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1	Title: Life Tables of <i>Bactrocera cucurbitae</i> (Coquillett) (Diptera: Tephritidae):
2	with a Mathematical Invalidation for Applying the Jackknife Technique to the
3	Net Reproductive Rate
4	
5	Running title: Life table and jackknife technique
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17 Abstract

18	1.	Life table data for the melon fly, Bactrocera cucurbitae (Coquillett), reared on
19		cucumber (Cucumis sativus L.) were collected under laboratory and simulated
20		field conditions.

- Means and standard errors of life table parameters were estimated for two
 replicates using the jackknife technique.
- 23 3. At 25°C, the intrinsic rates of increase (r) found for the two replicates were 24 0.1354 and 0.1002 day⁻¹, and the net reproductive rates (R_0) were 206.3 and 25 66.0 offspring, respectively.
- 4. When the cucumbers kept under simulated field conditions were covered with
 leaves, the *r* and *R*₀ for the two replicates were 0.0935 and 0.0909 day⁻¹, 17.5
 and 11.4 offspring, respectively. However, when similar cucumbers were left
 uncovered, the *r* and *R*₀ for the two replicates were 0.1043 and 0.0904 day⁻¹,
 and 27.7 and 10.1 offspring, respectively.
- 5. Our results revealed that considerable variability between replicates in both
 laboratory and field conditions is possible; this variability should be taken into
 consideration in data collection and application of life tables.
- 6. Mathematical analysis has demonstrated that applying the jackknife technique results in unrealistic pseudo- R_0 and overestimation of its variance.
- 36 7. We suggest that the jackknife technique should not be used for the estimation 37 of variability of R_0 .
- 38

Key words. *Bactrocera cucurbitae*, *Cucumis sativus*, life table, net reproductive
rate, jackknife method.

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41 Introduction

42	The melon fly, Bactrocera cucurbitae (Coquillett) (Diptera: Tephritidae), has been one
43	of the most important pests in Taiwan (Huang & Chi, 2011), and in many other
44	regions in Asia (Koyama et al., 2004; Dhillon et al., 2005) for several decades.
45	Although the agricultural agencies have invested heavily in research, workshops, and
46	control measures related to the fly, it remains a major pest in Taiwan (Huang & Chi,
47	2011). For sustainable pest management in organic farming, it is crucial to develop a
48	comprehensive understanding of the population ecology of the target pests. Life
49	table studies should be the first priority in ecologically sound pest management
50	programs because only life tables can provide the most detailed and correct
51	descriptions of the survival, stage differentiation, and reproduction of populations.
52	Age-specific female life tables of <i>B. cucurbitae</i> were developed by Vargas <i>et al.</i> (1996,
53	1997, 2000) and Yang et al. (1994). However, the theories relating to female
54	age-specific life tables (Lewis, 1942; Leslie, 1945; Birch, 1948) address only female
55	populations and ignore male populations. Chi & Liu (1985) and Chi (1988)
56	observed that female age-specific life tables cannot correctly describe the growth and
57	stage differentiation of insect and mite populations. Thus, although numerous
58	female life tables have been published for many insect species, their practical
59	applications are quite limited. Huang & Chi (2011) reported the first age-stage,
60	two-sex life table for <i>B. cucurbitae</i> under laboratory conditions with cucumber slices
61	as the rearing medium. They demonstrated that an erroneous relationship is obtained
62	if an age-specific female life table is applied to a two-sex population. Furthermore,
63	they indicated that the study of life tables constructed under field conditions can be
64	helpful by revealing differences between the values of population parameters in the
65	field and in the laboratory.

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66 Liquido (1991) demonstrated that fallen fruits on the ground act as a reservoir for 67 melon fly populations. To construct precise predictions of the dynamics of 68 populations in the field, it is necessary to identify the differences between life tables 69 collected in the laboratory and those actual life tables under field conditions. On the 70 other hand, due to the tedious and time-consuming work of life table studies, most life 71 table studies are carried out by using single cohort without replication. To estimate 72 the means and variances of population parameters obtained from a single cohort, 73 jackknife technique is widely used. Meyer et al. (1986) used jackknife and bootstrap 74 techniques in estimating uncertainty in intrinsic rate and concluded that jackknife was 75 more cost-effective based on simulation. Efron & Tibshirani (1993) discussed the 76 failure of jackknife. Chi & Yang (2003) pointed out that application of jackknife 77 will result in some degree of discrepancy between the estimated means of population 78 parameters and their theoretical definition. When we use the jackknife method to 79 estimate the mean value of the net reproductive rate, we often obtain some pseudo- R_0 80 value of zero. An mathematical explanation is needed to justify or falsify the use of 81 jackknife technique. In this study, eggs of melon flies were artificially introduced 82 into whole cucumbers (Cucumis sativus L.), then kept at 25°C and under field with 83 replications. Life tables were constructed and the population parameters were 84 measured for replicates. Furthermore, we derived a mathematical proof to 85 demonstrate the problem of the jackknife method for the estimation of the mean and 86 standard error of the net reproductive rate.

