# Vision in flying insects Martin Egelhaaf* and Roland Kern ${ }^{\dagger}$ 


#### Abstract

Vision guides flight behaviour in numerous insects. Despite their small brain, insects easily outperform current man-made autonomous vehicles in many respects. Examples are the virtuosic chasing manoeuvres male flies perform as part of their mating behaviour and the ability of bees to assess, on the basis of visual motion cues, the distance travelled in a novel environment. Analyses at both the behavioural and neuronal levels are beginning to unveil reasons for such extraordinary capabilities of insects. One recipe for their success is the adaptation of visual information processing to the specific requirements of the behavioural tasks and to the specific spatiotemporal properties of the natural input.


## Addresses

Lehrstuhl für Neurobiologie, Fakultät für Biologie, Universität Bielefeld, Postfach 100131, D-33501 Bielefeld, Germany
*e-mail: martin.egelhaaf@uni-bielefeld.de
†e-mail: roland.kern@uni-bielefeld.de

## Abbreviations <br> HSE horizontal system, equatorial cell <br> LGMD lobula giant movement detector <br> TC tangential cell

## Introduction

As an observer moves through a natural environment, his or her photoreceptors typically receive a wildly fluctuating pattern of image flow. It is the task of the brain to interpret this complex spatiotemporal input and to make use of it in guiding behaviour. Existing artificial vision systems are outperformed by nervous systems in many respects in their ability to process retinal image flow. This is most remarkable for insect brains, with their small number of neurons and the extraordinary speed with which retinal images are processed. Because of the relative ease with which their nervous systems can be examined electrophysiologically and using imaging techniques, insects have served for many years as model systems for analysing the processing of retinal image flow [1-7]. During the review period, there have been important new developments that aid understanding of not only insect vision but, quite generally, how behaviourally relevant visual information is processed.

## Behavioural tasks

To understand the mechanisms underlying visually guided behaviour, a causal relationship between visual input and behavioural output needs to be established. This has been possible for various behavioural contexts [4,8,9]. By artfully manipulating the environment, those spatiotemporal features of the visual input that are decisive for behavioural control
can be pinpointed. Only three particularly well analysed examples can be summarised here: visual course control; estimation of travelled distance; and pursuit of moving targets.

Visual course control: Optic flow is an important source of information used to control the course and velocity of locomotion. For example, regulating the angular velocity of the retinal image of the environment controls the flight speed of bees. Accordingly, bees decelerate when flying, for example, through a narrow gap [4]. When intending to move straight, it has been concluded that bees and flies equate the overall optic flow on their eyes [4,10]. If one eye is occluded, an optomotor equilibrium can be reached only on a slightly curved path $[11,12]$. Flies reach this equilibrium only on average, whereas individual paths of locomotion can deviate from it almost arbitrarily depending on the animals' momentary intentions. Optic flow not only helps mediate a straight course of locomotion but might also elicit turns, to prevent collision with an obstacle, for example. It was concluded that when a fly approaches a wall, the time at which a sharp saccade-like turn is elicited and the direction of the saccade depend on the characteristics of retinal image expansion [13 ${ }^{\circ}$ ].

Estimation of travelled distance: It is essential for a worker bee to determine the distance between a food source and her hive. This information, in addition to flight direction, is communicated to her hive mates by the waggle dance and allows them to find the food source. Distance estimation is hard to accomplish during flight, because many distance cues that can be used while walking are not available. By contrast to walking ants $\left[14,1^{\bullet}\right]$, flying bees gauge distance in terms of the optic flow experienced during flight to the food source (Figure 1; [4]). Since translational optic flow depends on the three-dimensional layout of the environment, distance information gathered in this way is ambiguous. Nevertheless, the ambiguities do not lead to problems, as long as the recruited bee tends to fly on the same route as the forager and if the environment does not change much between the flight of the forager and that of recruited bees. Whereas such changes of the environment were systematically made for experimental analysis $\left[16,17^{\bullet \bullet}, 18^{\bullet \bullet}\right]$, they occur only rarely in natural environments during a day or couple of days. Hence, visual estimation of flight distance is not reliable in all circumstances but is sufficient for specific needs under normal behavioural conditions.

