Bladder Morbidity and Hepatic Fibrosis in Mixed Schistosoma haematobium and S. mansoni Infections: A Population-Wide Study in Northern Senegal

Lynn Meurs^{1*}, Moustapha Mbow², Kim Vereecken¹, Joris Menten¹, Souleymane Mboup², Katja Polman¹

1 Institute of Tropical Medicine, Antwerp, Belgium, 2 Laboratory of Bacteriology and Virology, Centre Hospitalier Universitaire Aristide Le Dantec, Dakar, Senegal

Abstract

Background: The global distribution map of schistosomiasis shows a large overlap of *Schistosoma haematobium*- and *S. mansoni*-endemic areas in Africa. Yet, little is known about the consequences of mixed *Schistosoma* infections for the human host. A recent study in two neighboring co-endemic communities in Senegal indicated that infection intensities of both species were higher in mixed than in single infections. Here, we investigated the relationship between mixed *Schistosoma* infections and morbidity in the same population. So far, this has only been studied in children.

Methods: Schistosoma infection was assessed by microscopy. *Schistosoma*-specific morbidity was assessed by ultrasound according to WHO guidelines. Multivariable logistic regression models were used to identify independent risk factors for morbidity.

Principal Findings: Complete parasitological and morbidity data were obtained from 403 individuals. Schistosoma haematobium-specific bladder morbidity was observed in 83% and S. mansoni-specific hepatic fibrosis in 27% of the participants. Bladder morbidity was positively associated with S. haematobium infection intensity (OR = 1.9 (95% CI 1.3–2.9) for a 10-fold increase in intensity). Moreover, people with mixed infections tended to have less bladder morbidity than those with single S. haematobium infections (OR = 0.3 (95% CI 0.1–1.1)). This effect appeared to be related to ectopic S. mansoni egg elimination in urine. Hepatic fibrosis on the other hand was not related to S. mansoni infection intensity (OR = 0.9 (95% CI 0.6–1.3)), nor to mixed infections (OR = 1.0 (95% CI 0.7–1.7)).

Conclusions/Significance: This is the first population-wide study on the relationship between mixed *Schistosoma* infections and morbidity. Mixed infections did not increase the risk of *S. mansoni*-associated morbidity. They even tended to reduce the risk of *S. haematobium*-associated morbidity, suggesting a protective effect of *S. mansoni* infection on bladder morbidity. These unexpected results may have important consequences for schistosomiasis control in co-endemic areas and warrant further investigation.

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* E-mail: LMeurs@itg.be

Introduction

Worldwide more than 207 million people are infected with *Schistosoma*, 85% of whom live in Africa [1]. Due to the large overlap in *Schistosoma haematobium*- and *S. mansoni*-endemic areas [2–4] many people are at risk of co-infection. Yet, little is known about mixed *Schistosoma* infections and their impact on host morbidity.

Clinical manifestations of schistosomiasis are associated with the species-specific oviposition sites [2]. Both *S. haematobium* and *S. mansoni* mature in the portal vein of the human host and form male-female pairs. Subsequently, the *S. mansoni* male carries the female to the mesenteric plexus whereas the *S. haematobium* couple continues its way to the veins of the pelvis. In these respective sites, they lay eggs, which are eventually eliminated from the body via the urine (*S. haematobium*) or feces (*S. mansoni*). About half of the eggs, however, are

carried away with the blood stream and/or trapped in the tissues. These retained eggs provoke inflammatory and granulomatous immune responses [2]. For *S. haematobium*, this can lead to inflammation, ulceration and pseudopolyposis of bladder and ureteral walls, and in children this is often accompanied by hematuria. Chronic lesions may evolve to fibrotic and sandy patches with severe sequelae such as hydroureter, hydronephrosis, and squamous bladder cancer [2,5,6]. For *S. mansoni*, egg deposition can lead to inflammatory hepatic schistosomiasis and hepatosplenomegaly in children and adolescents [7]. Trapped schistosomal eggs are gradually replaced by fibrotic deposits, and give rise to chronic hepatic schistosomiasis [2].

In mixed *S. mansoni* and *S. haematobium* infections, the abovedescribed processes act in parallel, which may result in more severe or other abnormalities than in single infections. So far, only

Author Summary

In the developing world, over 207 million people are infected with parasitic Schistosoma worms. Schistosoma haematobium and S. mansoni are the most abundant species in Africa and many people carry both. Yet, little is known about the consequences of such mixed infections. In general, S. haematobium affects the urinary tract of the host and S. mansoni the liver. Here, we investigated the effect of mixed Schistosoma infection on these health problems. We examined 403 people from northern Senegal for Schistosoma infections as well as for abnormalities of the urinary bladder and liver. Recently, we observed that people with mixed Schistosoma infections had generally higher infection intensities than those with single infections. The present study showed that abnormalities of the urinary bladder were more common in heavy than in light S. haematobium infections. Also, they were more common in single S. haematobium than in mixed infections. So far, only two studies have looked into the relationship between mixed Schistosoma infection and abnormalities of the bladder and liver, but only investigated children. Our findings suggest a possible protective effect of S. mansoni on bladder disease, in children as well as in adults. This may have important consequences for schistosomiasis control in co-endemic areas.

two studies have investigated the relationship between mixed infections and host morbidity. In Zimbabwe, a positive association between *S. mansoni* egg output and liver size was found in the presence but not in the absence of *S. haematobium*. Yet, this effect was very little [8]. In a study in Mali, mixed infection was associated with reduced hepatic morbidity on one hand and increased bladder morbidity on the other [9]. Both studies were performed in schoolchildren, in whom severe hepatic schistosomiasis is unlikely to have developed already [2,7,10]. Communitywide studies would therefore be more appropriate to investigate mixed infections and morbidity.

