

Syddansk Universitet

## Ancient microbial activity in deep hydraulically conductive fracture zones within the forsmark target area for geological nuclear waste disposal, Sweden

Drake, Henrik; Ivarsson, Magnus ; Tillberg, Mikael; Whitehouse, Martin J.; Kooijman, Ellen

*Published in:*  
Geosciences (Switzerland)

*DOI:*  
[10.3390/geosciences8060211](https://doi.org/10.3390/geosciences8060211)

*Publication date:*  
2018

*Document version*  
Publisher's PDF, also known as Version of record

*Document license*  
CC BY

### *Citation for pulished version (APA):*

Drake, H., Ivarsson, M., Tillberg, M., Whitehouse, M. J., & Kooijman, E. (2018). Ancient microbial activity in deep hydraulically conductive fracture zones within the forsmark target area for geological nuclear waste disposal, Sweden. *Geosciences (Switzerland)*, 8(6), [211]. DOI: 10.3390/geosciences8060211

### **General rights**

Copyright and moral rights for the publications made accessible in the public portal are retained by the authors and/or other copyright owners and it is a condition of accessing publications that users recognise and abide by the legal requirements associated with these rights.


- Users may download and print one copy of any publication from the public portal for the purpose of private study or research.
- You may not further distribute the material or use it for any profit-making activity or commercial gain
- You may freely distribute the URL identifying the publication in the public portal ?

### **Take down policy**

If you believe that this document breaches copyright please contact us providing details, and we will remove access to the work immediately and investigate your claim.

Article

# Ancient Microbial Activity in Deep Hydraulically Conductive Fracture Zones within the Forsmark Target Area for Geological Nuclear Waste Disposal, Sweden

Henrik Drake <sup>1,\*</sup> , Magnus Ivarsson <sup>2</sup>, Mikael Tillberg <sup>1</sup>, Martin J. Whitehouse <sup>3</sup> and Ellen Kooijman <sup>3</sup>

<sup>1</sup> Department of Biology and Environmental Science, Linnaeus University, 392 31 Kalmar, Sweden; mikael.tillberg@lnu.se

<sup>2</sup> Department of Biology, University of Southern Denmark, Campusvej 55, 5230 Odense, Denmark; magnus.ivarsson@nrm.se

<sup>3</sup> Department of Geosciences, Swedish Museum of Natural History, Box 50 007, 104 05 Stockholm, Sweden; martin.whitehouse@nrm.se (M.J.W.); ellen.kooijman@nrm.se (E.K.)

\* Correspondence: henrik.drake@lnu.se; Tel.: +46-480-447-300

Received: 24 April 2018; Accepted: 7 June 2018; Published: 11 June 2018



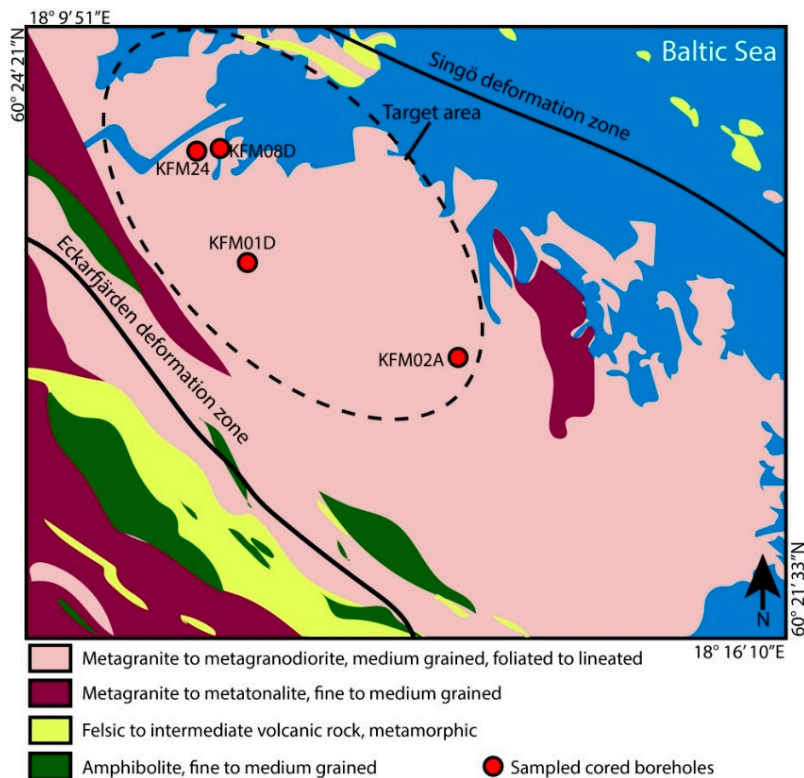
**Abstract:** Recent studies reveal that organisms from all three domains of life—Archaea, Bacteria, and even Eukarya—can thrive under energy-poor, dark, and anoxic conditions at large depths in the fractured crystalline continental crust. There is a need for an increased understanding of the processes and lifeforms in this vast realm, for example, regarding the spatiotemporal extent and variability of the different processes in the crust. Here, we present a study that set out to detect signs of ancient microbial life in the Forsmark area—the target area for deep geological nuclear waste disposal in Sweden. Stable isotope compositions were determined with high spatial resolution analyses within mineral coatings, and mineralized remains of putative microorganisms were studied in several deep water-conducting fracture zones (down to 663 m depth), from which hydrochemical and gas data exist. Large isotopic variabilities of  $\delta^{13}\text{C}_{\text{calcite}}$  (−36.2 to +20.2‰ V-PDB) and  $\delta^{34}\text{S}_{\text{pyrite}}$  (−11.7 to +37.8‰ V-CDT) disclose discrete periods of methanogenesis, and potentially, anaerobic oxidation of methane and related microbial sulfate reduction at several depth intervals. Dominant calcite–water disequilibrium of  $\delta^{18}\text{O}$  and  $^{87}\text{Sr}/^{86}\text{Sr}$  precludes abundant recent precipitation. Instead, the mineral coatings largely reflect an ancient archive of episodic microbial processes in the fracture system, which, according to our microscale Rb–Sr dating of co-genetic adularia and calcite, date back to the mid-Paleozoic. Potential Quaternary precipitation exists mainly at ~400 m depth in one of the boreholes, where mineral–water compositions corresponded.

**Keywords:** deep biosphere; crystalline crust; stable isotopes; methanogenesis; sulfate reduction; calcite; pyrite; bedrock fractures; nuclear waste disposal

## 1. Introduction

The deep continental biosphere is a potentially vast microbial habitat, and it has been estimated that 2–19% of the global biomass may be accumulated within it [1]. However, our knowledge of this biosphere is still scarce, largely owing to the difficulty of accessing samples (cost efficiently) from this realm. Observations have, therefore, been limited to a few sites with deep boreholes or to underground constructions, such as research tunnel facilities and deep mines. Studies of modern microbial activity in the continental deep biosphere include, but are not limited to, works in South

African mines [2–4], Canadian mines [5], deep research boreholes, and underground facilities in Sweden [6–10], Finland [11–15], and Japan [16,17], showing a high degree of site-specificity on the microbial community profiles. A diverse biosphere consisting of archaea, bacteria, and recently, also eukaryotes (mainly fungi), has been detected at considerable depths; several kilometers for prokaryotes [4,18], and at least 700–800 m for fungi [19,20]. In general, the metabolisms tend to shift from dominantly heterotrophic, at shallow depth in the crust, to autotrophic at greater depth [18,21,22]. In addition, compositions of deep gases (e.g., methane) and dissolved compounds in groundwater have been used as indicators for microbial activity and for related substrate sources [13,23–28]. Signs of ancient microbial activity have been surveyed by isotopic investigations of minerals formed in response to microbial activity in deep fracture systems [29–32], and by detection of fossilized microorganisms, and preserved organic remains [30,33,34] and their compound specific isotope signatures [35,36]. Diagnostic C isotope signatures for anaerobic oxidation of methane have been detected in calcite from several hundred meters depth at different Swedish sites using high-spatial resolution analysis within minerals, and these findings that date back to Paleozoic times [29,30]. Similarly, sulfur isotope signatures diagnostic for microbial sulfate reduction have been shown to be widespread in pyrite precipitated in the fracture systems at the Laxemar site in Sweden [37], and the Olkiluoto site in Finland [38]. Due to the overall few observations of ancient life signatures in the crystalline continental crust, there is a need for more observations, particularly regarding how widespread the different microbial processes are in space and time in this realm. Here, we present high spatial resolution isotope determinations in secondary minerals from deep water-conducting bedrock fractures in the Forsmark area, Sweden, an area dominated by Paleoproterozoic crystalline bedrock that has been subjected to numerous events of fracture reactivations [39]. This site is the target area for the deep repository for spent nuclear fuel in Sweden [40]. Our study is focused in packed off borehole sections with water-conducting fractures within the target bedrock volume for the repository (Figure 1), in contrast to earlier reconnaissance studies of regional character [32]. Drill core samples of mineral-coated water-conducting fractures were collected from borehole sections, where hydrochemical and gas compositions have been determined. The main focus was on the newly drilled deep pilot borehole for the future skip shaft (borehole KFM24, Figure 1). The objective of the study can be divided into a general aim: to provide new knowledge of microbial processes and their depth distribution over long time spans in the continental deep biosphere; and a site-specific aim: to determine how microbial processes and hydrochemical conditions have varied over time in deep water-conducting fractures within the target area for a nuclear waste repository. At this site, deep Quaternary infiltration of glacial and marine waters is evident [41]. Therefore, the study also explores whether there are any potentially Quaternary mineral precipitates in the deep fracture system, and whether the isotopic signatures in the mineralogical record match the hydrochemical and microbiological record in the waters currently residing in the fractures.



**Figure 1.** Map with sampled boreholes indicated. Geological units modified and slightly simplified from Sandström and Stephens [42].

## 2. Materials and Methods

### 2.1. Materials and Site

Samples were collected from four deep cored boreholes (KFM01D, KFM02A, KFM08D, and KFM24) drilled by the Swedish Nuclear Fuel and Waste Management Co (SKB) within the target repository area for spent nuclear fuel at the Forsmark site in eastern Sweden (Figure 1). The target area is located in a tectonic lens bordered by the regional Singö and Eckerfjärden deformation zones. The rock in the area is Proterozoic in age (crystallization age is 1.89–1.86 Ga [43]), and dominantly consisting of metamorphosed granitoids with subordinate amphibolite and volcanic rocks. Several events of Proterozoic and Paleozoic fracturing and fracture mineralization have been documented at the site [29,32,39,44,45]. The dominating rock type in the boreholes is a medium-grained and foliated metagranite–granodiorite. The fracture frequency for open fractures in the boreholes is 0.6–1.5 fractures/m, and for sealed fractures, 1.0–5.9 fractures/m [46,47].

Seventeen open fractures containing calcite and pyrite (Table 1) were sampled from seven hydraulically conductive fracture zones, where comprehensive hydrochemical characterization (and limited microbial and gas characterization) have been carried out by SKB in packed off borehole sections. Flow log measurements e.g., [48,49] were used to distinguish the most likely conductive fractures for mineral analysis in each packed off section. The quality controlled hydrochemical, gas, and microbiological data from the borehole sections have been extracted from the SKB site database (Sicada) for this study, and are presented in Table 2 and Table S1, and the methodology for sampling and analysis is described in detail elsewhere [50–52].

The groundwaters in the sampled sections are according to the site coding [50] of brackish non-marine origin (KFM01D:431, KFM08D:670 first sampling in time series, and KFM08D:830) or a mixture of the latter water type with glacial water (all sections in KFM24, and KFM08D:670 second sampling in time series). In KFM01D:316 there is a post-glacial brackish marine water

component (post-glacial Littorina sea stage) in addition to the glacial/brackish non-marine water. These compositions reveal that there has been Quaternary infiltration and mixing of different surficial water types at large depth in the fracture system, which may have triggered mineral supersaturation and precipitation. For the KFM02A samples, hydrochemical data from a nearby fracture is used. The Cl concentrations in the boreholes vary from 2000 mg/L in the shallow KFM24 section (195 m) to 8000 mg/L in the deep section in KFM08D (830 m). The dissolved inorganic carbon ( $\text{HCO}_3^-$ ) and sulfate concentrations are highest (75 and 273 mg/L, respectively) in the KFM01D:316 section, which has a Littorina Sea stage component, whereas the other sections have concentrations in the ranges 6–36 and 9–125 mg/L, respectively. Dissolved organic carbon (DOC) varied from below detection (1 mg/L) to 14.7 mg/L (Table 2).  $\text{Ca}^{2+}$  occurred at 400–3000 mg/L.  $\text{Mn}^{2+}$  and  $\text{Fe}^{2+}$  occurred in detectable amounts in most waters, which is in line with what is expected in deep anoxic waters. From a structural and hydrogeological perspective, the borehole sections with minor fracture zones and single conductive fractures sampled in KFM01D and KFM24 are most representative for the target area, whereas the samples from KFM08D are from a major fracture zone.

During sampling and further preparation, the samples were treated with gloved hands and sterile tools like tweezers and chisels to minimize introduction of post-drilling contaminations.

The number of cells of microorganisms in the waters were previously presented for three borehole sections [53], and were highest in KFM01D:431 (total number of 250,000 cells/mL, based on staining technique), whereas KFM08D:670 and 830 had lower counts (21,000 and 11,000, respectively) [8]. These estimates are based on most probable number method, and for comparison, more recent analysis by direct cell counts from boreholes with similar DOC levels at the Äspö Hard Rock Laboratory, Sweden, place the cell numbers in the range of 100s per mL [6]. Methanogens were around or below detection limit (0.2 cells/mL). Sulfate-reducing bacteria (SRB) were in the range 2–13 cells/mL, whereas acetogens, and bacteria that reduce nitrate, iron, and manganese had one to three orders of magnitude higher cell counts (all based on viable counts).

## 2.2. Sample Preparation and Scanning Electron Microscopy

Sample characterization and mineral identification was carried out directly on the fracture surfaces of the core samples using a Hitachi S-3400N SEM (Hitachi, Tokyo, Japan) equipped with an integrated energy dispersive spectroscopy (EDS) system under low-vacuum conditions. The pyrite and calcite crystals were then handpicked from the fracture surface under the microscope, embedded in epoxy mounts ( $\varnothing = 1$  inch), and polished to expose cross-sections of the crystal interiors. The polished crystals had sizes from 20 to 500  $\mu\text{m}$ , and were examined with SEM prior to isotopic analysis in order to detect zonations, impurities, and cracks. Low-temperature K-feldspar (adularia) crystals, being co-genetic with pyrite and calcite (as revealed by SEM investigation), were handpicked from a fracture surface and used for LA-ICP-MS in situ Rb–Sr dating. Characterization of fossilized microorganisms was carried out by environmental scanning electron microscopy (ESEM) directly on the fracture surface using a Philips XL30 microscope (Semtech Solutions, Billerica, MA, USA) with a field emission gun (XL30 ESEM-FEG) equipped with an Oxford x-act EDS, backscatter electron detector, and a secondary electron detector (Oxford corp., Oxford, England). The acceleration voltage was 20 kV, and low-vacuum conditions were used on uncoated samples. The instrument was calibrated with a cobalt standard. Peak and element analyses were made using the Aztec software (Oxford corp.). Since the cores can be colonized by microorganisms during storage, this characterization was only done on the newly retrieved core, KFM24, shortly after drilling. In addition, in order to minimize the potential influence from contamination and post-drilling colonization, characterization focused on partly sealed fractures that were mechanically opened with a chisel just prior to introduction into the SEM.

