



RESEARCH PAPER

The V-ATPase from etiolated barley (*Hordeum vulgare* L.) shoots is activated by blue light and interacts with 14-3-3 proteins

O. I. Klychnikov¹, K. W. Li², H. Lill³ and A. H. de Boer^{1,*}

¹ The 14-3-3 Biology Group, Department of Structural Biology, Faculty of Earth and Life Sciences, Vrije Universiteit, De Boelelaan 1087, 1081 HV Amsterdam, The Netherlands

² Department of Molecular and Cellular Neurobiology, Center of Neurogenomics and Cognitive Research, Faculty of Earth and Life Sciences, Vrije Universiteit, De Boelelaan 1085, 1081 HV Amsterdam, The Netherlands

³ Department of Structural Biology, Faculty of Earth and Life Sciences, Vrije Universiteit Amsterdam, The Netherlands

Received 1 September 2006; Revised 23 October 2006; Accepted 11 November 2006

Abstract

The vacuolar H⁺-ATPase (V-ATPase) is a key enzyme that controls the electrochemical proton potential across endomembranes. Although evidence suggests that V-ATPase is important for photo-morphogenesis, little is known about short-term regulation of V-ATPase upon initiation of the photo-morphogenetic programme by exposure of dark-grown plants to light. In this study, etiolated coleoptiles were given a short blue light treatment and V-ATPase characteristics were determined. The effectiveness of the light treatment was assessed by means of fusicoccin binding to the plasma membrane; this increased 5-fold. The short light treatment also induced a 2-fold to 3-fold increase in the hydrolytic activity of V-ATPase. Members of the 14-3-3 protein family are involved in both blue light perception and the subsequent activation of the P-type ATPase. We provide evidence that 14-3-3 proteins specifically interact with the catalytic A-subunit of the V-ATPase. First, the isolated V1-part of the V-ATPase co-purifies with 14-3-3 on a gel filtration column. Secondly, in an overlay experiment, 14-3-3 interacts with a 68 kDa band that was identified as the V1 A-subunit by mass spectrometry. Thirdly, in 14-3-3 affinity chromatography, both A- and B-subunits of the catalytic moiety of the V-ATPase were identified by matrix-assisted laser desorption ionization tandem time of flight mass spec-

trometry (MALDI TOF/TOF MS) as 14-3-3-interacting proteins. It was shown that the A-subunit can be phosphorylated *in vitro* by a tonoplast-bound kinase, whose properties are affected by blue light. Taken together, the data show that besides the P- and F-type H⁺-ATPases, the V-type H⁺-ATPase also interacts with 14-3-3 proteins.

Key words: 14-3-3 interaction, blue light, phosphorylation, V-ATPase.

Introduction

The vacuolar H⁺-ATPase (V-ATPase) is a proton-translocating enzyme, present in endomembranes of all eukaryotes. This includes the endoplasmic reticulum, the Golgi apparatus, lysosomes, the tonoplast, and intracellular vesicles that traffic between all these membranes (Dettmer *et al.*, 2006). The V-ATPase transports protons from the cytoplasm into the extraplasmic compartments, thereby creating the driving force for a variety of transport events of ions and metabolites. In this way, the pump has a general housekeeping function, but it is also very important for plant growth, development, and adaptation to environmental stress conditions (Schumacher *et al.*, 1999; Ratajczak, 2000; Dietz *et al.*, 2001). In view of its

* To whom correspondence should be addressed. E-mail: bert.de.boer@falw.vu.nl

Abbreviations: BSA, bovine serum albumin; DTT, dithiothreitol; FC, fusicoccin; [³H]FC, [³H]dihydrofusicoccin; HRP, horseradish peroxidase; MALDI TOF/TOF MS, matrix-assisted laser desorption ionization tandem time of flight mass spectrometry; Phot1, phototropin 1; PM, plasma membrane; PMSF, phenylmethylsulphonyl fluoride; PVPP, polyvinylpyrrolidene; TCA, trichloroacetic acid; V-ATPase, vacuolar H⁺-ATPase.

role in the regulation of cytoplasmic ion homeostasis and turgor pressure, it is not surprising that the activity of the pump changes in response to salt and drought stress, low and high temperature, acidification of the cytosol, and anoxia and plant hormone treatment (Dietz *et al.*, 2001).