In northern Senegal, many communities have in the past decades become co-endemic for *S. mansoni* and *S. haematobium* [11–16]. *Schistosoma mansoni* was introduced in Richard-Toll in 1988 upon construction of the Diama dam and rapidly spread throughout the region [17,18]. By 1994, virtually the entire Guiers Lake ('Lac de Guiers') area had become exposed to this species [19]. Today, both *S. mansoni* and *S. haematobium* are widely spread, resulting in a large number of people with mixed infections in the communities around the lake.

Recently, we reported on the distribution of and risk factors for mixed *Schistosoma* infections in two communities on the banks of Lake Guiers. Individuals with mixed infections were found to have higher infection intensities than those with single infections [20]. In the present study, we set out to investigate the relationship between mixed *Schistosoma* infections and morbidity in the same communities. We studied the patterns of *S. haematobium*-specific bladder morbidity and *S. mansoni*-specific hepatic morbidity in this co-endemic focus, and compared morbidity in people with mixed *Schistosoma* infections to those with single infections.

Methods

Ethics Statement

This study was part of a larger investigation on the epidemiology of schistosomiasis and innate immune responses (SCHIS-TOINIR: www.york.ac.uk/res/schistoinir) for which approval was obtained from the review board of the Institute of Tropical Medicine, the ethical committee of the Antwerp University Hospital and 'Le Comité National d'Ethique de la Recherche en Santé' in Dakar. Informed and written consent was obtained from all participants prior to inclusion into the study.

Study Area

This study was conducted from July until November 2009 in Ndieumeul (also known as Thiekène) and Diokhor Tack, two neighboring communities on the Nouk Pomo peninsula in Lake Guiers. Details on the study area have been described elsewhere [20].

Parasitology and Urine Dipstick

Two urine and two feces samples were collected from each participant on consecutive days for microscopic analysis [21,22]. Per feces sample, two Kato-Katz slides of 25 mg fecal material each were prepared and microscopically examined for Schistosoma species [22]. Schistosoma mansoni infection intensity was expressed as the number of eggs detected per gram of feces (epg). Urine filtration was performed using a filter of 12 µm pore-size (Isopore) according to standard procedures [21]. Schistosoma haematobium infection intensity was expressed as the number of eggs detected per 10 ml of urine (ep10ml). Ectopic eggs were measured qualitatively (positive/negative). Ectopic egg elimination refers to elimination of schistosomal eggs via the unusual route - i.e. S. mansoni eggs in urine or S. haematobium eggs in feces. Single infection was defined as passing eggs of only one species, and mixed infection as passing eggs of both S. mansoni and S. haematobium, regardless of the route of egg elimination [20]. Microhematuria was determined in a subsample using Combur 7 dipsticks (Roche) on the first urine sample. All community members were offered praziquantel (one dose of 40 mg/kg body weight) and mebendazole (one dose of 500 mg) treatment after the study according to WHO guidelines [23].

Ultrasound

Participants were examined using a portable ultrasonography device with convex transducer. Pathologic lesions associated with S. haematobium or S. mansoni infection were recorded according to the Niamey guidelines [10]. All examinations were performed by the same clinician who was blind to the participant's infection status. Participants with severe pathology that needed further treatment were referred to the appropriate health authority. For S. haematobium-specific morbidity, the urinary bladder score was determined [10]. A score of ≥ 1 was considered as S. haematobiumspecific urinary bladder morbidity in accordance with previous studies [9,24-30]. Individuals with a score of 0 were categorized as controls. For S. mansoni-specific morbidity, the liver image pattern was determined [10]. Additional measurement of periportal thickening was not included as this approach has been shown to be not reproducible [31]. Liver image patterns of C to F were categorized as S. mansoni-specific hepatic morbidity [31-35]. Individuals with liver image pattern A or B are not likely to have periportal fibrosis [10] and were therefore categorized as controls. Individuals with signs of hepatic morbidity that were not specific for S. mansoni (e.g. hepatitis, cirrhosis or fatty liver) were excluded [10].

Statistical Analysis

IBM SPSS 19.0 (SPSS, Inc.) was used for statistical analysis. Results were considered significant when the *p*-value was <0.05. As egg outputs showed skewed distributions, data were normalized

Table 1. Schistosoma haematobium-associated bladder morbidity, hematuria and S. haematobium infection in the two co-endemic communities studied.

Morbidity	Urinary bladder score	n	(%)	Microhematuria (%)	S. haematobium infection	
					%	GM (95%CI) ^a
Negative	0	69	(17)	21	33	3.9 (1.8–8.1) ep10ml
Positive	≥1	334	(83)	44	58	8.8 (6.7–11.5) ep10ml
	1	15	(4)	62	73	8.5 (2.2–31.5) ep10ml
	2	301	(75)	42	56	8.5 (6.4–11.3) ep10ml
	≥3	18	(4)	71	78	13.1 (4.2–39.5) ep10ml
Total		403	(100)	40	54	8.1 (6.2–10.4) ep10ml

^aGM) Geometric Mean; calculated for microscopically *S. haematobium*-positive individuals only. doi:10.1371/journal.pntd.0001829.t001

by log (base 10)-transformation after adding half of the detection limit to allow for zeros.

Differences between groups were determined by the Pearson Chisquare test for community and gender, and by the Mann-Whitney U test for age. Furthermore, the Pearson Chi-square test was used to determine the association between bladder morbidity and microhematuria, as well as between bladder and liver morbidity.

Because of the non-linear trend of morbidity over age, the population was divided into four age groups (0-9, 10-19, 20-39) and ≥ 40 years) for multivariable regression analysis. Multivariable logistic regression models were used to identify independent risk factors for *S. haematobium*-specific bladder morbidity and *S. mansoni*-specific hepatic fibrosis, respectively. Age, gender, community of residence, *S. haematobium* infection intensity and *S. mansoni* infection intensity were included as potential risk factors. Moreover, significant interaction terms with age (p < 0.05) were added.