Pursuit of moving targets: Some insects are able to chase moving targets, be it prey or a mate, in virtuosic aerial manoeuvres (Figure 2a; [19-23]). As the visual input is very complex during these manoeuvres, it has been hard to unravel the underlying control systems. By reducing the complexity of the visual input and using dummy targets

Figure 1


Honeybees measure distances in terms of optic flow and communicate this information to their hive mates by the waggle dance. Behavioural analysis of how honeybees estimate the distance travelled between their hive and a food source. (a) Layout for the experiments using tunnels and probabilities of waggle (W; green bars) and round dance ( R ; red bars) for the different experiments. A tunnel with a length of 6 min and a width of 11 cm was positioned either at a distance to the hive of 35 min (not drawn to scale) or at a distance of only 6 min . The walls of the tunnel were either covered with a texture that contained vertically oriented elements (Exp.1, Exp.2, Exp.4) or horizontally aligned stripes (Exp.3). When the food source was placed at the entrance of the tunnel (Exp.1), the bees performed mainly round dances, signalling a short distance to the food source. When the food source was placed at the end of the tunnel containing vertically oriented texture (Exp.2), the returning bees performed mainly waggle dances, signalling much larger distances to the hive, although the actual travel distance was not much increased. A food source at the same distance, however, located
in a tunnel with horizontally oriented stripes (Exp.3), again led mainly to round dances. The main difference between Exp. 2 and Exp. 3 is that in the former much optic flow is evoked on the eyes of the honeybee while flying along the tunnel, whereas in the latter case there is only little optic flow, because the contours are mainly oriented along the flight direction. When the tunnel covered with vertical contours and the food source close to its end is placed near to the hive (Exp.4), mainly waggle dances are performed, which are shorter than those performed in Exp. 2 (compare blue bars). These experiments show that travelled distance is measured in terms of optic flow. (b) Calibration of the odometer of the honeybee. Mean duration of waggle dances elicited by outdoor feeders at various distances to the hive. Also shown are the mean durations of waggle dances measured in Exp. 2 and Exp. 4 and their equivalent outdoor flight distances, as read from the regression line. At a mean distance between the honeybees and the tunnel wall of $5.5 \mathrm{~cm}, 1 \mathrm{~ms}$ of waggle in the dance corresponded to $17.7^{\circ}$ of image motion on the eyes. (Adapted from [17*•]).
moving on well-defined paths, however, it has been possible to analyse the chasing behaviour of male blowflies. It was concluded that flies employ similar viewing strategies to those of primates: spontaneous changes in gaze are done rapidly by saccades [24], whereas moving targets are followed by smooth pursuit [25*]. The pursuit system of flies is relatively simple but rapid and efficient. The forward velocity of the chasing fly is controlled by the angular size of the target, whereas its turning velocity depends on the angle by which the target is seen. During pursuit, primatelike catch-up saccades are observed only when the target changes its trajectory too rapidly to allow the pursuer to follow smoothly. Model simulations revealed that even these catch-up saccades, which occur frequently during chases of real fly targets [22], can be explained as a by-product of
the smooth pursuit system if neuronal latencies and the inertia of the chasing fly are taken into account [25•].

## Neuronal circuits for visual information processing

Which computational strategies allow insects to perform so extraordinarily well in visually guided orientation? Although researchers are far from being able to understand the functioning of the neural circuits mediating the various components in the behavioural repertoires of insects, it is becoming increasingly clear that visual information processing is exquisitely adapted to the operating conditions in a normal behavioural context. Such adaptations are present already at the level of photoreceptors and may be followed up to the highest levels of visual information processing.

One example is the visual mechanisms involved in chasing behaviour. In many insects, the design of the male visual system is highly specialised. Apparently, this male-specific specialisation has evolved to match the need for higher accuracy in the frontal part of the visual field, where the target is held in a relatively fixed position during chases [26]. Moreover, the temporal resolution has been found recently to be enhanced in this part of the eye in male blowflies (Figure 2) as a consequence of specific ionic currents and biophysical properties of the photoreceptors [27•, $\left.28^{\bullet}\right]$. These specialisations, together with a malespecific connection pattern between photoreceptors and second-order neurons [29], result in a more reliable performance of the frontal part of the eye [28•]. Although there is good anatomical evidence for sex-specific neuronal circuits at more central processing stages in male flies [30-32], it is not yet possible to assess the functional significance of these circuits for chasing behaviour.

An approaching object, such as a predator, is characterised by increasing retinal size. Neurons detecting looming stimuli have been characterised in moths [33] and in locusts [3,34,35•,36•]. In locusts, the lobula giant movement detector (LGMD) neurons respond best to objects on a collision course. By relating the neuronal activity to steering responses of tethered flying animals, it was concluded that this neuronal system plays a role in predator avoidance [ $36^{\bullet}$ ]. Despite disagreement as to the details $\left[3,34,35^{\bullet}, 36^{\bullet}\right]$, it appears that the LGMD response to looming stimuli is mediated by three distinct processes: motion-sensitive excitation impinging retinotopically onto a dendritic subfield of LGMDs; an inhibitory network acting presynaptically onto the motion-sensitive pathway; and feedforward inhibition impinging on two additional dendritic subfields of LGMDs. Many properties of LGMDs can be captured by network models of their input organisation [3,35*$]$.