**Table 1.** Isotopic variability of the calcite and pyrite samples.

Borehole	Sample	Vertical Depth (m)	$\delta^{13}\text{C}_{\text{min}}$	$\delta^{13}\text{C}_{\text{max}}$	$n_{\delta^{13}\text{C}}$	$\delta^{18}\text{O}_{\text{min}}$	$\delta^{18}\text{O}_{\text{max}}$	$n_{\delta^{18}\text{O}}$	$^{87}\text{Sr}/^{86}\text{Sr}_{\text{min}}$	$\pm$	$^{87}\text{Sr}/^{86}\text{Sr}_{\text{max}}$	$\pm$	$n_{^{87}\text{Sr}/^{86}\text{Sr}}$	$\delta^{34}\text{S}_{\text{min}}$	$\delta^{34}\text{S}_{\text{max}}$	$n_{\delta^{34}\text{S}}$
			‰, V-PDB	‰, V-PDB		‰, V-PDB	‰, V-PDB							‰, V-CDT	‰, V-CDT	
KFM01D	316	−253.27	−7.4	−2.8	15	−12.7	−5.9	15	0.7106	0.0016	0.7118	0.0012	2			0
KFM01D	431	−341.50	−10.9	−4.5	13	−13.4	−6.2	18	0.7054	0.0025			1			0
KFM02A	162	−155.29	0.0	11.3	10			0					0			0
KFM02A	175	−167.94	−31.1	−8.7	10	−12.9	−10.3	7	0.7142	0.0010			1			0
KFM08D	677	−544.25	−12.6	−0.5	21	−14.5	−8.0	21	0.7097	0.0011	0.7124	0.0012	10			0
KFM08D	678	−544.48	−8.9	−2.9	25	−8.0	−17.9	18					0			0
KFM08D	830	−662.86	−4.5	−4.2	3	−16.1	−8.3	5					0	−3.5	−1.3	13
KFM08D	832	−664.86			0			0					0	0.0	12.4	8
KFM24	193	−191.00	−9.1	18.5	4	−13.1	−8.2	7					0			0
KFM24	194	−192.00	−16.1	−4.9	13	−16.1	−7.4	21	0.7182	0.0012			1			0
KFM24	379	−376.00	−17.1	20.2	30	−13.1	−7.9	16	0.7176	0.0033			1	−11.7	37.8	24
KFM24	396	−392.00	−36.2	−10.4	24	−13.7	−9.8	11	0.7143	0.0016	0.7159	0.0023	2	−1.3	36.0	29
KFM24	398	−394.00	−32.6	−3.0	13	−13.9	−8.6	13	0.7114	0.0031			1	−2.4	21.2	27
KFM24	399	−395.00	−35.8	−10.6	23	−12.4	−7.1	16	0.7089	0.0031	0.7161	0.0025	4	−0.7	31.0	26
KFM24	410	−406.00	−29.3	17.6	17	−14.7	−7.0	19	0.7150	0.0033	0.7197	0.0024	3			0
KFM24	413	−409.53	−29.8	−9.9	16	−15.0	−6.2	18	0.7184	0.0034			1	−11.5	29.0	25
KFM24	414	−410.53	−17.4	−12.0	15	−14.8	−12.1	15	0.7105	0.0029	0.71603	0.00076	3	7.1	28.4	27
Total			−36.2	20.2	252	−16.1	−5.9	220	0.7054	0.0025	0.7197	0.0024	30	−11.7	37.8	179

V-PDB = Vienna Pee Dee Belemnite, V-CDT = Vienna Canyon Diablo Troilite.

**Table 2.** Chemical composition (concentrations and isotopes) of waters and gases.

Borehole	Sampling Date	Upper Packer	Lower Packer	Na	K	Ca	Mg	HCO <sub>3</sub>	Cl	SO <sub>4</sub>	Fe	Mn	Sr	DOC	pH (lab)	S <sub>2</sub> in HS	δ <sup>13</sup> C	δ <sup>34</sup> S <sub>SO4</sub>	<sup>87</sup> Sr/ <sup>86</sup> Sr	δ <sup>2</sup> H	δ <sup>18</sup> O	Temp	
<i>Groundwaters</i>				(m)	(m)	mg/L	mg/L	mg/L	mg/L	mg/L	mg/L	mg/L	mg/L	mg/L	mg/L	mg/L	‰, V-PDB	‰, V-CDT		‰, V-SMOW	‰, V-SMOW	°C	
KFM01D	2015	312.50	319.61	1790	17.4	1080	90.9	75.9	4466	273	0.804	0.344	11.4	3.3	7.73	0.29	−11.40	28.9	0.720596	−70.8	−10.01		
KFM01D	2006	428.50	435.64	1550	9.02	1430	19.5	35.8	4940	125	1.93	0.182	17.4	3.7	7.55	0.01	−13.20	26.7	0.720149	−74.9	−10.70		
KFM01D	2015	428.50	435.64	1750	7.20	1610	18.4	14.7	5272	67.2	0.0437	0.0772	19.1	4.6	7.81	3.22		42.9	0.720040	−72.6	−11.00		
KFM08D	2007	669.70	676.84	1900	5.42	2740	4.9	6.98	7460	101	0.0044	0.0498	29.3	b.d.	8.14	b.d.		31.6	0.717523	−84.0	−12.30		
KFM08D	2013	669.70	676.84	1780	7.23	2080	10.4	10.0	6455	75.4	b.d.	0.114	22.8	0.7	8.40	b.d.	−4.50	36.6	0.718009	−84.1	−13.11		
KFM08D	2007	828.40	835.54	1990	6.26	2960	8.5	9.07	8160	156	0.114	0.0725	33.1	b.d.	7.93	0.07		26.6	0.717524	−82.5	−11.60		
KFM24	2016	191.50	198.62	817	4.79	411	11.3	21.0	1965	44.0	0.789	0.119	4.5	9.3	7.20	0.17	−15.90	33.2	0.719201	−107.8	−14.40	15.7	
KFM24	2016	395.50	402.62	1500	3.08	1740	−0.5	6.80	5148	15.1	b.d.	0.00397	20.0	2.0	8.30	b.d.		48.5 *	0.719778 *	−93.4	−13.09	16.1	
KFM24	2016(nov)	410.00	417.12	1600	3.27	1930	−0.5	6.50	5564	8.94	0.0049	0.00071	22.4	2.0	8.83	0.29		57.5	0.719968	−91.8	−13.41	15.9	
KFM24	2016(dec) **	410.00	417.12	1460	3.60	1730	1.2	9.60	5032	15.8	b.d.	0.0066	19.9	14.7	8.83	0.14	−21.90	37.6	0.719916	−91.5	−3.21	15.4	
Borehole	Sampling Date	Upper Packer	Lower Packer	He	Ar	N <sub>2</sub>	CO <sub>2</sub>	O <sub>2</sub>	CH <sub>4</sub>	H <sub>2</sub>	C <sub>2</sub> H <sub>2</sub>	C <sub>2</sub> H <sub>4</sub>	C <sub>2</sub> H <sub>6</sub>	C <sub>3</sub> H <sub>4</sub>	C <sub>3</sub> H <sub>6</sub>	C <sub>3</sub> H <sub>8</sub>	C <sub>1</sub> /(C <sub>2</sub> +C <sub>3</sub> )						
<i>Gases, Concentrations</i>				(mL/L)	(mL/L)	(mL/L)	(mL/L)	(mL/L)	(mL/L)	(uL/L)	(uL/L)	(uL/L)	(uL/L)	(uL/L)	(uL/L)	(uL/L)	(uL/L)						
KFM01D *	2015	312.50	319.61	11	b.d.	133	0.32	b.d.	0.64	3.5	b.d.	b.d.	0.28	b.d.	b.d.	b.d.	2289						
KFM01D	2006	428.50	435.64	26	1.10	64	0.20	0.062	0.14	b.d.	b.d.	0.09	2.5	b.d.	b.d.	0.57	54						
KFM01D	2015	428.50	435.64	23	1.30	91	0.12	8.0	0.41	b.d.	b.d.	b.d.	3.0	b.d.	b.d.	0.62	137						
KFM08D	2007	669.70	676.84	23	1.70	73	0.020	0.035	0.090	b.d.	b.d.	0.10	0.49	b.d.	b.d.	0.20	153						
KFM08D	2013	669.70	676.84	22	1.20	77	0.040	0.25	0.10	b.d.	1.10	1.40	1.4	b.d.	b.d.	0.75	26						
KFM08D	2007	828.40	835.54	24	1.40	85	0.030	b.d.	0.060	b.d.	b.d.	0.11	0.55	b.d.	b.d.	0.22	91						
KFM24	2016	395.50	402.62	10	1.30	93	0.019	1.4	0.19	b.d.	b.d.	b.d.	1.1	b.d.	b.d.	b.d.	173						
KFM24	2016	410.00	417.12	10	1.30	121	0.018	3.7	0.18	b.d.	b.d.	b.d.	1.1	b.d.	b.d.	b.d.	164						
Borehole	Sampling Date	Upper Packer	Lower Packer	δ <sup>2</sup> H <sub>CH4</sub>		δ <sup>13</sup> C <sub>CO2</sub>		δ <sup>13</sup> C <sub>CH4</sub>		δ <sup>18</sup> O <sub>CO2</sub>													
<i>Gases, Isotopes</i>				(m)	(m)	‰, V-SMOW	± ‰, 1s	‰, V-PDB	± ‰, 1s	‰, V-PDB	± ‰, 1s	‰, V-SMOW	± ‰, 1s										
KFM24	2016	410.00	417.12	−227	10.0	−12.5	0.3	−49.3	1.5	36.1	0.2												
KFM24	2016 ***	410.00	417.12	−242	10.0	−13.4	0.3	−50.4	1.5	35.4	0.2												

These results have been extracted from SKB: s database Sicada, and the data from 2007 have previously been presented in Hallbeck and Pedersen, 2009 [53]. DOC = dissolved organic carbon. V-SMOW = Vienna Standard Mean Ocean Water. b.d. = below detection. \* From a separate sample in same monitoring time series. \*\* Indication of some degree of short circuiting. Gas sampling was done during this sampling in the monitoring time series. \*\*\* Duplicate analysis.

### 2.3. Secondary Ion Mass Spectrometry (SIMS) for Stable S, O, and C Isotopes

Intracrystal SIMS-analysis (10  $\mu\text{m}$  lateral beam dimension, 1–2  $\mu\text{m}$  depth dimension) of sulfur isotopes in pyrite and carbon and oxygen isotopes in calcite was performed on a CAMECA IMS1280 ion microprobe (CAMECA, Gennevilliers, France), following the analytical settings and tuning reported previously [30,37,54,55].

Sulfur was sputtered using a  $^{133}\text{Cs}^+$  primary beam with 20 kV incident energy (10 kV primary, –10 kV secondary) and a primary beam current of ~1.5 nA. A normal incidence electron gun was used for charge compensation. Analyses were performed in automated sequences, with each analysis comprising a 70 s pre-sputter to remove the gold coating over a rastered 15  $\mu\text{m}$   $\times$  15  $\mu\text{m}$  area, centering of the secondary beam in the field aperture to correct for small variations in surface relief and data acquisition in sixteen four second integration cycles. The magnetic field was locked at the beginning of the session using an NMR field sensor. Secondary ion signals for  $^{32}\text{S}$  and  $^{34}\text{S}$  were detected simultaneously using two Faraday detectors with a common mass resolution of 4860 ( $M/\Delta M$ ). Data were normalized for instrumental mass fractionation using matrix matched reference materials, which were mounted together with the sample mounts and analyzed after every sixth sample analysis. Results are reported as per mil (‰)  $\delta^{34}\text{S}$  based on the Vienna Canon Diablo Troilite (V-CDT)-reference value. Analytical transects of up to nine spots were made from core to rim in the crystals. Two to six crystals were analyzed from each fracture sample (median  $n = 5$ ). In total, 179 analyses were made for  $\delta^{34}\text{S}$  ( $^{34}\text{S}/^{32}\text{S}$ ) of pyrite from 37 crystals. The pyrite reference material S0302A with a conventionally determined value of  $0.0 \pm 0.2\text{‰}$  (R. Stern, University of Alberta, pers. comm.) was used. Typical precision on a single  $\delta^{34}\text{S}$  value, after propagating the within-run and external uncertainties from the reference material measurements was  $\pm 0.10\text{‰}$ .

For calcite, a total number of 252  $\delta^{13}\text{C}$  and 220  $\delta^{18}\text{O}$  SIMS-analyses were performed on the same CAMECA IMS1280 described above. Settings follow those described for S isotopes, with some differences: O was measured on two Faraday cages (FC) at mass resolution 2500, whereas C used a FC/EM combination, with mass resolution 2500 on the  $^{12}\text{C}$  peak and 4000 on the  $^{13}\text{C}$  peak to resolve it from  $^{12}\text{C}^1\text{H}$ . Influence of organic carbon was avoided in the SIMS-analyses by careful spot placement to areas in the crystals without microfractures or inclusions and at a sufficient distance from grain-boundaries, where fine-grained clusters of other minerals and remnants of organic material may appear. The uncertainty associated with potential organic inclusions and matrix composition is therefore considered to be insignificant compared to the isotopic variations. Calcite results are reported as per mil (‰)  $\delta^{13}\text{C}$  and  $\delta^{18}\text{O}$ , based on the Vienna Pee Dee Belemnite (V-PDB) reference value. Analyses were carried out running blocks of six unknowns bracketed by two standards. Isotope data from calcite were normalized using calcite reference material S0161 from a granulite facies marble in the Adirondack Mountains, kindly provided by R.A. Stern (Univ. of Alberta). The values used for instrumental mass fractionation correction were determined by conventional stable isotope mass spectrometry at Stockholm University on ten separate pieces, yielding  $\delta^{13}\text{C} = -0.22 \pm 0.11 \text{‰ V-PDB}$  (1 std. dev.) and  $\delta^{18}\text{O} = -5.62 \pm 0.11 \text{‰ V-PDB}$  (1 std. dev.). Precision was  $\delta^{18}\text{O}$ :  $\pm 0.2\text{--}0.3\text{‰}$  and  $\delta^{13}\text{C}$ :  $\pm 0.4\text{--}0.5\text{‰}$ . Values of the reference material measurements are listed in Tables S4 (C), S5 (O), and S7 (S).

### 2.4. LA-(MC)-ICP-MS Analyses for $^{87}\text{Sr}/^{86}\text{Sr}$ and Rb–Sr Dating

Following SEM-investigation of the fracture surface, fine-grained secondary adularia and calcite were handpicked and mounted in epoxy for Rb–Sr dating. The spot analyses were carried out following sample characterization of the polished crystals. Rb/Sr analysis of the outermost growth zone in the adularia (co-genetic with calcite) was performed at the Department of Earth Sciences, University of Gothenburg, Sweden, using an ESI 213NWR laser ablation system connected to an Agilent 8800QQQ ICP-MS with an ORS3 (octopole reaction system) reaction cell sandwiched between two quadrupoles. Following laser warm-up, ablation occurred with static spot mode in a constant He flow (800 mL/min). The ablated material was mixed with  $\text{N}_2$  and Ar before entering the ICP-MS



torch region, and ionized atoms reacted with N<sub>2</sub>O gas in the reaction cell to chemically separate <sup>87</sup>Rb from <sup>87</sup>Sr, and thereby enable calculation of <sup>87</sup>Rb/<sup>86</sup>Sr and <sup>87</sup>Sr/<sup>86</sup>Sr ratios [56]. While the octopole bias was set to negative voltage, N<sub>2</sub>O flow rates in the reaction cell were varied to obtain optimal SrO<sup>+</sup> production rates. In MS/MS mode, both quadrupoles were controlled while reactive gas was in the reaction cell with the quadrupoles set at different masses to measure reaction products in mass-shifted mode. On mass <sup>85</sup>Rb/mass-shifted <sup>86</sup>Sr and mass-shifted <sup>87</sup>Sr/mass-shifted <sup>86</sup>Sr raw ratios were used to calculate <sup>87</sup>Rb/<sup>86</sup>Sr and <sup>87</sup>Sr/<sup>86</sup>Sr, respectively. The raw ratios were converted by correction factors derived from repeated analysis reference materials NIST SRM 610, BCR-2G, and Mica-Mg. The reference materials were selected to ensure that the pulse/analog setting of each measured isotope was identical in samples and reference materials. The feasibility of using NIST SRM 610, BCR-2G and Mica-Mg for calibration of Rb–Sr isotopic data obtained by in situ LA-ICP-MS/MS has been discussed previously [56,57]. The NIST SRM 610 certified reference material with <sup>87</sup>Sr/<sup>86</sup>Sr of 0.7097 [58] was used for <sup>87</sup>Sr/<sup>86</sup>Sr ratio calibration of the sample data. <sup>87</sup>Rb/<sup>86</sup>Sr calibration was performed by using glass reference material BCR-2G [59]. Secondary reference material was Mica-Mg, a mineral powder of a phlogopite separate pressed to a nanopellet tablet [56]. The resulting ages of Mica-Mg in this study were 519 ± 26 Ma. This was comparable to the 519.4 ± 6.5 Ma age determining the <sup>87</sup>Rb/<sup>86</sup>Sr ratio of Mica-Mg to 154.00 ± 0.70 [56]. For <sup>87</sup>Rb/<sup>86</sup>Sr and <sup>87</sup>Sr/<sup>86</sup>Sr respectively, BCR-2G yielded precisions at 1.39% and 0.43%, while NIST SRM 610 yielded 1.43% and 0.40%, and Mica-Mg gave 1.74% and 0.92%. Average count rate calculation of the reference material data was conducted by Glitter©, while sample data reduction and within-run error calculation of selected elements and isotopic ratios were performed using an in-house spreadsheet. No error propagation from uncertainties in literature data or within-run errors of reference materials was applied to sample errors.