- 87
- 88 Materials and methods

89 *Life Table Study*

90 Melon flies were collected in a field used to grow vegetables and subsequently

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91 reared on cucumber (Cucumis sativus L.). The colony was maintained in the 92 laboratory of the Department of Entomology, National Chung Hsing University 93 (Taichung, Taiwan) for two generations before the beginning of the life table study. 94 For the life table study, eggs laid within 24 h were collected using piled cucumber 95 slices following the method of Huang & Chi (2011). For implanting eggs into the 96 cucumber, a pyramid-shaped hole with a rectangular base (1.5 cm each side, 1.5 cm 97 height) was cut with an arrowhead-shaped knife. Twenty eggs were placed in the 98 hole with a fine writing brush. Before the pyramid-shaped cucumber piece was 99 replaced, its tip was removed to leave a space for the eggs. To study the cohort life 100 tables at 25° C, five cucumbers with eggs were kept in a plastic jar (26 cm height, 23 101 cm diameter) with loamy soil. The mouth of the jar was covered with fine mesh net 102 and kept at a constant temperature of 25°C in a growth chamber under a photoperiod 103 of 12:12 (L:D) h. To study the life table under field conditions, five cucumbers with 104 eggs were placed in a jar, kept in a shaded area and covered with dried mango leaves. 105 Another five cucumbers with eggs were placed in a jar and kept under direct sunlight 106 in the field with no leaf cover. The field study was conducted from 5 June to 9 107 September 2006. The average field temperature was 28.1°C. Two replicates were 108 used for each treatment. The numbers of emerged adults were observed, and pairs of 109 adults were formed. The eggs laid daily by the melon flies were collected on sliced 110 cucumber as described in Huang & Chi (2011).

111

112 Demographic Analysis

113 The life history data were analyzed according to the age-stage, two-sex life table 114 theory (Chi & Liu, 1985) and the method described by Chi (1988). The means and 115 standard errors of the life table parameters were estimated with the jackknife method

116 (Sokal & Rohlf, 1995). The population parameters estimated were the intrinsic rate 117 of increase (r), the finite rate of increase (λ), the gross reproductive rate (*GRR*), the 118 net reproductive rate (R_0) and the mean generation time (T). In this paper, the 119 intrinsic rate of increase is estimated with the iterative bisection method from the 120 Euler-Lotka formula

121
$$\sum_{x=0}^{\infty} e^{-r(x+1)} l_x m_x = 1$$
(1)

122 with age indexed from 0 (Goodman, 1982). The mean generation time is defined as the length of time that a population needs to increase to R_0 -fold of its size (i.e., e^{rT} = 123 R_0 or $\lambda^T = R_0$) at the stable age-stage distribution and is calculated as $T = (\ln R_0)/r$. 124 125 The age-stage life expectancy (e_{xi}) is calculated according to Chi & Su (2006). To 126 facilitate the tedious process of raw data analysis, a computer program 127 TWOSEX-MSChart for the age-stage, two-sex life table analysis (Chi, 2010) in 128 Visual BASIC (version 6, service pack 6) for the Windows system is available at 129 http://140.120.197.173/ Ecology/ (Chung Hsing University) and at 130 http://nhsbig.inhs.uiuc.edu.tw/www/chi.html (Illinois Natural History Survey). We 131 used a Tukey-Kramer procedure (Dunnett, 1980) to compare the difference among 132 treatments following the description of Sokal & Rohlf (1995).

