



Sparse single-step genomic blup in crossbreeding schemes

Vandenplas, J., Calus, M. P. L., & ten Napel, J.

This is a "Post-Print" accepted manuscript, which has been published in "Journal of Animal Science"

This version is distributed under a non-commercial no derivatives Creative Commons



([CC-BY-NC-ND](https://creativecommons.org/licenses/by-nc-nd/4.0/)) user license, which permits use, distribution, and reproduction in any medium, provided the original work is properly cited and not used for commercial purposes. Further, the restriction applies that if you remix, transform, or build upon the material, you may not distribute the modified material.

Please cite this publication as follows:

Vandenplas, J., Calus, M. P. L., & ten Napel, J. (2018). Sparse single-step genomic blup in crossbreeding schemes. *Journal of Animal Science*, 96(6), 2060-2073. DOI: 10.1093/jas/sky136

You can download the published version at:

<https://doi.org/10.1093/jas/sky136>

1 Sparse single-step

2

3 **Sparse single-step genomic BLUP in crossbreeding** 4 **schemes¹**

5

6 **J. Vandenplas,^{*2} M.P.L. Calus,^{*} J. ten Napel^{*}**

7

8 ^{*}Animal Breeding and Genomics Centre, Wageningen UR Livestock Research, P.O. Box 338,
9 6700 AH Wageningen, the Netherlands

10

11 ¹This study was financially supported by the Dutch Ministry of Economic Affairs (TKI-
12 Toeslag; Public-private partnership “Breed4Food” code BO-22.04-011-001-ASG-LR-3) and
13 the Breed4Food partners Cobb Europe, CRV, Hendrix Genetics and Topigs Norsvin.

14 ²Corresponding author: jeremie.vandenplas@wur.nl

15 We declare that we do not have any competing interest in the matter and results covered by
16 this manuscript.

17

18

19

ABSTRACT

20 The algorithm for Proven and Young animals (**APY**) efficiently computes an approximated
21 inverse of the genomic relationship matrix, by dividing genotyped animals in so-called core
22 and non-core animals. The APY leads to computationally feasible single-step genomic Best
23 Linear Unbiased Prediction (**ssGBLUP**) with a large number of genotyped animals, and was
24 successfully applied to real single breed or line datasets. This study aimed to assess the
25 quality of genomic breeding values (**GEBV**) when using the APY (**GEBV_{APY}**), in comparison
26 to GEBV when using the directly inverted genomic relationship matrix (**GEBV_{DIRECT}**), for
27 situations based on crossbreeding schemes, including F1 and F2 crosses, such as the ones for
28 pigs and chickens. Based on simulations of a three-way crossbreeding program, we compared
29 different approximated inverses of a genomic relationship matrix, by varying the size and the
30 composition of the core group. We showed that **GEBV_{APY}** were accurate approximations of
31 **GEBV_{DIRECT}** for multivariate **ssGBLUP** involving different breeds and their crosses.
32 **GEBV_{APY}** as accurate as **GEBV_{DIRECT}** were obtained when the core groups included animals
33 from different breed compositions, and when the core groups had a size between the numbers
34 of the largest eigenvalues explaining 98% and 99% of the variation in the raw genomic
35 relationship matrix.

36

37 **Key words:** single-step, genomic evaluation, APY

38

INTRODUCTION

39

40 Single-step genomic Best Linear Unbiased Prediction (**ssGBLUP**) is currently the method of
41 choice to predict genomic breeding values in many species (Legarra et al., 2014). The main
42 reason is that ssGBLUP enables simultaneous use of phenotypes from genotyped and non-
43 genotyped animals by combining genomic and pedigree relationship matrices. An
44 inconvenience of ssGBLUP is that the inverse of a dense genomic relationship matrix (**G**) is
45 required, leading to a soft limit of approximately 100,000 genotyped animals for the currently
46 available computers (Misztal et al., 2014).

47 Recently, Misztal et al. (2014, 2016) proposed the so-called Algorithm for Proven and Young
48 animals (**APY**) to compute an approximated inverse of **G** (\mathbf{G}_{APY}^{-1}) for a large number of
49 genotyped animals. The computation of \mathbf{G}_{APY}^{-1} involves the inversion of a genomic relationship
50 submatrix among a limited number of genotyped animals, called core animals, and the
51 recursive computation of other coefficients for non-core animals. The APY was successfully
52 applied on (large) real datasets with animals originating from a single breed or line
53 (Fragomeni et al., 2015; Lourenco et al., 2015; Masuda et al., 2016; Ostersen et al., 2016;
54 Pocrnic et al., 2016b; Strandén et al., 2017). However, several livestock production systems,
55 such as the ones for pigs and chickens, are based on well-structured crossbreeding schemes,
56 generating production animals with a specific breed composition. In these cases, the
57 ssGBLUP may include non-genotyped and genotyped animals from different breeds, as well
58 as their crossbred progeny. Using the APY with such datasets is desirable for implementing
59 ssGBLUP in crossbreeding schemes efficiently.

60 The aim of this study was to assess the quality of genomic estimated breeding values (**GEBV**)
61 when using \mathbf{G}_{APY}^{-1} , in comparison to GEBV when using the direct inversion of **G** (\mathbf{G}_{direct}^{-1}), for
62 situations based on well-structured crossbreeding schemes that include genotyped animals

63 from a few different breeds and their F1 and F2 crosses. Influence of the selection strategy of
64 the core animals and of the number of core animals, were also investigated. All analyses were
65 based on simulated data.

66

67 MATERIALS AND METHODS

68 *Single-step genomic Best Linear Unbiased Prediction*

69 The ssGBLUP method replaces the inverse of the pedigree relationship matrix for all animals
70 (\mathbf{A}^{-1}) with the inverse of the combined pedigree-genomic relationship matrix (\mathbf{H}^{-1}), defined
71 as (Aguilar et al., 2010; Christensen and Lund, 2010):

$$72 \mathbf{H}^{-1} = \mathbf{A}^{-1} + \begin{bmatrix} \mathbf{0} & \mathbf{0} \\ \mathbf{0} & \mathbf{G}^{-1} - \mathbf{A}_{22}^{-1} \end{bmatrix} \quad (1)$$

73 where \mathbf{A}_{22} is the pedigree relationship matrix for the genotyped animals, $\mathbf{G} = (1 - w)\mathbf{G}_a +$
74 $w\mathbf{A}_{22}$ with \mathbf{G}_a being a genomic relationship matrix adjusted to be on the same scale as \mathbf{A}_{22} ,
75 and w being the weight on the pedigree relationship matrix. Several approaches for
76 computing \mathbf{G}_a by adjusting a raw genomic relationship matrix \mathbf{G}^* towards \mathbf{A}_{22} were proposed
77 in the literature (Powell et al., 2010; Vitezica et al., 2011; Christensen, 2012; Lourenco et al.,
78 2016).

79 Highest computational costs for creating \mathbf{H}^{-1} are the creation and the inversion of the dense
80 matrices \mathbf{G} and \mathbf{A}_{22} . Additional computational costs also appear during solving of the mixed
81 model equations due to an increase of non-zero elements in \mathbf{H}^{-1} , increasing the number of
82 operations per iteration, e.g., of the preconditioned conjugate gradient used to solve the mixed
83 model equations (Ostersen et al., 2016).

84 *Sparse inversion of G*

85 The matrix \mathbf{G} can be divided into four submatrices as:

86
$$\mathbf{G} = \begin{bmatrix} \mathbf{G}_{cc} & \mathbf{G}_{cn} \\ \mathbf{G}'_{cn} & \mathbf{G}_{nn} \end{bmatrix}$$

87 where the subscript c refers to a group of genotyped animals called hereafter “core animals”,
 88 and the subscript n refers to a second group of genotyped animals called hereafter “noncore
 89 animals”.

90 Following Misztal (Misztal et al., 2014; Misztal, 2016), the inverse of \mathbf{G} , \mathbf{G}^{-1} , can be
 91 approximated using the APY as follows:

92
$$\mathbf{G}_{APY}^{-1} = \begin{bmatrix} \mathbf{G}_{cc}^{-1} + \mathbf{G}_{cc}^{-1}\mathbf{G}_{cn}\mathbf{M}^{-1}\mathbf{G}'_{cn}\mathbf{G}_{cc}^{-1} & -\mathbf{G}_{cc}^{-1}\mathbf{G}_{cn}\mathbf{M}^{-1} \\ -\mathbf{M}^{-1}\mathbf{G}'_{cn}\mathbf{G}_{cc}^{-1} & \mathbf{M}^{-1} \end{bmatrix}$$

93 where the matrix \mathbf{M} is a diagonal matrix of size of the number of noncore animals and with a
 94 diagonal element for the i^{th} noncore animal equal to $\mathbf{M}_{ii} = \text{diag}(\mathbf{G}_{nn_{ii}} - \mathbf{G}'_{ci}\mathbf{G}_{cc}^{-1}\mathbf{G}_{ci})$ with
 95 \mathbf{G}_{ci} being the i^{th} column of \mathbf{G}_{cn} . It is worth noting that the matrix \mathbf{M} is an approximation of the
 96 Schur complement of \mathbf{G}_{cc} , i.e., $\mathbf{S} = \mathbf{G}_{nn} - \mathbf{G}'_{cn}\mathbf{G}_{cc}^{-1}\mathbf{G}_{cn}$. Replacing \mathbf{M} by \mathbf{S} in the formula of
 97 \mathbf{G}_{APY}^{-1} would lead to the computation of the inverse of \mathbf{G} , \mathbf{G}^{-1} .