Similar models were used to assess the independent effect of mixed infection (as compared to single infection) on *S. haematobium*-specific bladder morbidity and *S. mansoni*-specific hepatic fibrosis, respectively. Among *S. haematobium*-positive subjects, the association between bladder morbidity and mixed infection was investigated using a dummy variable for mixed infection (1 = mixed, 0 = single), and age, gender, community of residence and *S. haematobium* infection intensity as other determinants. Likewise, the association between hepatic fibrosis and mixed infection was investigated in *S. mansoni*-positive subjects with a dummy variable for mixed infection for age, gender, community of residence and *S. mansoni* infection intensity.

Results

Complete parasitological data were obtained from 857 individuals [20]. Ultrasound data were collected from a random subsample of 403 individuals. The latter group consisted of 207 males and 196 females with a median age of 16 (range 3–85) years. There were no significant dissimilarities between those who participated in the ultrasound examination and those who did not, except for a slightly lower percentage of individuals participating from the community of Ndieumeul as compared to Diokhor Tack (p = 0.005), and from the youngest as compared to the older age groups (p = 0.030). No significant differences in age or gender were observed between the two communities.

Morbidity Prevalences

Schistosoma haematobium-specific bladder morbidity was observed in 83% of the study population (334/403; Table 1). Most common lesions concerned multifocal or diffuse bladder wall thickening (n = 189), irregularities (n = 94) or a single mass (n = 19). Microhematuria was twice as prevalent among those with bladder morbidity as compared to those without (44% versus 21%, p = 0.002).

S. mansoni-specific fibrosis was present in 27% (109/403) of the population (Table 2). Liver image patterns up to F were observed, but the large majority had pattern C (89/109).

Morbidity of both liver and bladder was observed in 24% (93/ 391) of the study participants (Table 3). Those who had bladder morbidity tended to be more at risk for hepatic fibrosis and *vice versa* (odds ratio (OR) = 1.3 (95% confidence interval (CI) 0.7–2.4)).

Age-Related Patterns

Figure 1 shows that bladder morbidity was mainly observed in children (<20 years), with a peak in 10-to-19-year-olds. The agerelated distribution of bladder morbidity coincided with that of *S. haematobium* infection intensity and microhematuria, although the peak was slightly later in adolescence, and the subsequent decline in adults less pronounced.

Table 2. Schistosoma mansoni-associated hepatic fibrosis and

 S. mansoni infection in the two co-endemic communities

 studied.

Morbidity	Liver image pattern	n	(%)	5. m	ansoni infection
				%	GM (95%CI) ^a
Negative	А-В	282	(70)	72	123 (99–153) epg
	А	142	(35)	75	131 (98–175) epg
	В	140	(35)	69	115 (83–160) epg
Positive	C-F	109	(27)	61	113 (76–168) epg
	С	89	(22)	63	125 (81–193) epg
	D	10	(2)	40	113 (29–400) epg
	E	9	(2)	67	45 (10–160) epg
	F	1	(0.2)	0	N/A
Excluded		12	(3)		
Total		403	(100)	69	120 (99–145) epg

^aGM) Geometric Mean; calculated for microscopically *S. mansoni*-positive individuals only. N/A) Not Applicable. doi:10.1371/journal.pntd.0001829.t002

Table 3. Bladder and liver co-morbidity in the two coendemic communities studied.

Bladder morbidity	Hepatic fibr		
	Negative	Positive	Total
Negative	51	16	67
Positive	231	93	324
Total	282	109	391

Number of cases.

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The prevalence of hepatic fibrosis increased from 6% in 0-to-9year-olds to 44% in those ≥ 20 years old. The more severe forms of hepatic fibrosis (liver image patterns D, E and F) only became apparent in adults (≥ 20 years old). The peak in the age-related distribution of hepatic fibrosis occurred more than 10 years later in life than the *S. mansoni* infection intensity peak (Figure 1).

Risk Factors

Multivariable analysis showed age to be a significant risk factor for bladder morbidity as well as hepatic fibrosis (Table 4). However, agerelated patterns of hepatic fibrosis differed between the two communities (p = 0.033): while the ORs for hepatic fibrosis increased with age in Diokhor Tack (p < 0.001), they did not vary with age in Ndieumeul (data not shown). Individuals from Diokhor Tack were significantly more at risk for hepatic fibrosis but tended to be less at risk for bladder morbidity than their counterparts from Ndieumeul (Table 4). Furthermore, females were less at risk for both forms of morbidity than males. Neither *S. mansoni* intensity nor *S. haematobium* intensity was identified as an independent risk factor for hepatic fibrosis. On the other hand, *S. haematobium* (but not *S. mansoni*) infection intensity was a strong risk factor for bladder morbidity.

Effect of Mixed Infection

After including mixed infection into the multivariable model, the above-described trends remained the same (Table 5). The risk of hepatic fibrosis did not differ between subjects with single *S*. *mansoni* and those with mixed infections. Interestingly however, mixed infection tended to be negatively associated with *S. haematobium*-specific bladder morbidity, suggesting a protective effect of current *S. mansoni* infection (p = 0.068).

Ectopic Egg Elimination and Bladder Morbidity

Ectopic *S. haematobium* eggs were found in one (0.6%) and ectopic *S. mansoni* eggs in 23 (13%) out of 176 individuals with mixed infections. Table 6 illustrates the importance of the route of *S. mansoni* egg elimination in the development of bladder morbidity. Those who eliminated *S. mansoni* via both urine and feces (n = 17) had highest *S. haematobium* infection intensities and prevalences of bladder morbidity. Lowest prevalences of morbidity were observed in those who exclusively eliminated *S. mansoni* eggs via the urine (and not via the feces; n = 6), despite relatively high *S. haematobium* infection intensities.