V-ATPase is a tightly regulated proton pump. Several mechanisms at different levels and time spans have been recognized to control its functioning: regulation by gene expression, redistribution of the V-ATPase, modulation of the ATPase activity, and regulation of coupling of ATP hydrolysis to H⁺ pumping (Finbow and Harrison, 1997; Merzendorfer *et al.*, 1997; Dietz *et al.*, 2001). Adaptations on a long-term basis are achieved through variation of the V-ATPase gene expression, as is often seen in salt-stressed salt-tolerant plants, for example, salt-activated expression of subunits correlating with increased V-ATPase activity (Ratajczak *et al.*, 1994; Barkla *et al.*, 1995; Tsiantis *et al.*, 1996). For short-term variation, the V-ATPase is subjected to biochemical regulation by pH and ions (Cl⁻, NO₃⁻), ATP/ADP, and P_i, or oxidation/reduction. Relatively little is known thus far about another well-recognized mechanism to control enzyme activity on a short-term basis, i.e. phosphorylation/dephosphorylation. Liu *et al.* (2004) showed that phosphorylation increased the activity of the V-ATPase in vesicles isolated from maize root. The authors identified the A-subunit as being phosphorylated on a serine residue in a motif conserved in the A-subunit of many species (RLVSQK). In view of the amino acid sequence of this phosphorylated motif and the fact that the A-subunit of V-ATPases appears to be homologous with the β-subunit of the F-ATPase, it was hypothesized that the V-ATPase might interact with members of the so-called 14-3-3 family of proteins in a phosphorylation-dependent manner, just like the F-ATPase. Recently it was found, that the β-subunit of the plant F-ATPase can bind 14-3-3 proteins (Moorhead *et al.*, 1999; Bunney *et al.*, 2001). Both the mitochondrial and the chloroplast F-ATPase interact with 14-3-3, and the complex is more stable in the presence of phosphatase inhibitors. Binding of the 14-3-3 proteins to the F-ATPases resulted in inhibition of their activity (Bunney *et al.*, 2001). V-ATPases resemble F-ATPases; these pumps probably share common ancestors and show similarity in their complex structure and subunit composition (Nelson and Taiz, 1989; Nelson, 1992).

14-3-3 proteins are conserved regulatory proteins, ubiquitously present in the eukaryotic kingdom (Aitken, 2006). They are capable of binding to phosphorylated motifs, such as R/K-x-x(-x)-S/T-x-P, in target proteins (Muslin *et al.*, 1996; Yaffe *et al.*, 1997). In plants, 14-3-3 proteins also bind to the third type of H⁺-ATPases, the plasma membrane-located P-type ATPase, and form the so-called 'fusiccoccin receptor' (Korthout and de Boer, 1994; Marra *et al.*, 1994; Oecking *et al.*, 1994). Binding of 14-3-3 to the P-ATPase plays a role in plant adaptation to abiotic stresses such as low temperature and osmotic

stress via activation of the pump (Chelysheva *et al.*, 1999; Babakov *et al.*, 2000). Moreover, in a series of elegant experiments, it was shown that the P-ATPase is activated after a short (seconds) treatment by blue light (Kinoshita and Shimazaki, 1999, 2002; Kinoshita *et al.*, 2003). The activation mechanism involves (i) phosphorylation of the C-terminal domain of the P-ATPase; (ii) binding of 14-3-3 to that domain; and (iii) activation of the enzyme. It is worth noting that the blue light receptor involved in perception of the blue light signal and activation of the pump, phototropin 1 (Phot1), also binds 14-3-3 upon blue light activation (Kinoshita *et al.*, 2003). Intriguingly, the V-ATPase can also be involved in blue light responses. Mutation in one of the V-ATPase subunits in *Arabidopsis* (*det3*-mutation) results not only in lack of cell expansion and abolished stomata closure after blue light treatment, but also in a de-etiolated phenotype (Schumacher *et al.*, 1999; Allen *et al.*, 2000).