98 The APY only requires the computation of the submatrices \mathbf{G}_{cc} , \mathbf{G}_{cn} and of the diagonal
 99 elements of \mathbf{G}_{nn} , in addition to the inversion of the submatrix \mathbf{G}_{cc} . Thus, the computational
 100 costs of the APY are reduced in comparison to the setting up and the direct inversion of \mathbf{G} .
 101 Also, the memory costs of the APY are reduced because only submatrices, \mathbf{G}_{cc} and \mathbf{G}_{cn} ,
 102 must be stored and the matrix \mathbf{G}_{APY}^{-1} is sparse thanks to the diagonal matrix \mathbf{M}^{-1} .

103 ***Simulated data***

104 ***Populations.*** The assessment of the quality of the genomic predictions from a sparse
105 ssGBLUP in crossbreeding schemes was achieved by simulating a three-way crossbreeding
106 program with random selection (Figure 1). Simulations of historic, purebred and crossbred
107 recent populations were performed using the QMSim software (Sargolzaei and Schenkel,
108 2009). For the historic population, 70 discrete random mating generations (i.e., generations 1
109 to 70) with a constant size of 18,840 individuals with equal number of individuals from each
110 sex were simulated, followed by 10 generations (i.e., generations 71 to 80) in which the
111 effective population size was gradually reduced to 390 individuals. The next 20 generations
112 (i.e. generations 81 to 100) were simulated to gradually expand the population size to 18,840.
113 The last generation (i.e. generation 100) included 90 males and 18,750 females. Matings for
114 all generations were based on the random union of gametes, which were randomly sampled
115 from the pools of male and female gametes. To simulate the three breed populations (hereafter
116 referred to as breeds A, B, and C), three random samples were drawn from the generation 100
117 of the historic population, each including 30 males and 6,250 females. Subsequently, within
118 each breed, 100 generations (i.e. generations 101 to 200) of random mating were simulated
119 before starting the three-way crossbreeding program (Figure 1). In each of the simulated 100
120 generations of random mating, each female had one male and one female offspring.

121 In the second step, a three-way crossbreeding program was simulated (Figure 1). Purebred
122 (i.e., A, B, and C) animals that were used as founders of the pedigree (i.e., the first generation
123 of the pedigree) were from generations 200. For each breed, A, B, and C, the next 9 discrete
124 generations (i.e. generations 201 to 209) of purebred animals were simulated by means of
125 random selection and matings while maintaining a constant size of 30 males and 6,250
126 females. For mimicking a three-way crossbreeding program, from the generation 205 until the

127 generation 208, B and C purebred animals were randomly crossed to produce four generations
128 (i.e. generations 206 to 209) of F1 animals, that is 30 BC crossbred males and 6,250 BC
129 crossbred females. These BC crossbred animals were then randomly mated to males from
130 breed A to produce four generations (i.e. generations 206 to 209) of F2 animals, called A(BC)
131 crossbred animals. For each generation, 6,280 A(BC) crossbred animals were simulated
132 (Figure 1). Purebred animals that were used as parents of crossbred animals could also be
133 parents of purebred animals in the next generation. A total of 5 replicates were simulated
134 using the QMSim software.

135 **Genotypes.** The genome was simulated using the QMSim software, simultaneously with the
136 simulation of the historic, purebred and crossbred recent populations. The genome consisted
137 of 18 chromosomes designed to resemble the Sus Scrofa genome with a SNP density that was
138 comparable to that of a 60k SNP chip. The SNP positions were randomized across the
139 genome and a recurrent mutation rate of 2.5×10^{-5} , as well as 1 mean crossover per 1 Morgan,
140 were assumed. All SNPs that segregated in the last historical generation (i.e., generation 100)
141 and with a minor allele frequency (MAF) higher than or equal to 0.05 were selected and used
142 to simulate the genotypes of the purebred and crossbred animals. In addition to the SNPs,
143 4,500 QTL were simulated, and their positions were also randomized across the genome.
144 Mutation rate and MAF of the QTL were the same as the ones for the simulated SNPs.

145 **Phenotypes.** For all purebred and crossbred animals, phenotypes for the breed composition to
146 which they belonged were simulated under additive gene action using a custom Fortran
147 program. This resulted in five traits: one trait for each of the purebred performances A, B and
148 C, and one trait for each of the crossbred performances BC and A(BC). Genetic correlations
149 between traits were randomly sampled in the range [0.2-0.8] from a uniform distribution.
150 Simulated genetic correlations between purebred and crossbred traits were in the lowest range

151 of reported values in the literature as reviewed by Wientjes and Calus (2017) (Table 1).
152 Heritabilities (h_i^2) were randomly sampled in the range of [0.2-0.4] from a uniform
153 distribution. Residual covariances were set to zero, as they would be in practice, because each
154 animal has a phenotype for one of the five traits only. The same genetic correlations and
155 heritabilities were used in all replicates, and are reported in Table 1.

156 For each animal and for each of the five traits, a true breeding value (TBV) was simulated by
157 summing a polygenic effect and the multiplication of the simulated allele substitution effects
158 with the genotypes of the 4,500 QTL coded as 0, 1 and 2. This genotype multiplication
159 allowed different genetic levels across breeds for the same trait because QTL allele
160 frequencies differ across breeds. For each trait, the polygenic effect of each individual was
161 equal to the sum of the average of polygenic effects of the parent and a Mendelian sampling
162 term. The Mendelian sampling terms for the five traits were sampled from a multinormal
163 distribution with means of 0 and variances equal to the Mendelian sampling variances
164 (Mrode, 2005). Correlations between the simulated Mendelian sampling terms were assumed
165 to be equal to the genetic correlations. The variance of the polygenic effect of each i^{th} trait
166 was assumed to be equal to 5% of the total additive genetic variance (σ_{Ai}^2).

167 The allele substitution effects of QTLs were sampled from a multinormal distribution with
168 means of 0, and variances of 1. The correlations between allele substitution effects of the QTL
169 underlying the 5 traits were equal to the genetic correlations. For each trait, the genetic
170 variance explained by all QTLs was computed as the sum of the variances across all QTLs,
171 assuming no correlation between the QTLs. The simulated additive genetic variance of each
172 j^{th} QTL was calculated as $\sigma_{gj}^2 = 2p_j(1 - p_j)a_j^2$, where p_j is the allele frequency and a_j is the
173 allele substitution effect of j^{th} QTL. For each trait, the allele substitution effects were rescaled
174 to obtain an additive genetic variance explained by the QTLs (σ_g^2) equal to 1. The part of the

175 total additive genetic variance explained by the QTLs was assumed to be equal to 95% for
176 each i^{th} trait. Finally, the phenotypes for each trait for each animal were generated by
177 summing the TBV and a residual error sampled from a normal distribution with a mean 0 and
178 a variance equal to $\left(\frac{1}{h_i^2} - 1\right) * \sigma_{Ai}^2$.

179 **Datasets.** For all the analyses, the pedigree included all the animals simulated for the creation
180 of the three-way crossbreeding program. The phenotype dataset included 126,000 records.
181 Among all records, 100,000 records were associated with purebred (i.e. A, B, and C) animals
182 randomly sampled among all purebred animals from generations 204 until 208. A total of
183 16,000 records were associated with A(BC) crossbred animals randomly sampled among all
184 A(BC) crossbred animals from generations 206 until 209. Finally, 10,000 records were
185 associated with BC crossbred dams. Average numbers of purebred and crossbred animals per
186 generation with a phenotype are given in the E-Supplements Table S1.

187 The genotype dataset included 89,000 genotypes. This included all 26,000 phenotyped BC
188 and A(BC) crossbred animals. A total of 48,000 genotypes were from purebred (i.e. A, B, and
189 C) animals randomly sampled among all purebred animals from the generations 205 until 208,
190 regardless whether they had a phenotype or not. A total of 15,000 genotypes were from
191 purebred (i.e. A, B, and C) animals randomly sampled among all purebred animals from
192 generation 209. These 15,000 animals did not have phenotypes and are hereafter considered
193 as selection candidates. Average numbers of purebred and crossbred animals per generation
194 with a phenotype and a genotype are given in the E-Supplements Table S2.

195

196 ***Model and scenarios evaluated***

197 Five-trait ssGBLUP was performed. The model for the i^{th} trait ($i = A, B, C, BC, A(BC)$) was
 198 as follows:

$$199 \mathbf{y}_i = \mathbf{1}\mu_i + \mathbf{W}_i\mathbf{a}_i + \mathbf{e}_i$$

200 where, for the i^{th} trait, \mathbf{y}_i is the vector of records, μ_i is the general mean, \mathbf{a}_i is the vector of
 201 additive genetic effects, \mathbf{e}_i is the vector of residuals, the vector $\mathbf{1}$ is a vector of 1's relating the
 202 records to the general mean, and \mathbf{W}_i is an incidence matrix relating the records to the animals.
 203 The variance components used for the simulations were used for the five-trait ssGBLUP. The
 204 vector of additive genetic effects $\mathbf{a} = [\mathbf{a}'_A \quad \mathbf{a}'_B \quad \mathbf{a}'_C \quad \mathbf{a}'_{BC} \quad \mathbf{a}'_{A(BC)}]'$ followed a multivariate
 205 normal (MVN) distribution $MVN(\mathbf{0}, \mathbf{H}^{-1} \otimes \mathbf{\Gamma})$ where \otimes is the Kronecker product, $\mathbf{\Gamma}$ is the
 206 additive genetic (co)variance matrix, and the vector of residuals $\mathbf{e} =$
 207 $[\mathbf{e}'_A \quad \mathbf{e}'_B \quad \mathbf{e}'_C \quad \mathbf{e}'_{BC} \quad \mathbf{e}'_{A(BC)}]'$ followed a MVN distribution $MVN(\mathbf{0}, \mathbf{I} \otimes \mathbf{R})$ where \mathbf{R} is the
 208 residual (co)variance matrix.