The <sup>87</sup>Sr/<sup>86</sup>Sr values of the calcite crystals were determined by LA-MC-ICP-MS analysis at the Vegacenter, Swedish Museum of Natural History, Stockholm, Sweden using a Nu plasma (II) MC-ICP-MS, and an ESI NWR193 ArF eximer laser ablation system. Ablation frequency was 15 Hz, spot size 80 μm, and fluence 2.8 J/cm<sup>2</sup>, and the same crystal growth zones analyzed with SIMS for C and O isotopes were targeted. Wash-out and ablation times were both 45 s. The <sup>87</sup>Sr/<sup>86</sup>Sr analyses were normalized to an in-house brachiopod reference material “*Ecnomiosa gerda*” (linear drift and accuracy correction) using a value established by TIMS of 0.709181 (2 sd 0.000004, [60]). A modern oyster shell from Western Australia was used as a secondary reference material, and analyzed at regular intervals together with the primary reference. The accuracy of these analyses was quantified by comparison to the modern seawater value for <sup>87</sup>Sr/<sup>86</sup>Sr of 0.7091792 ± 0.0000021 [61]. Values of the reference material measurements are listed in Table S6.

### 3. Results

#### 3.1. Mineralogy

Calcite was more abundant on the fracture surfaces than pyrite. The crystal morphologies and paragenetic assemblages varied between the different fractures (Figure 2). The KFM01D samples and the shallowest KFM24 samples featured fine-grained subhedral calcite aggregates, the KFM02A calcite occurred as thin slickenfibres, and the KFM08D calcite as equant crystals on euhedral quartz. In the KFM24 samples from 379 to 414 m, euhedral calcite crystals (mostly of equant type with small c-axis to a-axes ratio) and cubic, octahedral, and pyritohedral pyrite were observed on top, and intergrown with euhedral quartz, and occasionally, euhedral adularia. Chlorite and clay minerals were usually present on the fracture surfaces. There were also small patches of organic material in the form of solid hydrocarbon (asphaltite) and filamentous material (described in more detail below) on the fracture surfaces, particularly in the KFM24 samples at 379 and 396–399 m. Accessory minerals in the latter samples included euhedral bastnäsite and scattered galena. In KFM24:412, pyrite occurred as cryptocrystalline aggregates in addition to larger single crystals.

### 3.2. Isotope Signatures in Calcite, Waters, and Gases

#### 3.2.1. Carbon Isotopes and Gas Compositions

The carbon isotope composition varied substantially between and within the calcite samples. The total span in  $\delta^{13}\text{C}_{\text{calcite}}$  was 56.4‰ (−36.2 to +20.2‰, span of each sample is listed in Table 1 and full SIMS results in Table S2). Four samples contained  $\delta^{13}\text{C}_{\text{calcite}}$  values  $\gg 0$ ‰ (KFM24: 193, 379 and 410, and KFM02A:162). In KFM24: 193 it was just one analysis out of seventeen showing these heavy values, in KFM24:379 the  $^{13}\text{C}$ -rich values were only found in the outermost rim of the crystals (Figure 3a) whereas crystals in the other two samples were dominantly  $^{13}\text{C}$ -rich (Figure 3b). The other samples can be divided into two groups based on the  $\delta^{13}\text{C}_{\text{calcite}}$  values; (i) KFM01D:316, 431 and KFM08D:677, 830, and 832, as a rule, had values in the range −10 to −3‰, with small variations within the crystals (Figure 3d,e), and (ii) the remainder of the KFM24 samples and the KFM02A sample mainly showed  $\delta^{13}\text{C}_{\text{calcite}}$  values from −36‰ to −10‰, and with large variation within the crystals (Figure 3c). This means that significant span in  $\delta^{13}\text{C}_{\text{calcite}}$  was limited to samples from boreholes KFM02A and KFM24 (Figure 4b).

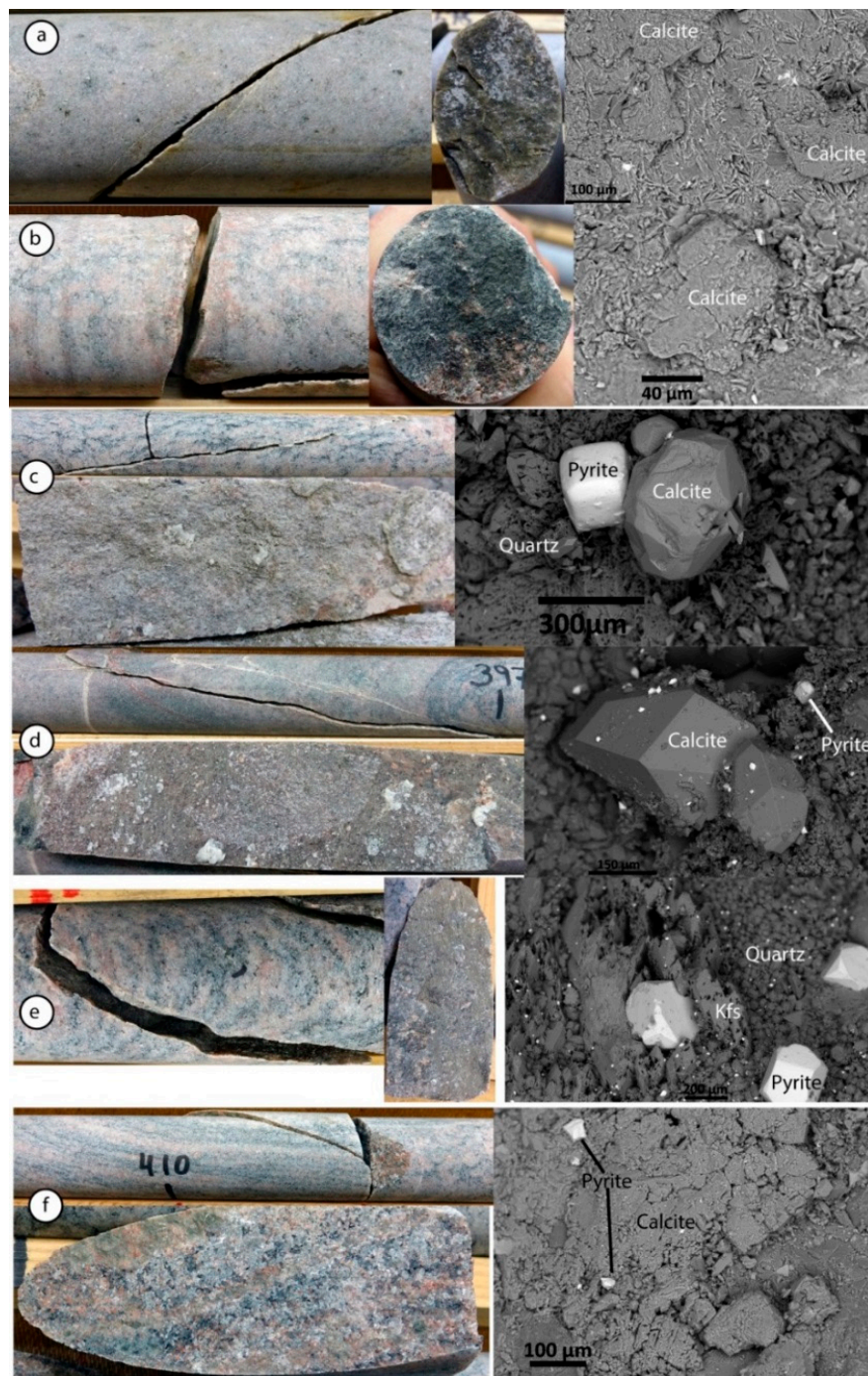
For the waters, the  $\delta^{13}\text{C}$  values of dissolved inorganic carbon ( $\delta^{13}\text{C}_{\text{DIC}}$ ) were in the −21.9 to −4.5‰ PDB range, but with a large variation between the different section (heaviest in KFM08D:677 and lightest in KFM24:410). The KFM24:410  $\delta^{13}\text{C}_{\text{DIC}}$  value is slightly uncertain, due to potential short circuiting at the end of a time series sampling (i.e., the hydrochemical composition gradually changed to that same as the section above, due to suspected interconnection between the conductive fractures in these two sections). The second lowest  $\delta^{13}\text{C}_{\text{DIC}}$  value of the sample set was −15.9‰ (KFM24:191). The KFM24:410 section was sampled for gases, and contained 0.18 mL/L  $\text{CH}_4$  and had a methane to higher hydrocarbon ratio,  $\text{C}_1/(\text{C}_2 + \text{C}_3)$ , of 170,  $\delta^{13}\text{C}_{\text{CH}_4}$ , and  $\delta^2\text{H}_{\text{CH}_4}$  values of −49.3‰ V-PDB (duplicate: −50.4‰) and −227‰ V-SMOW (Vienna Standard Mean Ocean Water, duplicate: −242‰), respectively (Table 2). With the exception of section KFM01D:316, which had higher methane concentrations (0.64 mL/L) and  $\text{C}_1/(\text{C}_2 + \text{C}_3)$  ratio (~2300), the gas samples from the other borehole sections had quite similar concentrations of methane (0.09–0.19 mL/L) and  $\text{C}_1/(\text{C}_2 + \text{C}_3)$  ratios (25–160), but no gas isotope analyses have been carried out from these samples.

#### 3.2.2. Oxygen Isotopes

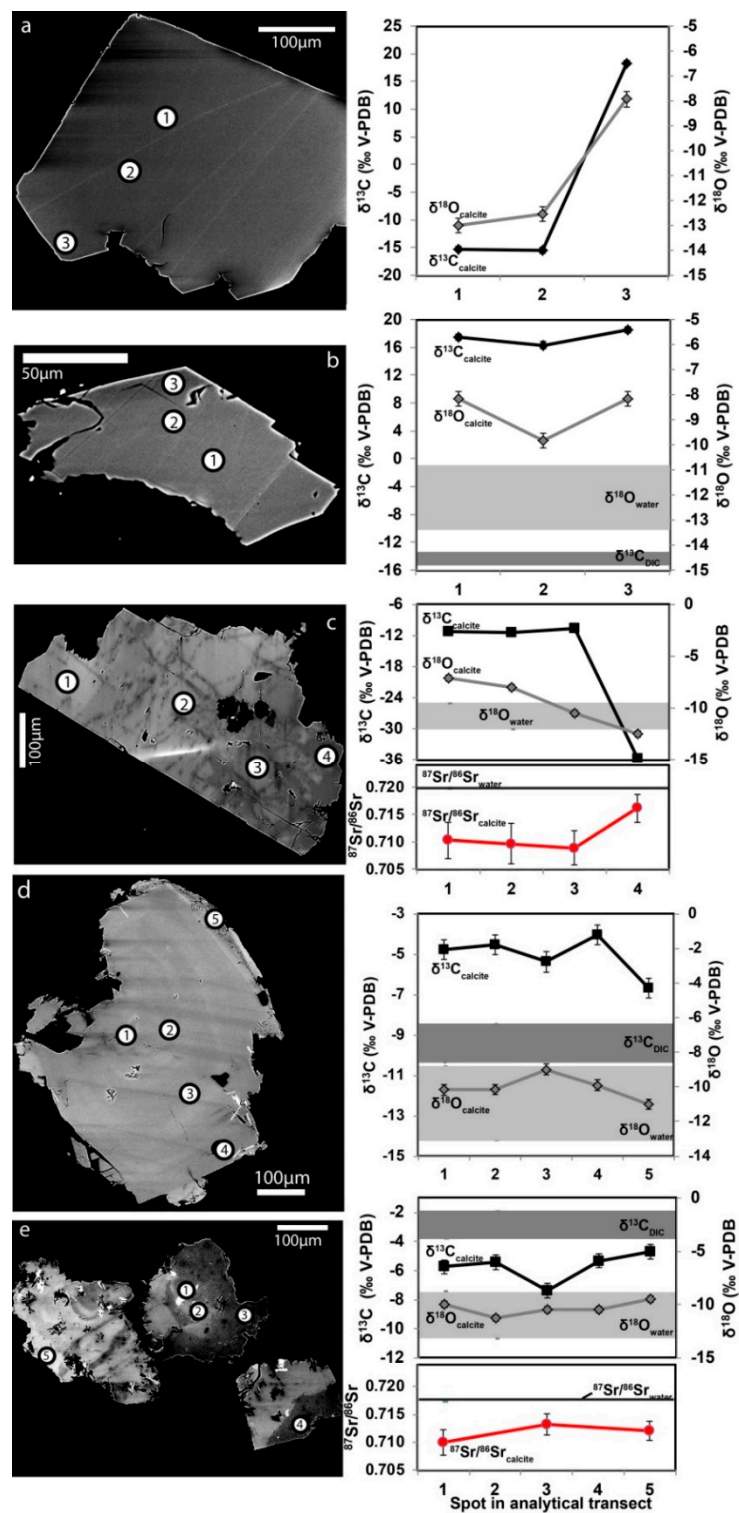
The  $\delta^{18}\text{O}_{\text{calcite}}$  values had a total span of 12‰ V-PDB (−17.9‰ to −5.9‰). Most values (percentiles 10 to 90,  $\text{P}_{10}\text{--}\text{P}_{90}$ ) were, however, in the −14 to −8‰ range, and heavier values than −7‰ were dominantly from KFM01D:316. Apart from this sample, there was no major depth variation in  $\delta^{18}\text{O}_{\text{calcite}}$  (Figure 4a). The maximum variation within a single crystal was 7‰, and temporal variation trends could be discerned within the crystals, e.g., increasing values in the rim of crystal compared to the core (Figure 3a) or the opposite (Figure 3c), in both cases, correlated with the evolution of  $\delta^{13}\text{C}_{\text{calcite}}$ . The  $\delta^{18}\text{O}$  values in the waters varied between −13.9‰ to −10.0‰ V-SMOW.

#### 3.2.3. Strontium Isotopes

Strontium isotope ratios within the crystals were overlapping with—but generally lower than—the corresponding values in the groundwaters (0.705–0.720,  $\text{P}_{10}\text{--}\text{P}_{90}$ :0.710–0.718 in the calcite vs 0.718–0.721 in the water, Figure 4c, Table 1, Table 2 and Table S2).  $^{87}\text{Sr}/^{86}\text{Sr}$  was generally elevated in the outermost growth zones compared to the crystal core (Figure 3c,e). It should be noted that the uncertainty of the Sr isotope analyses was relatively large due to the low concentrations of Sr in the calcite. The larger spot size of the Sr-isotope determinations (80  $\mu\text{m}$ ) compared to C and O (10  $\mu\text{m}$ ) inhibit full comparisons in finely zoned crystals. The fine outermost calcite growth zone could only be targeted in a small number of crystals, due to that the growth zone was smaller than the spot size, which inhibited direct  $^{87}\text{Sr}/^{86}\text{Sr}$  comparison of waters and the outermost calcite growth zones for some sections (Figure 4g).