133

134 **Results**

135 *Life Table of* B. cucurbitae

136 The developmental times for each stage are listed in Table 1. At 25 °C, the

- 137 duration of the preadult stage in whole cucumber was 17.8 and 18.5 d (two replicates).
- 138 This value was much greater than the corresponding value for growth in cucumber
- 139 kept under field conditions with or without leaf coverage. The adult

140 pre-ovipositional periods (APOP) in the different treatments ranged from 7.0 to 9.1 d. 141 There were no significant differences among these values. The total 142 pre-ovipositional period (TPOP) at 25°C was, however, significantly longer than those 143 found in the field. The adult longevities of both male and female adults at 25°C are 144 also longer than those observed under field conditions. The total fecundity varied 145 significantly among treatments (Table 2). Significantly higher fecundities (859 and 146 660 eggs/female) were observed in females reared at 25°C than in females emerged The high coefficients of variation (CV) of mean fecundities 147 under field conditions. 148 showed the high reproductive variability among individuals.

149 The detailed age-stage survival rates (s_{xi}) of *B. cucurbitae* for the different 150 treatments are plotted in Fig. 1. The parameter s_{xi} is the probability that a newborn 151 will survive to age x and stage j. The survival rate curves of B. cucurbitae cohorts 152 vary significantly between replicates for populations reared in whole cucumbers. In 153 general, the survival rate in the laboratory is higher than in the other treatments. At 154 25°C, cohorts in the laboratory survived longer than those in the field. This 155 difference is also evident from the longer developmental time of the preadult stage 156 and from the adult longevities (Table 1).

The daily mean number of offspring produced by individual B. cucurbitae of 157 158 age x and stage *j* per day is shown with the age-stage fecundity (f_{xi}) in Fig. 2. 159 Because only adult females produce offspring, there is only a single curve f_{x2} (i.e., the 160 adult female is the second life history stage). The age-specific survival rate (l_x) and 161 the age-specific fecundity (m_x) are also plotted in Fig. 2. The l_x curve describes the 162 change in the survival rate of the cohort with age. Significant variability can be observed between the two replicates. In one replicate at 25°C, more than 40% B. 163 164 *cucurbitae* survived to the adult stage, but the corresponding value in another

165	replicate was much smaller, approximately 20%.	However, at 25°C, the survival
166	rates in the laboratory are higher than those in the	field (Fig. 2).

167

168 Population Parameters

The means and standard errors of population parameters of B. cucurbitae in the 169 170 different treatments investigated are listed in Table 2. For the eggs artificially placed in cucumber and kept at 25°C, the intrinsic rates of increase (r) found for the 171 two replicates were 0.1354 and 0.1002 day⁻¹, the net reproductive rates (R_0) were 172 173 206.3 and 66.0 offspring, and the mean generation times (T) were 39.5 and 42.6 days, 174 respectively. For the cucumbers kept in the field and covered with leaves, the population parameters (r, R_0 and T) were 0.0935 and 0.0909 day⁻¹, 17.5 and 11.4 175 176 offspring, and 34.0 and 35.0 days, respectively. However, for the cucumbers kept in the field without leaves, the population parameters $(r, R_0 \text{ and } T)$ were 0.1043 and 177 0.0904 day⁻¹, 27.7 and 10.1 offspring, and 32.8 and 27.2 days, respectively. The 178 maximum intrinsic rate of increase (0.1354 d⁻¹) was obtained at 25°C in the 179 180 laboratory. All parameters have very high values of CV. The age-stage specific life expectancy (e_{xi}) (Fig. 3) is the lifespan remaining for 181 182 an individual of age x and stage *j*. The contribution of an individual of age x and stage *i* to the future population is described by the age-stage reproductive value (v_{xi}) 183 184 (Fig. 4). The reproductive value of a newborn (v_{01}) is exactly equal to the finite rate 185 of increase.