Discussion

The global distribution map of schistosomiasis shows a large overlap of *Schistosoma mansoni-* and *S. haematobium-*endemic areas in Africa [2–4]. Yet, little is known about the consequences of mixed *Schistosoma* infections for the human host. Here, we report on the relationship of mixed *Schistosoma* infections with *S. haematobium-*specific bladder morbidity and *S. mansoni-*specific hepatic morbidity in two co-endemic communities.

Risk factors for schistosomal morbidity in this co-endemic area i.e. age, gender, community of residence - were similar to those generally observed in Schistosoma-endemic areas. In line with other studies in mono-endemic areas, we observed that children are more at risk of bladder morbidity [36,37], while adults are more at risk for hepatic fibrosis [2,38]. Participants from one community tended to be less at risk for bladder morbidity but were significantly more at risk for hepatic fibrosis than participants from the other. Microgeographical differences (i.e. between neighboring communities) have been reported before [32-34,39], and are probably due to geographical heterogeneities in Schistosoma exposure (history) [20,40-42], although other factors (e.g. genetic or environmental factors) may also be involved [39]. Males were more at risk for bladder morbidity and hepatic fibrosis than females. Previous studies in either S. mansoni- or S. haematobium-endemic areas have found similar associations for both forms of morbidity [25,30-34,36,38,43-49]. Possibly, males



Figure 1. Age distribution of schistosomiasis morbidity in the two co-endemic communities studied. Colored stacks indicate morbidity prevalences and continuous black lines indicate mean 10log-transformed infection intensities among positive subjects with the standard error of the mean (whiskers). **Panel A:** Different forms of *S. haematobium*-specific bladder morbidity are denoted by a color gradient: light yellow stacks designate a urinary bladder score of 1, bright yellow a score of 2 and orange (3 and 4), red (5) and violet (6) indicate higher morbidity scores. The dotted red line indicates hematuria prevalence in a subsample (n=317). **Panel B:** The severity of *S. mansoni*-specific fibrosis is denoted by a color gradient. Yellow stacks designate liver image pattern C, orange pattern D, red pattern E, and violet stacks indicate pattern F. Striped stacks designate those with borderline liver morbidity (pattern B, not classified as morbidity). doi:10.1371/journal.pntd.0001829.q001

Table 4. Risk factors for schistosomiasis morbidity in the total study population.

Risk factors		S. hae	<i>matobium</i> -specific blad	der morbidity (n=403)	S. mansoni-specific hepatic fibrosis (n = 391)		
		n	Unadjusted OR (95% CI)	Adjusted OR (95% Cl)	n	Unadjusted OR (95%Cl)	Adjusted OR (95%Cl)
Age ^a	<10 y	102	0.3 (0.1–0.7)**	0.3 (0.1–0.8)*	102	0.1 (0.03–0.2)***	0.1 (0.01–1.3)
	10–19 y	121	Ref.	Ref.	119	0.4 (0.2–0.7)**	1.4 (0.4–5.1)
	20–39 y	109	0.3 (0.1–0.7)**	0.6 (0.3–1.3)	105	Ref.	Ref.
	≥40 y	71	0.4 (0.2–0.97)*	0.9 (0.3–2.3)	65	1.0 (0.6–1.9)	0.7 (0.1–3.8)
Gender	Male	207	Ref.	Ref.	200	Ref.	Ref.
	Female	196	0.4 (0.2–0.7)***	0.3 (0.2–0.6)***	191	0.8 (0.5–1.3)	0.5 (0.3–0.9)*
Community	Ndieumeul	100	Ref.	Ref.	99	Ref.	Ref.
	Diokhor Tack	303	0.4 (0.2–0.8)*	0.6 (0.3–1.3)	292	1.9 (1.1–3.3)*	5.1 (1.3–20.8)*
S. haematobium	Infection intensity ^b	403	2.0 (1.4–2.9)***	1.9 (1.3–2.9)**	391	0.6 (0.5–0.8)***	0.9 (0.7–1.3)
S. mansoni	Infection intensity ^b	403	1.5 (1.1–2.1)*	1.1 (0.8–1.6)	391	0.8 (0.6–1.0)	0.9 (0.6–1.3)
Interaction	Age×community	N/A	N/A	N/A	391	N/A	$p = 0.033^{a}$

OR) Odds Ratio; 95%CI) 95% Confidence Interval; Ref.) Reference category; N/A) Not Applicable; *) p<0.05; **) p<0.01; ***) p<0.001.

^aFor *S. haematobium*-specific bladder morbidity, the trend with age was significant at the level of p = 0.025 in the uni- and p = 0.043 in the multivariable analysis. For *S. mansoni*-specific hepatic fibrosis, the trend with age was significant in the crude analysis (p < 0.001). In the adjusted analysis the ORs for hepatic fibrosis increased with age in Diokhor Tack (p < 0.001) but they did not vary with age in Ndieumeul. ^bOR for a 10-fold increase in infection intensity.

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are more prone to *Schistosoma*-associated pathology because of hormonal and immunological differences. It has been proposed that hepatic fibrosis might be more pronounced in men because androgens can reduce interferon (IFN)- γ levels while estrogens have the opposite effect on this antifibrogenic cytokine [43]. Similarly, a study in Uganda showed that the risk of hepatic fibrosis was associated with different cytokine profiles in men and women [35].

Recently, we found infection intensities of both *S. mansoni* and *S. haematobium* to be elevated in mixed as compared to single infections [20]. In the present study, *S. haematobium* infection intensity was identified as an independent risk factor for bladder morbidity, in accordance with previous studies from mono-endemic areas [29,30,36,44–48,50]. However, subjects with mixed infections tended to have less bladder morbidity than those with single *S.*

Table 5. The effect of mixed Schistosoma infection on bladder morbidity and on hepatic fibrosis.