Optic flow elicited by self-motion is specified by global, rather than merely local, features. This implies that mechanisms extracting optic-flow information from the retinal input need to combine local motion measurements from large areas of the visual field. Accordingly, local motion information has been shown to be spatially pooled on the extended dendrites of the tangential cells (TCs) in flies [2,6,7]. The preferred directions of the local motion detectors that synapse onto a given TC appear to coincide with the directions of the velocity vectors characterising the optic flow induced during particular types of self-motion [37]. This sophisticated pattern of local preferred directions is not affected by sensory experience but is the consequence of phylogenetic adaptations [38]. In the fly, specificity for certain types of optic flow has been shown to be much enhanced by synaptic interactions between TCs in the ipsilateral and/or contralateral half of the visual system. As a consequence, individual TCs are thought to be tuned to certain types of self-motion or to the relative motion between objects and their background [39-41,42•,43•,44*॰].

Figure 2


Retinal adaptations of male blowflies to pursuit of female targets. (a) Example of a chasing manoeuvre as seen from above. The position and the orientation of the body long-axis of the leading fly (blue) and of the chasing fly (red) are shown every 20 ms . The chasing fly closely follows the leading fly, thereby fixating it in the dorso-frontal part of its visual field. During chases, the pursuer may reach angular velocities of more than 3000\%. (Data from Böddeker). (b) Spatial (left) and temporal sensitivities (right) of photoreceptors of the male fly subserving the frontal visual field are larger than those of photoreceptors looking to the side or backwards. The spatial sensitivity of photoreceptors was determined by light flashes directed at the eye from various angular positions in the cell's receptive field. Data are fitted by a Gaussian model. The temporal sensitivity of photoreceptors is given by their impulse response. It is shorter and faster for frontal photoreceptors than for sideways- and backwards-looking ones. (Adapted from [28]).

In addition to extending knowledge on the wiring diagrams of the neuronal circuits underlying optic-flow processing, it has been possible to account for some of the underlying computations in terms of biophysical properties of nerve cells and their synaptic interactions. For example, a synapse that conveys visual motion information gathered by TCs in one half of the brain via another TC to the contralateral visual system was analysed. Synaptic signal transfer was found to operate linearly and very reliably within a certain dynamic range. In this range, motion information is signalled by the visual system with a large gain $\left[45^{\bullet \bullet}, 46^{\bullet}\right]$.

Figure 3


Response of a blowfly motion-sensitive neuron to optic flow as experienced during a free-flight manoeuvre. (a) Section of a flight trajectory monitored in a cubic cage ( $40 \times 40 \times 40 \mathrm{~cm}$ ) covered on its side walls with a natural texture. The position of the fly is shown in yellow every 10 ms . The position and orientation of the body long-axis are shown in red every 130 ms ; the starting position is indicated in mauve, the end position in green. (b) OmniMax $X$, a high-speed panoramic visual stimulation device, consisting of 14 of the 20 triangles of an icosahedron, each of which forms a printed circuit board subserving 7168 light-emitting diodes in total. The device operates at 370 updates per second. In the foreground, the micromanipulators by which recording electrodes are inserted into the fly's brain can be seen. (c) Input organisation of the HSE-cell, the activity of which was recorded. The HSE-cell receives input from the eye ipsilateral to its main dendrite from many retinotopic motionsensitive elements. Consequently, it is depolarised by front-to-back motion and hyperpolarised by back-to-front motion. The HSE-cell receives additional input on its main dendrite from the H 1 -cell or close to its axon terminal from the H 2 -cell. The spike activity of H 1 and H 2 is increased
during back-to-front motion in the contralateral visual field and elicits EPSPs in HSE (see insets). As a consequence of its input organisation, the right HSE-cell can be expected to be depolarised during counterclockwise rotations of the fly and hyperpolarised during rotations in the opposite direction. (d) Responses to behaviourally generated stimuli. Upper trace: individual response; HSE responds to motion with graded depolarisations and hyperpolarisations; spikes superpose the graded potential changes; the resting potential is indicated by a dashed line. Middle trace: average response. Bottom trace: angular velocity. Sharp angular velocity peaks corresponding to saccade-like turns of the fly dominate the time-dependent angular velocity profile. By contrast to expectations based on the input organisation of the HSE-cell, there are no obvious depolarising response peaks during preferred direction motion evoked by counterclockwise saccades. However, there are brief pronounced hyperpolarisations going along with clockwise saccades; some of these hyperpolarizing peaks and the corresponding saccades are indicated by arrows (Data shown in (a), (b) and (d) from [57*•]; data shown in (c) from [41]).

