracts of G. lucida leaf, G. oligantha leaf possessed the same level repellency against L. bostrychophila at 2 h after exposure and the extracts of G. craibii Tanaka leaf, G. oligantha leaf, G. pentaphylla leaf, G. esquirolii leaf also displayed strong repellency against L. bostrychophila at 4 h after exposure (Table 3). While other Glycosmis species extracts possessed moderate repellency at 2 h and 4 h after exposure. With the concentration decreasing, the repellency of the six Glycosmis species extracts weakened gradually. Compared with the positive control, at the concentrations of 25.26,  $5.05 \text{ nL/cm}^2$ , the extracts of all species displayed lower level repellency against L.

*bostrychophila*. At the concentration of 5.05 nL/cm<sup>2</sup>, the extracts of G. pentaphylla leaf and G. esquirolii stem, also no repellency but some insect attractant properties against the booklouse were observed at 4 h after exposure (Table 3). Some extracts were evaluated for repellency against insects (Lou et al., 2013). In China, the extracts derived from Chinese medicinal herbs were also evaluated for insecticidal activity and repellency against the red flour beetle and other insects (e.g., Huang et al., 2007; Yao et al., 2007; Tang et al., 2009a; Tang et al., 2009b; Tang et al., 2009c; Huang et al., 2010; Wei et al., 2014). However, no reports are on bioactivity to the booklouse. In this article, the extracts of 6 Glycosmis species were evaluated for repellency against the two grain storage insects, T. castaneum and L. bostrychophila.

$\begin{array}{c c c c c c c } \label{eq:Term} \begin{tabular}{ c c c c c } \hline Pint & Pint $	Iribolium castaneum after two exposure times							
$\begin{array}{ c c c c c c c c c c c c c c c c c c c$	Treatment	Plant	2h			4h		
$ \begin{array}{c ccccccccccccccccccccccccccccccccccc$		part	157.26*	31.45*	6.29*	157.26*	31.45*	6.29*
$ \begin{array}{c ccccccccccccccccccccccccccccccccccc$	G lucida	leaf	90±6ab	86±10abc	94±6ab	88±9ab	76±21ab	84±13a
$ \begin{array}{cccccccccccccccccccccccccccccccccccc$			V	V	V	V	IV	V
$ \begin{array}{cccccccccccccccccccccccccccccccccccc$	G craibii var. glabra	leaf	94±9ab	88±10ab	60±19cd	100±0a	74±16ab	58±14bc
$ \begin{array}{c} G\ craibii\ Tanaka \\ G\ craibii\ Tanaka \\ eaf \\ \  \  \  \  \  \  \  \  \  \  \  \  \$			V	V	111	V	IV	111
$ \begin{array}{c} G \ oligan tha \\ Ha \\ $	G craibii Tanaka	leaf	58±18c	90±8ab	60±18cd	72±20c	88±11a	34±19c
$ \begin{array}{c} G \ oligan tha \\ G \ pentaphylla \\ G \ pent$			III	V	III	IV	V	II
$ \begin{array}{cccccccccccccccccccccccccccccccccccc$	G oligantha	leaf	80±15bc	76±16bc	76±21bc	86±14b	68±18b	64±23b
$ \begin{array}{c} G \ oliganiha \\ Stem \\ Stem \\ G \ pentaphylla \\ G \ pentap$		icai	IV	IV	IV	$\mathbf{V}$	IV	IV
$ \begin{array}{cccccccccccccccccccccccccccccccccccc$		stom	84±14ab	82±18abc	42±21d	92±9ab	72±16ab	30±21c
$ \begin{array}{c} G \ pentaphylla \\ G \ pentaphylla \\ G \ esquirolii \\ G \ esquirolii \\ DEET \\ \end{array} \begin{array}{c}  eaf \\  $		Stem	$\mathbf{V}$	V	III	$\mathbf{V}$	IV	II
$ \begin{array}{cccccccccccccccccccccccccccccccccccc$		loof	54±19c	76±16bc	-58±17e	48±21d	66±19b	-8±13d
$ \begin{array}{c} G \ pentaphylla \\ stem \\ G \ esquirolii \\ \end{array} \begin{array}{c} -36\pm 20d \\ stem \\ -36\pm 20d \\ -62\pm 16d \\ -76\pm 15e \\ V \\ IV \\ IV \\ IV \\ IV \\ IV \\ IV \\ IV$		lear	III	IV	-	III	IV	-
G esquirolii     leaf     82±18ab     68±15c     64±9cd     76±15c     90±12a     42±11bc       Stem     IV     IV     IV     IV     V     III       DEET     IV     V     III     IV     IV     IV	G pentaphylla	atom	-36±20d	-62±16d	-76±15e	-64±12e	-76±21c	-84±10e
$\begin{array}{c} G \ esquirolii \\ G \ esquirolii \\ DEET \end{array} \begin{array}{c ccccccccccccccccccccccccccccccccccc$		stem	-	-	-	-	-	-
$\begin{array}{cccccccccccccccccccccccccccccccccccc$	G. esquirolii	16	82±18ab	68±15c	64±9cd	76±15c	90±12a	42±11bc
Ce esquiroiti       78±18bc       88±18ab       52±12d       80±13bc       80±8a       44±17bc         IV       V       III       IV       IV       III       IV       III         DEET       100±0a       98±3a       78±14bc       96±3a       82±8a       68±5b         V       V       IV       V       IV       V       IV		lear	$\mathbf{V}$	IV	IV	IV	V	III
DEET         IV         V         III         IV         IV         III           00±0a         98±3a         78±14bc         96±3a         82±8a         68±5b           V         V         IV         V         V         IV		a <b>t</b> a	78±18bc	88±18ab	52±12d	80±13bc	80±8a	44±17bc
DEET         100±0a         98±3a         78±14bc         96±3a         82±8a         68±5b           V         V         IV         V         V         IV		stem	IV	$\mathbf{V}$	III	IV	IV	III
V V IV V IV	DEET		100±0a	98±3a	78±14bc	96±3a	82±8a	68±5b
			V	V	IV	V	V	IV

