NASA GeneLab Project: Bridging Space Radiation Omics with Ground Studies

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ABSTRACT

Accurate assessment of risk factors for long-term space missions is critical for human space exploration: therefore it is essential to have a detailed understanding of the biological effects on humans living and working in deep space. Ionizing radiation from Galactic Cosmic Rays (GCR) is one of the major risk factors factor that will impact health of astronauts on extended missions outside the protective effects of the Earth's magnetic field. Currently there are gaps in our knowledge of the health risks associated with chronic low dose, low dose rate ionizing radiation, specifically ions associated with high (H) atomic number (Z) and energy (E). The GeneLab project (genelab.nasa.gov) aims to provide a detailed library of Omics datasets associated with biological samples exposed to HZE. The GeneLab Data System (GLDS) currently includes datasets from both spaceflight and ground-based studies, a majority of which involve exposure to ionizing radiation. In addition to detailed information for ground-based studies, we are in the process of adding detailed, curated dosimetry information for spaceflight missions. GeneLab is the first comprehensive Omics database for space related research from which an investigator can generate hypotheses to direct future experiments utilizing both ground and space biological radiation data. In addition to previously acquired data, the GLDS is continually expanding as Omics related data are generated by the space life sciences community. Here we provide a brief summary of space radiation related data available at GeneLab.

INTRODUCTION

The radiation biology community has a need for data to assess the impact of exposure to chronic low dose, low dose-rate highly-ionizing radiation. Acquisition of such data for animals or cell cultures is possible only at a small number of facilities world-wide (1-7).

NASA's GeneLab Data Systems (GLDS) (genelab.nasa.gov) have included a steadily growing library of Omics datasets with associated detailed curated dosimetry information since 2015 (**Fig** 1). This repository is helping bridge the gap between irradiation studies performed on earth and biological experiments conducted in space since the early 1990's. In this commentary, we provide a quick summary of the content of the repository in terms of species, radiation quality and dosimetry method, as well as the types of omics data being collected. We also include a comparison of dose and dose rates for samples used in ground studies versus spaceflight specimens.

Most of these studies were funded by the NASA Human Research Program, which has supported

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numerous ground-based studies with high-LET radiation generated at the NASA Space Radiation Laboratory (NSRL) (3); the DOE low dose program providing primarily low-LET data; and the NASA Space Biology Program providing primarily data from specimens flown in space.

There is a diverse set of space related Omics data available on GeneLab. A total of 154 datasets from space related Omics research are currently available. These entries include data from a wide variety of different organisms including: rodents, human cells, plants, microbes, invertebrates, drosophila, and fish (**Fig 2A**). 53% of the datasets were gathered from spaceflight experiments and 47% from ground related space studies (which includes radiation studies done at NSRL) (**Fig 2B**).

GeneLab currently hosts datasets employing with a variety of Omics assay objectives (**Fig 2C**). with the majority being transcription profiling, and a majority of these (77%) using oligonucleotide microarray techniques, but a growing fraction (currently 23%) employing RNA sequencing techniques. We strongly encourage readers with published or unpublished Omics data related to space research to join our effort by contributing their datasets (instructions for which are on the web site).

RADIATION GROUND STUDIES CURRENTLY IN GENELAB

There are a total of 69 GeneLab datasets which are ground (earth-based) space research with 28 datasets specifically related to radiation work and the majority of these were radiation experiments done at NSRL (**Fig 3A**) (3). Investigations of simulated microgravity comprise the rest of the ground experiments (8-16), together with a small number studying atmospheric pressure (17), clinical treatment and exercise/nutrition (18), freezing (19), genotype (20-27), hypergravity (8, 28-30), sample collection protocols (31), temperature (8, 32, 33), or timepoints (34). The majority of the ground-based radiation experiments were done using mice (**Fig 3B**). In vitro experiments involving human tissues make up most of the rest the experiments with one study each of microbes, plants, and rats. Gamma exposure experiments comprise the majority of the radiation datasets available in the repository (**Fig 3C**). These datasets were mainly done by investigators for comparisons to HZE radiation. Proton and ⁵⁶Fe radiation experiments (27, 35-41) comprise the majority of simulated space radiation-related datasets available on GeneLab with a few experiments related to ¹⁶O (42), ¹²C (43), ²⁸Si (44), and neutrons (45) (**Fig 3C**).