Hence, synaptic signal transfer appears to be adapted to convey motion information reliably and without distortion in the behaviourally relevant operating range.

## Representation of behaviourally generated optic flow

Knowing about the wiring of a neuronal circuit does not allow us to infer safely how efficiently and reliably information is processed and represented in natural behavioural situations. Two constraints are particularly relevant. The first pertains to the reliability of neural coding. Neurons are inherently noisy elements; accordingly, neuronal responses to identical stimuli may be highly variable. This variability limits the precision and timescale of representing motion information. This important and still-controversial topic has been reviewed recently $[6,47,48]$. The other constraint pertains to the peculiar dynamics of natural optic flow, which are largely determined by the dynamics of the animal's self-motion. The direction of self-motion may change rapidly, such as during saccadic turns during flight [24,49], or an order of magnitude more slowly, as during walking [ $50^{\bullet}$ ]. Natural stimuli thus differ largely from those usually used for the analysis of visual information processing, be it constant velocity stimuli or white-noise velocity fluctuations. Therefore, the neuronal representation of natural optic flow is currently being analysed using different approaches.

Because it is not yet possible to record from neurons in freely moving insects, indirect approaches have been used in flies to assess the responses of TCs to behaviourally generated optic flow. Recordings have been made from the brain of flies that were rotated in a natural outdoor setting about their vertical body-axis [51•], partly with a dynamic mimicking the rotational selfmotion component experienced in free flight [52 ${ }^{\bullet}$ ]. In another approach, the optic flow experienced by moving flies was reconstructed and subsequently replayed to animals during nerve-cell recordings. This approach has been employed for various behavioural situations during tethered flight in a flight simulator [53-55], during unrestrained walking $\left[50^{\bullet}, 56\right]$ and most recently during free flight in a three-dimensional environment [57••]. The simulation of free flight has been possible thanks to the development of sophisticated techniques (Figure 3a,b). First, free-flight behaviour can be monitored with unprecedented resolution by means of sensor-coils [24,49]. Second, a panoramic visual stimulator for presentation of optic flow that is sufficiently fast for visual stimuli as experienced by a fly during rapid saccade-like turns has been designed [57*•]. Although it would be premature to draw general conclusions on the basis of the studies on processing of behaviourally generated optic flow, it might be safe to conclude that the neuronal responses to complex optic flow as experienced during unrestrained locomotion can be understood only partly in terms of the concepts that were established on the basis of experiments done with conventional motion stimuli.

One example is shown in Figure 3. As judged from its input organisation, the HSE-cell (horizontal system equatorial cell), a major output element of the fly visual-motion pathway, is expected to respond best during rotations of the animal about its vertical body-axis (Figure 3c; [41]). The responses to optic flow experienced by free-flying flies only partly fit these expectations [570॰]. The cell responds with graded depolarisations superimposed by spikes almost during the entire flight sequence. Saccades going along with optic flow in the cell's null direction lead to brief pronounced hyperpolarisations. By contrast, saccades eliciting preferred direction motion do not lead to corresponding depolarisations. This is not merely because of saturation of the response, because saccades fail to elicit depolarisations even during phases of the flight sequence in which the overall depolarisation of the cell is much smaller than the maximal depolarisation level that can be evoked by visual motion (Figure 3d). This example illustrates that it is difficult to predict the performance of the system during complex flight manoeuvres, even when wiring diagrams and responses to simplified optic-flow stimuli are well established.

Understanding these complex responses might be further complicated, because the properties of fly TCs were shown to change as a result of stimulus history [58-62,63•, $64^{\bullet}$ ]. Although the functional significance of these adaptive processes is still debated, they might play a role in adjusting the operating range of the mechanisms underlying opticflow processing in different behavioural contexts.

## Conclusions and perspectives

In spite of their tiny brains, insects are able to solve sophisticated visual orientation problems sufficiently fast and reliably to make them the most successful phylum in terms of number of species and biomass. This might have been accomplished because the insect brain appears to be no general-purpose information-processing device, but is kept as simple as possible by adapting the visual system to the specific needs encountered in normal life. During the review period, it became increasingly clear that, on its own, systems analysis of the processing of visual information with conventional stimuli is not sufficient to assess which aspects of the environment, as well as the animal's own behaviour, are encoded by neuronal circuits. Rather, the functional significance of neural computations might become evident only if visual information processing is viewed also from the perspective of sensory and behavioural ecology. To learn how nervous systems solve visual orientation problems in an efficient and parsimonious way, it is necessary to know about both the neuronal circuits and the conditions under which they operate.