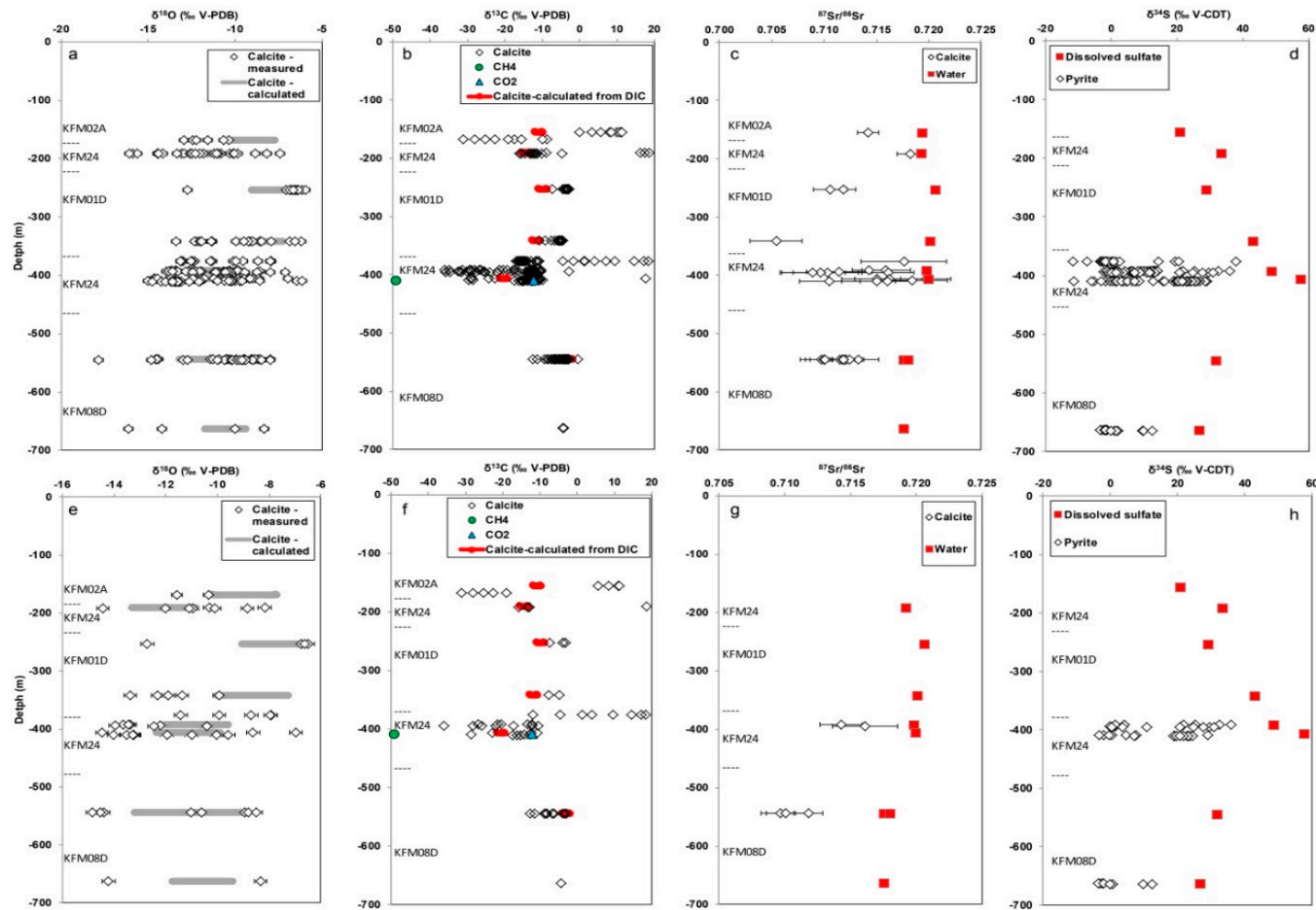


**Figure 2.** Mineral coating descriptions. Photos of the drill core samples are on the left, including a separate photo of the sampled fracture surface. Backscattered SEM images are on the right-hand side. (a) KFM08D:677 m; subhedral coating of calcite on a chlorite-rich fracture coating. (b) KFM24:193; subhedral coating of calcite on a chlorite-rich fracture coating. (c) KFM24:379; euhedral pyrite (cubic with rounded edges) and calcite (equant) on euhedral quartz. (d) KFM24:396; euhedral calcite and pyrite on a coating of euhedral quartz. Calcite habit is scalenohedral, with relatively small *c*-axis to *a*-axes ratio, and obtuse crystal tip termination. Bright euhedral bastnäsite crystals occur on the calcite surface. (e) KFM24:399; intergrown K-feldspar (adularia, partly altered) and cubic pyrite on top and partly intergrown with euhedral quartz. (f) KFM24:410 m; subhedral calcite coating with scattered intergrown pyrite crystals.



**Figure 3.** Microanalytical transects within calcite. Backscattered SEM images of polished crystal cross sections are shown to the left, with spot locations indicated (for closely spaced 10 µm  $\delta^{13}\text{C}$  and  $\delta^{18}\text{O}$  SIMS spots and in (c,e) also for subsequent 80 µm  $^{87}\text{Sr}/^{86}\text{Sr}$  LA-MC-ICP-MS spots). Corresponding isotopic values are shown on the graphs. Spans of calculated values for hypothetical calcite are also shown, and are based on the groundwater composition in the same section. For this comparison, temperature dependent fractionation factors for calcite precipitation at ambient temperatures are used for  $\delta^{13}\text{C}$  [62] and  $\delta^{18}\text{O}$  [63]. Due to the geothermal gradient, the deeper borehole sections (>500 m) have

been assigned a larger temperature span (5–20 °C) than the shallower samples (~0–500 m; 5–15 °C). The lower temperature limit of 5 °C includes the possibility of rapid infiltration of glacial water to a great depth. Specific features include (a) spatiotemporal intracrystal variation in  $\delta^{13}\text{C}$  and  $\delta^{18}\text{O}$  values in sample KFM24:379. Both values increase substantially in the rim of the crystal compared to the crystal core. No groundwater data exists from this fracture, but the 15 m deeper section show hypothetical calcite  $\delta^{18}\text{O}$  values of  $-12$  to  $-9.6\text{‰}$  V-PDB (Vienna Pee Dee Belemnite) that are neither corresponding to the rim nor to the core of the crystal. (b)  $^{13}\text{C}$ -enrichment throughout the crystal. The  $\delta^{18}\text{O}$  and  $\delta^{13}\text{C}$  values of the calcite are higher than the values calculated from the groundwater (KFM24:193). (c) Example of substantial  $^{13}\text{C}$ -depleted calcite, in this case, in the outermost rim of the crystal (KFM24:399, same feature exists in e.g., KFM24:410). The value of the calcite crystal rim neither has  $\delta^{18}\text{O}$  nor  $^{87}\text{Sr}/^{86}\text{Sr}$  in equilibrium with the groundwater, although the outer part has elevated  $^{87}\text{Sr}/^{86}\text{Sr}$  compared to the rest of the crystal. Similar  $\delta^{18}\text{O}$  and  $^{87}\text{Sr}/^{86}\text{Sr}$  discrepancy between calcite and water occurs in KFM24:414 and KFM01D:431. (d) Calcite with moderate  $^{13}\text{C}$ -depletion (not in equilibrium with  $\delta^{13}\text{C}_{\text{DIC}}$ ) and  $\delta^{18}\text{O}$  values in correspondence with the groundwater (KFM08D:678, similar features occur in KFM24:413 and KFM24:194). (e) Aggregate of fine-grained crystals showing zonation (KFM08D:677). Microanalyses targeting the different crystal zones show overall correspondence of  $\delta^{18}\text{O}$  with the groundwater. By contrast, both  $\delta^{13}\text{C}$  and  $^{87}\text{Sr}/^{86}\text{Sr}$  are overall higher in the groundwater than in calcite.



**Figure 4.** Isotope values ( $\delta^{18}\text{O}$ ,  $\delta^{13}\text{C}$ ,  $^{87}\text{Sr}/^{86}\text{Sr}$ , and  $\delta^{34}\text{S}$ ) in calcite, pyrite, groundwaters and gases vs depth. Top row (a–d) shows all SIMS spots analyzed (V-CDT = Vienna Canyon Diablo Troilite). Lower row (e–h) shows only the outermost crystal growth zone in contact with the modern groundwater. The graphs also show ranges for calculated values for hypothetical calcite based on the groundwater composition in the same borehole section (see caption to Figure 3 for details). For KFM02A, groundwater data from a slightly shallower zone is shown for comparison.

### 3.2.4. Sulfur Isotopes

The  $\delta^{34}\text{S}_{\text{pyrite}}$  values of the samples varied between  $-11.7\text{‰}$  to  $+37.8\text{‰}$  V-CDT (span of each sample is listed in Table 1 and full SIMS results in Table S3). Pyrite was not as abundant and evenly distributed as calcite, and consequently, the depth distribution pattern is not as complete (Figure 4d). The variability was larger in borehole KFM24 than in KFM08D. Microanalytical transects of  $\delta^{34}\text{S}_{\text{pyrite}}$  within individual crystals showed two distinct groups: (1) relatively small variation between different growth zones (Figure 5a, in the deepest KFM24 section at variable  $\delta^{34}\text{S}$  in different crystals, and in KFM08D), and (2) significant increase in  $\delta^{34}\text{S}_{\text{pyrite}}$  in the outermost growth zone(s) (Figure 5b–d), which dominated in the middle section in KFM24, and existed in the deeper section of the same borehole. The increase from crystal core to rim exceeded 40‰.

The groundwater  $\delta^{34}\text{S}_{\text{sulfate}}$  values varied between  $+20.8\text{‰}$  to  $+57.5\text{‰}$  (Figure 4d), and was particularly elevated in deeper sections of KFM24 (deepest section:  $+57.5\text{‰}$  and middle section  $+48.5\text{‰}$ , Table 2).

### 3.2.5. Rb–Sr Dating

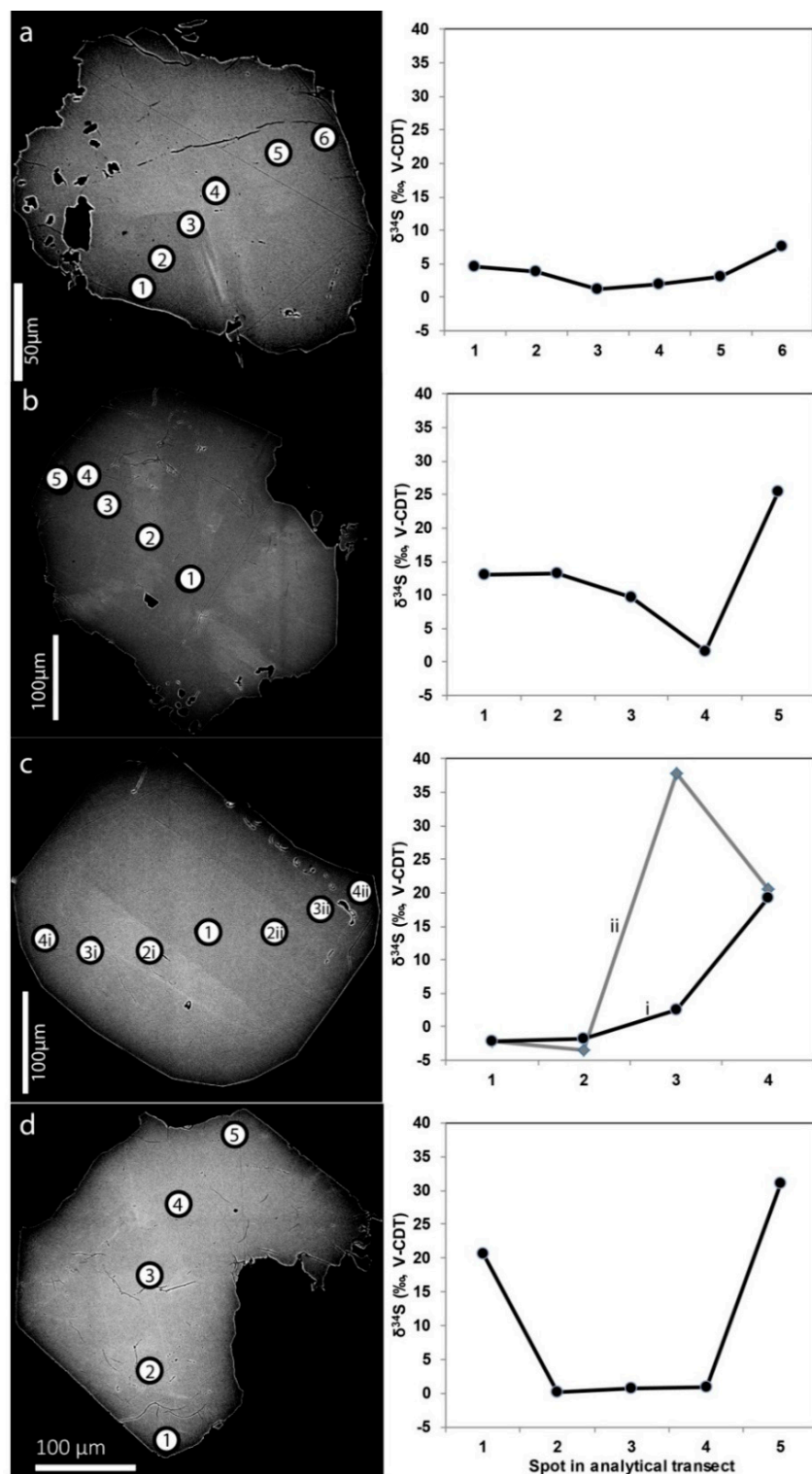
The secondary adularia–calcite assemblage in sample KFM24:399 gave an age of  $396 \pm 7$  Ma (Figure 6, isotopic ratios in Table S8). The isochron consisted of nine adularia spots and a calcite spot for initial  $^{87}\text{Sr}/^{86}\text{Sr}$ . The isochron gave a mean square weighted deviation (MSWD) of 0.52 and a probability of fit of 0.84 (maximum = 1).

### 3.2.6. Putative Fossilized Microorganisms

Samples KFM24:379 and 399 featured abundant carbonaceous material (CM) on mineral surfaces, incorporated in between calcite crystals and as components in clay minerals (Figure 7). The CM occurred as biofilm-like sheets (Figure 7A), carbonaceous or partly mineralized by clay in nature (Figure 7B–D). Transitions between carbonaceous portions and mineralized portions were visible (Figure 7A,C,D), and it is evident that the CM has acted as nucleation sites for mineral growth including clay, oxides, calcite, and other Si-rich phases. The CM also occurred in between crystals and as a dominant constituent in clay-like phases in between calcite crystals, thus, CM was an integrated substance of the vein mineralogy (Figure 7E).

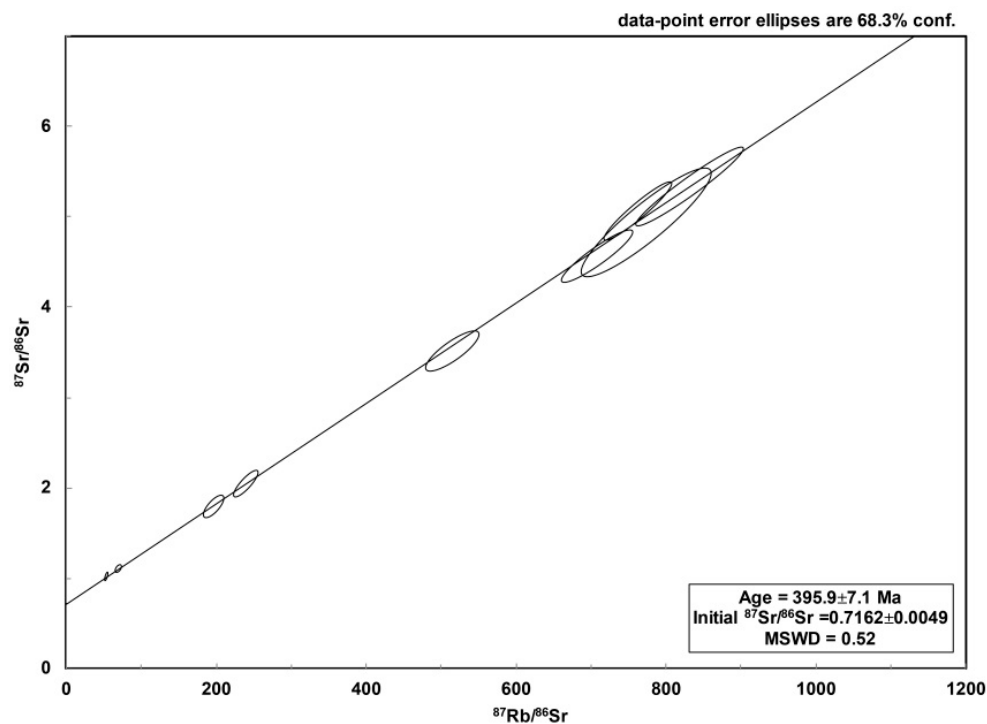
Filamentous structures protrude from the biofilm-like CM with the same composition as the CM, either completely carbonaceous in nature or partly mineralized by Fe-oxides, calcite, clay minerals and other Si-rich phases, along with some pyrite (Figure 8A–D). The diameter of the filaments ranged from 5 to 40  $\mu\text{m}$ , and the lengths were up to 1 mm. They occurred in entangled networks with frequent branching and occasional loops (Figure 8C).