The GLDS also contains extensive information on parameters related to the ionizing radiation experiments/datasets. Most datasets include the following information: ion, total dose, dose rate, LET, and energies associated with the radiation. The distribution of dose rates vs total dose (**Fig 4A**) illustrates the wide range of radiation-related datasets available to the public ranging from low doses (0.1Gy) to high doses (30Gy) with dose rates ranging from 0.001Gy/min to 5.66Gy/min. Typically, microbes and plants were exposed to higher total doses, while the majority of the rodent research used doses between 0.1 Gy and 3 Gy (with some gamma experiments done at higher doses). The linear energy transfer (LET) for these experiments ranged from 0.24 keV/µm to 150 keV/µm (**Fig 4B**). Since GeneLab relies on the investigators to provide these metadata, and not all datasets are complete: there are 13 datasets with missing dose rate information. It is critical for all investigators submitting data to GeneLab to provide as much information as possible about irradiation conditions.

SPACE RADIATION DOSIMETRY

GeneLab has recently begun documenting the absorbed radiation dose received by samples flown in space for experiments primarily testing the effects of microgravity, and, only incidentally examining space-radiation effects. To date OMICS data is available on GeneLab which includes experiments conducted on three types of vehicles that have carried samples in space: the "Space Shuttle" (or Space Transportation System, STS), free flying satellites (BION, FOTON) (46, 47), and the International Space Station (ISS). The dosimetry measuring techniques varied from vehicle to vehicle and, for ISS, from module to module. Because the primary purpose of on-board dosimetry on crewed vehicles is monitoring astronaut health and, on free flyers, wholeenvironmental monitoring, biology experiments typically have not flown with "dedicated" dosimeters (i.e., dosimeters integrated into experiment platform housing). Therefore, doses to which study samples are exposed frequently must be interpolated and/or extrapolated from the closest dosimeters. Both passive and active dosimetry have been used on these vehicles. For STS, 3 passive dosimeter packages were fixed in locations on the shuttle middeck, where biological samples were located (Fig 5A). In the case of BION (46, 47) (Fig 5B) and ISS, both passive (thermoluminescent dosimeters, TLD, plastic nuclear track detectors, PNTD) and active (solid state, tissue equivalent proportional counters (TEPC)) were used. Two qualities of radiation are considered: low-LET (photons and electrons) and high-LET (charged nuclei). For passive dosimeters, TLD are sensitive to low-LET charged particles (< 10 keV/µm) and PNTD to high-LET (> 10 keV/μm). Active dosimeters are sensitive to a wider range in LET and, depending on the detector, can provide time resolution, LET spectra and some particle identification. By integrating the dose from the time-resolved data over the duration of the experiment, the total absorbed dose can be calculated. Depending on the configuration of dosimeters in the vicinity of the samples, absorbed dose may be reported as averaged with other detectors, or individually.

Taking into account the wide variety of dosimetry scenarios, we have chosen to report the "lowest common denominator" data: the data that are common to most if not all dosimeters, whether passive or active. Therefore, each omics dataset in the GeneLab repository with samples flown in space has a corresponding metadata set which includes the exposure duration, and the average, minimum and maximum absorbed dose received, broken out into low LET and high LET charged particles (when LET resolution is available). The duration of the exposure is defined as the time a sample was in space and biologically active, *i.e.* when the sample has returned to Earth or when it is chemically fixed or frozen in space.