Because of the efficiency of visually guided orientation in insects, there is great interest in applying principles of insect motion information processing to autonomous artificial systems. Although this has been successful for some behavioural components [65-70], many biomorphic
autonomous robots appear still to be dull compared with the original after which they are modelled. By contrast to man-made systems, however, natural vision systems have been tested and improved on a much longer timescale by many millions of years of evolution.

## Update

In a recent paper [ $71^{\bullet \bullet}$ ] a novel flight simulator is described that combines realistic, interactive visual environments with mechanosensory and olfactory stimuli in conjunction with multichannel neurophysiological recording. Initial experiments reveal that this system, used on tethered flying insects, is potentially very useful to examine activity from groups of neurons during realistic closed-loop behaviour in virtual environments. As the visual image flow is presented at a frame rate of only 60 frames $/ \mathrm{s}$, the system is suitable for insects with relatively slow photoreceptors. Moreover, image motion should not be too fast to avoid aliasing problems. All these conditions appear to be perfectly satisfied for the nocturnal moths for which the flight simulator has been developed.

## Acknowledgements

We are grateful to N Böddeker, J Grewe, K Karmeier and R Kurtz for discussing the paper critically. The work done in the authors' laboratory is supported by the Deutsche Forschungsgemeinschaft (DFG).

## References and recommended reading

Papers of particular interest, published within the annual period of review, have been highlighted as:

- of special interest
$\bullet \bullet$ of outstanding interest

1. Srinivasan MV, Poteser M, Kral K: Motion detection in insect orientation and navigation. Vis Res 1999, 39:2749-2766.
2. Egelhaaf M, Warzecha A-K: Encoding of motion in real time by the fly visual system. Curr Opin Neurobiol 1999, 9:454-460.
3. Rind FC, Simmons PJ: Seeing what is coming: building collisionsensitive neurones. Trends Neurosci 1999, 22:215-220.
4. Srinivasan MV, Zhang S-W: Visual navigation in flying insects. Int Rev Neurobiol 2000, 44:67-92.
5. Frye MA, Dickinson MH: Fly flight: a model for the neural control of complex behavior. Neuron 2001, 32:385-388.
6. Egelhaaf M, Kern R, Krapp HG, Kretzberg J, Warzecha A-K: Neural encoding of behaviourally relevant motion information in the fly. Trends Neurosci 2002, 25:96-102.
7. Borst A, Haag J: Neural networks in the cockpit of the fly. J Comp Physiol [A] 2002, 188:419-437.
8. Collett TS, Zeil J: Flights of learning. Curr Dir Psychol Sci 1996, 5:149-155.
9. Collett TS, Collett M: Memory use in insect visual navigation. Nature Rev Neurosci 2002, 3:542-552.
10. Götz KG: The optomotor equilibrium of the Drosophila navigation system. J Comp Physiol 1975, 99:187-210.
11. Kern R, Egelhaaf M: Optomotor course control in flies with largely asymmetric visual input. J Comp Physiol [A] 2000, 186:45-55.
12. Kern R, Lutterklas M, Egelhaaf M: Neural representation of optic flow experienced by unilaterally blinded flies on their mean walking trajectories. J Comp Physiol [A] 2000, 186:467-479.
13. Tammero LF, Dickinson MH: The influence of visual landscape on

- the free flight behavior of the fruit fly Drosophila melanogaster. J Exp Biol 2002, 205:327-343.
It is shown that behaviour of Drosophila is directly affected by the optic flow induced by self-motion. Both saccadic turns and gradual turns of the animal
are evoked by asymmetries in the global optic-flow pattern. Interestingly, the behavioural responses are in contrast to what is expected from the optomotor turning response that can be recorded in tethered animals, but also affects the behaviour of freely walking blowflies [11,12].