Risk factors		Blado subje	ler morbidity in <i>S. </i> cts (n=216)	aematobium-infected	Hepatic fibrosis in <i>S. mansoni</i> -infected subjects (n = 270)		
		n	Unadjusted OR (95% CI)	Adjusted OR (95% CI)	n	Unadjusted OR (95%CI)	Adjusted OR (95%Cl)
Age ^a	<10 y	63	0.3 (0.1–1.1)	0.2 (0.1–0.9)*	65	0.1 (0.03–0.3)***	0.1 (0.02–0.2)***
	10–19 y	88	Ref.	Ref.	102	0.4 (0.2–0.8)**	0.3 (0.2–0.7)**
	20–39 y	44	0.2 (0.1–0.8)*	0.3 (0.1–1.2)	66	Ref.	Ref.
	≥40 y	21	0.3 (0.1–1.4)	0.5 (0.1–3.0)	37	0.8 (0.3–1.8)	0.8 (0.4–1.9)
Gender	Male	109	Ref.	Ref.	140	Ref.	Ref.
	Female	107	0.3 (0.1–0.8)*	0.3 (0.1–0.9)*	130	1.0 (0.6–1.8)	0.6 (0.3–1.2)
Community	Ndieumeul	70	Ref.	Ref.	87	Ref.	Ref.
	Diokhor Tack	146	0.2 (0.04-0.8)*	0.2 (0.04–0.7)*	183	1.5 (0.8–2.8)	1.6 (0.8–3.2)
S. haematobium	Infection intensity ^b	216	1.8 (0.96–3.3)	1.8 (0.9–3.7)		N/A	N/A
S. mansoni	Infection intensity ^b		N/A	N/A	270	0.9 (0.6–1.4)	1.0 (0.7–1.7)
Mixed infection	No	40	Ref. ^c	Ref. ^c	97	Ref. ^d	Ref. ^d
	Yes	176	0.6 (0.2–2.2)	0.3 (0.1–1.1)	173	0.8 (0.6–1.1)	1.1 (0.8–1.5)

OR) Odds Ratio; 95%CI) 95% Confidence Interval; Ref.) Reference category; N/A) Not Applicable; *) p<0.05; **) p<0.01; ***) p<0.01.

^aThe trends with age were not significant for *S. haematobium*-specific bladder morbidity, but for *S. mansoni*-specific hepatic fibrosis, they were at the level of *p*<0.001 in both analyses.

^bOR for a 10-fold increase in infection intensity.

^cMixed infections as compared to single S. haematobium infections.

^dMixed infections as compared to single *S. mansoni* infections.

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Table 6. S. mansoni egg elimination, infection intensity and bladder morbidity in subjects passing S. haematobium eggs in urine.

			S. haematobium infection intensity	
Route of <i>S. mansoni</i> egg	l elimination	n	(GM (95% CI)) ^a	Bladder morbidity (%)
No elimination	(single S. haematobium infection)	39	3.1 (2.0–4.7) ep10ml	92
Exclusively via feces	(mixed infection)	153	8.0 (6.0–10.8) ep10ml	89
Via feces & urine	(mixed infection)	17	60.1 (27.2–132.3) ep10ml	100
Exclusively via urine	(mixed infection)	6	12.0 (1.5–86.5) ep10ml	50
Total		215	8.1 (6.2–10.4) ep10ml	89

^aGM) Geometric Mean; 95%Cl) 95% Confidence Interval. doi:10.1371/journal.pntd.0001829.t006

haematobium infections, independently of infection intensity. This would suggest a protective effect of *S. mansoni* on bladder morbidity.

The observed reduction in bladder morbidity in mixed infections appeared to be related to ectopic S. mansoni egg elimination. Lowest prevalences of bladder morbidity were observed in S. haematobium-positive individuals who also eliminated S. mansoni eggs in urine (but not in feces), despite the relatively high S. haematobium infection intensities in this group. Experimental models have shown that S. mansoni and S. haematobium can form heterologous male-female pairs. As the S. haematobium male is assumed to be competitively stronger than S. mansoni [51,52], this would result in more heterologous pairs in the vesical than in the mesenteric plexus and thus in more ectopic S. mansoni than ectopic S. haematobium egg elimination in mixed foci [12,20,51-55]. Nothing is known yet about the pathogenicity of eggs from heterologous pairs [54,56]. It could be speculated however, that these heterologous pairs would produce less (pathogenic) eggs than homologous S. haematobium worm pairs, or that their eggs would deviate to other sites [57,58]. Competition between S. mansoni and S. haematobium females for S. haematobium males would then lead to a reduction of bladder morbidity in mixed as opposed to single infections. Other factors may also underlie the potential protective effect of S. mansoni infection. As our study population has been exposed to S. mansoni for a longer period of time than to S. haematobium [19], one could argue that S. mansoni-induced crossresistance to S. haematobium might have played a role as well [59]. Obviously, more research is needed to confirm the observed relation between mixed Schistosoma infection and bladder morbidity, and to understand the underlying mechanisms.

Only one epidemiological study has looked into mixed infections and bladder morbidity before. In contrast to our study, the authors reported a positive association between mixed infection and bladder morbidity in Malian subjects [9]. However, they studied only schoolchildren (7–14 years) and did not take ectopic egg elimination into account. Restricting our analysis to children between 5 and 14 years (the limited sample size did not allow us to perform the analysis on a smaller age range), the association between mixed infection and bladder morbidity still tended to be negative (OR = 0.5 (95%CI 0.1– 3.4)). Disregarding ectopic eggs as well, the association became positive (OR = 1.7 (95%CI 0.4–7.6)), which is in line with the Malian study (data not shown). These discrepancies clearly demonstrate the importance of considering ectopic egg elimination in mixed *Schistosoma* infections and morbidity.

