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# Effect of Manganese in Test Media on *In Vitro* Susceptibility of *Enterobacteriaceae* and *Acinetobacter baumannii* to Tigecycline

J. Veenemans,<sup>a</sup> J. W. Mouton,<sup>b,c</sup> J. A. J. W. Kluytmans,<sup>a</sup> R. Donnelly,<sup>c</sup> C. Verhulst,<sup>a</sup> and P. H. J. van Keulen<sup>a</sup>

Laboratory for Microbiology and Infection Control, Amphia Hospital, Breda, The Netherlands<sup>a</sup>; Regional Laboratory for Public Health, Canisius Wilhelmina Hospital, Nijmegen, The Netherlands<sup>b</sup>; and Department of Medical Microbiology, Radboud University Nijmegen Medical Centre, Nijmegen, The Netherlands<sup>c</sup>

**We assessed the effect of increasing manganese concentrations in test media (0.001 to 1,024 mg/liter) on MICs of tigecycline. For both broth microdilution (BMD) and Etests, this effect was negligible for physiological concentrations, but MICs increased when concentrations exceeded 8 mg/liter. Susceptibility testing should be performed on media with standardized low manganese content.**

Tigecycline is one of the few antimicrobial drugs that can be used to treat infections with highly resistant organisms (7, 8). Several studies reported that *in vitro* bacterial susceptibility to tigecycline varies by test medium and condition (3, 4, 10, 13, 14), and a recent study showed that addition of manganese (Mn) to the test medium led to increased MICs of tigecycline, as determined by an Etest for *Enterobacteriaceae* and staphylococci. (5) Variability in Mn concentrations in standard media may thus result in falsely elevated MICs for tigecycline. A concentration-effect relationship was not established, however, and the threshold concentration of Mn below which MICs are unaffected remains unknown. Because such a threshold may depend on the test method used, we evaluated the effect of increasing manganese concentrations in the test medium on MICs, as determined by broth microdilution, Etest, and disk diffusion.

The study included strains of five species of *Enterobacteriaceae* (*Escherichia coli*, *Klebsiella pneumoniae*, *Citrobacter freundii*, *Enterobacter cloacae*, and *Proteus mirabilis*) and *Acinetobacter baumannii*, which were collected and stored at  $-80$  degrees in a prospective cohort study in 18 Dutch hospitals as described previously (15). An ATCC control strain (Table 1) was included for each species.

In the first experiment, we determined the effect of increasing medium Mn concentrations on MICs, as measured by Etest (bioMérieux), and on zone diameters, as measured by disk (15  $\mu$ g tigecycline; Oxoid, United Kingdom) diffusion. Strains were thawed and inoculated on Columbia agar plates with 5% sheep blood and incubated for 18 to 24 h at 35°C. Subsequently, a suspension of a 0.5 McFarland standard (in 0.9% saline) was used to inoculate freshly prepared Iso-Sensitest agar plates (Oxoid, United Kingdom). The Iso-Sensitest medium has been developed specifically to overcome the variability in mineral concentrations observed in Mueller-Hinton medium and has a stable mineral content with an MnCl<sub>2</sub> concentration of 2 mg/liter (2). Media were freshly prepared to avoid effects of oxygenation (1, 9) and supplemented with MnCl<sub>2</sub> (VWR, The Netherlands) in 2-fold serial dilutions to achieve Mn concentrations ranging between 2 mg/liter and 1,024 mg/liter. Plates were incubated for 18 to 24 h at 35°C before values were read.

Because reference values for Mn concentrations in human serum (0.2 to 1.4  $\mu$ g/liter) (12) are substantially lower than concentrations found in standard susceptibility test media ( $\geq 2$  mg/liter), a second experiment in which the tested Mn concentration range

also included physiological concentrations was performed. MICs were measured using broth microdilution (BMD), and MnCl<sub>2</sub> was added to a manganese-free medium (synthetic amino acid medium [SAAM-B]; U.S. Biological, MA; catalog number S9270-02) in a series of 21 2-fold dilutions to achieve concentrations in the test wells ranging between 1,024 and 0.001 mg/liter. Tests were performed according to guidelines from the International Organization for Standardization (ISO; guideline 20776) (6). Tigecycline was provided by Pfizer. MICs were read visually as the lowest concentrations that completely inhibited growth.