14. Ronacher B, Gallizzi K, Wohlgemuth S, Wehner R: Lateral optic flow does not influence distance estimation in the desert ant cataglyphis fortis. J Exp Biol 2000, 203:1113-1121.
15. Wohlgemuth S, Ronacher B, Wehner R: Ant odometry in the third dimension. Nature 2001, 411:795-798.
Desert ants were trained within an artificial landscape of uphill and downhill channels and later tested on a flat terrain, or vice versa. Ants indicated distances to their nest that did not agree with the distances actually travelled but to the corresponding ground distances.
16. Esch HE, Burns JM: Distance estimation by foraging honeybees. $J$ Exp Biol 1996, 199:155-162.
17. Srinivasan MV, Zhang S, Altwein M, Tautz J: Honeybee navigation:
-• nature and calibration of the "odometer". Science 2000, 287:851-853.
Honeybees were trained to fly through narrow tunnels to reach a food source. These bees communicate by their waggle dance an exaggerated distance to their hive mates. This finding provides strong evidence that travelled distance is gauged in terms of optic flow experienced while flying from the hive to a food source. Although this distance measure is ambiguous, navigational errors are avoided, because recruited bees tend to fly the same route as the dancers.
18. Esch HE, Zhang S, Srinivasan MV, Tautz J: Honeybee dances
-• communicate distances measured by optic flow. Nature 2001, 411:581-583.
See annotation to $\left[17^{\circ}{ }^{\circ}\right]$.
19. Land MF: Chasing and pursuit in the dolichopodid fly Poecilobothrus nobilitatus. J Comp Physiol [A] 1993, 173:605-613.
20. Collett TS, Land MF: Visual control of flight behaviour in the hoverfly Syritta pipiens L. J Comp Physiol 1975, 99:1-66.
21. Land MF, Collett TS: Chasing behaviour of houseflies (Fannia canicularis). A description and analysis. J Comp Physiol 1974, 89:331-357.
22. Wagner H : Flight performance and visual control of the flight of the free-flying housefly (Musca domestica). II. Pursuit of targets. Phil Trans R Soc Lond B 1986, 312:553-579.
23. Olberg RM, Worthington AH, Venator KR: Prey pursuit and interception in dragonflies. J Comp Physiol [A] 2000, 186:155-162.
24. Schilstra C, Hateren JHv: Blowfly flight and optic flow. I. Thorax kinematics and flight dynamics. J Exp Biol 1999, 202:1481-1490.
25. Boeddeker N, Kern R, Egelhaaf M: Finding Ms Right: smooth

- pursuit guides male blowflies to their target. Proc R Soc Lond B 2002, in press.
The control system underlying the chasing behaviour of male blowflies was unravelled at a phenomenological level by reducing the complexity of real flights and by using black spheres as dummy targets. Most importantly, it was shown that flies are equipped with a smooth pursuit system that is similar, in many respects, to that of primates.

26. Land MF: Variations in the structure and design of compound eyes. In Facets of Vision. Edited by Stavenga D, Hardie RC. Berlin: Springer; 1989:90-111.
27. Hornstein EP, O'Carroll DC, Anderson JC, Laughlin SB: Sexual

- dimorphism matches photoreceptor performance to behavioural requirements. Proc $R$ Soc Lond Ser B 2000, 267:2111-2117.
By measuring both the spatial and the temporal properties of photoreceptors at different positions across the compound eye of male and female blowflies, it was shown that frontal photoreceptors of males have a better spatiotemporal resolution than that of lateral ones, as well as those of females. The authors conclude that this feature is an adaptation to the visual control of chasing behaviour.

28. Burton BG, Tatler BW, Laughlin SB: Variations in photoreceptor

- response dynamics across the fly retina. J Neurophysiol 2001, 86:11.
See annotation to [27•].

29. Hardie RC: The photoreceptor array of the dipteran retina. Trends Neurosci 1986, 9:419-423.
30. Hausen K, Strausfeld NJ: Sexually dimporphic interneuron arrangements in the fly visual system. Proc $R$ Soc Lond Ser $B$ 1980, 208:57-71.
31. Strausfeld NJ: Structural organization of male-specific visual neurons in calliphorid optic lobes. J Comp Physiol [A] 1991, 169:379-393.
32. Gilbert C, Strausfeld NJ : The functional organization of malespecific visual neurons in flies. J Comp Physiol [A] 1991, 169:395-411.
33. Wicklein M, Strausfeld NJ: Organization and significance of neurons that detect change of visual depth in the hawk moth Manduca sexta. J Comp Neurol 2000, 424:356-376.
34. Gabbiani F, Krapp HG, Laurent G: Computation of object approach by a wide-field, motion-sensitive neuron. J Neurosci 1999, 19:1122-1141.
35. Gabbiani F, Mo C, Laurent G: Invariance of angular threshold - computation in a wide-field looming-sensitive neuron. J Neurosci 2001, 21:314-329.
This paper proposes a computational model of the responses of an identified wide-field looming-sensitive neuron of locusts. The model reproduces the invariance of angular threshold computation accomplished by the neuron.
36. Gray JR, Lee JK, Robertson RM: Acitivity of descending - contralateral movement detector neurons and collision avoidance behaviour in response to head-on visual stimuli in locusts. $J$ Comp Physiol [A] 2001, 187:115-129.
This study compares the performance of an identified wide-field loomingsensitive neuron of locusts with behavioural reactions recorded in tethered flight. It is concluded that the neuron is better suited to predator evasion than to collision avoidance. Although the paper leaves several questions open, it is one of the rare attempts to relate the performance of this well-known neuronal system to visually guided behaviour.
37. Krapp HG: Neuronal matched filters for optic flow processing in flying insects. In Neuronal Processing of Optic Flow. Edited by Lappe M. San Diego: Academic Press; 2000:93-120.
38. Karmeier K, Egelhaaf M, Krapp HG: Early visual experience and receptive field organization of the optic flow processing interneurons in the fly motion pathway. Vis Neurosci 2001, 18:1-8.
39. Hausen K: Monocular and binocular computation of motion in the lobula plate of the fly. Verh Dtsch Zool Ges 1981, 74:49-70.
40. Warzecha A-K, Egelhaaf M, Borst A: Neural circuit tuning fly visual interneurons to motion of small objects. I. Dissection of the circuit by pharmacological and photoinactivation techniques. J Neurophysiol 1993, 69:329-339.
41. Horstmann W, Egelhaaf M, Warzecha A-K: Synaptic interactions increase optic flow specificity. Europ J Neurosci 2000, 12:2157-2165.
42. Krapp HG, Hengstenberg R, Egelhaaf M: Binocular contribution to