Thus, keeping in mind that P- and V-ATPases are involved in similar processes in the cell, such as generation of electrochemical potential differences across the plasma membrane and tonoplast, stabilization of cytoplasmic pH, turgor pressure control, and plant cell expansion, and the possibility of analogous regulation of the F- and V-ATPase, the question was addressed of whether 14-3-3 might interact with the V-type ATPase. Since it is well-known that 14-3-3 can act as an 'alarm regulator', changing enzymatic activity at short notice after a given stimulus (Kinoshita and Shimazaki, 1999; Sinnige *et al.*, 2005), blue light was used as a signal to change the activity of the V-ATPase. The P-type ATPase was monitored as a positive control for the action of blue light and regulation by 14-3-3 (Kinoshita and Shimazaki, 2002).

In this study, it is demonstrated that short treatment of etiolated barley coleoptiles with blue light results in activation of specific ATP hydrolysis by the V-ATPase. The results indicate that the V-ATPase hydrolytic part, V1, co-purifies with 14-3-3 in gel filtration experiments and can be pulled-down by 14-3-3 affinity chromatography. The interaction between 14-3-3 and the A-subunit of the V1 part of the ATPase was confirmed in overlay experiments. In an *in vitro* experiment, it is demonstrated that the A-subunit of the V-ATPase can be phosphorylated and the level of kinase activity increases after the blue light treatment.

Materials and methods

Plant material

Barley (*H. vulgare* L. cv. *Alexis*) seeds (300 g) were washed with tap water for 1 h, germinated overnight on wet tissue at 22 °C, then sown in soil and grown in the dark for 4 d at 20 °C. After that, the pots covered in aluminium foil were transferred to a room, illuminated with red light (Kodak GBX-2 safe light filter, intensity of 100 mE m⁻² s⁻¹). Coleoptiles (120 g) were cut with a razor

blade, gently mixed, split into two samples, and spread out in two Petri dishes. While one Petri dish was covered in aluminium foil, the second sample received a 10 min blue light treatment (Strand Filters Cinelux No 419 Dark Blue Primary λ_{\max} 450 nm) at a photon intensity of 50 mE m⁻² s⁻¹ superimposed on the background of red light.

Microsomal and tonoplast vesicles isolation

Tonoplast vesicles were isolated according to the protocol of Maeshima and Yoshida (1989) with minor modifications. In brief, 60 g of coleoptiles were homogenized in 120 ml of ice-cold buffer A [50 mM MES/TRIS pH 7.6, 250 mM sorbitol, 10 mM EDTA, 5 mM K₂S₂O₅, 5 mM dithiothreitol (DTT), 1 mM phenylmethylsulphonyl fluoride (PMSF), 1% polyvinylpolypyrrolidone (PVPP), 10 μ M cantharidin] with a Waring blender at the highest speed, three times for 30 s each with a 1 min interval. The homogenate was filtered through four layers of Miracloth, centrifuged at 6000 g for 10 min at 4 °C, the supernatant was centrifuged at 100 000 g for 30 min at 4 °C, and the pellet was resuspended in 15 ml of buffer B (10 mM K₂HPO₄/KOH pH 7.8, 500 mM sucrose, 1 mM EDTA, 2 mM DTT, 10 μ M cantharidin). The resuspended microsomes (5 ml) were layered under 5 ml of buffer C (5 mM MES/TRIS pH 7.3, 250 mM sorbitol, 1 mM DTT) in tubes for a swing-out rotor. Samples were centrifuged at 120 000 g for 35 min at 4 °C. The white disc of the interphase was carefully collected, diluted five times in buffer C, and pelleted by centrifugation at 100 000 g for 35 min at 4 °C. Samples were stored at -80 °C in aliquots.

Fusicoccin binding assay

Binding of [³H]dihydrofusicoccin ([³H]FC) to 50 μ g of the microsomal membrane fraction was carried out in 100 μ l of binding buffer (20 mM MES/TRIS pH 7.0, 5 mM MgCl₂, 1 mM CaCl₂, 2 mM DTT) at a concentration 60 nM [³H]FC. Microsomal membranes were incubated with [³H]FC for 30 min at room temperature, then samples were filtered through GF-C Millipore filters pre-charged with 1% polyethylenimine, and washed three times (5 ml each) with ice-cold 20 mM glycine/KOH (pH 9.5), dried in air and counted on an Rack Beta 1219 LKB counter in a liquid scintillator. Unspecific binding was measured in the presence of 10 μ M of unlabelled fusicoccin (FC).