186

187 Discussion

188 Life Table of B. cucurbitae

189 The shorter preadult stage in the treatment under field conditions with leaf

190	coverage might be due to the higher temperature and the higher humidity. These					
191	conditions can promote the decay of cucumber and thereby generate conditions					
192	favorable for flies. Vayssières et al. (2008) reported that the total preadult					
193	development time of <i>B. cucurbitae</i> on cucumber at 25 and 30°C was 17.2 and 13.2					
194	days, respectively. Huang & Chi (2011) reported that the total preadult development					
195	time of <i>B. cucurbitae</i> was 15.1 days at 25°C. These studies show that the preadult					
196	development time of <i>B. cucurbitae</i> decreases as the temperature increases. Under					
197	field conditions, melon flies in different fallen fruits may experience different					
198	micro-environments and may result in higher variations in developmental rate,					
199	survival and reproduction.					
200	Because the variable developmental rate among individuals is incorporated in					
201	the age-stage, two-sex life table, the overlap between stages can be observed in Fig. 1.					
202	If the survival curves were constructed based on the means of each stage or adult age					
203	(e.g., Marcic, 2003, 2005; Legaspi, 2004; Legaspi & Legaspi, 2005; Lin & Ren, 2005;					
204	Liu, 2005; Kivan & Kilic, 2006; Kontodimas & Stathas, 2005; Tsoukanas et al.,					
205	2006), the stage overlap would not have been observed and would have resulted in					
206	errors in the survival curves as well as the fecundity curves. Liu (2005) noticed the					
207	overlap of the stages of Delphastus catalinae (Coleoptera: Coccinellidae).					
208	Nevertheless, he ignored the variable developmental rate and constructed					
209	age-specific fecundity schedules based on adult age. Yu et al. (2005) and Chi & Su					
210	(2006) gave detailed explanations and a mathematical proof to address the errors in					
211	life tables based on adult age.					
212	In Vargas <i>et al.</i> (1997), the fecundity of <i>B. cucurbitae</i> at 24°C was 578.6 eggs.					
213	In Huang & Chi (2011), the mean fecundity of melon flies reared on cucumber at					
214	25°C was 341 eggs. Jiang et al. (2006) reported that the mean fecundity of melon					

215 flies reared on cucumber at 30°C was 895.65 eggs. In this study, the mean 216 fecundity of *B. cucurbitae* reared on whole cucumber at 25°C was higher than the 217 fecundity given in Huang & Chi (2011). If the survival rate and fecundity are 218 constructed based solely on the adult age, the differences in preadult development are 219 ignored, and it is assumed that all adults emerge on the same day. These artificial 220 manipulations and assumptions will not only falsely diminish the real variability 221 among individuals, but also consequently result in errors in the survival and 222 fecundity curves (Chi, 1988; Yu et al., 2005; Chi & Su, 2006; Huang & Chi, 2011). 223

224 Population Parameters

225 Due to the problems associated with the female age-specific life table (Huang & 226 Chi, 2011), we used the age-stage, two-sex life table to calculate the population 227 parameters of *B. cucurbitae*. The intrinsic rate of increase (*r*) ranged from 0.0904 to 0.1354 days⁻¹. The treatments did not differ significantly based on the estimated 228 means and standard errors obtained by using the jackknife technique and 229 230 Tukey-Kramer procedure. The net reproductive rate (R_0) of melon flies reared in 231 the laboratory at 25°C was higher than the corresponding rate under field conditions. 232 The relationship between the net reproductive rate R_0 and the mean female fecundity F was given by Chi (1988) for the two-sex life table as 233

$$R_0 = F \cdot \left(\frac{N_f}{N}\right) \tag{2}$$

where *N* is the total number of eggs used for the life table study at the beginning and N_f is the number of female adults emerged. Yu *et al.* (2005) gave the relationship among the gross reproductive rate (*GRR*), the net reproductive rate (*R*₀) and the preadult survivorship (*l_a*) as

 $GRR > l_a \cdot GRR > R_0 \tag{3}$

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All of our results for *B. cucurbitae* at different treatments are consistent with the relationships given by equations 2 and 3. If a life table is constructed based on adult age and ignores the preadult mortality, an erroneous relationship between the mean fecundity and the net reproductive rate will be obtained. Yu *et al.* (2005) and Chi & Su (2006) discussed this problem in detail.

245 The shorter preoviposition period will cause a higher intrinsic rate of increase if 246 fecundity remains the same (Lewontin, 1965). In the study of Huang & Chi (2011), 247 the TPOP of B. cucurbitae reared on cucumber at 25°C was 23.1 d. In our study, the 248 TPOP, i.e., the duration from egg to first oviposition, of melon flies reared in the 249 laboratory at 25°C was longer than that under field conditions. This result might be 250 explained by the higher field temperature (28°C) and humidity. At 25°C, the 251 age-stage life expectancy gradually decreases with age because no other adverse 252 effects occur in the laboratory. Under field conditions, however, the life expectancies were lower and varied significantly due to the variable abiotic factors. 253 254 The life expectancy is calculated using the age-stage specific survival rate (s_{xi}) 255 without assuming that the population reaches the stable age-stage distribution (Chi & 256 Thus, it can be used to predict the survival of a population under those Su, 2006). 257 conditions. For example, at 25°C both newly emerged female and male adults can 258 be expected to remain alive, on average, more than two months. The life 259 expectancy based on the age-stage, two-sex life table reveals the difference among 260 individuals of the same age but of different stages or different sexes. Chi (1988), 261 Chi & Yang (2003) and Chi & Su (2006) discussed in detail the differences between 262 the traditional female age-specific life table and the age-stage, two-sex life table and identified possible errors in the survival and fecundity curves based on the adult age. 263 264 Fisher (1930) defined the reproductive value as the contribution of an individual