- optic flow processing in the fly visual system. J Neurophysiol 2001, 85:724-734.
The specificity of motion-sensitive neurons for certain types of optic flow is surveyed on the basis of a close inspection of their local receptive field properties, and in the light of heterolateral connections (see also [41,43•]).


## 43. Haag J, Borst A: Recurrent network interactions underlying flow

 - field selectivity of visual interneurons. J Neurosci 2001, 21:5685-5692.Dual recordings lead the authors a step further in the understanding of the connectivity between visual motion-sensitive neurons in the fly. As is also pointed out in [41] and [42*], the connections within one circuit seem to be dedicated to increasing the specificity of its output element for rotatory versus translatory optic flow.
44. Haag J, Borst A: Dendro-dendritic interactions between motion -. sensitive large-field neurons in the fly. J Neurosci 2002, 22:3227-3233.
The characteristics of synaptic connections between visual motion-sensitive neurons in the fly brain are analysed. The methodologically challenging combination of dual recordings with optical imaging helped not only to demonstrate the existence of connections but also to uncover their spatial arrangement.
45. Kurtz R, Warzecha A-K, Egelhaaf M: Transfer of visual information -. via graded synapses operates linearly in the natural activity range. J Neurosci 2001, 21:6957-6966.
Synaptic transfer of visual motion information is analysed by dual recording, partly combined with calcium imaging [44*•]. The analysed synaptic connection that conveys visual motion information gathered by motion-sensitive neurons in one half of the brain via another neuron to the contralateral visual system was found to operate linearly and very reliably within a certain dynamic range. This range corresponds to the one in which the gain of the
visual motion pathway for signalling motion information is large [see also 46•].
46. Warzecha A-K, Kurtz R, Egelhaaf M: Synaptic transfer of dynamical

- motion information between identified neurons in the visual system of the blowfly. J Neurosci 2002, (submitted).
See annotation to [ $45^{\bullet \bullet}$ ].

47. Warzecha A-K, Egelhaaf M: Neuronal encoding of visual motion in real-time. In Processing Visual Motion in the Real World: A Survey of Computational, Neural, and Ecological Constraints. Edited by Zanker JM, Zeil J. Berlin: Springer; 2001:239-277.
48. Ruyter van Steveninck Rd, Borst A, Bialek W: Real-time encoding of motion: answerable questions and questionable answers from the fly's visual system. In Motion Vision. Edited by Zanker JM, Zeil J. Berlin: Springer; 2001:279-306.
49. Hateren JH, Schilstra C: Blowfly flight and optic flow. II. Head movements during flight. J Exp Biol 1999, 202:1491-1500.
50. Kern R, Petereit C, Egelhaaf M: Neural processing of naturalistic - optic flow. J Neurosci 2001, 21:RC139 1-5.

In flies, responses of a visual motion-sensitive neuron to optic-flow stimuli generated during walking were analysed. The neuron signals turning direction largely independently of specific environmental characteristics. This invariance might result from the spatiotemporal properties of natural optic flow evoked during walking, because when simpler stimuli were used, the response of the neuron was influenced by the specific textural properties of the stimuli.
51. Egelhaaf M, Grewe J, Kern R, Warzecha A-K: Outdoor performance

- of a motion-sensitive neuron in the blowfly. Vis Res 2001, 41:3627-3637.
The activity of a fly neuron is shown to remain constant over the broad range of lighting conditions as they occur during the day in a natural environment, but to depend considerably on ambient temperature.