It is important to note that the absorbed doses we provide in these metadata are an approximation, due to several limiting factors. First, there is known contribution of sensitivity in charge and LET for each detector being used. For example, even though TLDs detect low-LET radiation, the detected dose also includes some contribution from charged nuclei depending on the charge and speed of the nuclei traversing the detector. Similarly, active detectors even if tuned to specific energies and charges can still have traces dose from low-LET particles and neutrons. Second, reported dosimetry does not take into account the additional shielding provided by the sample enclosure. For low energy particles or low-LET this hardware could have significant attenuating effect and would be unique for each mission and experiment. Therefore, in addition to the radiation metadata for individual dataset, GeneLab also reports dosimetry measurements for all detectors available for each mission, in a "reference information" section of the GeneLab database. A glance at these data show that a wide range of total absorbed dose over the various spaceflight experimental environments (**Fig 5C**). For example, rodents flown on the BION-M1 missions (48-50) obtained the largest dose of ionizing radiation (30.2 mGy), while the rodents on-board STS missions only received in the range of 1 mGy to 4.7 mGy. GeneLab is currently assembling all the

dosimetry information for the ISS related datasets and will have it available in the near future. Our aim is that by providing these data, we will facilitate more detailed analyses of these kinds of environments by other investigators.

SUMMARY

The goal of GeneLab is to provide users with a data repository and analytical and visualization tools for molecular signals relating to a host of factors known to influence biological response, including radiation dose, dose rate, and quality. Standardizing the dosimetry information in the entire repository is an essential first step towards accomplishing this task and we hope the radiation biology community will be excited about this new source of information and will visit the database often.

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CONFLICTS OF INTEREST

All authors declare no competing financial interests and conflict of interest with the data and information in this manuscript.

CONTRIBUTIONS

A.B. Analyzed all data, made figures, and wrote manuscript. Y.K. did radiation data acquisition and annotation. D.B. provided exchange of ideas and edited the manuscript. S.G. provided exchange of ideas and edited the manuscript. J.M. acquired radiation dosimetry information, provided exchange of ideas, and edited the manuscript. S.V.C. wrote manuscript, provided exchange of ideas, and edited the manuscript.

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REFERENCES

- 1. Ohnishi T, Life science experiments performed in space in the ISS/Kibo facility and future research plans. Journal of radiation research 2016; 57 Suppl 1, i41-i46.
- 2. Nelson GA, Green LM, Gridley DS, Archambeau JO, Slater JM, Research activities at the Loma Linda University and Proton Treatment Facility--an overview. Physica medica: PM: an international journal devoted to the applications of physics to medicine and biology: official journal of the Italian Association of Biomedical Physics 2001; 17 Suppl 1, 30-2.
- 3. Schimmerling W, Genesis of the NASA Space Radiation Laboratory. Life sciences in space research 2016; 9, 2-11.

- 4. Li M, Gonon G, Buonanno M, Autsavapromporn N, de Toledo SM, Pain D, et al., Health risks of space exploration: targeted and nontargeted oxidative injury by high-charge and high-energy particles. Antioxidants & redox signaling 2014; 20, 1501-23.
- 5. Miller J, Proton and heavy ion acceleration facilities for space radiation research. Gravitational and space biology bulletin: publication of the American Society for Gravitational and Space Biology 2003; 16, 19-28.
- 6. Durante M, Heavy ion radiobiology for hadrontherapy and space radiation protection. Radiotherapy and oncology: journal of the European Society for Therapeutic Radiology and Oncology 2004; 73 Suppl 2, S158-60.
- 7. Gressier V, Review of neutron calibration facilities and monitoring techniques: new needs for emerging fields. Radiation protection dosimetry 2014; 161, 27-36.
- 8. Herranz R, Hill RJ, Dijkstra CE, Eaves L, van Loon JJ, Medina FJ, The behavioural-driven response of the Drosophila imago transcriptome to different types of modified gravity. Genomics Discovery 2013; 1.
- 9. Arunasri K, Adil M, Venu Charan K, Suvro C, Himabindu Reddy S, Shivaji S, Effect of simulated microgravity on E. coli K12 MG1655 growth and gene expression. PloS one 2013; 8, e57860.
- 10. Casaburi G, Goncharenko-Foster I, Duscher AA, Foster JS, Transcriptomic changes in an animal-bacterial symbiosis under modeled microgravity conditions. Sci Rep 2017; 7, 46318.
- 11. Feger BJ, Thompson JW, Dubois LG, Kommaddi RP, Foster MW, Mishra R, et al., Microgravity induces proteomics changes involved in endoplasmic reticulum stress and mitochondrial protection. Sci Rep 2016; 6, 34091.
- 12. Girardi C, De Pitta C, Casara S, Calura E, Romualdi C, Celotti L, et al., Integration analysis of microRNA and mRNA expression profiles in human peripheral blood lymphocytes cultured in modeled microgravity. Biomed Res Int 2014; 2014, 296747.
- 13. Girardi C, De Pitta C, Casara S, Sales G, Lanfranchi G, Celotti L, et al., Analysis of miRNA and mRNA expression profiles highlights alterations in ionizing radiation response of human lymphocytes under modeled microgravity. PloS one 2012; 7, e31293.
- 14. Patel MJ, Liu W, Sykes MC, Ward NE, Risin SA, Risin D, et al., Identification of mechanosensitive genes in osteoblasts by comparative microarray studies using the rotating wall vessel and the random positioning machine. J Cell Biochem 2007; 101, 587-99.
- 15. Vidyasekar P, Shyamsunder P, Arun R, Santhakumar R, Kapadia NK, Kumar R, et al., Genome Wide Expression Profiling of Cancer Cell Lines Cultured in Microgravity Reveals Significant Dysregulation of Cell Cycle and MicroRNA Gene Networks. PloS one 2015; 10, e0135958.
- 16. Ward NE, Pellis NR, Risin SA, Risin D, Gene expression alterations in activated human T-cells induced by modeled microgravity. J Cell Biochem 2006; 99, 1187-202.
- 17. Zhou M, Callaham JB, Reyes M, Stasiak M, Riva A, Zupanska AK, et al., Dissecting Low Atmospheric Pressure Stress: Transcriptome Responses to the Components of Hypobaria in Arabidopsis. Frontiers in plant science 2017; 8, 528.