A simple linear regression model (SPSS v15.0 for Windows; SPSS, Chicago, IL) was used to model the relationship between manganese concentrations in the test medium and MIC values. MIC values were log<sub>2</sub> transformed so that a 1-unit increment in log values corresponded to a doubling of the MIC.

Figure 1 shows the MICs for tigecycline for each species, determined by broth microdilution (solid line) or Etest (dotted line), as a function of the concentration of manganese in the medium. For all species tested, the effect of manganese on geometric mean MICs was negligible for medium concentrations of less than 8 mg/liter. MIC values increased substantially, however, when medium manganese concentrations exceeded 64 mg/liter. The threshold whereby this increase occurred did not differ substantially between species except for with *P. mirabilis*, for which the effect seemed less pronounced. Compared to values measured on a Mn-free medium, MICs doubled at medium Mn concentrations of 220, 240, 200, 230, and 136 mg/liter for *E. coli*, *K. pneumoniae*, *C. freundii*, *E. cloacae*, and *A. baumannii*, respectively.

The effect of medium manganese concentrations on MIC values did not differ between BMD and Etests ( $P = 0.20$ ). Independent of medium manganese concentrations, MIC values determined by Etest were consistently higher than those determined by broth microdilution (geometric mean difference, 0.8 doubling dilutions; 95% confidence interval [CI], 0.7 to 0.9). Similar patterns

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Address correspondence to J. A. J. W. Kluytmans, Jankluytmans@gmail.com.

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TABLE 1 MICs for the ATCC control strains with each medium manganese concentration<sup>a</sup>

Strain	MIC ( $\mu\text{g/ml}$ ) with each added concn of Mn (mg/liter)										
	0	2	4	8	16	32	64	128	256	512	1,024
<i>Escherichia coli</i> ATCC 29522	0.09	0.19	0.19	0.25	0.25	0.38	0.5	0.5	0.5	1	4
<i>Klebsiella pneumoniae</i> ATCC 700603	3	4	4	4	6	6	6	8	8	16	48
<i>Enterobacter cloacae</i> ATCC 23355	0.75	0.75	0.75	1	1	1	1.5	1.5	2	4	12
<i>Citrobacter freundii</i> ATCC 8090	0.38	0.38	0.38	0.38	0.38	0.38	0.5	0.75	1	2	4
<i>Acinetobacter baumannii</i> ATCC 19606	1	0.75	0.75	1	1	1.5	3	4	4	8	8
<i>Proteus mirabilis</i> ATCC 43071	2	1.5	2	2	2	2	3	2	3	6	16

<sup>a</sup> MICs were tested on Iso-Sensitest agar with a standard manganese concentration of 2 mg/liter. Data are from Etests only.

were observed for the influence of medium manganese concentrations on inhibitory zone diameters (Fig. 2).

These data show that *in vitro* bacterial susceptibility to tigecycline is influenced by manganese concentrations in the test medium, whereby the effect of variations in manganese concentrations appears negligible when concentrations in the test medium are below 8 mg/liter. This suggests that the activity of tigecycline will be unaffected in human serum, which has a considerably lower concentration of manganese. Our findings emphasize the need to use test media with standardized low-Mn concentrations

for tigecycline susceptibility testing. Due to poor reproducibility of its mineral content, the Mueller-Hinton medium may be unsuitable for this purpose. Whereas most commonly available test media probably have concentrations below the critical range, considerable variability in the concentration of divalent cations in Mueller-Hinton broth or agar from different manufacturers has been reported (11), and manganese concentrations in Mueller-Hinton agar that are well above 500 mg/liter have been found (5).