209 Using all the 89,000 genotypes, the matrix \mathbf{G} required for the computation of \mathbf{H}^{-1} was
 210 computed without breed-specific adjustments, as suggested by Lourenco et al. (2016). This
 211 matrix was equal to $\mathbf{G} = 0.95\mathbf{G}_a + 0.05\mathbf{A}_{22}$ with the adjusted genomic relationship matrix \mathbf{G}_a
 212 computed as follows:

$$213 \mathbf{G}_a = (1 - \bar{f}_p)\mathbf{G}^* + 2\bar{f}_p\mathbf{J}$$

214 where \mathbf{G}^* is a raw genomic relationship matrix computed following the first method of
 215 VanRaden (2008) using current allele frequencies computed from all genotyped animals, \mathbf{J} is a
 216 matrix of ones, and \bar{f}_p is the average pedigree inbreeding coefficient across (core) genotyped
 217 animals. The matrix \mathbf{H}^{-1} was constructed in two different ways. First, the complete \mathbf{G} was
 218 directly inverted to obtain \mathbf{G}_{direct}^{-1} . Second, \mathbf{G}_{direct}^{-1} was replaced by \mathbf{G}_{APY}^{-1} . Because the APY

219 relies on the size and the composition of the set of core animals (Misztal et al., 2014), we
220 investigated different numbers of core animals and different strategies to select the core
221 animals. For all the strategies, the selection candidates were allowed to be considered as core
222 animals. The number of core animals were 4,000, 6,000, 8,000, 10,000, and 13,000. For each
223 size, four different strategies were applied to select the core animals. The core animals were
224 randomly sampled 1) among all breed A genotyped animals (called “Breed A”), 2) among all
225 purebred genotyped animals (called “Purebred”), or 3) among all purebred and crossbred
226 genotyped animals (called “Purebred + Crossbred”). For the fourth strategy, a QR
227 decomposition with pivoting of the transposed genotype matrix was applied to the animals.
228 The QR decomposition with pivoting returns a permutation matrix such that the diagonal
229 elements of the upper triangular matrix \mathbf{R} are decreasing (Golub and Van Loan, 1996). The
230 genotyped animals corresponding to the highest diagonal elements of the matrix \mathbf{R} were
231 chosen as core animals (called “QR”). The aim of this fourth strategy was to select core
232 animals such that the conditioning of the mixed model equations was improved, resulting in
233 faster convergence, in comparison to the other three strategies (Fernando et al., 2016). All
234 computations and analyses were run using our own custom programs for QR decomposition
235 and statistical analyses, `calc_grm` (Calus and Vandenplas, 2016) for the computation of the
236 different relationship matrices (i.e., \mathbf{G}_{direct}^{-1} , \mathbf{G}_{APY}^{-1} , and \mathbf{A}_{22}^{-1}), and MiXBLUP (ten Napel et al.,
237 2016) for predicting the different GEBV. The matrices $(\mathbf{G}_{direct}^{-1} - \mathbf{A}_{22}^{-1})$ and $(\mathbf{G}_{APY}^{-1} - \mathbf{A}_{22}^{-1})$
238 were provided to MiXBLUP as external matrices.

239 *Criteria*

240 We evaluated the prediction of GEBV of genotyped selection candidates for the purebred A,
241 B, and C performances and the crossbred A(BC) performances, for each set of core animals
242 and each breed separately. Three criteria were computed from the GEBV of the selection

243 candidates. First, the ratios between the accuracies of $GEBV_{APY}$ from alternative core groups
244 and the accuracies of $GEBV_{DIRECT}$ (i.e., from \mathbf{G}_{direct}^{-1}), were computed. Accuracies were
245 computed as the Pearson correlation between GEBV and TBV. A ratio of accuracies smaller
246 than 1 means that $GEBV_{APY}$ is less accurate than $GEBV_{DIRECT}$. Second, regression
247 coefficients of TBV on $GEBV_{APY}$ and on $GEBV_{DIRECT}$ (hereafter called bias) were computed.
248 Third, ratios between mean squares errors (MSE) of $GEBV_{APY}$ and MSE of $GEBV_{DIRECT}$,
249 were computed. The MSE were computed as the mean of the squared differences between
250 GEBV and TBV. All results were averaged across five replicates. Tukey's honest significant
251 difference test (Tukey, 1949) was used to assess significance of differences between scenarios
252 at a 5% significance level.

253 For situations with single breeds, the number of required core animals that gives accurate
254 GEBV, can be determined as the number of largest eigenvalues explaining 98-99% of the
255 variation in \mathbf{G}^* (Misztal, 2016; Pocrnic et al., 2016a; b). For investigating this relationship in
256 situations involving multiple breeds and their F1 and F2 crosses, we computed the numbers of
257 eigenvalues that explained 98% and 99% of the variation in \mathbf{G}^* that included all the 89,000
258 genotyped purebred and crossbred animals. Computations were performed with `calc_grm`
259 (Calus and Vandenplas, 2016). For each scenario, the number of eigenvalues were compared
260 to the number of core animals needed such that the accuracies of $GEBV_{APY}$ were equal to or
261 higher than 99% of the accuracy for $GEBV_{DIRECT}$ for both purebred and crossbred
262 performance traits.

263

264

RESULTS

265 *Characteristics of simulated data*

266 The simulation yielded three breeds, A, B, and C, that were highly separated, as shown by the
267 projections of genomic relationships into the two first eigenvectors for the first replicate
268 (Figure 2). The estimated global Wright's F_{st} statistics, that is a measure to quantify the level
269 of genetic differentiation between the breeds, was equal to 0.35 on average across the five
270 replicates. The global Wright's F_{st} statistics were estimated from the genotypes of all purebred
271 animals of the generation 204 with the software Genepop (4.2) (Raymond and Rousset, 1995;
272 Rousset, 2008). The mean absolute difference in allele frequencies between breeds was about
273 0.34 on average across the five replicates. All these observations suggest three genetically
274 divergent populations. The average linkage disequilibrium, expressed as r^2 (Hill and
275 Robertson, 1968), between adjacent SNP pairs with $MAF > 0.05$ and across chromosomes,
276 was 0.25 for the three breeds on average across the five replicates. Genomic relationship
277 matrices required for the singular value decomposition and genomic predictions were based
278 on 52,518 SNPs on average across the five replicates.

279 *Composition of the core groups*

280 Four selection strategies were applied to compose the core groups: (1) the core animals were
281 randomly selected among only breed A animals, (2) the core animals were randomly selected
282 among purebred animals of breed A, B, and C, (3) the core animals were randomly selected
283 among purebred animals of breed A, B, and C, and crossbred BC and A(BC) animals, and (4)
284 the core animals were selected based on a QR decomposition of the genotype matrix. For the
285 four selection strategies, Figure 3 shows the proportions of core animals across the
286 generations and across the breed compositions of a randomly chosen replicate for the scenario
287 with 8,000 core animals. Similar results were obtained for the other replicates and sizes of
288 core groups. Proportions of core animals were similar across the generations, and across the
289 breed compositions for the first three selection strategies. For the selection strategy based on

290 QR decomposition, core animals were unequally spread across all generations and breed
291 compositions: the highest proportions of core animals selected within a generation and a breed
292 composition were observed among the crossbred A(BC) animals and the first generation of
293 genotyped purebred animals (Figure 3).

294 *Quality of GEBV with G_{direct}^{-1}*

295 On average 5,000 genotyped selection candidates per breed were considered for computing
296 accuracy, bias, and MSE (Table 2). For purebred performance, the accuracies were between
297 0.79 and 0.81. For crossbred performance, the accuracies were between 0.63 and 0.71. All
298 sets of GEBV were almost unbiased (i.e., values for bias were close to 1) and had values of
299 MSE close to 0 (Table 2).

300 *Quality of GEBV with only breed A core animals*

301 When the core groups included only breed A animals, $GEBV_{APY}$ were predicted as accurately
302 as $GEBV_{DIRECT}$ for the breed A selection candidates for both purebred and crossbred
303 performance traits, as shown by the ratios between the accuracies of $GEBV_{APY}$ and of
304 $GEBV_{DIRECT}$ (Figure 4). In addition, $GEBV_{APY}$ were unbiased, and MSE was close to 0
305 (Figure 4; Table 3; Table 4; E-Supplements Tables S3-S6). However, $GEBV_{APY}$ were less
306 accurate and more biased than $GEBV_{DIRECT}$ for the breed B and breed C selection candidates,
307 as shown by low ratios of accuracies, and high values for bias and ratios of MSE of
308 $GEBV_{APY}$ (Figure 4; Table 3; Table 4; E-Supplements Tables S3-S6). Across core groups,
309 $GEBV_{APY}$ were from 18% to 40% less accurate than $GEBV_{DIRECT}$, and MSE of $GEBV_{APY}$
310 were between 16 and 81% higher than the corresponding MSE of $GEBV_{DIRECT}$.