In contrast to our findings for bladder morbidity, hepatic fibrosis was neither associated with current *S. mansoni* infection intensity nor with mixed infections. Hepatic fibrosis only develops after 5-15 years of exposure to *S. mansoni* [2]. This was illustrated by the >10 years' time lag between the *S. mansoni* infection and *S. mansoni*-associated hepatic fibrosis peaks in the respective age-related curves (Figure 1).

While inflammatory hepatic morbidity is generally positively associated with current *S. mansoni* infection [2,60,61], the progression of morbidity into chronic schistosomiasis is driven by cumulative exposure to *S. mansoni* eggs rather than current infection [7,62,63].

So far, only two other studies have investigated the relationship between mixed infections and liver morbidity, with contrasting results. A Zimbabwean study found a slight positive association between mixed infections and liver size [8]. The afore-mentioned Malian study observed a negative association between mixed infections and hepatic fibrosis [9]. However, the latter classified liver image pattern B as abnormal [24], which is not according to the Niamey guidelines [10,31-35], and hampers an adequate comparison with our findings. Adopting the criteria of the Malian study, we still found a positive association between mixed infections and liver fibrosis in children (7-14 years), although not significant (data not shown). The divergences between the Malian study and our study - for bladder as well as liver morbidity - could furthermore be due to differences in the distribution of S. haematobium and S. mansoni infection status and intensity between the three Malian study areas, which were not accounted for [9]. Also other differences in e.g. transmission dynamics [39], between the Malian and present study cannot be excluded.

Conclusion

Up to now, the relationship between mixed *Schistosoma* infection and morbidity has only been studied in schoolchildren. This populationwide study is the first to include adults. Mixed infections were not associated with an increased risk of *S. mansoni*-associated morbidity and even tended to reduce the risk of bladder morbidity. These unexpected results warrant further investigation of a possible protective effect of *S. mansoni* on bladder morbidity. Especially the role of interspecies interactions and ectopic *S. mansoni* egg elimination should be studied in more detail, as these phenomena may have important consequences for schistosomiasis morbidity and control in co-endemic areas.

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Author Contributions

Conceived and designed the experiments: KP SM LM. Performed the experiments: LM MM KV. Analyzed the data: LM KP JM. Contributed reagents/materials/analysis tools: KP SM. Wrote the paper: LM KP.

References

- WHO (2010 February) Schistosomiasis fact sheet. Available: http://www.who. int/mediacentre/factsheets/fs115/en/index.html. Accessed 2012 Aug 30.
- Gryseels B, Polman K, Clerinx J, Kestens L (2006) Human schistosomiasis. Lancet 368: 1106–1118.
- Montgomery S (2011) Infectious Diseases Related To Travel Schistosomiasis. In: Brunette GW, editors. Yellow Book. New York: Oxford University Press.
- Doumenge JP, Mott KE, Cheung C, Villenave D, Chapuis O, et al. (1987) Atlas of the global distribution of schistosomiasis. Bordeaux: Presses Universitaires de Bordeaux.
- King CH (2002) Ultrasound monitoring of structural urinary tract disease in Schistosoma haematobium infection. Mem Inst Oswaldo Cruz 97 Suppl 1: 149– 152.
- Smith JH, Christie JD (1986) The pathobiology of Schistosoma haematobium infection in humans. Hum Pathol 17: 333–345.
- Wilson S, Vennervald BJ, Dunne DW (2011) Chronic hepatosplenomegaly in african school children: a common but neglected morbidity associated with schistosomiasis and malaria. PLoS Negl Trop Dis 5: e1149.
- Friis H, Ndhlovu P, Kaondera K, Franke D, Vennervald BJ, et al. (1996) Ultrasonographic assessment of Schistosoma mansoni and S. haematobium morbidity in Zimbabwean schoolchildren. Am J Trop Med Hyg 55: 290–294.
- Koukounari A, Donnelly CA, Sacko M, Keita AD, Landoure A, et al. (2010) The impact of single versus mixed schistosome species infections on liver, spleen and bladder morbidity within Malian children pre- and post-praziquantel treatment. BMC Infect Dis 10: 227.
- Richter J, Hatz C, Campagne N, Bergquist R, Jenkins JM (1996) Ultrasound in schistosomiasis: A practical guide to the standardized use of ultrasonography for the assessment of schistosomiasis-related morbidity. TDR/STR/SCH/00.1.
- De Clercq D, Vercruysse J, Picquet M, Shaw DJ, Diop M, et al. (1999) The epidemiology of a recent focus of mixed Schistosoma haematobium and Schistosoma mansoni infections around the 'Lac de Guiers' in the Senegal River Basin, Senegal. Trop Med Int Health 4: 544–550.
- Ernould JC, Ba K, Sellin B (1999) Increase of intestinal schistosomiasis after praziquantel treatment in a Schistosoma haematobium and Schistosoma mansoni mixed focus. Acta Trop 73: 143–152.
- Huyse T, Webster BL, Geldof S, Stothard JR, Diaw OT, et al. (2009) Bidirectional introgressive hybridization between a cattle and human schistosome species. PLoS Pathog 5: e1000571.
- Southgate V, Tchuem Tchuente LA, Sene M, De Clercq D, Theron A, et al. (2001) Studies on the biology of schistosomiasis with emphasis on the Senegal river basin. Mem Inst Oswaldo Cruz 96 Suppl: 75–78.
- Ten Hove RJ, Verweij JJ, Vereecken K, Polman K, Dieye L, et al. (2008) Multiplex real-time PCR for the detection and quantification of Schistosoma mansoni and S. haematobium infection in stool samples collected in northern Senegal. Trans R Soc Trop Med Hyg 102: 179–185.
- Van der Werf MJ, Mbaye A, Sow S, Gryseels B, de Vlas SJ (2002) Evaluation of staff performance and material resources for integrated schistosomiasis control in northern Senegal. Trop Med Int Health 7: 70–79.
- Talla I, Kongs A, Verle P (1992) Preliminary study of the prevalence of human schistosomiasis in Richard-Toll (the Senegal river basin). Trans R Soc Trop Med Hyg 86: 182.
- Talla I, Kongs A, Verle P, Belot J, Sarr S, et al. (1990) Outbreak of intestinal schistosomiasis in the Senegal River Basin. Ann Soc Belg Med Trop 70: 173– 180.
- Picquet M, Ernould JC, Vercruysse J, Southgate VR, Mbaye A, et al. (1996) Royal Society of Tropical Medicine and Hygiene meeting at Manson House, London, 18 May 1995. The epidemiology of human schistosomiasis in the Senegal river basin. Trans R Soc Trop Med Hyg 90: 340–346.
- Meurs L, Mbow M, Vereecken K, Menten J, Mboup S, et al. (2012) Epidemiology of mixed Schistosoma mansoni and Schistosoma haematobium infections in northern Senegal. Int J Parasitol 42: 305–311.
- WHO (1991) Basic laboratory methods in medical parasitology. Geneva: World Health Organization.
- Katz N, Chaves A, Pellegrino J (1972) A simple device for quantitative stool thick-smear technique in schistosomiasis mansoni. Rev Inst Med Trop Sao Paulo 14: 397–400.
- WHO (2006) Preventive chemotherapy in human helminthiasis Coordinated use of anthelminthic drugs in control interventions: a manual for health professionals and programme managers.
- 24. Koukounari A, Sacko M, Keita AD, Gabrielli AF, Landoure A, et al. (2006) Assessment of ultrasound morbidity indicators of schistosomiasis in the context of large-scale programs illustrated with experiences from Malian children. Am J Trop Med Hyg 75: 1042–1052.
- Hatz C, Savioli L, Mayombana C, Dhunputh J, Kisumku UM, et al. (1990) Measurement of schistosomiasis-related morbidity at community level in areas of different endemicity. Bull World Health Organ 68: 777–787.
- King CL, Malhotra I, Mungai P, Wamachi A, Kioko J, et al. (2001) Schistosoma haematobium-induced urinary tract morbidity correlates with increased tumor necrosis factor-alpha and diminished interleukin-10 production. J Infect Dis 184: 1176–1182.
- 27. Shiff C, Veltri R, Naples J, Quartey J, Otchere J, et al. (2006) Ultrasound verification of bladder damage is associated with known biomarkers of bladder