The filaments were characterized by a flattened appearance, probably due to dehydration (Figure 8B). However, the flattened filaments are partly mineralized, indicating that the dehydration occurred pre-drilling, and not as a result of the vacuum in the ESEM chamber. According to the EDS analyses, the carbonaceous material consist of O, C, Fe, Si, Mg, Al, and minor amounts of Ca, K, and S (see examples in Supplementary Material 1). The element concentrations vary depending on the grade of mineralization. Carbon makes up about 5–15 wt %, and decreases with increased mineralization. The opposite trend is seen for the other elements.

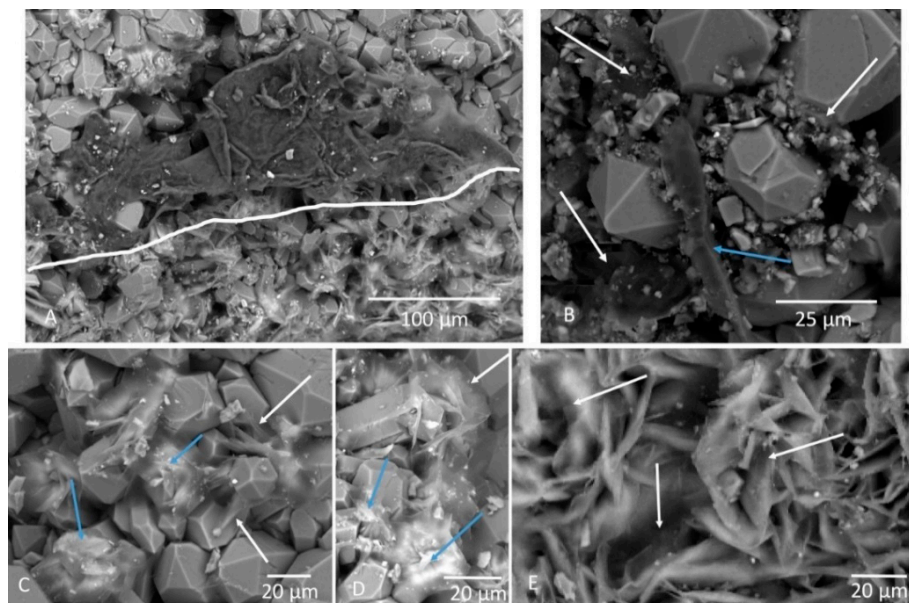


**Figure 5.** S isotope microanalytical transects within individual pyrite crystals. (a) KFM24:412 m, no major  $\delta^{34}\text{S}_{\text{pyrite}}$  variation with growth. (b–d) Increase in  $\delta^{34}\text{S}_{\text{pyrite}}$  in the rim zone of the crystals. (b) KFM24:396, (c) KFM24:379 divided into two transects from core to rim (i and ii), (d) KFM24:399; several other crystals on the same fracture (and in sample KFM24:398) showed identical pattern. Errors are within size of the symbols. The axes of the different graphs have the same range, for comparison purposes.

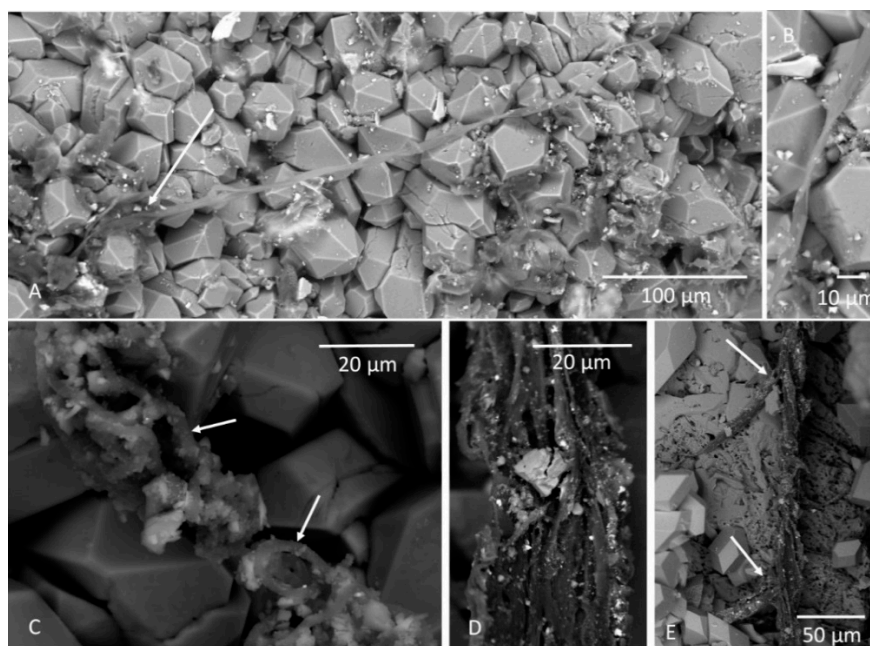




**Figure 6.** Rb–Sr dating of co-genetic adularia–calcite from sample KFM24:399 m.



**Figure 7.** Carbonaceous matter as biofilms on, and incorporated in, minerals from sample KFM24:398. (A) A biofilm with characteristic “sheet-like” appearance. The top part is carbonaceous, and the lower part mineralized. The dashed line marks the transition from carbonaceous to mineralized portions of the biofilm. The appearance of the carbonaceous material (CM) is characterized by dark, patchy areas slightly stretched in the margins. (B) Abundant CM in between calcite and quartz crystals. The CM is partly mineralized (marked with white arrows). Remnant of associated putative fungal hyphae is marked by blue arrow. (C,D) CM with typical elongated and stretched appearance distributed on and in between calcite and quartz crystals, marked with white arrows. The CM has acted as nucleation site for mineralization, marked with blue arrows. (E) CM, as part of clay lining the sheet-like crystal lattice and voids in between the sheets as a dark patchy substance, marked by white arrows.



**Figure 8.** Putative fungal hyphae from sample KFM24:398 (A–C) and KFM24:379 (D,E) preserved as CM or partly mineralized. (A) A long hypha originating in a biofilm (marked with a white arrow), which is incorporated in between calcite and quartz crystals. (B) Close up of a hypha that is flattened due to dehydration. (C) Hyphae in a chaotic and entangled mycelium-like network. Note the partial mineralization of the hyphae. Individual hyphae are marked with white arrows. (D) Close up of a hypha partly mineralized by calcite. (E) A hypha with multiple branching. The points of branching are marked with white arrows.

#### 4. Discussion

Although it is commonly assumed, based on experimental observations, that calcite crystals will reprecipitate and homogenize during long-term interaction with fluids, observations from deep crystalline rock fractures show that zonation is sustained within crystals [64], with examples of >400 Ma old growth zonation [29] that has not been affected by dissolution-reprecipitation. The isotopic inventory of calcite crystals therefore serves as archives for processes in the fracture system over long time periods. In addition, the crystals (both calcite and pyrite) in the current study show dominantly sharp borders between different growth zones, which means that influence from dissolution-reprecipitation and large-scale homogenization due to fluid–mineral interaction can be regarded as minor. In the following sections, we discuss the various microbial processes that can be responsible for the observed isotope signatures, and compare the mineral signatures with water and gas data. However, because the fractures potentially host a complex community of different metabolic types, all which have different isotope discrimination values, various degrees of mixed signals from different processes can be expected in the studied minerals, which complicates certain process determinations. The highly variable  $\delta^{13}\text{C}_{\text{calcite}}$  values between different fractures show that the processes leading to calcite precipitation have been fracture specific. There is usually also distinct temporal  $\delta^{13}\text{C}_{\text{calcite}}$  variability within the single fractures, due to the episodic microbial processes. In addition, we discuss the findings of putative fossilized microorganisms.

##### 4.1. Episodic Microbial Methanogenesis

Microbial methane usually is  $^{13}\text{C}$ -depleted compared to other carbon compounds [65]. As a consequence of the fractionation occurring during methanogenesis, which discriminates against  $^{13}\text{C}$ , the residual  $\text{CO}_2$  becomes  $^{13}\text{C}$ -rich [66]. Subsequent involvement of the residual C into precipitating

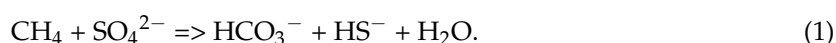
carbonate minerals is, therefore, a useful diagnostic C-isotope tracer for methanogenesis [67], as shown by  $^{13}\text{C}$ -rich secondary carbonates in sedimentary basins [68], or in fractured crystalline rocks in the Fennoscandian shield [29,30,32,69–71]. Isotopic signatures of methane from Outokumpu, Finland, have also been indicative of microbial methane production at great depth [72]. The significantly  $^{13}\text{C}$ -enriched calcite ( $\delta^{13}\text{C}_{\text{calcite}}$  values as heavy as +20.2‰, Figure 4b) observed in the present study, is thus a strong line of evidence for microbial methanogenesis in situ in the deep fracture systems, especially since potential abiotic methane forming chemical processes, such as serpentinization, graphite metamorphism, or Fischer–Tropsch type reactions [73], are unlikely under the local physicochemical and geological conditions. The distribution of the  $^{13}\text{C}$ -rich calcite marks methanogenesis in the depth interval 155–406 m, both as the dominant origin of the bicarbonate that was incorporated into calcite (Figure 3b), and late in the precipitation history (Figure 3a). In anaerobic environments, sulfate reducers and methanogens compete for the same substrates [74]. Since sulfate reducers have a higher affinity for acetate and hydrogen than methanogens, sulfate reducers may outcompete methanogens at high sulfate concentrations. The experiments of Lovley & Klug [74] showed that this can occur at as low sulfate concentrations as 60–105  $\mu\text{M}$  sulfate (=10 mg/L). However, there are also observations of co-existence of SRB and methanogens at high sulfate concentrations, up to 10 mM in sulfate-rich estuarine sediments [75], and even up to 60 mM in organic-rich coastal sediments [76,77]. Nevertheless, the concentrations of dissolved sulfate can be an important aspect that explains why methanogenesis- and anaerobic oxidation of methane (AOM)-related calcite is localized to borehole KFM24, where sulfate concentrations are relatively low (9–16 mg/L in the lower sections, and 45 mg/L in the uppermost section), and not in the other sections, where concentrations are higher (KFM01D and KFM08D: 67–273 mg/L). Detected autotrophic and heterotrophic methanogens and scattered elevated methane concentrations suggest microbial methanogenesis at least in a few deep groundwater sections in the Forsmark area, at present [53]. Unfortunately, none of the sections with  $^{13}\text{C}$ -rich calcite have been investigated for microorganisms. Overall, the methane concentrations are, however, generally low in the sampled sections (<1 mL/L, Table 2), suggesting that microbial methanogenesis is not a significant process at present. The ratio of methane to higher hydrocarbons,  $\text{C}_1/(\text{C}_2 + \text{C}_3)$ , points to a biogenic source of the methane in one of the borehole sections (KFM01D:316), where the ratio is >1000 (Table 2, cf. [78]). In addition, the only  $\delta^{13}\text{C}_{\text{CH}_4}$  measurement carried out (KFM24:410) shows a value,  $\sim -50\text{‰}$ , that is close to the border zone between biogenic and abiotic methane [65]. It is, however, lighter than  $\delta^{13}\text{C}_{\text{CH}_4}$  in abiotic methane in volcanic-hydrothermal systems, serpentinized ultramafic rock systems, and deep boreholes in crystalline igneous rocks in South Africa, Canada, and Scandinavia [72,73]. The  $\delta^2\text{H}_{\text{CH}_4}$  value is relatively heavy ( $-222\text{‰}$  to  $-247\text{‰}$ ), which places the gas in the microbial gas field on the border to mixed origin in a  $\delta^{13}\text{C}_{\text{CH}_4}$  vs  $\delta^2\text{H}_{\text{CH}_4}$  scatter plot [73,79] (not shown due to few data points). Taken together, a gas of mixed origin with a significant microbial component is the most likely origin for the KFM24 gas, in line with the support for microbial methanogenesis in situ offered by  $\delta^{13}\text{C}_{\text{calcite}}$ . Microbial methanogenesis occurs either by organotrophic methanogens utilizing methyl group compounds (e.g., acetate fermentation [80]), or by chemolithoautotrophic carbonate ( $\text{CO}_2$ ) reduction with  $\text{H}_2$  [81]. The  $\delta^{13}\text{C}_{\text{CH}_4}$  values of the KFM24 gas fit best with microbial gas formed via the acetate fermentation pathway, whereas the carbonate reduction pathway usually features even more  $^{13}\text{C}$ -depleted gas [65,73,82]. There are, to the best of our knowledge, no DNA-based data from this site that could give more information about which of these pathways is dominant. An additional substrate in these fractures is asphaltite, which is believed to originate from Paleozoic black shale that once covered the bedrock in the area [29,83]. These organics were most likely introduced to the deep fracture system during an extensional tectonic episode when elevated temperatures in the bottom of the sedimentary pile made the solid hydrocarbons viscous [83]. Asphaltite is highly reduced organic matter that also contains sulfate and can be utilized by the microorganisms (e.g., SRB or acetogens) in the presence of an electron acceptor.

The calcite samples with heavy  $\delta^{13}\text{C}$  dominantly had  $\delta^{18}\text{O}$  and/or  $^{87}\text{Sr}/^{86}\text{Sr}$  out of equilibrium with modern groundwater when temperature dependent fractionation is taken into account (Figures 3b

and 4e–g) suggesting that the heavy  $\delta^{13}\text{C}_{\text{calcite}}$  reflects mainly ancient methanogenesis, which may have occurred at different salinity and sulfate concentrations than at present. At around 400 m depth in KFM24, overlap in  $\delta^{18}\text{O}$  and/or  $^{87}\text{Sr}/^{86}\text{Sr}$  between calcite and groundwater occurred in some crystals. These are, thus, the best candidates for modern precipitation.

#### 4.2. Microbial Anaerobic Oxidation of Methane and Oxidation of Organic Matter

In marine seabed systems, AOM coupled with microbial sulfate reduction (MSR) is a major process that mitigates methane transport to the atmosphere [84–86]. At a depth horizon where surficial marine sulfate mixes with deeper seated methane, the methane is oxidized by methanotrophs (archaea), whereas sulfate is reduced by sulfate reducers via the simplified reaction:



After reaction of the produced bicarbonate and sulfide with dissolved  $\text{Ca}^{2+}$  and  $\text{Fe}^{2+}$ , carbonate (mainly calcite) and sulfide (pyrite) minerals precipitate, respectively. During recent years, this AOM–MSR process has been observed in deep crystalline bedrock settings as well, both by ancient findings-based isotopic signatures in secondary minerals in fractures [29,30,66], and by microbiological and geochemical observations from modern groundwaters and gases residing in the deep fractured bedrock [2,3,16–18,87]. Significant  $^{13}\text{C}$ -depletion is inherited from the source methane during AOM, and can subsequently be detected in the  $\delta^{13}\text{C}$  signature of authigenic carbonate. This diagnostic  $\delta^{13}\text{C}_{\text{calcite}}$  signature is one of the most widely used tools to distinguish ancient and modern AOM [35,36,88,89]. Although AOM involves fractionation that results in an even more  $^{13}\text{C}$ -depleted product than the source methane [84,90,91], the  $\delta^{13}\text{C}$  signature of the carbon originating from oxidized methane is usually diluted by other relatively  $^{13}\text{C}$ -rich dissolved carbon species (e.g., DIC) prior to incorporation in calcite. Consequently, authigenic carbonate in well-studied sedimentary systems is, as a rule, less  $^{13}\text{C}$ -depleted than the source methane. Typical  $\delta^{13}\text{C}$  values of AOM-related authigenic carbonate are in the  $-70\text{‰}$  to  $-30\text{‰}$  range [36,89,92], and in some cases, significantly heavier [93]. Although  $\delta^{13}\text{C}_{\text{calcite}}$  values in the  $-35$  to  $-25\text{‰}$  can theoretically reflect oxidation of degraded organic matter e.g., in the form of oil and related substances which have  $\delta^{13}\text{C}$  values in the  $-35\text{‰}$  to  $-25\text{‰}$  range [94], the  $^{13}\text{C}$ -depleted calcite from Forsmark ( $\delta^{13}\text{C}$  down to  $-36.2\text{‰}$ ) is proposed to originate from AOM, especially when taking into account dilution of the  $\delta^{13}\text{C}$  signature by other C-sources after AOM and prior to calcite precipitation, as described above. Since the  $\delta^{13}\text{C}$  values overlap with both microbial and abiotic methane [65,73], the origin for the source methane cannot be distinguished. The calcite crystals with moderately  $^{13}\text{C}$ -depleted values down to  $\sim -20\text{‰}$ , although overlapping with values of abiotic methane [73], can, however, not be distinguished from bicarbonate formed by e.g., microbial oxidation of organic matter by SRB or other microorganisms [65], and can alternatively be of a mixed origin. Also, masking of process-specific C-isotope signatures is most likely to be expected in fractures with high bicarbonate contents (e.g., 76 mg/L in KFM01D). The  $^{13}\text{C}$ -depleted calcite occurs in middle section of KFM24 (396–399 m), where all fractures feature calcite with minimum values lighter than  $-32.5\text{‰}$  suggesting that AOM has occurred in all fractures in this section. In the deepest KFM24 section (410–414 m), there are also  $^{13}\text{C}$ -depleted values in the calcite (down to  $-29.8\text{‰}$ ) which suggest that AOM may have occurred there as well (Figure 4b). In the latter section, there is also one very heavy  $\delta^{13}\text{C}_{\text{calcite}}$  value marking methanogenesis in situ, but there is no indicator of how methanogenesis and AOM are temporally related. It suggests that the position of the sulfate–methane transition zone (SMTZ) has changed with time, in accordance with findings from Olkiluoto, Finland, where a modern SMTZ is indicated at 300–400 m [13–15,87], but the calcite record suggests a more shallow position for a fossil SMTZ [69]. In the middle section of KFM24, there are no positive  $\delta^{13}\text{C}_{\text{calcite}}$  values, showing that methanogenesis has not occurred there. Extensive precipitation of  $^{13}\text{C}$ -rich calcite is, instead, observed in shallower samples (e.g., 194 and 379 m in the same borehole). This indicates a reverse SMTZ at depths of roughly 395–415 m in KFM24, at which methane formed in shallower

fractures mixes with deeper seated sulfate-rich water, in similarity with  $\delta^{13}\text{C}_{\text{calcite}}$  distributions in the crystalline basement fracture system at Laxemar, Sweden [30]. In contrast to the hydrochemical situation at Laxemar, where the sulfate concentrations increase with depth, the Forsmark waters show overall decreasing sulfate concentrations beneath 300–500 m ([50] and Table 2). In the KFM24 borehole sections, the dissolved sulfate concentrations are overall low, and decrease slightly with depth (195 m: 44 mg/L; 395 m: 15 mg/L and 415 m: 9 mg/L). In addition, the depleted  $\delta^{13}\text{C}_{\text{calcite}}$  values in the outermost calcite growth zones are lower than the  $\delta^{13}\text{C}_{\text{DIC}}$  ( $-21.9\%$ ) of modern groundwater (Figures 3c and 4f) and the  $^{87}\text{Sr}/^{86}\text{Sr}$  and  $\delta^{18}\text{O}$  do not generally correspond. The SMTZ indicated by the  $\delta^{13}\text{C}_{\text{calcite}}$  values is therefore proposed to be ancient. A Devonian precipitation age of a calcite–adularia assemblage was obtained by the Rb–Sr dating at these depths (Figure 7), and Devonian–Carboniferous microbial activity, such as AOM, has earlier been recorded from outside the target area in Forsmark and in the Laxemar area [29]. In KFM02A, the same depth relation as in KFM24 occurs, with methanogenesis-related  $\delta^{13}\text{C}_{\text{calcite}}$  values in the shallower sample (up to  $+11.3\%$  at 162 m), and proposed AOM-related values in the deeper sample (down to  $-1.2\%$  at 175 m).

#### 4.3. Microbial Sulfate Reduction

Pyrite is the most common non-carbonate mineral at marine methane-seeps. It forms as a result of the AOM–MSR reaction (1), and inherits the S isotopic composition of the hydrogen sulfide [95], and because the MSR metabolism produces hydrogen sulfide strongly depleted in  $^{34}\text{S}$ ,  $\delta^{34}\text{S}_{\text{pyrite}}$  serves as a marker for MSR.

The low minimum  $\delta^{34}\text{S}_{\text{pyrite}}$  values in the deep and middle sections of KFM24, at lightest  $-11.5\%$  CDT, reflect  $^{32}\text{S}$  enrichment in the produced sulfide during MSR. The initial sulfate is anticipated to originate dominantly from infiltration of marine waters. The brackish marine waters in the deep fracture system show relatively homogeneous  $\delta^{34}\text{S}_{\text{SO}_4}$  values between 24–26‰ [50], which infers an isotope enrichment,  $^{34}\epsilon$  ( $\delta^{34}\text{S}_{\text{SO}_4} - \delta^{34}\text{S}_{\text{pyrite}}$ ), of up to 40‰, when comparing the outermost pyrite growth zonation and the dissolved sulfate in the same fracture. This is a fully possible isotope enrichment, as experiments of pure culture MSR had resulted in  $^{34}\epsilon$  of up to 66‰ [96], and exclude abiotic sulfide production through thermochemical sulfate reduction, which results in more moderate  $^{34}\epsilon$  [97]. These  $\delta^{34}\text{S}_{\text{pyrite}}$  values, together with the  $^{13}\text{C}$ -depleted calcite in the same fractures, support coupled AOM–MSR at the proposed fossil SMTZ at around 395–415 m depth. During a closed system Rayleigh fractionation cycle, the  $\delta^{34}\text{S}_{\text{SO}_4}$  values will increase gradually as the sulfate pool is being exhausted, and consequently, the  $\delta^{34}\text{S}_{\text{pyrite}}$  values in pyrite will show a large span in values and increase from core to rim of the pyrite crystals [98]. Therefore, we propose that the large  $\delta^{34}\text{S}_{\text{pyrite}}$  variability spanning almost 50‰ (Figure 4d) within the individual fractures marks Rayleigh isotope reservoir effects in the fracture system. The increase in  $\delta^{34}\text{S}_{\text{pyrite}}$  values across the crystal is rather abrupt, and occurs mainly close to the crystal rims. This suggests that the pyrite in the rim has precipitated in relation to MSR of a dissolved sulfate pool that was already enriched in  $^{34}\text{S}$  when reaching the fracture, and that the system undergoing Rayleigh fractionation is larger than the individual sampled borehole intercepts. The observed spatiotemporal relation between calcite growth zones depleted in  $^{13}\text{C}$  and pyrite growth zones enriched in  $^{34}\text{S}$  (outermost parts of the crystals, Figures 3c and 5d) is similar to observations at SMTZ:s in seabed systems [99,100]. It is intriguing that the  $\delta^{34}\text{S}_{\text{SO}_4}$  values are elevated (up to  $+57.5\%$ ) in the fractures with highest  $\delta^{34}\text{S}_{\text{pyrite}}$  values. This shows that the modern dissolved sulfate in the fracture system is affected by MSR-related Rayleigh isotope fractionation, which is plausible considering that sulfide is currently produced [101,102], and sulfate-reducing bacteria are detected in modern waters [8], although presence of viable cells is not direct proof for active metabolism. Pyrite with MSR-related  $\delta^{34}\text{S}$  values also occurs together with calcite with methanogenesis-related  $\delta^{13}\text{C}$  values, despite that there may be competition between sulfate reducers and methanogens for substrates. Explanations for this may be that methanogenesis is from non-competitive substrates, or an excess of substrates keeps concentrations above the threshold for methanogenesis (DOC presence in the water is locally at fairly high concentrations;  $\sim 15$  mg/L).

The deepest pyrite sample (KFM08D) showed a smaller  $\delta^{34}\text{S}_{\text{pyrite}}$  range (−2 to +12‰), and may theoretically be a result of thermochemical reduction.

#### 4.4. Putative Microfossils

The abundance of CM as biofilms and filaments in the KFM24:379 and KFM24:395–399 sections suggests previous presence of microorganisms in the fractures. Considering the immediate sampling of the samples after drilling, the sealed veins they occurred in, and the partly mineralized nature of the remnants, the biofilms and associated preserved microorganisms are likely to be primary, and not modern contaminants. Microbial remains of similar modes of preservation with carbonaceous and partly mineralized portions have previously been reported from fractures in granitic bedrock [20,103]. The widths of the filaments are 10–40  $\mu\text{m}$ , which exclude most prokaryotes and suggest that the filaments are remains of fungal hyphae. Size-wise, cyanobacteria and giant sulfate bacteria are the only possible prokaryotic candidates, but both can be ruled out considering the anoxic and dark environment. Fungi, on the other hand, have been shown to thrive in such deep settings, feeding on biomass from prokaryotes, and upon death, become preserved in an identical manner as the current fossils [20,103,104]. The branching and entangled appearance is characteristic for fungal hypha and mycelium, as well, even though it is not diagnostic for fungi alone [103]. Observations of fungi in the deep biosphere have, until recently, been more or less exclusively from the marine or sub-seafloor realm [105]. Recent detections of fungi in deep waters from Olkiluoto, Finland [19], and fossilized remains at great depth in Laxemar, Sweden [20] suggest that fungi play a key role in this biosphere [106], where they seem to consort with prokaryotes [103,107]. Our current findings from the deep bedrock fractures Forsmark of putative partly mineralized fungal hyphae occurring together with minerals with microbial activity-specific stable isotope signatures (e.g., MSR) are in agreement with the recent observations from the deep continental biosphere. Heterotrophic fungi require a continuous growth of associated prokaryotic biomass available for their metabolism. In deep anoxic systems, it has been suggested that obligate anaerobic eukaryotes, including fungi, equipped with hydrogenosomes may produce  $\text{H}_2$  at a rate that makes continuous growth of  $\text{H}_2$ -dependent prokaryotes possible, whereas rates of  $\text{H}_2$  formation by radiolysis is probably too low for that [20,107]. In addition, microbial activity in combination with mechanochemical reactions may also enhance  $\text{H}_2$  production [108].

#### 4.5. Potential Quaternary Mineral Precipitation

The correlation between isotopic signatures of the outermost mineral growth zones and the waters in some of the sections means that Quaternary precipitation cannot be completely ruled out. The best candidates are around 400 m depth in KFM24, where there are overlaps in  $^{87}\text{Sr}/^{86}\text{Sr}$  between calcite and water, corresponding to  $\delta^{18}\text{O}$  and heavy  $\delta^{34}\text{S}$  values, both in the outermost growth zones of pyrite crystals and in the dissolved sulfate. Additional indications of relatively recent microbial activity are the remains of carbonaceous matter on the fracture surfaces here. However, overall, it can be concluded that most of the calcite formation is pre-Quaternary, and at least in the inner parts of the crystals, most likely can be linked to Paleozoic and Mesozoic precipitation events, which have been documented in other parts of the Forsmark area [29,39], and by our single Rb–Sr dating (Figure 7). Microbial activity has apparently been active over several hundred million years in this area. Extensive mineral precipitation related to microbial activity has, however, been episodic in nature.

## 5. Conclusions

This comprehensive isotope study of minerals, waters, and gases from the target area for a spent nuclear fuel repository at Forsmark, Sweden shows long-term evidence of microbial processes in the deep fracture system. The transects of microscale isotope analyses provided unique information about the evolution of the deep fracture system in the Forsmark area during ambient temperature conditions, and allowed discrimination of processes that would not have been possible using conventional isotope analysis of bulk samples. The large isotope variabilities of  $\delta^{13}\text{C}_{\text{calcite}}$  (−36.2‰ to +20.2‰)

and  $\delta^{34}\text{S}_{\text{pyrite}}$  (−11.7‰ to +37.8‰) mark microbial methanogenesis, potential anaerobic oxidation of methane, and coupled microbial sulfate reduction in the fracture system. A potential sulfate–methane transition zone was detected at 395–415 m in the shaft pilot borehole. Despite the potential triggers for carbonate precipitation, i.e., (1) active modern microbial processes, such as MSR (shown by isotopic signatures and detected microorganisms), and (2) large degree of water mixing during the Quaternary, potential modern mineral precipitates are few, with the best potential in the shaft pilot borehole at around 400 m depth. The large temporal isotopic variability within the minerals and radiometric dating of one of the fracture coatings point to episodic microbial activity-related mineral precipitation that dates back, at least, to the Devonian time period.

**Supplementary Materials:** The following are available online at <http://www.mdpi.com/2076-3263/8/6/211/s1>, Supplementary Material 1 consists of a PDF-file of ESEM images and corresponding EDS-spectra. Supplementary tables are enclosed as a spreadsheet (xlsx-file). These tables contain microorganism data (Table S1), isotope data (spot-by-spot) of calcite (Table S2) and pyrite (Table S3), (spot-by-spot) reference material analysis for C isotopes (Table S4), O isotopes (Table S5), Sr isotopes (Table S6) and S isotopes (Table S7). Rb/Sr isotope (spot-by-spot) data are presented in Table S8.

**Author Contributions:** H.D. conceived and designed the study and wrote the paper; M.I. and H.D. performed the SEM-analyses and M.I. wrote the part about the fossilized microorganisms; H.D. performed the SIMS and LA-MC-ICP analyses together with M.J.W. and E.K., respectively; M.T. and H.D. performed the Rb–Sr dating and M.T. performed corresponding data handling.

**Acknowledgments:** We acknowledge the following grants: Swedish research council (grant #2017-05186, to H.D., and grant#2015-04129 to M.I.), Formas (grant #2017-00766, to H.D.), SKB (grant #19216, to H.D.) and a Villum Investigator grant to Donald Canfield. Kenneth Åkerström (Sweco AB), Kerstin Lindén (Swedish Museum of Natural History), Thomas Zack (University of Gothenburg), Johan Hogmalm (University of Gothenburg) and Melanie Schmitt (Swedish Museum of Natural History) are thanked for assistance during sampling, sample preparation and analysis and Eva-Lena Tullborg (Terralogica AB), Ann-Chatrin Nilsson (Geosigma AB), Mats Tröjbom and Cecilia Berg (SKB) are thanked for providing quality controlled hydrochemical and gas data from the Forsmark site. This is NordSIM publication 562 and Vegacenter publication #9. All cited SKB reports can be downloaded from [www.skb.se](http://www.skb.se).

**Conflicts of Interest:** The authors declare no conflict of interest and the founding sponsors had no role in the design of the study; in the collection, analyses, or interpretation of data; in the writing of the manuscript, and in the decision to publish the results.