 Table 2

 Percentage repellency of the *Glycosmis* species extracts against *Tribolium castaneum* after two exposure times

\* Concentration (nL/cm<sup>2</sup>);

a-e Means in the same column followed by the same letters do not differ significantly (P < 0.05) in ANOVA and LSD tests. PR was subjected to an arcsine square-root transformation before ANOVA and LSD tests.

To our knowledge, the repellent activity of G. lucida, G. craibii var. glabra, G. craibii Tanaka, G. oligantha, G. pentaphylla and G. esquirolii has not been reported before. Previous studies have isolated carbazole alkaloid, acridone alkaloids from Gpentaphylla (Govindachari et al., 1966; Yang et al., 2012), hydroquinone diglycoside acyl esters, isoflavone diglycosides from G. pentaphylla (Wang et al., 2006a; Wang et al., 2006b) and flavanones, flavanonols from Glycosmis species (Lukaseder et al., 2009). Some compounds from Glycosmis species with antitumor. antifungal, antibacterial and insecticidal have been observed (Bhattacharyya et al., 1985; Greger et al., 1996; Quader et al., 1999; Ito et al., 2004; Sripisut et al., 2012). However, their association with repellency activity is currently

unknown. The results pointed out the potential value of the compounds from the extracts used to prevent the stored products insects.

#### CONCLUSIONS

The results have shown that the extracts of *G. lucida*, *G. craibii* var. glabra, *G. craibii* Tanaka, *G. oligantha* and *G. esquirolii* exhibit strong repellent activity against *T. castaneum*. They show potential to be developed as possible natural repellent for control of stored product insects. However, for the practical application of the extracts, further studies on the safety of the extracts to humans and on the effect of the compounds from the plant extracts are necessary to improve the efficacy.

after two exposure times							
Treatment	Plant	2h					
	part	126.32*	25.26*	5.05*	126.32*	25.26*	5.05*
G. lucida	leaf	84±10ab	58±8b	34±12bc	76±6cd	48±18bc	20±12cd
		$\mathbf{V}$	III	II	IV	III	Ι
<i>G. craibii</i> var.	leaf	78±17bc	70±8ab	38±11bc	78±10bcd	68±12abc	18±13cd
glabra	lear	IV	IV	II	IV	IV	Ι
G. craibii	loof	76±14bc	66±13ab	34±11bc	86±13bc	76±13ab	46±12bc
Tanaka	lear	IV	IV	II	$\mathbf{V}$	IV	III
G oligantha	leaf	88±8ab	58±11b	40±12bc	100±5a	74±11ab	32±18c
		$\mathbf{V}$	III	II	$\mathbf{V}$	IV	II
	stem	80±9abc	62±15ab	48±8bc	62±12d	56±15bc	60±8b
		IV	IV	III	IV	III	III
	leaf	70±17bc	52±6b	16±19de	84±14bc	58±15bc	-24±24e
G pentaphylla		IV	III	Ι	$\mathbf{V}$	III	-
	stem	70±13bc	50±7b	50±9bc	76±10cd	38±6c	10±14cd
		IV	III	III	IV	II	Ι
G. esquirolii	leaf	74±12bc	54±13b	68±17ab	88±10bc	76±13ab	36±18bc
		IV	III	IV	$\mathbf{V}$	IV	II
	stem	66±13c	28±20c	6±7e	58±14d	38±8c	-6±12de
		IV	II	Ι	III	II	-
DEET		94±6a	82±5a	86±8a	92±5ab	84±3a	82±8a
		V	V	V	V	V	V

 Table 3

 Percentage repellency of the Glycosmis species extracts against Liposcelis bostrychophila

 after two exposure times

\* Concentration (nL/cm<sup>2</sup>);

a-e Means in the same column followed by the same letters do not differ significantly (P < 0.05) in ANOVA and LSD tests. PR was subjected to an arcsine square-root transformation before ANOVA and LSD tests.

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