cancer in adults chronically infected with Schistosoma haematobium in Ghana. Trans R Soc Trop Med Hyg 100: 847–854.

- Kouriba B, Traore HA, Dabo A, Sangare L, Guindo H, et al. (2005) Urinary disease in 2 Dogon populations with different exposure to Schistosoma haematobium infection: progression of bladder and kidney diseases in children and adults. J Infect Dis 192: 2152–2159.
- Leutscher PD, Reimert CM, Vennervald BJ, Ravaoalimalala VE, Ramarokoto CE, et al. (2000) Morbidity assessment in urinary schistosomiasis infection through ultrasonography and measurement of eosinophil cationic protein (ECP) in urine. Trop Med Int Health 5: 88–93.
- Medhat A, Zarzour A, Nafeh M, Shata T, Sweifie Y, et al. (1997) Evaluation of an ultrasonographic score for urinary bladder morbidity in Schistosoma haematobium infection. Am J Trop Med Hyg 57: 16–19.
- 31. King CH, Magak P, Salam EA, Ouma JH, Kariuki HC, et al. (2003) Measuring morbidity in schistosomiasis mansoni: relationship between image pattern, portal vein diameter and portal branch thickness in large-scale surveys using new WHO coding guidelines for ultrasound in schistosomiasis. Trop Med Int Health 8: 109–117.
- Malenganisho WL, Magnussen P, Friis H, Siza J, Kaatano G, et al. (2008) Schistosoma mansoni morbidity among adults in two villages along Lake Victoria shores in Mwanza District, Tanzania. Trans R Soc Trop Med Hyg 102: 532–541.
- Boisier P, Ramarokoto CE, Ravoniarimbinina P, Rabarijaona L, Ravaoalimalala VE (2001) Geographic differences in hepatosplenic complications of schistosomiasis mansoni and explanatory factors of morbidity. Trop Med Int Health 6: 699–706.
- Booth M, Vennervald BJ, Kabatereine NB, Kazibwe F, Ouma JH, et al. (2004) Hepatosplenic morbidity in two neighbouring communities in Uganda with high levels of Schistosoma mansoni infection but very different durations of residence. Trans R Soc Trop Med Hyg 98: 125–136.
- Booth M, Mwatha JK, Joseph S, Jones FM, Kadzo H, et al. (2004) Periportal fibrosis in human Schistosoma mansoni infection is associated with low IL-10, low IFN-gamma, high TNF-alpha, or low RANTES, depending on age and gender. J Immunol 172: 1295–1303.
- Garba A, Pion S, Cournil A, Milet J, Schneider D, et al. (2010) Risk factors for Schistosoma haematobium infection and morbidity in two villages with different transmission patterns in Niger. Acta Trop 115: 84–89.
- King CH, Blanton RE, Muchiri EM, Ouma JH, Kariuki HC, et al. (2004) Low heritable component of risk for infection intensity and infection-associated disease in urinary schistosomiasis among Wadigo village populations in Coast Province, Kenya. Am J Trop Med Hyg 70: 57–62.
- Berhe N, Myrvang B, Gundersen SG (2007) Intensity of Schistosoma mansoni, hepatitis B, age, and sex predict levels of hepatic periportal thickening/fibrosis (PPT/F): a large-scale community-based study in Ethiopia. Am J Trop Med Hyg 77: 1079–1086.
- Butterworth AE, Curry AJ, Dunne DW, Fulford AJ, Kimani G, et al. (1994) Immunity and morbidity in human schistosomiasis mansoni. Trop Geogr Med 46: 197–208.
- Pinot de Moira A, Fulford AJ, Kabatereine NB, Kazibwe F, Ouma JH, et al. (2007) Microgeographical and tribal variations in water contact and Schistosoma mansoni exposure within a Ugandan fishing community. Trop Med Int Health 12: 724–735.
- Robert CF, Bouvier S, Rougemont A (1989) Epidemiology of schistosomiasis in the riverine population of Lagdo Lake, northern Cameroon: mixed infections and ethnic factors. Trop Med Parasitol 40: 153–158.
- Anderson RM, May RM (1985) Helminth infections of humans: mathematical models, population dynamics, and control. Adv Parasitol 24: 1–101.
 Mohamed-Ali Q, Elwali NE, Abdelhameed AA, Mergani A, Rahoud S, et al.
- 43. Mohamed-Ali Q, Elwali NE, Abdelhameed AA, Mergani A, Rahoud S, et al. (1999) Susceptibility to periportal (Symmers) fibrosis in human Schistosoma mansoni infections: evidence that intensity and duration of infection, gender, and inherited factors are critical in disease progression. J Infect Dis 180: 1298– 1306.
- Brouwer KC, Ndhlovu PD, Wagatsuma Y, Munatsi A, Shiff CJ (2003) Epidemiological assessment of Schistosoma haematobium-induced kidney and bladder pathology in rural Zimbabwe. Acta Trop 85: 339–347.
- El Khoby T, Galal N, Fenwick A, Barakat R, El Hawey A, et al. (2000) The epidemiology of schistosomiasis in Egypt: summary findings in nine governorates. Am J Trop Med Hyg 62: 88–99.
- Traore M, Traore HA, Kardorff R, Diarra A, Landoure A, et al. (1998) The public health significance of urinary schistosomiasis as a cause of morbidity in two districts in Mali. Am J Trop Med Hyg 59: 407–413.
- Serieye J, Boisier P, Ravaoalimalala VE, Ramarokoto CE, Leutscher P, et al. (1996) Schistosoma haematobium infection in western Madagascar: morbidity determined by ultrasonography. Trans R Soc Trop Med Hyg 90: 398–401.
- 48. Heurtier Y, Lamothe F, Develoux M, Docquier J, Mouchet F, et al. (1986) Urinary tract lesions due to Schistosoma haematobium infection assessed by ultrasonography in a community based study in Niger. Am J Trop Med Hyg 35: 1163–1172.
- 49. Wagatsuma Y, Aryeetey ME, Sack DA, Morrow RH, Hatz C, et al. (1999) Resolution and resurgence of Schistosoma haematobium-induced pathology after community-based chemotherapy in ghana, as detected by ultrasound. J Infect Dis 179: 1515–1522.