- 18. Chopard A, Lecunff M, Danger R, Lamirault G, Bihouee A, Teusan R, et al., Large-scale mRNA analysis of female skeletal muscles during 60 days of bed rest with and without exercise or dietary protein supplementation as countermeasures. Physiological genomics 2009; 38, 291-302.
- 19. Choi S, Ray HE, Lai SH, Alwood JS, Globus RK, Preservation of Multiple Mammalian Tissues to Maximize Science Return from Ground Based and Spaceflight Experiments. PloS one 2016; 11, e0167391.
- Nguyen DH, Oketch-Rabah HA, Illa-Bochaca I, Geyer FC, Reis-Filho JS, Mao JH, et al., Radiation acts on the microenvironment to affect breast carcinogenesis by distinct mechanisms that decrease cancer latency and affect tumor type. Cancer cell 2011; 19, 640-51.
- 21. Missirian V, Conklin PA, Culligan KM, Huefner ND, Britt AB, High atomic weight, high-energy radiation (HZE) induces transcriptional responses shared with conventional stresses in addition to a core "DSB" response specific to clastogenic treatments. Frontiers in plant science 2014; 5, 364.
- 22. Camirand A, Goltzman D, Gupta A, Kaouass M, Panda D, Karaplis A, The Role of Parathyroid Hormone-Related Protein (PTHrP) in Osteoblast Response to Microgravity: Mechanistic Implications for Osteoporosis Development. PloS one 2016; 11, e0160034.
- 23. Neutelings T, Nusgens BV, Liu Y, Tavella S, Ruggiu A, Cancedda R, et al., Skin physiology in microgravity: a 3-month stay aboard ISS induces dermal atrophy and affects cutaneous muscle and hair follicles cycling in mice. NPJ microgravity 2015; 1, 15002.
- 24. Kwon T, Sparks JA, Nakashima J, Allen SN, Tang Y, Blancaflor EB, Transcriptional response of Arabidopsis seedlings during spaceflight reveals peroxidase and cell wall remodeling genes associated with root hair development. American journal of botany 2015; 102, 21-35.
- 25. Goossens KV, Ielasi FS, Nookaew I, Stals I, Alonso-Sarduy L, Daenen L, et al., Molecular mechanism of flocculation self-recognition in yeast and its role in mating and survival. mBio 2015; 6.
- 26. Visscher AM, Paul AL, Kirst M, Guy CL, Schuerger AC, Ferl RJ, Growth performance and root transcriptome remodeling of Arabidopsis in response to Mars-like levels of magnesium sulfate. PloS one 2010; 5, e12348.
- 27. Bhattacharya S, Srinivasan K, Abdisalaam S, Su F, Raj P, Dozmorov I, et al., RAD51 interconnects between DNA replication, DNA repair and immunity. Nucleic acids research 2017; 45, 4590-605.
- 28. Aceto J, Nourizadeh-Lillabadi R, Maree R, Dardenne N, Jeanray N, Wehenkel L, et al., Zebrafish Bone and General Physiology Are Differently Affected by Hormones or Changes in Gravity. PloS one 2015; 10, e0126928.
- 29. Casey T, Patel OV, Plaut K, Transcriptomes reveal alterations in gravity impact circadian clocks and activate mechanotransduction pathways with adaptation through epigenetic change. Physiological genomics 2015; 47, 113-28.
- *30.* Hateley S, Hosamani R, Bhardwaj SR, Pachter L, Bhattacharya S, Transcriptomic response of Drosophila melanogaster pupae developed in hypergravity. Genomics 2016; 108, 158-67.