311 *Quality of GEBV with core animals of different breed compositions*

312 Based on the three performance criteria, ratios of accuracies, bias, and ratios of MSE, the
313 scenarios with core animals of different breed compositions outperformed the scenarios with
314 only breed A core animals for both purebred and crossbred performance traits. Use of core
315 groups with core animals of different breed compositions allowed the prediction of $GEBV_{APY}$
316 that were unbiased, and (almost) as accurate as $GEBV_{DIRECT}$, for all selection candidates and
317 performance traits. Indeed, the regression coefficients of TBV on $GEBV_{APY}$ were close to 1
318 (Table 3); the ratios of accuracies were higher than 0.97 for the purebred performance trait,
319 and higher than 0.94 for the crossbred performance trait (Figure 5; Figure 6; E-Supplements
320 Table S3); and the MSE of $GEBV_{APY}$ were similar to MSE of $GEBV_{DIRECT}$ (Table 4; E-
321 Supplements Table S6). Ratios of accuracies close to, or higher than, 0.99 were then obtained
322 for both traits when at least 8,000 core animals were used. The corresponding Pearson
323 correlations between $GEBV_{APY}$ and $GEBV_{DIRECT}$, which is usually used as criteria in studies
324 on real datasets (e.g., Ostensen et al., 2016; Strandén et al., 2017), were about 0.995 (E-
325 Supplements). It is worth noting that the core size of 8,000 animals is between the numbers of
326 eigenvalues that explained 98% and 99% of the variation in \mathbf{G}^* , that is about 6,498 and 9,213
327 eigenvalues on average across the five replicates, respectively (Figure 4-Figure 6).

328 Comparison of the three performance criteria for the purebred performance trait showed no
329 difference among the three core selection strategies involving core animals of different breed
330 compositions (Figure 5; Table 3; Table 4; E-Supplements Tables S3-S6). For the crossbred
331 performance trait, the scenarios with purebred and crossbred core animals, either randomly
332 chosen or chosen based on a QR decomposition, slightly outperformed the scenarios with
333 only purebred core animals (Figure 6). However, these outperformances were not always
334 significant (E-Supplements).

335 *Quality of GEBV for core and non-core selection animals*

336 Table 5 shows ratios of accuracies and of MSE, and the regression coefficients for the
337 scenario using 8000 core animals randomly selected among purebred and crossbred animals.
338 The regression coefficients and ratios of MSE for $GEBV_{APY}$ of core selection candidates and
339 of non-core selection candidates were similar. Ratios of accuracies for non-core selection
340 candidates were slightly lower than the corresponding ratios for the core selection candidates,
341 meaning that $GEBV_{APY}$ of non-core selection candidates were slightly less accurate than those
342 of core selection candidates, in comparison to $GEBV_{DIRECT}$. However, the differences
343 between accuracies of $GEBV_{APY}$ of core and of non-core selection candidates were not
344 significant following a Welch's t-test (Welch, 1947) with a 5% significance level.

345 *Convergence of ssGBLUP with alternative core groups*

346 Convergence of ssGBLUP with alternative core groups of 8,000 animals were compared
347 against ssGBLUP using G_{direct}^{-1} . Number of iterations of ssGBLUP using G_{APY}^{-1} were
348 expressed as the ratio to the number of iterations of ssGBLUP using G_{direct}^{-1} . Average values
349 of this ratio across the 5 replicates (SD within brackets), were 0.85 (0.39) using breed A core
350 animals, 1.05 (0.33) using purebred core animals, 0.95 (0.31) using purebred and crossbred
351 animals, and 0.94 (0.30) using core animals selected based on a QR decomposition of the
352 genotype matrix. In comparison to ssGBLUP with G_{direct}^{-1} , use of the APY led to similar
353 number of iterations to reach convergence. The selection strategy based on the QR
354 decomposition led to similar convergence as the other selection strategies.

355

356

DISCUSSION

357 In this study, we showed that $GEBV_{APY}$ were accurate approximations of $GEBV_{DIRECT}$ for
358 multivariate ssGBLUP involving multiple breeds and their crosses. $GEBV_{APY}$ as accurate as

359 GEBV_{DIRECT} were obtained when the core groups included animals from different breed
360 compositions, and when the core groups had a size between the numbers of the largest
361 eigenvalues explaining 98% and 99% of the variation in the raw (i.e., before blending with the
362 pedigree relationship matrix) genomic relationship matrix (\mathbf{G}^*).

363 *Composition of the core groups and selection strategies*

364 The quality of the GEBV_{APY} for both purebred and crossbred performance traits was close to
365 the GEBV_{DIRECT} as long as all classes of purebred and crossbred animals were well
366 represented in the core group. This was not the case if not all breeds were included in the core
367 group. Such a situation where core animals are only from one breed, could be obtained with a
368 naive random selection strategy on a large genotype dataset that is dominated by one breed.
369 Due to the properties of the simulated datasets, e.g, similar numbers of genotyped animals per
370 breed and per generation, a random selection of core animals across the full dataset led to
371 similar proportions of core animals per breed composition and per generation. Based on a
372 study involving single breed ssGBLUP, Ostensen et al. (2016) advised that core groups should
373 represent all generations. Including animals from each generation in the core group was also
374 recommended by Bradford et al. (2017), especially when genotyped animals had incomplete
375 pedigree, such as unknown parents. Incomplete pedigree could be common in crossbreeding
376 schemes, because pedigree data for crossbred animals in field conditions is difficult to collect
377 (Ibánñez-Escriche et al., 2009). From our results with the selection strategy based on QR
378 decomposition with pivoting, it seems that all generations, and all breed compositions, do not
379 have to be similarly represented in core groups. Indeed, in comparison to a random selection,
380 the selection strategy based on QR decomposition included higher proportions of crossbred
381 A(BC) animals and of the first generation of genotyped purebred animals selected as core
382 animals. One possible explanation is that genotypes of the crossbred A(BC) animals and of

383 the first generation of genotyped purebred animals include a large proportion of the
384 independent chromosome segments from all the genotyped purebred and crossbred animals.
385 However, core groups including animals that were randomly selected and that represented
386 similarly all generations and all breed composition gave results similar to the numerical
387 strategy based on QR decomposition, which is computationally expensive. Therefore, a
388 random selection of core animals by ensuring that core animals represent similarly all
389 generations and all breed compositions is advisable for the implementation of the APY in
390 well-structured crossbreeding schemes as investigated in this study. More complex situations,
391 such as multibreed (beef) cattle populations with a large variation in the observed breed
392 compositions, would probably benefit from more advanced APY core selection approaches
393 (Mäntysaari et al., 2017), such as the proposed numerical strategy based on QR
394 decomposition.

395 *Size of the core groups*

396 For single breed ssGBLUP, Pocrnic et al. (2016a; b) showed that the size of the core groups
397 required to predict $GEBV_{APY}$ at least as accurate as $GEBV_{DIRECT}$ was related to the
398 dimensionality of the genomic information. In their studies, the most accurate $GEBV_{APY}$ were
399 obtained when the core size was at least equal to the number of largest eigenvalues that
400 explained 98% of the variation in the raw genomic relationship matrix \mathbf{G}^* . In this study,
401 $GEBV_{APY}$ as accurate as $GEBV_{DIRECT}$ (i.e., with correlations between them ≥ 0.995) were
402 obtained when the core sizes were between the numbers of largest eigenvalues that explained
403 98% and 99% of the variation in the raw genomic relationship matrix \mathbf{G}^* , provided that the
404 composition of the core group represented the variation in all the breeds and crosses. Using a
405 multibreed beef cattle population, Mäntysaari et al. (2107) also showed that a core size larger
406 than the number of largest eigenvalues that explained 98% of the variation in \mathbf{G}^* was needed

407 to get correlations between $GEBV_{APY}$ and $GEBV_{DIRECT}$ close to 1. Furthermore, Mäntysaari et
408 al. (2107) observed that the correlation between $GEBV_{APY}$ and $GEBV_{DIRECT}$ depended on the
409 composition of the core groups, even with a core size close to the number of largest
410 eigenvalues that explained 98% of the variation in \mathbf{G}^* . All these results suggest that the core
411 size involving multiple breeds and crosses can be also approximated based on the
412 dimensionality of the genomic information of all breeds and crosses together to ensure that
413 the core size is optimal. It should be noted, however, that in crossbreeding situations
414 relationships between the core size, the dimensionality of the genomic information, and some
415 population parameters (e.g., number of independent segments, effective population size) is not
416 as straightforward in as in single breed situations (Pocrnic et al., 2016a; b).

417

418 **CONCLUSIONS**

419 We showed that the APY algorithm gives results equivalent to those obtained with the direct
420 inversion of the genomic relationship matrix when genotyped animals belong to a few
421 different breeds and their F1 and F2 crosses, such as commonly observed in pig and poultry
422 breeding programs. For such situations, we suggest that core animals could be randomly
423 selected among all purebred and crossbred genotyped animals, while ensuring that they
424 represent all generations and all breed compositions. It was also shown that selecting a
425 number of core animals equal to the number of largest eigenvalues needed to explain 98-99%
426 of the variation on the raw genomic relationship matrix, is sufficient to achieve good quality
427 of GEBV in crossbreeding schemes.