- Brouwer KC, Munatsi A, Ndhlovu PD, Wagatsuma Y, Shiff CJ (2004) Urinary schistosomiasis in Zimbabwean school children: predictors of morbidity. Afr Health Sci 4: 115–118.
- Webster BL, Southgate VR, Tchuem Tchuente LA (1999) Mating interactions between Schistosoma haematobium and S. mansoni. J Helminthol 73: 351–356.
- Cunin P, Tchuem Tchuente LA, Poste B, Djibrilla K, Martin PM (2003) Interactions between Schistosoma haematobium and Schistosoma mansoni in humans in north Cameroon. Trop Med Int Health 8: 1110–1117.
- Ratard RC, Ndamkou CN, Kouemeni LE, Ekani Bessala MM (1991) Schistosoma mansoni eggs in urine. J Trop Med Hyg 94: 348–351.
- Khalil SB, Mansour NS (1995) Worm development in hamsters infected with unisex and cross-mated Schistosoma mansoni and Schistosoma haematobium. J Parasitol 81: 8–11.
- Southgate VR, Jourdane J, Tchuente LA (1998) Recent studies on the reproductive biology of the schistosomes and their relevance to speciation in the Digenea. Int J Parasitol 28: 1159–1172.
- Jourdane J, Imbert-Establet D, Tchuente LA (1995) Parthenogenesis in schistosomatidae. Parasitol Today 11: 427–430.

- Mansour NS, Soliman GN, El-Assal FM (1984) Studies on experimental mixed infections of Schistosoma mansoni and S. haematobium in hamsters. Z Parasitenkd 70: 345–357.
- Soliman GN, el Assal FM, Mansour NS, Garo K (1986) Comparison of two Egyptian strains of Schistosoma mansoni in hamsters. Z Parasitenkd 72: 353– 363.
- Webbe G, James C, Nelson GS, Ismail MM, Shaw JR (1979) Cross resistance between Schistosoma haematobium and S. mansoni in the baboon. Trans R Soc Trop Med Hyg 73: 42–54.
- Guyatt H, Gryseels B, Smith T, Tanner M (1995) Assessing the public health importance of Schistosoma mansoni in different endemic areas: attributable fraction estimates as an approach. Am J Trop Med Hyg 53: 660–667.
- Kardorff R, Stelma FF, Vocke AK, Yazdanpanah Y, Thomas AK, et al. (1996) Ultrasonography in a Senegalese community recently exposed to Schistosoma mansoni infection. Am J Trop Med Hyg 54: 586–590.
- 62. Mahmoud, A A. (2001) Schistosomiasis. London: Imperial College Press.
- Chan MS, Guyatt HL, Bundy DA, Medley GF (1996) Dynamic models of schistosomiasis morbidity. Am J Trop Med Hyg 55: 52–62.