- 31. Rettig TA, Ward C, Bye BA, Pecaut MJ, Chapes SK, Characterization of the naive murine antibody repertoire using unamplified high-throughput sequencing. PloS one 2018; 13, e0190982.
- 32. Herranz R, Larkin OJ, Hill RJ, Lopez-Vidriero I, van Loon JJ, Medina FJ, Suboptimal evolutionary novel environments promote singular altered gravity responses of transcriptome during Drosophila metamorphosis. BMC evolutionary biology 2013; 13, 133.
- 33. Herranz R, Larkin OJ, Dijkstra CE, Hill RJ, Anthony P, Davey MR, et al., Microgravity simulation by diamagnetic levitation: effects of a strong gradient magnetic field on the transcriptional profile of Drosophila melanogaster. BMC genomics 2012; 13, 52.
- 34. Samanta MP, Tongprasit W, Sethi H, Chin CS, Stolc V, Global identification of noncoding RNAs in Saccharomyces cerevisiae by modulating an essential RNA processing pathway. Proceedings of the National Academy of Sciences of the United States of America 2006; 103, 4192-7.
- 35. Coleman MA, Sasi SP, Onufrak J, Natarajan M, Manickam K, Schwab J, et al., Low-dose radiation affects cardiac physiology: gene networks and molecular signaling in cardiomyocytes. American journal of physiology Heart and circulatory physiology 2015; 309, H1947-63.
- 36. Beheshti A, Peluso M, Lamont C, Hahnfeldt P, Hlatky L, Proton irradiation augments the suppression of tumor progression observed with advanced age. Radiation research 2014; 181, 272-83.
- 37. Wage J, Ma L, Peluso M, Lamont C, Evens AM, Hahnfeldt P, et al., Proton irradiation impacts age-driven modulations of cancer progression influenced by immune system transcriptome modifications from splenic tissue. Journal of radiation research 2015; 56, 792-803.
- 38. Ritchie LE, Taddeo SS, Weeks BR, Lima F, Bloomfield SA, Azcarate-Peril MA, et al., Space Environmental Factor Impacts upon Murine Colon Microbiota and Mucosal Homeostasis. PloS one 2015; 10, e0125792.
- 39. Meador JA, Ghandhi SA, Amundson SA, p53-independent downregulation of histone gene expression in human cell lines by high- and low-let radiation. Radiation research 2011; 175, 689-99.
- 40. Wu F, Zhang R, Burns FJ, Gene expression and cell cycle arrest in a rat keratinocyte line exposed to 56Fe ions. Journal of radiation research 2007; 48, 163-70.
- 41. Beheshti A, Sachs RK, Peluso M, Rietman E, Hahnfeldt P, Hlatky L, Age and space irradiation modulate tumor progression: implications for carcinogenesis risk. Radiation research 2013; 179, 208-20.
- 42. Casero D, Gill K, Sridharan V, Koturbash I, Nelson G, Hauer-Jensen M, et al., Space-type radiation induces multimodal responses in the mouse gut microbiome and metabolome. Microbiome 2017; 5, 105.
- 43. Moritake T, Fujita H, Yanagisawa M, Nakawatari M, Imadome K, Nakamura E, et al., Strain-dependent damage in mouse lung after carbon ion irradiation. International journal of radiation oncology, biology, physics 2012; 84, e95-e102.