52. Lewen GD, Bialek W, Ruyter van Steveninck Rd: Neural coding of - naturalistic motion stimuli. Network: Comput Neural Syst 2001, 12:317-329.
Electrophysiological analysis of the reliability with which naturalistic body rotations are encoded by a fly motion-sensitive neuron. The fly was rotated at different times of the day in an outdoor environment with an electrode inserted into its brain. Although some of the conclusions are controversial [ $51^{\circ}$ ], the study represents a methodological breakthrough.
53. Warzecha A-K, Egelhaaf M: Intrinsic properties of biological motion detectors prevent the optomotor control system from getting unstable. Phil Trans R Soc Lond B 1996, 351:1579-1591.
54. Warzecha A-K, Egelhaaf M: How reliably does a neuron in the visual motion pathway of the fly encode behaviourally relevant information? Europ J Neurosci 1997, 9:1365-1374.
55. Kimmerle B, Egelhaaf M: Performance of fly visual interneurons during object fixation. J Neurosci 2000, 20:6256-6266.
56. Kern R, Lutterklas M, Petereit C, Lindemann JP, Egelhaaf M: Neuronal processing of behaviourally generated optic flow: experiments and model simulations. Network: Computations in Neural Systems 2001, 12:351-369.
57. Lindemann JP, Kern R, Petereit C, Meyer P, van Hateren JH,
-. Egelhaaf M: Flimax, a novel stimulus device for panoramic and highspeed presentation of behaviourally generated optic flow. Vision Res 2002, (submitted).
This is the first study in which the optic flow as seen by a fly during flight is used for visual stimulation in electrophysiological experiments. For this to be possible, a high-speed visual stimulator had to be developed. The study shows that it is not easily possible to derive how this circuit represents complex visual information characteristic of natural conditions from the knowledge of the wiring diagram of a neuronal circuit.
58. Maddess T, Laughlin SB: Adaptation of the motion-sensitive neuron H1 is generated locally and governed by contrast frequency. Proc R Soc Lond Ser B 1985, 225:251-275.
59. de Ruyter van Steveninck R, Zaagman WH, Mastebroek HA: Adaptation of transient responses of a movement-sensitive neuron in the visual system of the blowfly, Calliphora erythrocephala. Biol Cybern 1986, 54:223-236.
60. Borst A, Egelhaaf M: Temporal modulation of luminance adapts time constant of fly movement detectors. Biol Cybern 1987, 56:209-215.
61. Harris RA, O'Carroll DC, Laughlin SB: Adaptation and the temporal delay filter of fly motion detectors. Vis Res 1999, 39:2603-2613.
62. Harris RA, O'Carroll DC, Laughlin SB: Contrast gain reduction in fly motion adaptation. Neuron 2000, 28:595-606.
63. Fairhall AL, Lewen GD, Bialek W: Ruyter van Steveninck Rd:

- Efficiency and ambiguity in an adaptive neural code. Nature 2001, 412:787-792.
Elaborated systems analysis on a motion-sensitive neuron of the fly. Potentially adaptive changes in the input/output relation of the neuron are investigated on a range of different timescales.

64. Harris RA, O'Carroll DC: Afterimages in fly motion vision. Vision - Res 2002, 42:1701-1714.

Afterimage-like effects shape the responses of fly motion-sensitive neurons when motion stimuli are preceded by slowly moving or static patterns. The authors show that some of these effects can be replicated by a simple modification of the classical correlation detector model of motion vision.
65. Srinivasan MV, Chahl JS, Zhang SW: Embodying natural vision into machines. In From Living Eyes to Seeing Machines. Edited by Srinivasan MV, Venkatesh S. Oxford: Oxford University Press; 1997:249-265.
66. Huber SA, Franz MO, Bülthoff HH: On robots and flies: modeling the visual orientation behavior of flies. Robotics and Autonomous Systems 1999, 29:227-242.
67. Franz MO, Mallot HA: Biomimetic robot navigation. Robots and Autonomous Systems 2000, 30:133-153.
68. Harrison RR, Koch C: A silicon implementation of the fly's optomotor control system. Neural Comput 2000, 12:2291-2304.
69. Srinivasan MV, Zhang S, Chahl JS: Landing strategies in honeybees, and possible applications to autonomous airborne vehicles. Biol Bull 2001, 200:216-221.
70. Rind FC: Motion detectors in the locust visual system: from biology to robot sensors. Microsc Res Tech 2002, 56:256-269.
71. Gray JR, Pawlowski V, Willis MA: A method for recording behavior
-. and multichannel CNS activity from tethered insects flying in virtual space. J Neurosci Methods 2002, 120:211-223.