428

429

LITERATURE CITED

- 430 Aguilar, I., I. Misztal, D.L. Johnson, A. Legarra, S. Tsuruta, and T.J. Lawlor. 2010. Hot topic:
431 A unified approach to utilize phenotypic, full pedigree, and genomic information for
432 genetic evaluation of Holstein final score. *J. Dairy Sci.* 93:743–752.
- 433 Bradford, H. I., I. Pocrnić, B. o. Fragomeni, D. a. I. Lourenco, and I. Misztal. 2017. Selection
434 of core animals in the Algorithm for Proven and Young using a simulation model. *J.*
435 *Anim. Breed. Genet.* 134:545–552. doi:10.1111/jbg.12276.
- 436 Calus, M.P.L., and J. Vandenplas. 2016. Calc_grm – a Program to Compute Pedigree,
437 Genomic, and Combined Relationship Matrices. ABGC, Wageningen UR Livestock
438 Research.
- 439 Christensen, O.F. 2012. Compatibility of pedigree-based and marker-based relationship
440 matrices for single-step genetic evaluation. *Genet. Sel. Evol.* 44:37.
- 441 Christensen, O.F., and M.S. Lund. 2010. Genomic prediction when some animals are not
442 genotyped. *Genet. Sel. Evol.* 42:2.
- 443 Fernando, R.L., H. Cheng, and D.J. Garrick. 2016. An efficient exact method to obtain
444 GBLUP and single-step GBLUP when the genomic relationship matrix is singular.
445 *Genet. Sel. Evol.* 48:80. doi:10.1186/s12711-016-0260-7.
- 446 Fragomeni, B.O., D.A.L. Lourenco, S. Tsuruta, Y. Masuda, I. Aguilar, A. Legarra, T.J.
447 Lawlor, and I. Misztal. 2015. Hot topic: Use of genomic recursions in single-step
448 genomic best linear unbiased predictor (BLUP) with a large number of genotypes. *J.*
449 *Dairy Sci.* 98:4090–4094.

450 Golub, G., and C.F. Van Loan. 1996. *Matrix Computations*. third ed. Johns Hopkins
451 University Press, Baltimore, MD, USA.

452 Hill, W.G., and A. Robertson. 1968. Linkage disequilibrium in finite populations. *Theor.*
453 *Appl. Genet.* 38:226–231. doi:10.1007/BF01245622.

454 Ibánñez-Escriche, N., R.L. Fernando, A. Toosi, and J.C. Dekkers. 2009. Genomic selection of
455 purebreds for crossbred performance. *Genet. Sel. Evol.* 41:12. doi:10.1186/1297-
456 9686-41-12.

457 Legarra, A., O.F. Christensen, I. Aguilar, and I. Misztal. 2014. Single Step, a general
458 approach for genomic selection. *Livest. Sci.* 166:54–65.

459 Lourenco, D.A.L., S. Tsuruta, B.O. Fragomeni, C.Y. Chen, W.O. Herring, and I. Misztal.
460 2016. Crossbreed evaluations in single-step genomic best linear unbiased predictor
461 using adjusted realized relationship matrices. *J. Anim. Sci.* 94:909–19.
462 doi:10.2527/jas.2015-9748.

463 Lourenco, D.A.L., S. Tsuruta, B.O. Fragomeni, Y. Masuda, I. Aguilar, A. Legarra, J.K.
464 Bertrand, T.S. Amen, L. Wang, D.W. Moser, and I. Misztal. 2015. Genetic evaluation
465 using single-step genomic best linear unbiased predictor in American Angus. *J. Anim.*
466 *Sci.* 93:2653–2662. doi:10.2527/jas.2014-8836.

467 Mäntysaari, E.A., R.D. Evans, and I. Strandén. 2017. Efficient single-step genomic evaluation
468 for a multibreed beef cattle population having many genotyped animals. *J. Anim. Sci.*
469 95:4728–4737. doi:10.2527/jas2017.1912.

470 Masuda, Y., I. Misztal, S. Tsuruta, A. Legarra, I. Aguilar, D.A.L. Lourenco, B.O. Fragomeni,
471 and T.J. Lawlor. 2016. Implementation of genomic recursions in single-step genomic

472 best linear unbiased predictor for US Holsteins with a large number of genotyped
473 animals. *J. Dairy Sci.* 99:1968–1974. doi:10.3168/jds.2015-10540.

474 Misztal, I. 2016. Inexpensive computation of the inverse of the genomic relationship matrix in
475 populations with small effective population size. *Genetics* 202:401–409.

476 Misztal, I., A. Legarra, and I. Aguilar. 2014. Using recursion to compute the inverse of the
477 genomic relationship matrix. *J. Dairy Sci.* 97:3943–3952.

478 Mrode, R.A. 2005. *Linear Models for the Prediction of Animal Breeding Values*. 2nd ed.
479 CABI Publishing, Wallingford, UK.

480 ten Napel, J., M.P.L. Calus, M. Lidauer, I. Stradén, E.A. Mäntysaari, H.A. Mulder, and R.F.
481 Veerkamp. 2016. *MiXBLUP, User-Friendly Software for Large Genetic Evaluations*
482 *Systems*. Version 2.0. Wageningen, the Netherlands.

483 Ostersen, T., O.F. Christensen, P. Madsen, and M. Henryon. 2016. Sparse single-step method
484 for genomic evaluation in pigs. *Genet. Sel. Evol.* 48:48. doi:10.1186/s12711-016-
485 0227-8.

486 Pocrnic, I., D.A.L. Lourenco, Y. Masuda, A. Legarra, and I. Misztal. 2016a. The
487 dimensionality of genomic information and its effect on genomic prediction. *Genetics*
488 203:573–581.

489 Pocrnic, I., D.A.L. Lourenco, Y. Masuda, and I. Misztal. 2016b. Dimensionality of genomic
490 information and performance of the Algorithm for Proven and Young for different
491 livestock species. *Genet. Sel. Evol.* 48:82. doi:10.1186/s12711-016-0261-6.

492 Powell, J.E., P.M. Visscher, and M.E. Goddard. 2010. Reconciling the analysis of IBD and
493 IBS in complex trait studies. *Nat. Rev. Genet.* 11:800–805. doi:10.1038/nrg2865.

494 Raymond, M., and F. Rousset. 1995. GENEPOP (Version 1.2): Population genetics software
495 for exact tests and ecumenicism. *J. Hered.* 86:248–249.

496 Rousset, F. 2008. genepop'007: a complete re-implementation of the genepop software for
497 Windows and Linux. *Mol. Ecol. Resour.* 8:103–106. doi:10.1111/j.1471-
498 8286.2007.01931.x.

499 Sargolzaei, M., and F.S. Schenkel. 2009. QMSim: a large-scale genome simulator for
500 livestock. *Bioinformatics* 25:680–681.

501 Strandén, I., K. Matilainen, G.P. Aamand, and E.A. Mäntysaari. 2017. Solving efficiently
502 large single-step genomic best linear unbiased prediction models. *J. Anim. Breed.*
503 *Genet.* 134:264–274. doi:10.1111/jbg.12257.

504 Tukey, J.W. 1949. Comparing individual means in the analysis of variance. *Biometrics* 5:99–
505 114.

506 Vitezica, Z.G., I. Aguilar, I. Misztal, and A. Legarra. 2011. Bias in genomic predictions for
507 populations under selection. *Genet. Res.* 93:357–366.

508 Welch, B.L. 1947. THE GENERALIZATION OF “STUDENT”’S’ PROBLEM WHEN
509 SEVERAL DIFFERENT POPULATION VARLANCES ARE INVOLVED.
510 *Biometrika* 34:28–35. doi:10.1093/biomet/34.1-2.28.

511 Wientjes, Y.C.J., and M.P.L. Calus. 2017. BOARD INVITED REVIEW: The purebred-
512 crossbred correlation in pigs: A review of theory, estimates, and implications. *J. Anim.*
513 *Sci.* 95:3467–3478.

514

516 **E-Supplements**

517 **Table S1** Number of purebred and crossbred animals with a phenotype per generation

518 (average for the 5 replicates; SD within brackets).

519

520 **Table S2** Number of purebred and crossbred animals with a phenotype and a genotype per

521 generation (average for the 5 replicates; SD within brackets).

522

523 **Table S3.** Relative accuracies (average for the 5 replicates; SD within brackets) of GEBV

524 from alternative core groups for the purebred (PB) and crossbred (CB) performance for

525 genotyped selection candidates.

526

527 **Table S4.** Pearson correlations (average for the 5 replicates; SD within brackets) between

528 GEBV for genotyped selection candidates from alternative core groups1 and GEBV from the

529 direct inversion of **G**.

530

531 **Table S5.** Regression coefficients (average for the 5 replicates; SD within brackets) of TBV

532 on GEBV from alternative core groups and the direct inversion of G for genotyped selection

533 candidates.