The mechanisms by which manganese influences *in vitro* activity of tigecycline require further investigation. These may involve

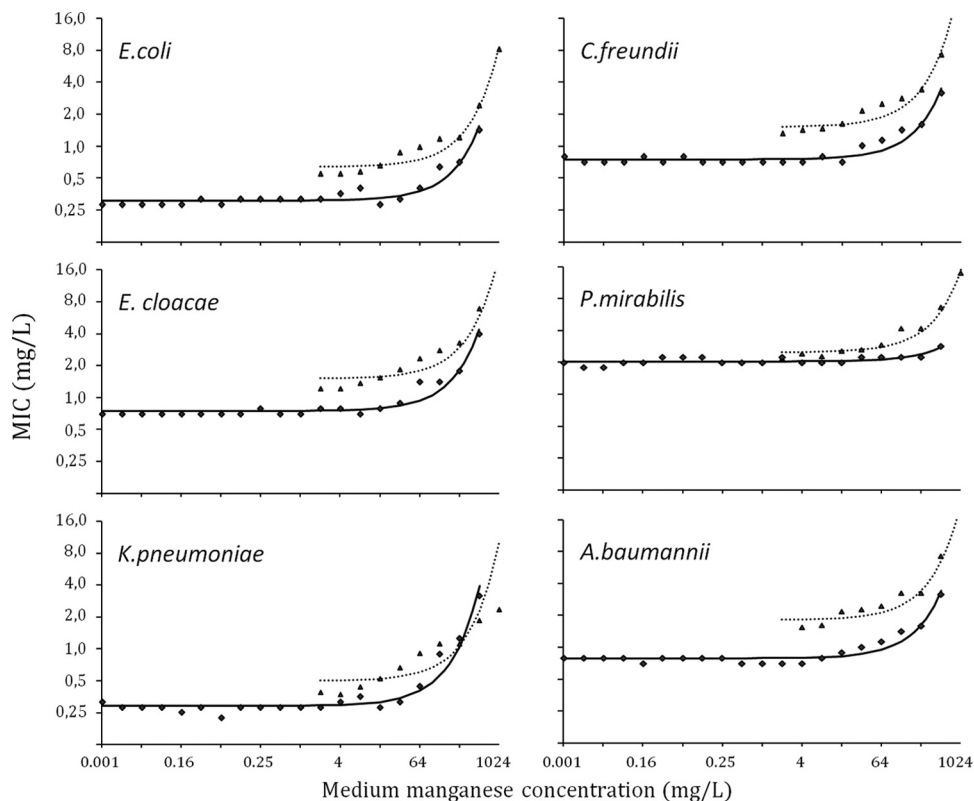


FIG 1 Effect of increasing medium manganese concentrations on MIC (geometric mean) by bacterial species. Medium manganese concentrations are presented on a log-transformed x axis. The regression line models the relationship between medium manganese concentrations and MICs for broth microdilution (solid line) and Etest (dotted line).

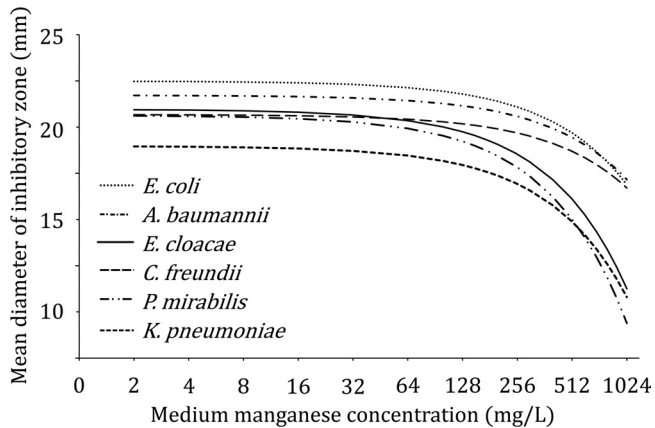


FIG 2 Effect of increasing medium manganese concentrations on average zone diameters, measured by disk diffusion.

the induction of bacterial resistance by manganese or the inactivation of tigecycline due to the formation of complexes between manganese and tigecycline (5). Other divalent cations may have similar effects on susceptibility test results, and because we did not use the same medium for the Etests and for the BMD, it is possible that differences in the concentrations of minerals other than manganese may partly explain the observed differences in MICs between these 2 methods. Further studies are needed to identify causal factors involved. Meanwhile, results of tigecycline susceptibility testing by Etest should be interpreted with caution.

Lastly, because Mn concentrations in human serum are substantially lower than those found in standard susceptibility test media, Mn is unlikely to interfere with tigecycline antibacterial activity under physiological conditions.

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