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265 to the future population. The reproductive value significantly increases at the time 266 of emergence of the adult females. For example, when a female adult emerges at 267 age 15 d at 25°C (Fig. 1), the reproductive value increases from a value of less than 268 10 for a nymph to 36 for a female (Fig. 4). The contribution of males to the future 269 population is not defined by Fisher (1930), and there is no curve for males. 270 The research reported here demonstrates that only life table study can 271 completely depict the development, stage differentiation, and reproduction of B. 272 *cucurbitae* and the variability of these processes in whole cucumber. Moreover, it 273 revealed significant differences between life tables collected in the laboratory and the 274 Thus, computer simulations of the growth of field populations should field. 275 incorporate considerations of these differences. Chi (1990) noted that a simulation 276 based on the age-stage, two-sex life table can be used to time pest management by 277 taking the stage-specific susceptibility to pesticide applications into consideration. 278 Chi & Getz (1988) constructed a mass-rearing model based on the age-stage, two-sex 279 life table. For an ecology-oriented integrated pest management of *B. cucurbitae*, 280 life tables collected under different conditions should play important roles in the 281 However, because a variety of wild cucurbits serve as a host for the melon future. fly and form a reservoir for this fly (Uchida et al., 1990), it might be necessary to 282 283 understand the life table of the fly on the major wild cucurbits.

284

285 Using the Jackknife Method to Estimate of the Net Reproductive Rate

Our results showed high values of CV in female mean fecundity and population parameters. The high CV in mean fecundity is calculated by using basic descriptive statistical method and they reflect the differences among female individuals. The high CVs of population parameters are, however, estimated by using the jackknife 290 technique. The jackknife technique is a resampling method which is usually used 291 when replication is impossible or difficult. Because life table studies are time- and labor-consuming, replication is in general impractical in most cases. The jackknife 292 293 method is thus used to estimate the means and standard errors of population 294 parameters (Chi & Getz, 1988; Maia et al., 2000; Huang & Chi, 2011). In the 295 jackknife method, we first use data on all individuals (n) to calculate the intrinsic rate 296 of increase of the whole cohort (r_{all}) . We then calculate the intrinsic rate r_i by 297 omitting individual *i*. The pseudo-value $r_{i-pseudo}$ is then calculated as:

298
$$r_{i-pseudo} = n \cdot r_{all} - (n-1)r_i \tag{4}$$

where *n* is the total number of individuals used at the beginning of the life table study. The mean value of all $r_{i-pseudo}$ is the estimated mean value of the intrinsic rate of increase of the cohort:

$$r = \frac{\sum_{i=1}^{n} r_{i-pseudo}}{n}$$
(5)

303 Similarly, if we use the jackknife method to calculate the mean value of the net 304 reproductive rate, we first use data on all individuals in the cohort to calculate $R_{0,all}$:

305
$$R_{0,all} = \sum_{x=0}^{\infty} l_x m_x$$
. (6)

306 If the total number of eggs laid by all surviving individuals at age x is F_x , the total 307 eggs laid by the whole cohort from birth to death is F_{total} and can be calculated as 308 $\sum_{x=0}^{\infty} F_x$. Then, the $R_{0,all}$ can also be calculated as

309
$$R_{0,all} = \sum_{x=0}^{\infty} l_x m_x = \sum_{x=0}^{\infty} \frac{n_x}{n} \cdot \frac{F_x}{n_x} = \sum_{x=0}^{\infty} \frac{F_x}{n} = \frac{1}{n} \sum_{x=0}^{\infty} F_x = \frac{F_{total}}{n}$$
(7)

310 where n_x is the number of surviving individuals at age x. Equation 7 shows that the 311 net reproductive rate is F_{total} divided by the total number of individuals n used at the beginning of the life table study. If the omitted individual *i* is type N (those dying at immature stages) or M (male), we define the total eggs laid by *n*-1 individuals at age $x ext{ as } F_{x,i}$. It is clear that $F_{x,i} = F_x$ for all ages, because types N and M do not lay eggs.