## References

1. McMahon, S.; Parnell, J. Weighing the deep continental biosphere. *FEMS Microbiol. Ecol.* **2014**, *87*, 113–120. [[CrossRef](#)] [[PubMed](#)]
2. Lau, M.C.Y.; Kieft, T.L.; Kuloyo, O.; Linage-Alvarez, B.; van Heerden, E.; Lindsay, M.R.; Magnabosco, C.; Wang, W.; Wiggins, J.B.; Guo, L.; et al. An oligotrophic deep-subsurface community dependent on syntrophy is dominated by sulfur-driven autotrophic denitrifiers. *Proc. Natl. Acad. Sci. USA* **2016**, *113*, E7927–E7936. [[CrossRef](#)] [[PubMed](#)]
3. Simkus, D.N.; Slater, G.F.; Lollar, B.S.; Wilkie, K.; Kieft, T.L.; Magnabosco, C.; Lau, M.C.Y.; Pullin, M.J.; Hendrickson, S.B.; Wommack, K.E.; et al. Variations in microbial carbon sources and cycling in the deep continental subsurface. *Geochim. Cosmochim. Acta* **2016**, *173*, 264–283. [[CrossRef](#)]
4. Chivian, D.; Brodie, E.L.; Alm, E.J.; Culley, D.E.; Dehal, P.S.; DeSantis, T.Z.; Gihring, T.M.; Lapidus, A.; Lin, L.-H.; Lowry, S.R.; et al. Environmental Genomics Reveals a Single-Species Ecosystem Deep Within Earth. *Science* **2008**, *322*, 275–278. [[CrossRef](#)] [[PubMed](#)]
5. Onstott, T.C.; McGown, D.J.; Bakermans, C.; Ruskeeniemi, T.; Ahonen, L.; Telling, J.; Soffientino, B.; Piffner, S.M.; Sherwood-Lollar, B.; Frappe, S.; et al. Microbial Communities in Subpermafrost Saline Fracture Water at the Lupin Au Mine, Nunavut, Canada. *Microb. Ecol.* **2009**, *58*, 786–807. [[CrossRef](#)] [[PubMed](#)]
6. Wu, X.; Holmfeldt, K.; Hubalek, V.; Lundin, D.; Åström, M.; Bertilsson, S.; Dopson, M. Microbial metagenomes from three aquifers in the Fennoscandian shield terrestrial deep biosphere reveal metabolic partitioning among populations. *ISME J.* **2016**, *10*, 1192–1203. [[CrossRef](#)] [[PubMed](#)]
7. Pedersen, K. Subterranean microbial populations metabolize hydrogen and acetate under in situ conditions in granitic groundwater at 450 m depth in the Aspo Hard Rock Laboratory, Sweden. *FEMS Microbiol. Ecol.* **2012**, *81*, 217–29. [[CrossRef](#)] [[PubMed](#)]

8. Hallbeck, L.; Pedersen, K. Culture-dependent comparison of microbial diversity in deep granitic groundwater from two sites considered for a Swedish final repository of spent nuclear fuel. *FEMS Microbiol. Ecol.* **2012**, *81*, 66–77. [[CrossRef](#)] [[PubMed](#)]
9. Hallbeck, L.; Pedersen, K. Characterization of microbial processes in deep aquifers of the Fennoscandian Shield. *Appl. Geochem.* **2008**, *23*, 1796–1819. [[CrossRef](#)]
10. Pedersen, K. Microbial life in granitic rock. *FEMS Microbiol. Rev.* **1997**, *20*, 399–414. [[CrossRef](#)]
11. Haveman, S.A.; Pedersen, K.; Ruotsalainen, P. Distribution and metabolic diversity in of microorganisms in deep igneous rock aquifers in Finland. *Geomicrobiology* **1999**, *16*, 277–294.
12. Purkamo, L.; Bomberg, M.; Nyyssönen, M.; Kukkonen, I.; Ahonen, L.; Kietäväinen, R.; Itävaara, M. Dissecting the deep biosphere: Retrieving authentic microbial communities from packer-isolated deep crystalline bedrock fracture zones. *FEMS Microbiol. Ecol.* **2013**, *85*, 324–337. [[CrossRef](#)] [[PubMed](#)]
13. Pedersen, K.; Bengtsson, A.F.; Edlund, J.S.; Eriksson, L.C. Sulphate-controlled Diversity of Subterranean Microbial Communities over Depth in Deep Groundwater with Opposing Gradients of Sulphate and Methane. *Geomicrobiol. J.* **2014**, *31*, 617–631. [[CrossRef](#)]
14. Pedersen, K. Metabolic activity of subterranean microbial communities in deep granitic groundwater supplemented with methane and H<sub>2</sub>. *ISME J.* **2012**, *7*, 839. [[CrossRef](#)] [[PubMed](#)]
15. Pedersen, K.; Arlinger, J.; Hallbeck, A.; Hallbeck, L.; Eriksson, S.; Johansson, J. Numbers, biomass and cultivable diversity of microbial populations related to depth and borehole-specific conditions in groundwater from depths of 4 to 450 m in Olkiluoto, Finland. *ISME J.* **2008**, *2*, 760–775. [[CrossRef](#)] [[PubMed](#)]
16. Ino, K.; Hermsdorf, A.W.; Konno, U.; Kouduka, M.; Yanagawa, K.; Kato, S.; Sunamura, M.; Hirota, A.; Togo, Y.S.; Ito, K.; et al. Ecological and genomic profiling of anaerobic methane-oxidizing archaea in a deep granitic environment. *ISME J.* **2017**, *12*, 31–47. [[CrossRef](#)] [[PubMed](#)]
17. Ino, K.; Konno, U.; Kouduka, M.; Hirota, A.; Togo, Y.S.; Fukuda, A.; Komatsu, D.; Tsunogai, U.; Tanabe, A.S.; Yamamoto, S.; et al. Deep microbial life in high-quality granitic groundwater from geochemically and geographically distinct underground boreholes. *Environ. Microbiol. Rep.* **2016**, *8*, 285–294. [[CrossRef](#)] [[PubMed](#)]
18. Nyyssönen, M.; Hultman, J.; Ahonen, L.; Kukkonen, I.; Paulin, L.; Laine, P.; Itävaara, M.; Auvinen, P. Taxonomically and functionally diverse microbial communities in deep crystalline rocks of the Fennoscandian shield. *ISME J.* **2014**, *8*, 126–138. [[CrossRef](#)] [[PubMed](#)]
19. Sohlberg, E.; Bomberg, M.; Miettinen, H.; Nyyssönen, M.; Salavirta, H.; Vikman, M.; Itävaara, M. Revealing the unexplored fungal communities in deep groundwater of crystalline bedrock fracture zones in Olkiluoto, Finland. *Front. Microbiol.* **2015**, *6*, 573. [[CrossRef](#)] [[PubMed](#)]
20. Drake, H.; Ivarsson, M.; Bengtson, S.; Heim, C.; Siljeström, S.; Whitehouse, M.J.; Broman, C.; Belivanova, V.; Åström, M.E. Anaerobic consortia of fungi and sulfate reducing bacteria in deep granite fractures. *Nat. Commun.* **2017**, *8*, 55. [[CrossRef](#)] [[PubMed](#)]
21. Hubalek, V.; Wu, X.; Eiler, A.; Buck, M.; Heim, C.; Dopson, M.; Bertilsson, S.; Ionescu, D. Connectivity to the surface determines diversity patterns in subsurface aquifers of the Fennoscandian shield. *ISME J.* **2016**, *10*, 2447–2458. [[CrossRef](#)] [[PubMed](#)]
22. Chapelle, F.H.; O'Neill, K.; Bradley, P.M.; Methe, B.A.; Ciuffo, S.A.; Knobel, L.L.; Lovley, D.R. A hydrogen-based subsurface microbial community dominated by methanogens. *Nature* **2002**, *415*, 312–315. [[CrossRef](#)] [[PubMed](#)]
23. Kietäväinen, R.; Purkamo, L. The origin, source, and cycling of methane in deep crystalline rock biosphere. *Front. Microbiol.* **2015**, *6*, 725. [[CrossRef](#)] [[PubMed](#)]
24. Onstott, T.C.; Lin, L.H.; Davidson, M.; Mislouck, B.; Borcsik, M.; Hall, J.; Slater, G.; Ward, J.; Lollar, B.S.; Lippmann-Pipke, J.; et al. The Origin and Age of Biogeochemical Trends in Deep Fracture Water of the Witwatersrand Basin, South Africa. *Geomicrobiol. J.* **2006**, *23*, 369–414. [[CrossRef](#)]
25. Kotelnikova, S. Microbial production and oxidation of methane in deep subsurface. *Earth-Sci. Rev.* **2002**, *58*, 367–395. [[CrossRef](#)]
26. Stevens, T.O.; McKinley, J.P. Lithoautotrophic Microbia, Ecosystems in Deep Basalt Aquifers. *Science* **1995**, *270*, 450–454. [[CrossRef](#)]
27. Sherwood Lollar, B.; Frape, S.K.; Fritz, P.; Macko, S.A.; Welhan, J.A.; Blomqvist, R.; Lahermo, P.W. Evidence for bacterially generated hydrocarbon gas in Canadian shield and fennoscandian shield rocks. *Geochim. Cosmochim. Acta* **1993**, *57*, 5073–5085. [[CrossRef](#)]



28. Kieft, T.L.; Walters, C.C.; Higgins, M.B.; Mennito, A.S.; Clewett, C.F.M.; Heuer, V.; Pullin, M.J.; Hendrickson, S.; van Heerden, E.; Sherwood Lollar, B.; et al. Dissolved organic matter compositions in 0.6–3.4 km deep fracture waters, Kaapvaal Craton, South Africa. *Org. Geochem.* **2018**, *118*, 116–131. [[CrossRef](#)]
29. Drake, H.; Heim, C.; Roberts, N.M.W.; Zack, T.; Tillberg, M.; Broman, C.; Ivarsson, M.; Whitehouse, M.J.; Åström, M.E. Isotopic evidence for microbial production and consumption of methane in the upper continental crust throughout the Phanerozoic eon. *Earth Planet. Sci. Lett.* **2017**, *470*, 108–118. [[CrossRef](#)]
30. Drake, H.; Åström, M.E.; Heim, C.; Broman, C.; Åström, J.; Whitehouse, M.J.; Ivarsson, M.; Siljeström, S.; Sjövall, P. Extreme <sup>13</sup>C-depletion of carbonates formed during oxidation of biogenic methane in fractured granite. *Nat. Commun.* **2015**, *6*, 7020. [[CrossRef](#)] [[PubMed](#)]
31. Drake, H.; Tullborg, E.-L. Paleohydrogeological events recorded by stable isotopes, fluid inclusions and trace elements in fracture minerals in crystalline rock, Simpevarp area, SE Sweden. *Appl. Geochem.* **2009**, *24*, 715–732. [[CrossRef](#)]
32. Sandström, B.; Tullborg, E.-L. Episodic fluid migration in the Fennoscandian Shield recorded by stable isotopes, rare earth elements and fluid inclusions in fracture minerals at Forsmark, Sweden. *Chem. Geol.* **2009**, *266*, 126–142. [[CrossRef](#)]
33. Drake, H.; Heim, C.; Høgmalm, K.J.; Hansen, B.T. Fracture zone-scale variation of trace elements and stable isotopes in calcite in a crystalline rock setting. *Appl. Geochem.* **2014**, *40*, 11–24. [[CrossRef](#)]
34. Heim, C.; Lausmaa, J.; Sjövall, P.; Toporski, J.; Dieing, T.; Simon, K.; Hansen, B.T.; Kronz, A.; Arp, G.; Reitner, J.; et al. Ancient microbial activity recorded in fracture fillings from granitic rocks (Åspö Hard Rock Laboratory, Sweden). *Geobiology* **2012**, *10*, 280–297. [[CrossRef](#)] [[PubMed](#)]
35. Ziegenbalg, S.B.; Birgel, D.; Hoffmann-Sell, L.; Pierre, C.; Rouchy, J.M.; Peckmann, J. Anaerobic oxidation of methane in hypersaline Messinian environments revealed by <sup>13</sup>C-depleted molecular fossils. *Chem. Geol.* **2012**, *292–293*, 140–148. [[CrossRef](#)]
36. Peckmann, J.; Thiel, V. Carbon cycling at ancient methane-seeps. *Chem. Geol.* **2004**, *205*, 443–467. [[CrossRef](#)]
37. Drake, H.; Åström, M.; Tullborg, E.-L.; Whitehouse, M.J.; Fallick, A.E. Variability of sulphur isotope ratios in pyrite and dissolved sulphate in granitoid fractures down to 1 km depth—Evidence for widespread activity of sulphur reducing bacteria. *Geochim. Cosmochim. Acta* **2013**, *102*, 143–161. [[CrossRef](#)]
38. Sahlstedt, E.; Karhu, J.A.; Pitkänen, P.; Whitehouse, M. Implications of sulfur isotope fractionation in fracture-filling sulfides in crystalline bedrock, Olkiluoto, Finland. *Appl. Geochem.* **2013**, *32*, 52–69. [[CrossRef](#)]
39. Sandström, B.; Tullborg, E.-L.; Larson, S.Å.; Page, L. Brittle tectonothermal evolution in the Forsmark area, central Fennoscandian Shield, recorded by paragenesis, orientation and <sup>40</sup>Ar/<sup>39</sup>Ar geochronology of fracture minerals. *Tectonophysics* **2009**, *478*, 158–174. [[CrossRef](#)]
40. Andersson, J.; Skagius, K.; Winberg, A.; Lindborg, T.; Ström, A. Site-descriptive modelling for a final repository for spent nuclear fuel in Sweden. *Environ. Earth Sci.* **2013**, *69*, 1045–1060. [[CrossRef](#)]
41. Laaksoharju, M.; Smellie, J.; Tullborg, E.-L.; Gimeno, M.; Molinero, J.; Gurban, I.; Hallbeck, L. Hydrogeochemical evaluation and modelling performed within the Swedish site investigation programme. *Appl. Geochem.* **2008**, *23*, 1761–1795. [[CrossRef](#)]
42. Sandström, B.; Stephens, M.B. *Mineralogy, Geochemistry, Porosity and Redox Properties of Rocks from Forsmark. Compilation of Data from the Regional Model Volume for SR-Site; SKB-R-09-51*; Swedish Nuclear Fuel and Waste Management Co. (SKB): Stockholm, Sweden, 2009.
43. Stephens, M.B.; Fox, A.; La Pointe, P.; Simeonov, A.; Isaksson, H.; Hermanson, J.; Öhman, J. *Geology Forsmark. Site Descriptive Modelling Forsmark Stage 2.2; R-07-45*; Swedish Nuclear Fuel and Waste Management Co.: Stockholm, Sweden, 2007.
44. Drake, H.; Sandström, B.; Tullborg, E.-L. *Mineralogy and Geochemistry of Rocks and Fracture Fillings from Forsmark and Oskarshamn: Compilation of Data for SR-Can*; SKB Report R-06-109; SKB: Stockholm, Sweden, 2006; p. 105.
45. Saintot, A.; Stephens, M.B.; Viola, G.; Nordgulen, O. Brittle tectonic evolution and paleostress field reconstruction in the southwestern part of the Fennoscandian Shield, Forsmark, Sweden. *Tectonics* **2011**, *30*. [[CrossRef](#)]
46. Dahlin, P.; Maskenskaya, O. *Boremap Mapping of Core Drilled Borehole KFM24*; SKB-Report P-16-28; Swedish Nuclear Fuel and Waste Management Co. (SKB): Stockholm, Sweden, 2017.