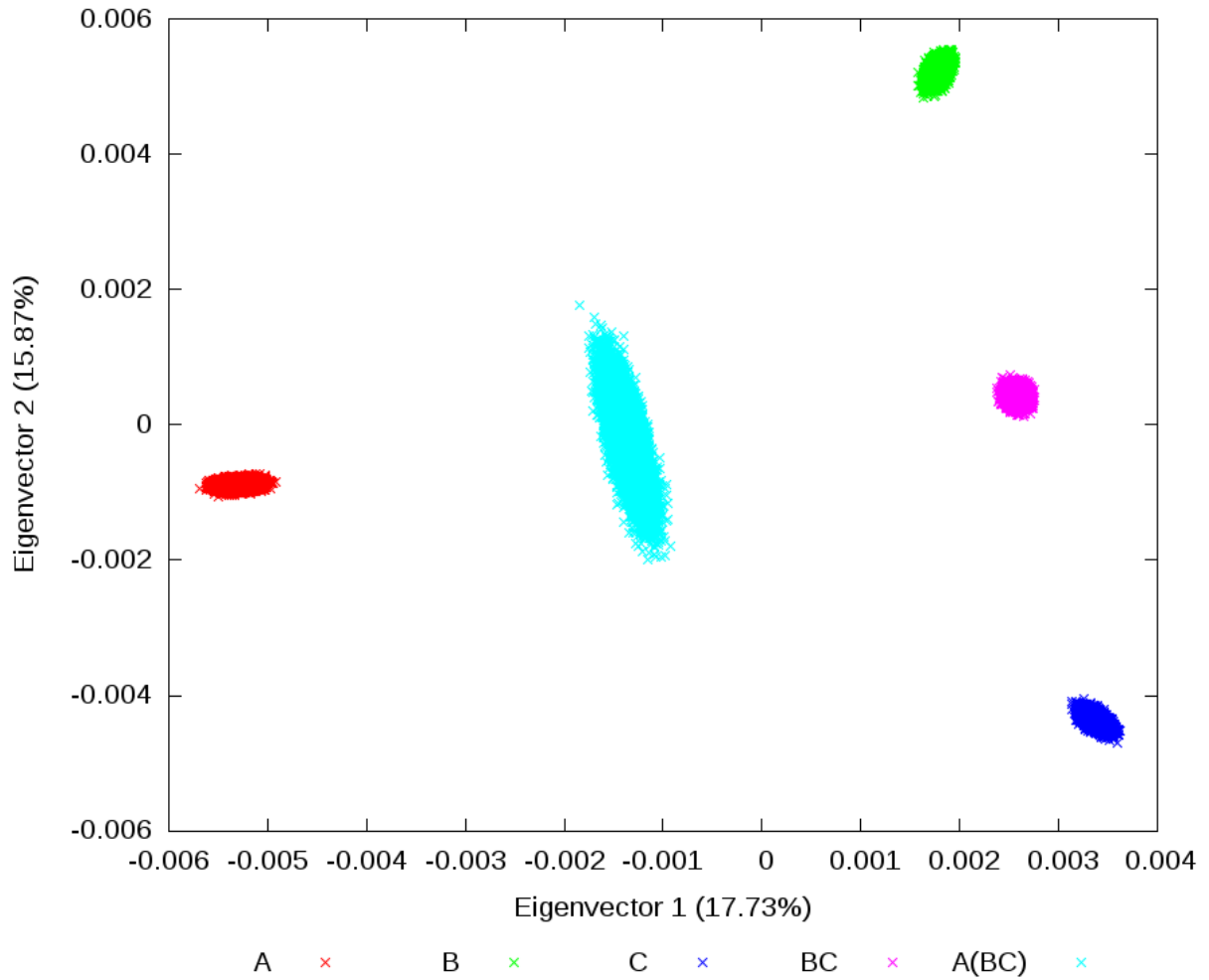
534

535 **Table S6.** Relative mean squares errors (average for the 5 replicates; SD within brackets) of
536 GEBV from alternative core groups for genotyped selection candidates.