315 The net reproductive rate with individual *i* omitted, i.e., $R_{0,i}$, can be calculated as

316
$$R_{0,i} = \sum_{x=0}^{\infty} \frac{n_{x,i}}{n-1} \cdot \frac{F_{x,i}}{n_{x,i}} = \sum_{x=0}^{\infty} \frac{F_{x,i}}{n-1} = \sum_{x=0}^{\infty} \frac{F_x}{n-1} = \frac{1}{n-1} \sum_{x=0}^{\infty} F_x$$
(8)

317 where $n_{x,i}$ is the number of surviving individuals at age x if individual *i* is omitted.

318 The pseudo-value for the omission of individual *i* is calculated analogously to

319 Equation 4:

320
$$R_{0,i-pseudo} = n \cdot R_{0,all} - (n-1) \cdot R_{0,i}$$
(9)

321 Replacing $R_{0,i}$ according to the proofs of Equation 7 and 8, we find

322
$$R_{0,i-pseudo} = n \left(\frac{1}{n} \sum_{x=0}^{\infty} F_x \right) - \left(n - 1 \right) \left(\frac{1}{n-1} \sum_{x=0}^{\infty} F_x \right)$$
(10)

323 Consequently, we obtain

324
$$R_{0,i-pseudo} = \sum_{x=0}^{\infty} F_x - \sum_{x=0}^{\infty} F_x = 0$$
(11)

325 Thus, we prove that if the omitted individual *i* is type N or M, the pseudo-value

326 $R_{0,i-pseudo}$ will always be zero.

327 If the omitted individual *i* is a female and can produce $b_{x,i}$ eggs at age *x*, the total

328 number of eggs laid by this female during its life span can be calculated as

329
$$B_i = \sum_{x=0}^{\infty} b_{x,i}$$
 (12)

330 If individual *i* is omitted, then the total eggs produced by the remaining individuals in

331 cohort at age x is $F_{x,t}$. It is clear that

332
$$F_{x,i} = F_x - b_{x,i}$$
 or $F_x = F_{x,i} + b_{x,i}$ (13)

333 According to Equation 8, we have

334
$$R_{0,i} = \frac{1}{n-1} \sum_{x=0}^{\infty} F_{x,i} = \frac{1}{n-1} \sum_{x=0}^{\infty} \left(F_x - b_{x,i} \right)$$
(14)

335 The pseudo-value for the omission of individual *i* is

336
$$R_{0,i-pseudo} = n \cdot R_{0,all} - (n-1) \cdot R_{0,i}$$
(15)

Replacing $R_{0,i}$ of Equation 15 with its value in Equation 14, we can simplify

338 Equation 15 to 16.

339
$$R_{0,i-pseudo} = n \cdot \left(\frac{1}{n} \sum_{x=0}^{\infty} F_x\right) - (n-1) \left[\frac{1}{n-1} \sum_{x=0}^{\infty} (F_x - b_{x,i})\right]$$

340
$$R_{0,i-pseudo} = \sum_{x=0}^{\infty} F_x - \sum_{x=0}^{\infty} F_x + \sum_{x=0}^{\infty} b_{x,i} = \sum_{x=0}^{\infty} b_{x,i} = B_i$$
(16)

341 It is clear that if the omitted individual *i* is a female, the pseudo-value of the net
342 reproductive rate is exactly the total fecundity of individual *i* itself,

343
$$R_{0,i-pseudo} = \sum_{x=0}^{\infty} b_{x,i} = B_i$$
(17)

This analysis shows that if the jackknife method is used, the pseudo-value of the net reproductive rate obtained by omitting individual *i* is exactly the total number of eggs laid by individual *i*. It is exactly the fecundity of individual *i*. The mean of all pseudo-values is the total number of eggs laid by all individuals divided by *n*:

348
$$\hat{R}_{0} = \frac{\sum_{i=1}^{n} R_{0,i-pseudo}}{n} = \frac{\sum_{i=1}^{n} B_{i}}{n}$$
(18)

349 By definition, it is clear that
$$\sum_{i=1}^{n} B_i = \sum_{x=0}^{\infty} F_x$$
.

350 The mean of all $R_{0,i-pseudo}$ is then

351
$$\hat{R}_{0} = \frac{\sum_{i=1}^{n} B_{i}}{n} = \frac{\sum_{i=0}^{\infty} F}{n} = R_{0,all}.$$
 (19)

352 The above proof can be concluded by making the following four observations: 1)

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the mean value of the net reproductive rate estimated with the jackknife method is exactly the same as the $R_{0,all}$ without the use of the jackknife method; 2) the net reproductive rate equals the total eggs of the cohort divided by n, i.e., the total number of newborns used for the life table study; 3) if the omitted individual is one of the males or one of those that died at an immature stage, the pseudo-value is zero; and 4) if the omitted individual is female, the pseudo-value is the fecundity of that omitted female.