- 44. Tang J, Fernandez-Garcia I, Vijayakumar S, Martinez-Ruis H, Illa-Bochaca I, Nguyen DH, et al., Irradiation of juvenile, but not adult, mammary gland increases stem cell self-renewal and estrogen receptor negative tumors. Stem cells 2014; 32, 649-61.
- 45. Mizukami-Murata S, Iwahashi H, Kimura S, Nojima K, Sakurai Y, Saitou T, et al., Genomewide expression changes in Saccharomyces cerevisiae in response to high-LET ionizing radiation. Applied biochemistry and biotechnology 2010; 162, 855-70.
- 46. Ambrožová I, Brabcová KP, Kubančák J, Šlegl J, Tolochek RV, Ivanova OA, et al., Cosmic radiation monitoring at low-Earth orbit by means of thermoluminescence and plastic nuclear track detectors. Radiat Meas 2017; 106, 262-66.
- 47. Dachev TP, Tomov BT, Matviichuk YN, Dimitrov PG, Bankov NG, Shurshakov VV, et al., "BION-M" No. 1 spacecraft radiation environment as observed by the RD3-B3 radiometer-dosimeter in April–May 2013. J Atmospheric Sol-Terr Phys 2015; 123, 82-91.
- 48. Gambara G, Salanova M, Ciciliot S, Furlan S, Gutsmann M, Schiffl G, et al., Gene Expression Profiling in Slow-Type Calf Soleus Muscle of 30 Days Space-Flown Mice. PloS one 2017; 12, e0169314.
- 49. Gambara G, Salanova M, Ciciliot S, Furlan S, Gutsmann M, Schiffl G, et al., Microgravity-Induced Transcriptome Adaptation in Mouse Paraspinal longissimus dorsi Muscle Highlights Insulin Resistance-Linked Genes. Frontiers in physiology 2017; 8, 279.
- 50. Tascher G, Brioche T, Maes P, Chopard A, O'Gorman D, Gauquelin-Koch G, et al., Proteome-wide Adaptations of Mouse Skeletal Muscles during a Full Month in Space. Journal of proteome research 2017; 16, 2623-38.

FIGURE LEGENDS

- **Figure 1. The number of datasets on GeneLab over time.** A scatter plot representing the number of datasets on GeneLab (left y-axis) that increase over time starting from the creation of GeneLab in 2014. The right axis and red data points represent the number terrabytes (TB) per study for the datasets on GeneLab. The lines represent exponential fits to the data.
- **Figure 2. The type of data that is available on GeneLab. A**) The distribution of organism datasets available on GeneLab. **B**) The representation of ground and spaceflight experiments on GeneLab. **C**) The types of Omics assays that are available on GeneLab.
- **Figure 3.** The distribution of ground studies on GeneLab. A) The type of data on GeneLab associated with ground studies. B) The organism distribution of radiation ground study datasets available on GeneLab. C) The number of datasets associated with different ionizing radiation available on GeneLab.
- Figure 4. Distribution dose rate, dose, and LET for ionizing radiation ground studies on GeneLab. A) Distribution of dose rate (Gy/min) vs total dose (Gy) for all radiation ground studies available on GeneLab. Some GeneLab radiation datasets have missing dose rate information due to the lack of information available from the investigator. B) LET (KeV/ μ m) vs Total Dose (Gy) for all radiation ground study datasets on GeneLab.
- **Figure 5. Summary of spaceflight datasets available on GeneLab. A)** Image of the placement of the dosimeters on the space shuttle missions. **B)** Image of placement of dosimeters on BION-M1 mission. **C)** Distribution of total dose absorbed (mGy) per spaceflight mission for each

organism. This includes both space shuttle missions (STS) and BION-M1 missions. The mission number and time in space is provided next to each datapoint.