537

538 **Figures**

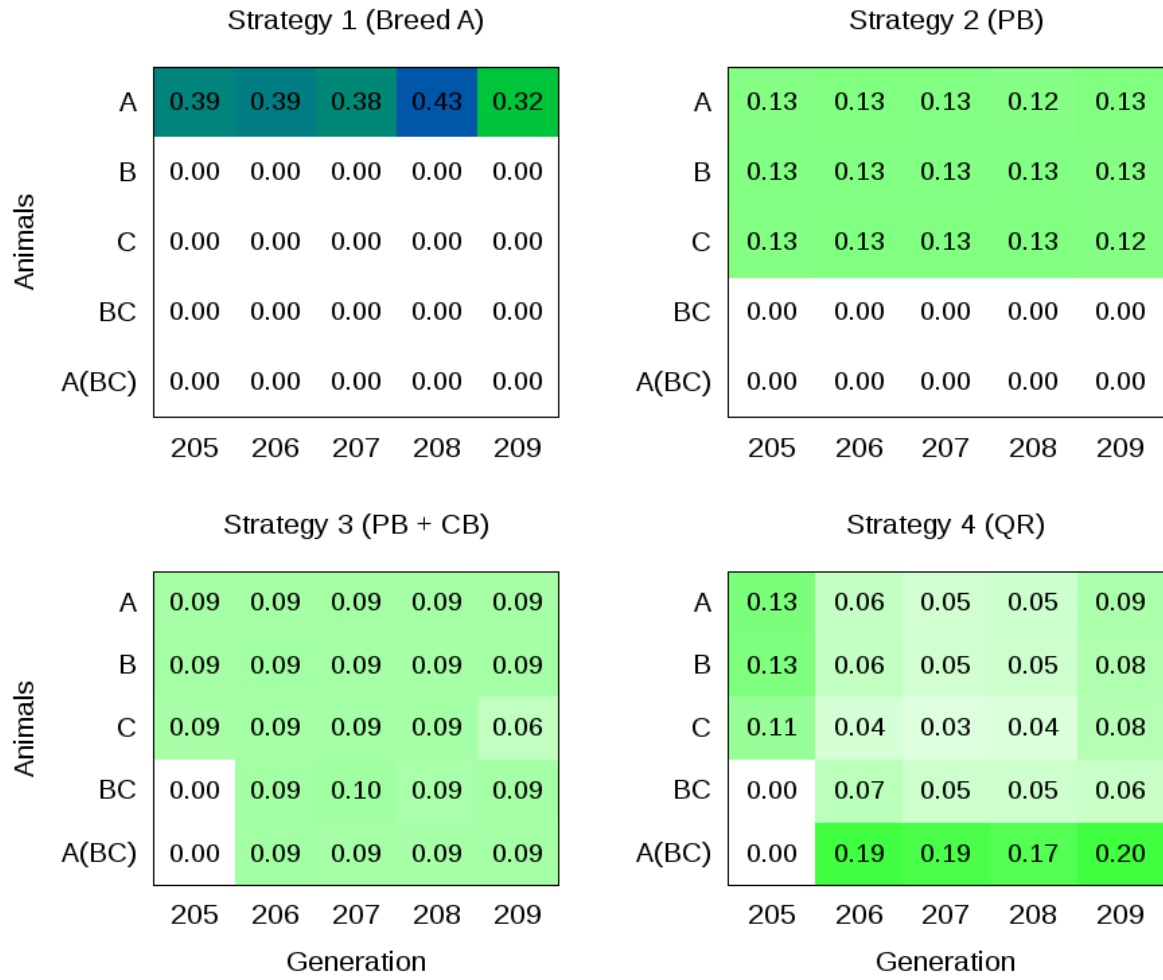
539



540

541 **Figure 2.**

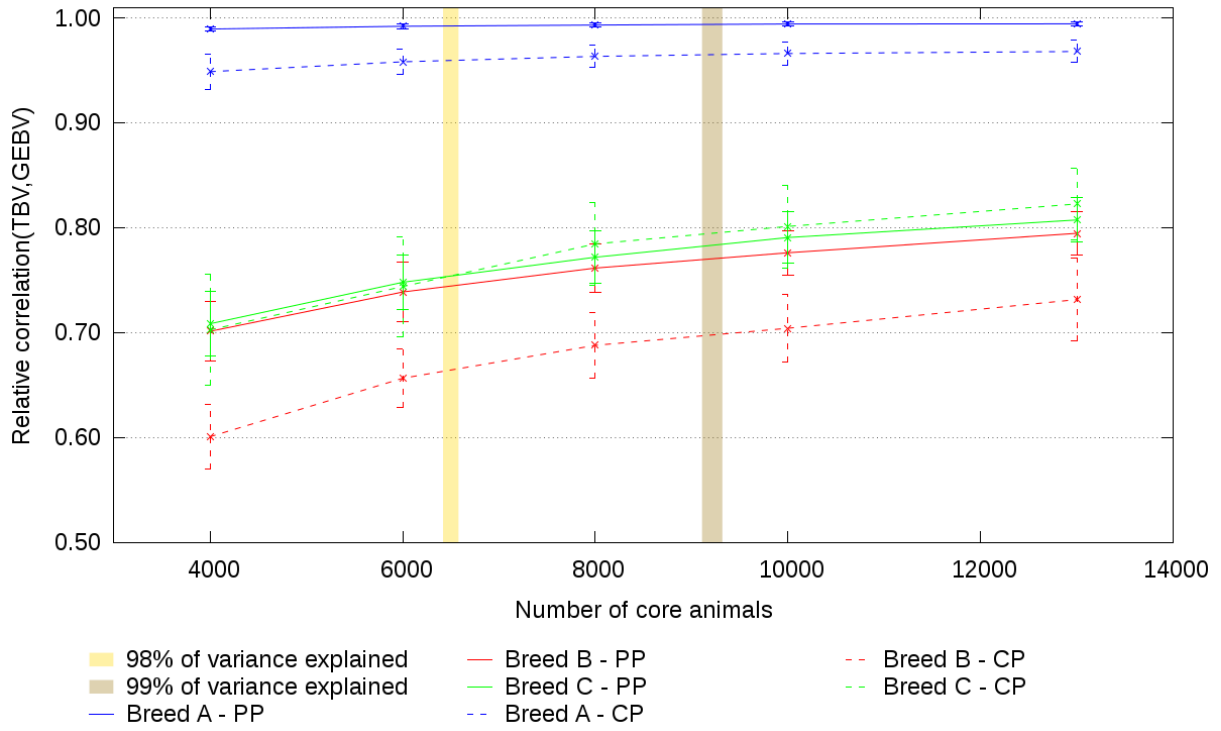
542



543

544 **Figure 3.**

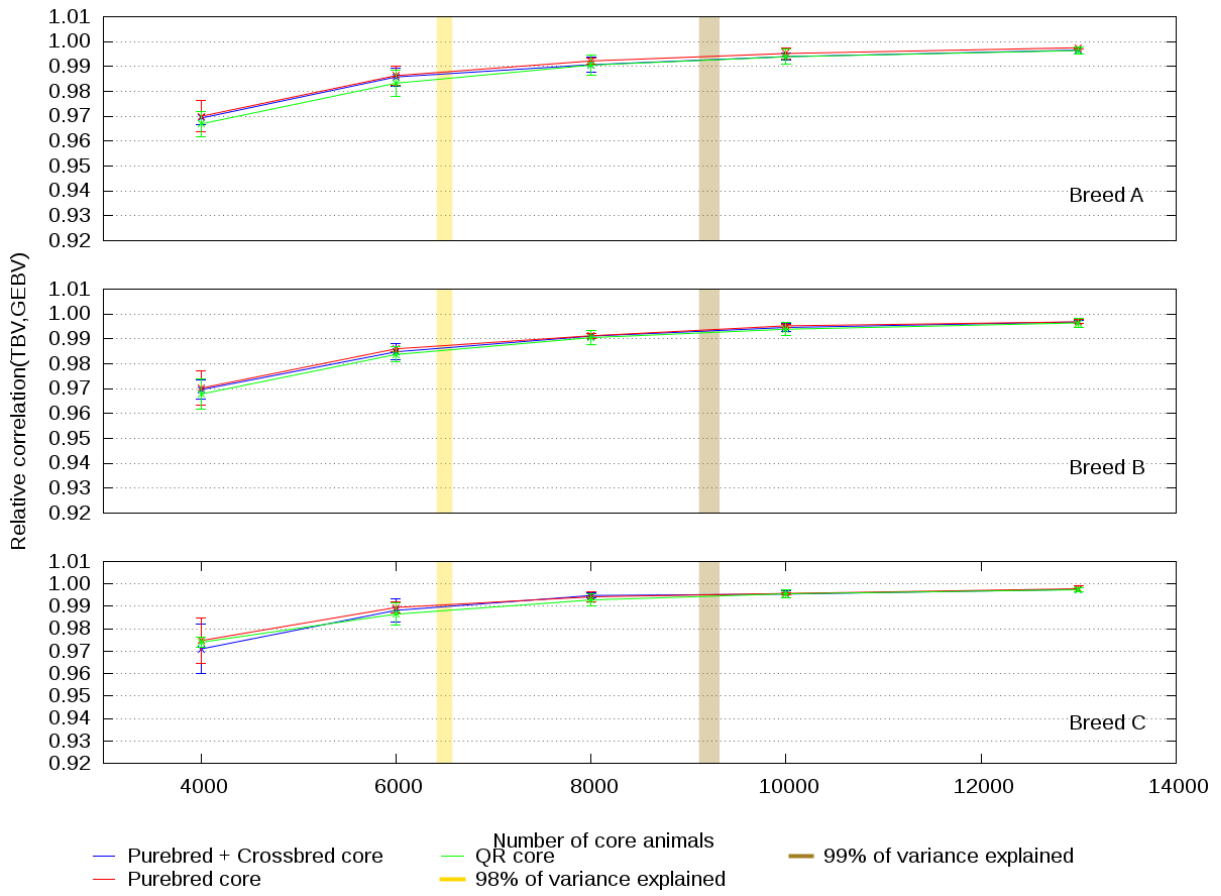
545



546

547 **Figure 4.**

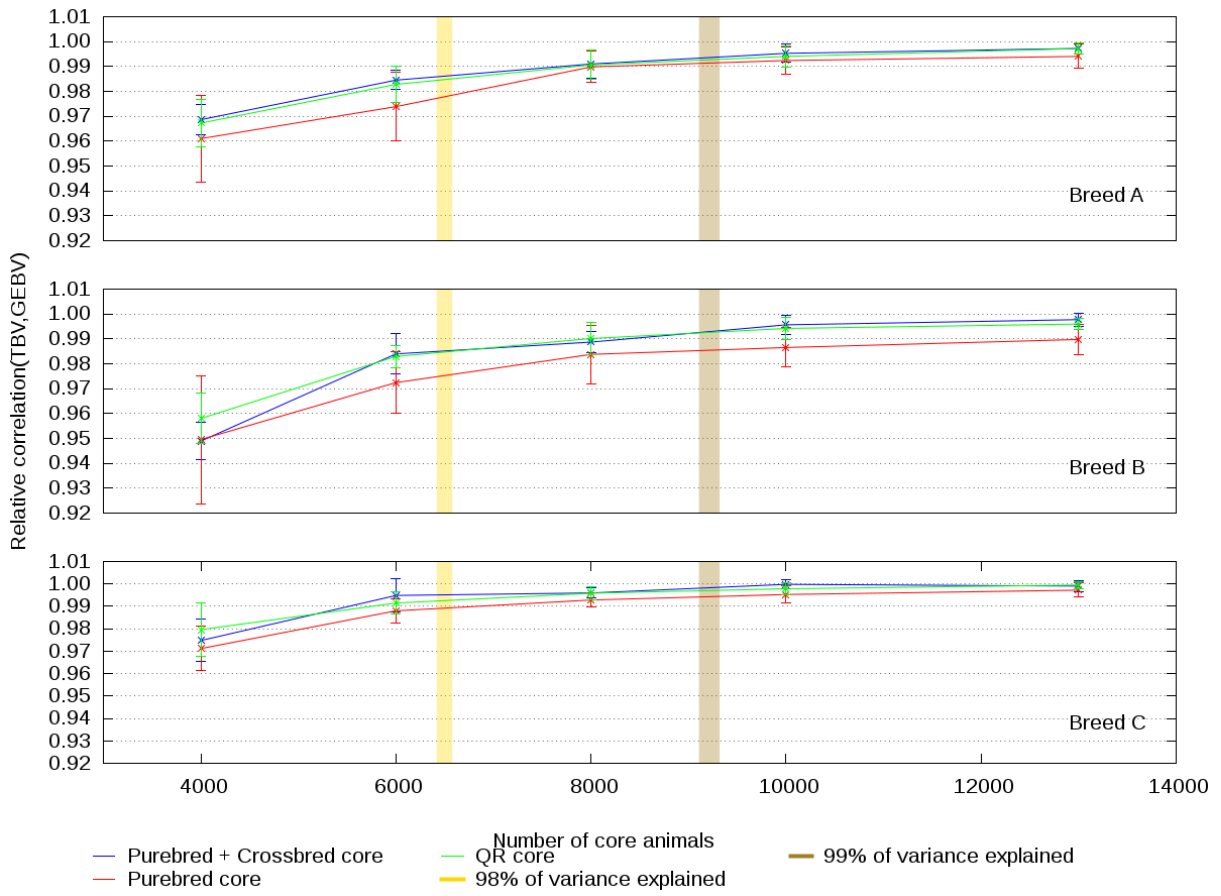
548



549

550 **Figure 5.**

551



552

553 **Figure 6.**

554

555 **Figure 1.** Schematic representation of the simulation. The crossbreeding program started at
556 generation 200 (generation numbers in bold). The number of males (M) and females (F) per
557 generation and per breed (A, B, and C), or per cross (BC, and A(BC)), are reported within
558 brackets. Blue arrows denote the sires and dams of the next generation; red arrows denote the
559 dams of the next generation; green arrows denote the sires of the next generation.

560

561 **Figure 2.** Projections of genomic relationships for purebred (A, B, and C) and crossbred (BC
562 and A(BC)) genotyped animals into the two first eigenvectors for the first replicate.

563

564 **Figure 3.** Proportions of core animals per generation and breed composition of one replicate
565 for the scenario using 8,000 core animals. Core animals were selected using four different
566 strategies: 1) only from breed A animals (Breed A), 2) from purebred animals of breed A, B
567 and C (PB), 3) from purebred animals of breed A, B and C, and crossbred BC and A(BC)
568 animals (PB + CB), and (4) chosen based on a QR decomposition of the genotype matrix
569 (QR). Darker colours represent higher proportions of core animals per generation and breed
570 composition.

571

572 **Figure 4.** Relative correlations of GEBV from different sizes of core groups with only breed
573 A animals. Relative correlations for the purebred performance (PP) and crossbred
574 performance (CP) traits are defined as the ratio between the accuracies of GEBV from
575 alternative core groups and the corresponding accuracies of GEBV from \mathbf{G}_{direct}^{-1} . Vertical
576 columns depict the number of eigenvalues that explained 98% and 99% of the variation in \mathbf{G}^* .
577 Results are averages for the 5 replicates.