360 In Fig. 5, the frequency distributions of pseudo- R_0 values of three treatments 361 showed the zeros obtained by using the jackknife technique. It is clear that the 362 omission of a single individual of type N or M will generate a pseudo- R_0 of zero. 363 The higher the preadult mortality or proportion of male, the higher the zero 364 pseudo- R_0 bar. Because there is generally preadult mortality, the bar of zero 365 pseudo- R_0 will be an important factor determining the frequency distribution of all 366 This is also the reason why statistical software shows the pseudo- R_0 life table data. 367 failed the normality test and instead suggests Mann-Whitney Rank Sum test or others. The omission of a single individual of type N or M caused the pseudo- R_0 of the 368 369 resampled population to zero. If we carry out a true replication of life table study as 370 we did in this study, however, we will generally not get a population with zero net 371 reproductive rate, i.e., all individuals are either type N or M. This shows the 372 jackknife technique will generate biologically unrealistic pseudo- R_0 , which results in 373 an overestimation of variances and standard errors of the net reproductive rates. 374 The overestimation of variances and standard errors consequently make significant 375 differences between treatments undetectable by using statistical tests. 376 Variance analysis is important for revealing the variability of experimental

The question of the suitability of the jackknife method for the estimation of

377

results.

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378 the mean and standard errors of the net reproductive rate is not the only difficulty 379 associated with life table analysis. The sample size must be sufficiently large to prevent inaccurate estimation of the standard errors. Because there are many 380 381 problems associated with female life tables and analyses based on adult age (Chi & 382 Liu, 1985; Chi, 1988; Yu et al., 2005; Chi & Su, 2006; Huang & Chi, 2011), the 383 application of the jackknife method to female life tables (Leslie, 1945; Birch, 1948; 384 Maia et al., 2000) or in analyses based on female population and adult age (Maia et 385 al., 2000) will not produce correct estimates.

386 The significant differences between replicates in this study showed, however, 387 that the variability in developmental rate, survival, and reproduction of a life table 388 could not be properly described and estimated with the jackknife method. For this 389 reason and many others, the prediction of population dynamics under field conditions 390 is difficult. In this paper, we limit our discussion to the application of jackknife 391 method to the net reproductive rate. There are other resampling methods, e.g., 392 bootstrapping, permutation test, cross validation, etc. Similar analysis is required to 393 re-evaluate their application in the estimation of means and variances of population 394 parameters. Despite these difficulties and problems, the life table is the only solid 395 theory which can correctly describe the survival, stage differentiation, and 396 reproduction in detail. The necessity and the difficulties associated with life table 397 study demonstrate that we need to draw the attention of scientists to life table theory 398 and data analysis in insect ecology, integrated pest management, as well as biological 399 control.

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521

522 Table 1. Means and standard errors of the developmental time, longevity, adult preoviposition period (APOP) and total preoviposition period

523 (TPOP) of *Bactrocera cucurbitae* for different treatments

	Stage	25°C		Field conditions			
Parameter				Without leaf coverage		With leaf coverage	
		Rep. 1	Rep. 2	Rep. 1	Rep. 2	Rep. 1	Rep. 2
Developmental time (days)	Preadult	17.8 ± 0.2 a	$18.5\pm0.2~\text{b}$	11.4 ± 0.2 c	$11.4 \pm 0.1 \text{ c}$	11.0 ± 0.0 c	11.2 ± 0.2 c
Adult longevity	Male	74.8 ± 7.1 a	63.2 ± 9.5 a	$33.6 \pm 13.1 \text{ b}$	$34.7\pm11.8~\text{b}$	$63.9 \pm 9.8 a$	$13.2\pm8.7~b$
(days)	Female	58.9 ± 6.5 a	45.6 ± 12.0 a	55.4 ± 11.6 a	$16.3\pm4.7~b$	28.5 ± 8.9 a	22.6 ± 14.5 a
APOP (days)	Female	8.7 ± 0.3 a	8.9 ± 0.6 a	9.1 ± 0.3 a	8.6 ± 0.4 a	$9.0 \pm 0.6 a$	7.0 ± 2.0 a
TPOP (days)	Female	26.7 ± 0.3 a	$28.0\pm0.8\;b$	$20.7\pm0.3\ c$	$20.0\pm0.3~\mathrm{c}$	$20.0\pm0.6\ c$	18.5 ± 1.5 c

524 Means in the same row followed by the same letter are not significantly different (P > 0.05) using the Tukey-Kramer procedure.

