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Complete Genome Sequence of *Mycobacterium fortuitum* subsp. *fortuitum* Type Strain DSM46621

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Mycobacterium fortuitum is a member of the rapidly growing nontuberculous mycobacteria (NTM). It is ubiquitous in water and soil habitats, including hospital environments. *M. fortuitum* is increasingly recognized as an opportunistic nosocomial pathogen causing disseminated infection. Here we report the genome sequence of *M. fortuitum* subsp. *fortuitum* type strain DSM46621.

ycobacterium fortuitum is a nonpigmented rapidly growing Mycobacterium classified in Runyon group IV (5), first isolated from an amphibian source in 1905 and subsequently identified as the cause of a human cutaneous infection in a patient in 1938 (3). Like many other nontuberculous mycobacteria (NTM), M. fortuitum is found worldwide in natural and processed water sources, including chlorine-treated water, as well as in sewage, dirt, and hospital environments (4, 8). Major types of disease caused by *M. fortuitum* include, in decreasing order of frequency, infections of postsurgical wounds, soft tissue, skin, and lung (3, 5, 11). Occasionally reported miscellaneous infections include keratitis, endocarditis, lymphadenitis, meningitis, hepatitis, peritonitis, catheter-related sepsis, and disseminated infections (3, 6, 11). The M. fortuitum group is attracting attention due to its increasing number of cases, its virulence, and its emerging resistance to antibiotics. However, in the genome database, there is no whole genome of this species present until now. To facilitate a more reliable genetic identification in the M. fortuitum complex, we have characterized the complete genome sequence of M. fortuitum subsp. fortuitum type strain DSM46621.

To sequence the genome of *M. fortuitum* subsp. *fortuitum* type strain DSM46621, we used a shotgun sequencing method and Illumina HiSeq 2000 technology using a paired-end library, with a read length of 100 bp and an insert size of 500 bp. A total of 2.95 million Illumina sequencing reads were generated. These short sequence reads were first quality trimmed before *de novo* assembled using Velvet (12). The draft genome was further improved with iCORN (7) and IMAGE (10), as described in PAGIT (9), before it was scaffolded with SSPACE (2). The final assembly has 82 supercontigs and an N_{50} of 159,889 bp. The genome annotation was performed using the NCBI Prokaryotic Genomes Automatic Annotation Pipeline (PGAAP).

The *M. fortuitum* subsp. *fortuitum* type strain DSM46621 genome sequence is 6,349,738 bp in length, with 6,241 predicted coding sequences. The overall GC content of the chromosome was 66%. There are 54 tRNA genes and four sets of rRNA operons as predicted by the PGAAP pipeline. It was possible to assign a biological function to 67.7% (4,225) of the coding sequences on the *M. fortuitum* subsp. *fortuitum* chromosome.

The RAST server annotation pipeline (1) revealed that *M. fortuitum* subsp. *fortuitum* has the highest similarity with *Mycobac*- *terium smegmatis* strain MC2 155, among all mycobacteria with complete genome sequences determined. The *M. fortuitum* subsp. *fortuitum* genome was found to be smaller (6.38 Mb) than the genome of *M. smegmatis* MC2 155 (6.99 Mb) and to contain fewer genes (6,076 versus 6,816). Comparative genomic analysis with *M. smegmatis* MC2 155 revealed that *M. fortuitum* subsp. *fortuitum* contains more virulence-associated mammalian cell entry (MCE) operons. In addition, several methyltransferases, glycosyltransferases, and dehydrogenases, which probably participate in the cell wall synthesis and modification, were present in the *M. fortuitum* genome. We expect that this genome sequence will serve as a valuable reference for understanding the disparity in virulence and epidemiologic traits in the *M. fortuitum* complex.

Nucleotide sequence accession numbers. This whole-genome shotgun project has been deposited at DDBJ/EMBL/GenBank under the accession number ALQB00000000. The version described in this paper is the first version, ALQB00000000.

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REFERENCES

- 1. Aziz RK, et al. 2008. The RAST server: rapid annotations using subsystems technology. BMC Genomics 9:75.
- 2. Boetzer M, Henkel CV, Jansen HJ, Butler D, Pirovano W. 2011. Scaffolding pre-assembled contigs using SSPACE. Bioinformatics 27: 578–579.
- 3. Brown TH. 1985. The rapidly growing mycobacteria—Mycobacterium fortuitum and Mycobacterium chelonei. Infect. Control 6:283–288.

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- 4. Galassi L, et al. 2003. Nontuberculous mycobacteria in hospital water systems: application of HPLC for identification of environmental mycobacteria. J. Water Health 1:133–139.
- 5. Muthusami JC, et al. 2004. Mycobacterium fortuitum: an iatrogenic cause of soft tissue infection in surgery. ANZ J. Surg. 74:662–666.
- 6. Olalla J, et al. 2002. Mycobacterium fortuitum complex endocarditis case report and literature review. Clin. Microbiol. Infect. 8:125–129.
- Otto TD, Sanders M, Berriman M, Newbold C. 2010. Iterative Correction of Reference Nucleotides (iCORN) using second generation sequencing technology. Bioinformatics 26:1704–1707.
- Petrini B. 2006. Non-tuberculous mycobacterial infections. Scand. J. Infect. Dis. 38:246–255.
- Swain MT, et al. 2012. A post-assembly genome-improvement toolkit (PAGIT) to obtain annotated genomes from contigs. Nat. Protoc. 7:1260–1284.
- Tsai IJ, Otto TD, Berriman M. 2010. Improving draft assemblies by iterative mapping and assembly of short reads to eliminate gaps. Genome Biol. 11:R41.
- 11. Wallace RJ, Jr, et al. 1983. Spectrum of disease due to rapidly growing mycobacteria. Rev. Infect. Dis. 5:657–679.
- 12. Zerbino DR, Birney E. 2008. Velvet: algorithms for de novo short read assembly using de Bruijn graphs. Genome Res. 18:821–829.