578

579

580 **Figure 5.** Relative correlations of GEBV from alternative core groups for the purebred
581 performance traits. Core groups include randomly selected purebred and crossbred animals
582 (Purebred + Crossbred core), randomly selected purebred animals (Purebred core), and
583 animals selected based on a QR decomposition of the genotype matrix (QR core). Relative
584 correlations are defined as the ratio between the accuracies of GEBV from alternative core
585 groups and the corresponding accuracies of GEBV from \mathbf{G}_{direct}^{-1} . Vertical columns depict the
586 number of eigenvalues that explained 98% and 99% of the variation in \mathbf{G}^* . Results are
587 averages for the 5 replicates.

588

589 **Figure 6.** Relative correlations of GEBV from alternative core groups for the crossbred
590 performance trait. Core groups include randomly selected purebred and crossbred animals
591 (Purebred + Crossbred core), randomly selected purebred animals (Purebred core), and
592 animals selected based on a QR decomposition of the genotype matrix (QR core). Relative
593 correlations are defined as the ratio between the accuracies of GEBV from alternative core
594 groups and the corresponding accuracies of GEBV from \mathbf{G}_{direct}^{-1} . Vertical columns depict the
595 number of eigenvalues that explained 98% and 99% of the variation in \mathbf{G}^* . Results are
596 averages for the 5 replicates.

597

598 **Tables**

599 **Table 1.** Heritabilities (diagonal) and genetic correlations (off-diagonal) among the five
 600 simulated traits.

Trait	Purebred A	Purebred B	Purebred C	Crossbred BC	Crossbred A(BC)
Purebred A	0.28				
Purebred B	0.46	0.39			
Purebred C	0.27	0.80	0.22		
Crossbred BC	0.33	0.58	0.30	0.36	
Crossbred A(BC)	0.55	0.31	0.26	0.69	0.23

601

602 **Table 2.** Accuracies, bias, and mean square errors (MSE) of GEBV from the direct inversion
 603 of **G** (average for the 5 replicates; SD within brackets).

Selection candidates	Number	Purebred performance			Crossbred performance		
		Accuracy	Bias	MSE	Accuracy	Bias	MSE
Breed A	5010	0.81	1.04	1.11	0.68	0.98	0.68
	(24)	(0.02)	(0.05)	(0.69)	(0.04)	(0.08)	(0.51)
Breed B	4975	0.85	1.06	1.16	0.63	0.95	0.90
	(30)	(0.01)	(0.03)	(0.81)	(0.02)	(0.04)	(0.43)
Breed C	5016	0.79	1.04	1.42	0.71	1.04	1.35
	(45)	(0.04)	(0.03)	(0.74)	(0.04)	(0.07)	(1.18)

604

605 **Table 3.** Regression coefficients (average for the 5 replicates; SD within brackets) of TBV on
 606 GEBV from alternative core groups¹ for genotyped selection candidates.

Number of core animals	Purebred performance				Crossbred performance			
	Breed A	PB	PB+CB	QR	Breed A	PB	PB+CB	QR
Breed A selection candidates								
4000	1.04 (0.06)	1.05 (0.06)	1.06 (0.06)	1.06 (0.06)	0.90 (0.08)	0.96 (0.08)	0.99 (0.09)	0.99 (0.08)
8000	1.04 (0.05)	1.05 (0.06)	1.05 (0.06)	1.05 (0.06)	0.92 (0.08)	0.97 (0.08)	0.99 (0.08)	0.99 (0.08)
13000	1.04 (0.05)	1.05 (0.05)	1.05 (0.05)	1.05 (0.05)	0.93 (0.08)	0.97 (0.08)	0.98 (0.08)	0.98 (0.08)
Breed B selection candidates								
4000	1.49 (0.08)	1.06 (0.03)	1.06 (0.02)	1.07 (0.03)	1.62 (0.11)	0.91 (0.08)	0.93 (0.05)	0.94 (0.06)
8000	1.46 (0.08)	1.06 (0.02)	1.06 (0.03)	1.06 (0.03)	1.58 (0.13)	0.93 (0.06)	0.95 (0.05)	0.95 (0.04)
13000	1.43 (0.09)	1.06 (0.03)	1.06 (0.03)	1.06 (0.03)	1.54 (0.14)	0.93 (0.05)	0.95 (0.05)	0.95 (0.04)
Breed C selection candidates								
4000	1.69 (0.15)	1.05 (0.04)	1.06 (0.04)	1.05 (0.03)	2.41 (0.30)	1.07 (0.08)	1.09 (0.07)	1.08 (0.07)
8000	1.62 (0.12)	1.04 (0.04)	1.05 (0.04)	1.04 (0.03)	2.27 (0.19)	1.06 (0.07)	1.06 (0.07)	1.06 (0.07)
13000	1.58 (0.11)	1.04 (0.03)	1.04 (0.04)	1.04 (0.03)	2.14 (0.14)	1.06 (0.07)	1.05 (0.07)	1.05 (0.07)

607 1 Core groups include 1) randomly selected breed A animals only (Breed A), 2) randomly selected purebred
 608 animals (PB), 3) randomly selected purebred and crossbred animals (PB+CB), and 4) animals selected based on
 609 a QR decomposition of the genotype matrix (QR).

610

611 **Table 4.** Relative mean squares errors¹ (average for the 5 replicates; SD within brackets) of
 612 GEBV from alternative core groups² for genotyped selection candidates.

Number of core animals	Purebred performance				Crossbred performance			
	Breed A	PB	PB+CB	QR	Breed A	PB	PB+CB	QR
Breed A selection candidates								
4000	1.09 (0.33)	1.05 (0.08)	1.05 (0.04)	1.04 (0.06)	1.14 (0.29)	1.14 (0.20)	1.04 (0.04)	1.03 (0.04)
8000	1.11 (0.34)	0.98 (0.05)	0.98 (0.06)	0.99 (0.07)	1.20 (0.28)	1.11 (0.22)	1.11 (0.23)	1.11 (0.22)
13000	1.12 (0.33)	1.04 (0.19)	0.98 (0.06)	0.98 (0.06)	1.20 (0.28)	1.11 (0.22)	1.12 (0.22)	1.10 (0.23)
Breed B selection candidates								
4000	1.75 (0.93)	1.12 (0.16)	1.05 (0.06)	1.08 (0.08)	1.34 (0.30)	0.99 (0.15)	1.04 (0.08)	1.02 (0.07)
8000	1.81 (1.04)	1.09 (0.18)	1.09 (0.17)	1.09 (0.18)	1.27 (0.21)	0.99 (0.15)	0.95 (0.10)	0.95 (0.10)
13000	1.77 (0.95)	1.15 (0.23)	1.08 (0.18)	1.08 (0.18)	1.24 (0.21)	1.00 (0.14)	0.93 (0.11)	0.94 (0.09)
Breed C selection candidates								
4000	1.29 (0.43)	0.97 (0.08)	1.02 (0.05)	1.00 (0.05)	1.20 (0.44)	1.00 (0.16)	1.00 (0.08)	1.02 (0.07)
8000	1.28 (0.37)	0.96 (0.08)	0.97 (0.09)	0.96 (0.09)	1.19 (0.38)	0.99 (0.16)	0.94 (0.09)	0.95 (0.09)
13000	1.24 (0.34)	0.88 (0.16)	0.96 (0.08)	0.96 (0.09)	1.16 (0.35)	1.00 (0.16)	0.95 (0.09)	0.95 (0.08)

613 ¹ Results are expressed as the ratio between MSE of GEBV from alternative core groups and MSE of GEBV
 614 from the direct inversion of **G**.

615 ² Core groups include 1) randomly selected breed A animals only (Breed A), 2) randomly selected purebred
 616 animals (PB), 3) randomly selected purebred and crossbred animals (PB+CB), and 4) animals selected based on
 617 a QR decomposition of the genotype matrix (QR).

618

619

620 **Table 5.** Quality of GEBV using APY for the core and non-core selection candidates.¹

Selection candidates	Number	Purebred performance			Crossbred performance		
		Accuracy ²	Reg. coef.	MSE ²	Accuracy ²	Reg. coef.	MSE ²
A core	453 (19)	0.999 (0.001)	1.02 (0.05)	0.960 (0.058)	0.997 (0.001)	0.98 (0.07)	1.087 (0.203)
A non-core	4557 (32)	0.990 (0.003)	1.06 (0.06)	0.980 (0.066)	0.990 (0.006)	0.99 (0.08)	1.112 (0.232)
B core	456 (23)	0.998 (0.002)	1.02 (0.06)	1.076 (0.169)	0.995 (0.004)	0.88 (0.09)	0.946 (0.091)
B non-core	4519 (37)	0.991 (0.001)	1.07 (0.03)	1.093 (0.170)	0.988 (0.004)	0.95 (0.04)	0.949 (0.096)
C core	322 (43)	0.998 (0.001)	1.07 (0.07)	0.961 (0.082)	0.999 (0.004)	1.03 (0.06)	0.939 (0.089)
C non-core	4694 (27)	0.994 (0.001)	1.04 (0.04)	0.966 (0.087)	0.996 (0.003)	1.06 (0.07)	0.942 (0.093)

621 ¹ Results (average for the 5 replicates; SD within brackets) are shown for the scenario using 8000 core animals
622 randomly selected among purebred and crossbred animals.

623 ² Results for accuracies and mean square errors (MSE) are expressed as the ratio between accuracies (MSE) of
624 GEBV using APY and accuracies (MSE) of GEBV using the direct inversion of **G**.

625