525 Table 2. Means, standard errors, and coefficients of variation (CV) (in parentheses) of the population parameters of *Bactrocera cucurbitae* for

526 different treatments

	25°C		Field conditions				
Population parameters			Without lea	af coverage	With leaf coverage		
	Rep. 1	Rep. 2	Rep. 1	Rep. 2	Rep. 1	Rep. 2	
Mean fecundity (F)	859.5 ± 107.8 a	660.1 ± 179.9 a	345.9 ± 92.5 b	112.1 ± 42.8 b	218.5 ± 115.5 b	227.6 ± 162.3 b	
(eggs/female)	(61.4%)	(86.2%)	(75.6%)	(114.4%)	(149.5%)	(159.5%)	
The intrinsic rate of increase r (days ⁻¹)	0.1354 ± 0.0060 a	0.1002 ± 0.0116 a	0.1043 ± 0.0151 a	0.0904 ± 0.0197 a	0.0935 ± 0.0120 a	0.0909 ± 0.0380 a	
	(44.0%)	(116.2%)	(145.2%)	(217.3%)	(224.3%)	(417.3%)	
The finite rate of increase λ (days ⁻¹)	1.145 ± 0.007 a	1.105 ± 0.013 a	1.110 ± 0.017 a	1.094 ± 0.021 a	1.098 ± 0.023 a	1.094 ± 0.040 a	
	(6%)	(11.6%)	(15.1%)	(19.5%)	(20.7%)	(36.9%)	
Gross reproductive rate (GRR) (offspring)	636.3 ± 129.6 a	426.5 ± 159.5 a	322.4 ± 120.0 a	119.3 ± 56.4 a	146.5 ± 94.7 a	868.61 ± 484.84 a	
	(203.6%)	(374.6%)	(372.1%)	(473.0%)	(646.3%)	(558.2%)	
The net reproductive rate R ₀ (offspring/individual)	206.3 ± 44.8 a (217.3%)	66.0 ± 26.3 b (398.0%)	27.7 ± 11.7 b (423.5%)	10.1 ± 4.9 b (482.4%)	17.5 ± 10.5 b (602.5%)	11.4 ± 8.8 b (776.4%)	
The mean generation time T (days)	39.5 ± 0.8 a	42.6 ± 1.5 a	32.8 ± 1.5 a	27.2 ± 2.3 b	34.0 ± 3.9 a	35.0 ± 7.4 a	
	(19.1%)	(34.3%)	(45.7%)	(83.2%)	(113.4%)	(212.2%)	

527 Means in the same row followed by the same letter are not significantly different (P > 0.05) using the Tukey-Kramer procedure.

528 Figure captions

529 Fig. 1. Age-stage specific survival rate (s_{xj}) of *Bactrocera cucurbitae* for 530 different treatments.

Fig. 2. Age-specific survival rate (l_x) , female age-specific fecundity (f_{x2}) , age-specific fecundity (m_x) and age-specific maternity (l_xm_x) of *Bactrocera cucurbitae* for different treatments.

Fig. 3. Age-stage specific life expectancy (e_{xj}) of *Bactrocera cucurbitae* for different treatments.

Fig. 4. Age-stage specific reproductive value (v_{xj}) of *Bactrocera cucurbitae* for different treatments.

538 Fig. 5. Frequency distribution of pseudo- R_0 grouped for different treatments.

Each bar represents the number of pseudo- R_0 between two ticks. The bar at zero

540 represents the frequency of pseudo- R_0 zero.





543 544

Fig. 1. Age-stage specific survival rate (s_{xj}) of *Bactrocera cucurbitae* for

545 different treatments.

546





age-specific fecundity (m_x) and age-specific maternity $(l_x m_x)$ of *Bactrocera cucurbitae* for different treatments.

548



554 Fig. 3. Age-stage specific life expectancy (e_{xj}) of *Bactrocera cucurbitae* for



557



559 Fig. 4. Age-stage specific reproductive value (v_{xj}) of *Bactrocera cucurbitae* for

560 different treatments.

558



561 Fig. 5. Frequency distribution of pseudo- R_0 grouped for different treatments. 563 Each bar represents the number of pseudo- R_0 between two ticks. The bar at zero

564 represents the frequency of pseudo- R_0 zero.