47. Samuelsson, E.; Rauséus, G. *Boremap Mapping of Telescopic Drilled Borehole KFM08D. Forsmark Site Investigation*; SKB-Report P-07-103; Swedish Nuclear Fuel and Waste Management Co. (SKB): Stockholm, Sweden, 2007.
48. Hurmerinta, E. *Forsmark Site Investigations. Difference Flow Logging in Borehole KFM24*; SKB-Report P-16-27; Swedish Nuclear Fuel and Waste Management Co. (SKB): Stockholm, Sweden, 2017.
49. Teurneau, B.; Forsmark, T.; Forssman, I.; Rhén, I.; Zinn, E. *Correlation of Posiva Flow Log Anomalies to Core Mapped Features in KFM01D, KFM07C, KFM08A, KFM08C and KFM10A. Forsmark Site Investigation*; SKB-Report P-07-127; Swedish Nuclear Fuel and Waste Management Co. (SKB): Stockholm, Sweden, 2008.
50. Laaksoharju, M.; Smellie, J.; Tullborg, E.-L.; Gimeno, M.; Hallbeck, L.; Molinero, J.; Waber, N. *Bedrock hydrogeochemistry Forsmark. Site Descriptive Modelling. SDM-Site Forsmark*; SKB Report R-08-47; Swedish Nuclear Fuel and Waste Management Co. (SKB): Stockholm, Sweden, 2008.
51. Pedersen, K. *Microorganisms in Groundwater from Boreholes KFM10A, KFM11A and KFM08D—Numbers, Viability, and Metabolic Diversity. Forsmark Site Investigation; Results from Five Sections 298.0-305.1 m and 478.0-487.5 m in KFM10A, 447.5-454.6 m in KFM11A, and 669.7-676.8 m and 828.4-835.5 m in KFM08D*; SKB-Report P-07-198; Swedish Nuclear Fuel and Waste Management Co. (SKB): Stockholm, Sweden, 2007.
52. Bergelin, A.; Lindquist, A.; Nilsson, K.; Wacker, P.; Nilsson, A.-C. *Hydrochemical Characterisation in Borehole KFM08D. Forsmark Site Investigation; Results from Two Investigated Borehole Sections at 669.7-676.8 m and 828.4-835.5 m*; SKB-Report P-07-190; Swedish Nuclear Fuel and Waste Management Co. (SKB): Stockholm, Sweden, 2007.
53. Hallbeck, L.; Pedersen, K. *Explorative Analysis of Microbes, Colloids and Gases. SDM-Site Forsmark*; SKB Report R-08-85; Swedish Nuclear Fuel and Waste Management Co. (SKB): Stockholm, Sweden, 2008.
54. Kamber, B.S.; Whitehouse, M.J. Micro-scale sulphur isotope evidence for sulphur cycling in the late Archean shallow ocean. *Geobiology* **2007**, *5*, 5–17. [[CrossRef](#)]
55. Drake, H.; Tullborg, E.-L.; Sandberg, B.; Blomfeldt, T.; Åström, M.E. Extreme fractionation and micro-scale variation of sulphur isotopes during bacterial sulphate reduction in Deep groundwater systems. *Geochim. Cosmochim. Acta* **2015**, *161*, 1–18. [[CrossRef](#)]
56. Hogmalm, K.J.; Zack, T.; Karlsson, A.K.O.; Sjoqvist, A.S.L.; Garbe-Schonberg, D. In situ Rb-Sr and K-Ca dating by LA-ICP-MS/MS: An evaluation of N<sub>2</sub>O and SF<sub>6</sub> as reaction gases. *J. Anal. At. Spectrom.* **2017**, *32*, 305–313. [[CrossRef](#)]
57. Zack, T.; Hogmalm, J. Laser ablation Rb/Sr dating by online chemical separation of Rb and Sr in an oxygen-filled reaction cell. *Chem. Geol.* **2016**, *437*, 120–133. [[CrossRef](#)]
58. Jochum, K.P.; Weis, U.; Stoll, B.; Kuzmin, D.; Yang, Q.; Raczek, I.; Jacob, D.E.; Stracke, A.; Birbaum, K.; Frick, D.A.; et al. Determination of reference values for NIST SRM 610–617 glasses following ISO guidelines. *Geostand. Geoanal. Res.* **2011**, *35*, 397–429. [[CrossRef](#)]
59. Elburg, M.; Vroon, P.; van der Wagt, B.; Tchalikian, A. Sr and Pb isotopic composition of five USGS glasses (BHVO-2G, BIR-1G, BCR-2G, TB-1G, NKT-1G). *Chem. Geol.* **2005**, *223*, 196–207. [[CrossRef](#)]
60. Kiel, S.; Glodny, J.; Birgel, D.; Bulot, L.G.; Campbell, K.A.; Gaillard, C.; Graziano, R.; Kaim, A.; Lazăr, I.; Sandy, M.R.; et al. The Paleocology, Habitats, and Stratigraphic Range of the Enigmatic Cretaceous Brachiopod *Peregrinella*. *PLoS ONE* **2014**, *9*, e109260. [[CrossRef](#)] [[PubMed](#)]
61. Mokadem, F.; Parkinson, I.J.; Hathorne, E.C.; Anand, P.; Allen, J.T.; Burton, K.W. High-precision radiogenic strontium isotope measurements of the modern and glacial ocean: Limits on glacial–interglacial variations in continental weathering. *Earth Planet. Sci. Lett.* **2015**, *415*, 111–120. [[CrossRef](#)]
62. Emrich, K.; Ehhalt, D.H.; Vogel, J.C. Carbon isotope fractionation during the precipitation of calcium carbonate. *Earth Planet. Sci. Lett.* **1970**, *8*, 363–371. [[CrossRef](#)]
63. Kim, S.-T.; O’Neil, J.R. Equilibrium and nonequilibrium oxygen isotope effects in synthetic carbonates. *Geochim. Cosmochim. Acta* **1997**, *61*, 3461–3475. [[CrossRef](#)]
64. Drake, H.; Mathurin, F.A.; Zack, T.; Schäfer, T.; Roberts, N.M.W.; Whitehouse, M.; Karlsson, A.; Broman, C.; Åström, M.E. Incorporation of metals into calcite in a deep anoxic granite aquifer. *Environ. Sci. Technol.* **2018**, *52*, 493–502. [[CrossRef](#)] [[PubMed](#)]
65. Whiticar, M.J. Carbon and hydrogen isotope systematics of bacterial formation and oxidation of methane. *Chem. Geol.* **1999**, *161*, 291–314. [[CrossRef](#)]
66. Boehme, S.E.; Blair, N.E.; Chanton, J.P.; Martens, C.S. A mass balance of <sup>13</sup>C and <sup>12</sup>C in an organic-rich methane-producing marine sediment. *Geochim. Cosmochim. Acta* **1996**, *60*, 3835–3848. [[CrossRef](#)]

67. Irwin, H.; Cultis, C.; Coleman, M. Isotopic evidence for source of diagenetic carbonates formed during burial of organic-rich sediments. *Nature* **1977**, *269*, 209–213. [[CrossRef](#)]
68. Budai, J.M.; Martini, A.M.; Walter, L.M.; Ku, T.C.W. Fracture-fill calcite as a record of microbial methanogenesis and fluid migration; a case study from the Devonian Antrim Shale, Michigan Basin. *Geofluids* **2002**, *2*, 163–183. [[CrossRef](#)]
69. Sahlstedt, E.; Karhu, J.A.; Pitkänen, P.; Whitehouse, M. Biogenic processes in crystalline bedrock fractures indicated by carbon isotope signatures of secondary calcite. *Appl. Geochem.* **2016**, *67*, 30–41. [[CrossRef](#)]
70. Sahlstedt, E.; Karhu, J.A.; Pitkänen, P. Indications for the past redox environments in deep groundwaters from the isotopic composition of carbon and oxygen in fracture calcite, Olkiluoto, SW Finland. *Isotopes Environ. Health Stud.* **2010**, *46*, 370–391. [[CrossRef](#)] [[PubMed](#)]
71. Clauer, N.; Frapé, S.K.; Fritz, B. Calcite veins of the Stripa granite (Sweden) as records of the origin of the groundwaters and their interactions with the granitic body. *Geochim. Cosmochim. Acta* **1989**, *53*, 1777–1781. [[CrossRef](#)]
72. Kietäväinen, R.; Ahonen, L.; Niinikoski, P.; Nykänen, H.; Kukkonen, I.T. Abiotic and biotic controls on methane formation down to 2.5 km depth within the Precambrian Fennoscandian Shield. *Geochim. Cosmochim. Acta* **2017**, *202*, 124–145. [[CrossRef](#)]
73. Etiopé, G.; Sherwood Lollar, B. Abiotic methane on Earth. *Rev. Geophys.* **2013**, *51*, 276–299. [[CrossRef](#)]
74. Lovley, D.R.; Klug, M.J. Sulfate Reducers Can Outcompete Methanogens at Freshwater Sulfate Concentrations. *Appl. Environ. Microbiol.* **1983**, *45*, 187–192. [[PubMed](#)]
75. Sela-Adler, M.; Ronen, Z.; Herut, B.; Antler, G.; Vigderovich, H.; Eckert, W.; Sivan, O. Co-existence of Methanogenesis and Sulfate Reduction with Common Substrates in Sulfate-Rich Estuarine Sediments. *Front. Microbiol.* **2017**, *8*, 766. [[CrossRef](#)] [[PubMed](#)]
76. Holmer, M.; Kristensen, E. Coexistence of sulfate reduction and methane production in an organic-rich sediment. *Mar. Ecol. Prog. Ser.* **1994**, *107*, 177–184. [[CrossRef](#)]
77. Jørgensen, B.B.; Parkes, R.J. Role of sulfate reduction and methane production by organic carbon degradation in eutrophic fjord sediments (Limfjorden, Denmark). *Limnol. Oceanogr.* **2010**, *55*, 1338–1352. [[CrossRef](#)]
78. Bernard, B.B.; Brooks, J.M.; Sackett, W.M. Light hydrocarbons in recent Texas continental shelf and slope sediments. *J. Geophys. Res. Oceans* **1978**, *83*, 4053–4061. [[CrossRef](#)]
79. Etiopé, G.; Schoell, M. Abiotic Gas: Atypical, But Not Rare. *Elements* **2014**, *10*, 291–296. [[CrossRef](#)]
80. Garcia, J.-L.; Patel, B.K.C.; Ollivier, B. Taxonomic, Phylogenetic, and Ecological Diversity of Methanogenic Archaea. *Anaerobe* **2000**, *6*, 205–226. [[CrossRef](#)] [[PubMed](#)]
81. Thauer, R.K.; Kaster, A.-K.; Seedorf, H.; Buckel, W.; Hedderich, R. Methanogenic archaea: Ecologically relevant differences in energy conservation. *Nat. Rev. Microbiol.* **2008**, *6*, 579–591. [[CrossRef](#)] [[PubMed](#)]
82. Whiticar, M.J.; Faber, E.; Schoell, M. Biogenic methane formation in marine and freshwater environments: CO<sub>2</sub> reduction vs. acetate fermentation—Isotope evidence. *Geochim. Cosmochim. Acta* **1986**, *50*, 693–709. [[CrossRef](#)]
83. Sandström, B.; Tullborg, E.-L.; de Torres, T.; Ortiz, J.E. The occurrence and potential origin of asphaltite in bedrock fractures, Forsmark, central Sweden. *GFF* **2006**, *128*, 234–242. [[CrossRef](#)]
84. Knittel, K.; Boetius, A. Anaerobic Oxidation of Methane: Progress with an Unknown Process. *Ann. Rev. Microbiol.* **2009**, *63*, 311–334. [[CrossRef](#)] [[PubMed](#)]
85. Michaelis, W.; Seifert, R.; Nauhaus, K.; Treude, T.; Thiel, V.; Blumenberg, M.; Knittel, K.; Gieseke, A.; Peterknecht, K.; Pape, T.; et al. Microbial reefs in the Black Sea fueled by anaerobic oxidation of methane. *Science* **2002**, *297*, 1013–1015. [[CrossRef](#)] [[PubMed](#)]
86. Orphan, V.J.; Hinrichs, K.U.; Ussler, W.; Paull, C.K.; Taylor, L.T.; Sylva, S.P.; Hayes, J.M.; Delong, E.F. Comparative Analysis of Methane-Oxidizing Archaea and Sulfate-Reducing Bacteria in Anoxic Marine Sediments. *Appl. Environ. Microbiol.* **2001**, *67*, 1922–1934. [[CrossRef](#)] [[PubMed](#)]
87. Bomberg, M.; Nyssönen, M.; Pitkänen, P.; Lehtinen, A.; Itävaara, M. Active Microbial Communities Inhabit Sulphate-Methane Interphase in Deep Bedrock Fracture Fluids in Olkiluoto, Finland. *BioMed Res. Int.* **2015**, *2015*, 979530. [[CrossRef](#)] [[PubMed](#)]
88. Natalicchio, M.; Birgel, D.; Dela Pierre, F.; Martire, L.; Clari, P.; Spötl, C.; Peckmann, J. Polyphasic carbonate precipitation in the shallow subsurface: Insights from microbially-formed authigenic carbonate beds in upper Miocene sediments of the Tertiary Piedmont Basin (NW Italy). *Palaeogeogr. Palaeoclimatol. Palaeoecol.* **2012**, *329–330*, 158–172. [[CrossRef](#)]

89. Feng, D.; Chen, D.; Peckmann, J.; Bohrmann, G. Authigenic carbonates from methane seeps of the northern Congo fan: Microbial formation mechanism. *Mar. Pet. Geol.* **2010**, *27*, 748–756. [[CrossRef](#)]
90. Reeburgh, W.S. Oceanic methane biogeochemistry. *Chem. Rev.* **2007**, *107*, 486–513. [[CrossRef](#)] [[PubMed](#)]
91. Whiticar, M.J.; Faber, E. Methane oxidation in sediment and water column environments—Isotope evidence. *Org. Geochem.* **1986**, *10*, 759–768. [[CrossRef](#)]
92. Campbell, K.A.; Farmer, J.D.; des Marais, D. Ancient hydrocarbon seeps from the Mesozoic convergent margin of California: Carbonate geochemistry, fluids and palaeoenvironments. *Geofluids* **2002**, *2*, 63–94. [[CrossRef](#)]
93. Peckmann, J.; Goedert, J.L.; Thiel, V.; Michaelis, W.; Reitner, J. A comprehensive approach to the study of methane-seep deposits from the Lincoln Creek Formation, western Washington State, USA. *Sedimentology* **2002**, *49*, 855–873. [[CrossRef](#)]
94. Roberts, H.H.; Aharon, P. Hydrocarbon-derived carbonate buildups of the northern Gulf of Mexico continental slope: A review of submersible investigations. *Geo-Mar. Lett.* **1994**, *14*, 135–148. [[CrossRef](#)]
95. Böttcher, M.E.; Smock, A.M.; Cypionka, H. Sulfur isotope fractionation during experimental precipitation of iron(II) and manganese(II) sulfide at room temperature. *Chem. Geol.* **1998**, *146*, 127–134. [[CrossRef](#)]
96. Sim, M.S.; Bosak, T.; Ono, S. Large sulfur isotope fractionation does not require disproportionation. *Science* **2011**, *333*, 74–77. [[CrossRef](#)] [[PubMed](#)]
97. Kiyosu, Y.; Krouse, H.R. The role of organic acid in the abiogenic reduction of sulfate and the sulfur isotope effect. *Geochem. J.* **1990**, *24*, 21–27. [[CrossRef](#)]
98. Kohn, M.J.; Riciputi, L.R.; Stakes, D.; Orange, D.L. Sulfur isotope variability in biogenic pyrite; reflections of heterogeneous bacterial colonization? *Am. Mineral.* **1998**, *83*, 1454–1468. [[CrossRef](#)]
99. Borowski, W.S.; Rodriguez, N.M.; Paull, C.K.; Ussler, W., III. Are <sup>34</sup>S-enriched authigenic sulfide minerals a proxy for elevated methane flux and gas hydrates in the geologic record? *Mar. Pet. Geol.* **2013**, *43*, 381–395. [[CrossRef](#)]
100. Jørgensen, B.B.; Böttcher, M.E.; Lüschen, H.; Neretin, L.N.; Volkov, I.I. Anaerobic methane oxidation and a deep H<sub>2</sub>S sink generate isotopically heavy sulfides in Black Sea sediments 1. *Geochim. Cosmochim. Acta* **2004**, *68*, 2095–2118. [[CrossRef](#)]
101. Drake, H.; Hallbeck, L.; Pedersen, K.; Rosdahl, A.; Tullborg, E.-L.; Wallin, B.; Sandberg, B.; Blomfeldt, T. *Investigation of Sulphide Production in Core-Drilled Boreholes in Äspö Hard Rock Laboratory. Boreholes KA3110A, KA3385A and KA3105A*; Swedish Nuclear Fuel and Waste Management Co. (SKB): Stockholm, Sweden, 2013.
102. Tullborg, E.-L.; Smellie, J.; Nilsson, A.-C.; Gimeno, M.J.; Auqué, L.F.; Brüchert, V.; Molinero, J. *SR-Site—Sulphide Content in the Groundwater at Forsmark*; SKB Report TR-10-39; Swedish Nuclear Fuel and Waste Management Co. (SKB): Stockholm, Sweden, 2010.
103. Ivarsson, M.; Bengtson, S.; Drake, H.; Francis, W. Fungi in Deep Subsurface Environments. In *Advances in Applied Microbiology*; Academic Press: Cambridge, MA, USA, 2017.
104. Bengtson, S.; Rasmussen, B.; Ivarsson, M.; Muhling, J.; Broman, C.; Marone, F.; Stampanoni, M.; Bekker, A. Fungus-like mycelial fossils in 2.4-billion-year-old vesicular basalt. *Nat. Ecol. Evol.* **2017**, *1*, 0141. [[CrossRef](#)] [[PubMed](#)]
105. Ivarsson, M.; Holm, N.G.; Neubeck, A. The Deep Biosphere of the Subseafloor Igneous Crust. In *Trace Metal Biogeochemistry and Ecology of Deep-Sea Hydrothermal Vent Systems*; Demina, L.L., Galkin, V.S., Eds.; Springer International Publishing: Cham, Switzerland, 2016; pp. 143–166.
106. Drake, H.; Ivarsson, M. The role of anaerobic fungi in fundamental biogeochemical cycles in the deep biosphere. *Fungal Biol. Rev.* **2018**, *32*, 20–25. [[CrossRef](#)]
107. Ivarsson, M.; Schnürer, A.; Bengtson, S.; Neubeck, A. Anaerobic fungi: A potential source of biological H<sub>2</sub> in the oceanic crust. *Front. Microbiol.* **2016**, *7*, 674. [[CrossRef](#)] [[PubMed](#)]
108. Parkes, R.J.; Linnane, C.D.; Webster, G.; Sass, H.; Weightman, A.J.; Hornibrook, E.R.C.; Horsfield, B. Prokaryotes stimulate mineral H<sub>2</sub> formation for the deep biosphere and subsequent thermogenic activity. *Geology* **2011**, *39*, 219–222. [[CrossRef](#)]