Dissociation of V1-ATPase and gel chromatography

Dissociation of the soluble V1 domain of the V-ATPase was carried out according to the protocol of Muller *et al.* (1996). Tonoplast vesicles (230 μ g) were resuspended in 200 ml of dissociation buffer (10 mM MES/BTP pH 7.6, 20 mM KCl, 1 mM EDTA, 2 mM DTT, 0.1 mM PMSF, 5 mM ATP, 7 mM MgSO₄, 500 mM KNO₃, and 10 μ M cantharidin). Samples were incubated on ice for 1 h and spun down for 15 min at 400 000 g at 4 °C. The supernatant was applied on a Superdex-200 chromatography column equilibrated with gel filtration buffer (20 mM HEPES/KOH pH 7.5, 0.5 M NaCl, 1 mM MgCl₂, 10 μ M cantharidin). Fractions between the void volume and the salt peak were collected and precipitated overnight at 4 °C with 10% trichloroacetic acid (TCA), washed twice with cold acetone, and loaded on an SDS-polyacrylamide gel for western blotting of 14-3-3s and the E-subunit of the V-ATPase.

SDS-PAGE and western blotting assay

Tonoplast proteins (5 μ g) were separated by 12% SDS-PAGE (Laemmli, 1970) and transferred during 140 min from the gel onto a polyvinylidene fluoride (PVDF) membrane (Bio-Rad) in a semi-dry Transfer Cell (Bio-Rad) at 2 mA cm⁻² at room temperature (Towbin *et al.*, 1979). Next, the membrane was blocked overnight

at room temperature by 5% skimmed milk ('Marvel') dissolved in TRIS-buffered saline-Tween (TBS-T; 0.05% Tween-20). Antibodies against barley 14-3-3 isoforms A, B, C, D, and E were applied at a 1:50 000 dilution. As common 14-3-3 antibody, K-19 (Santa Cruz) at a dilution of 1:10 000 was used; antibodies against the E-subunit of the V-ATPase were diluted at 1:5000 (the antibodies were kindly provided by Dr KJ Dietz) in TBS-T buffer with 1% bovine serum albumin (BSA) (Sigma) and incubated for 1 h at room temperature with constant agitation; secondary anti-rabbit horseradish peroxidase (HRP) conjugate (Promega) at 1:3000 dilution was applied in TBS-T. The blot was developed with an ECL kit (Amersham), and ECL film (Amersham) was used for detection of the signal with an exposure time of 1.5 min.

V-ATPase activity assay

Hydrolysis of ATP by V-ATPase was measured according to LeBel *et al.* (1978) in 1 ml of reaction solution: 50 mM Tricine/KOH pH 7.5, 2 mM MgSO₄, 2 mM ATP. For each measurement, 10 μ g of tonoplast protein was used. The blank sample contained all components except the tonoplast membrane. Samples were incubated for 10 min at 30 °C. The reaction was stopped by adding 1 ml of LeBel stop mixture; the colour developed for 5 min at room temperature and development was stopped by adding 100 μ l of 34% Na₂-citrate. Samples were centrifuged at 10 000 g for 4 min, and absorption was measured at 750 nm. Bafilomycin A (50 nM) was added specifically to inhibit the V-ATPase. All measurements were done in triplicate.

Production of 14-3-3Cbio and 14-3-3CHis

The N-six-histidine or N-biotin-tagged 14-3-3 barley isoform C (accession number Y14200) was expressed in *Escherichia coli*. The bacterial cells were induced overnight with 1 mM isopropyl- β -D-thiogalactopyranoside (IPTG). Bacteria were disrupted with a French press in the presence of complete EDTA-free protease inhibitor cocktail (Roche). The lysate was centrifuged at 100 000 g for 35 min at 4 °C and the supernatant was filtered through a 0.45 μ m filter. 14-3-3CHis was purified on a HiTrap chelating column (Amersham) with a 20–500 mM imidazole gradient according to the manufacturer's protocol. 14-3-3Cbio was purified on a HiTrap-Q column (Amersham). Proteins were aliquoted and stored at -20 °C in the presence of the protease inhibitor cocktail.

14-3-3 affinity chromatography

A 10 mg aliquot of 14-3-3CHis was immobilized on a 5 ml Ni-NTA column. The column was equilibrated in running buffer 20 mM HEPES/KOH with 50 mM imidazole at pH 7.5. A fraction with soluble proteins from 140 g of coleoptiles was obtained according to the protocol of Moorhead *et al.* (1999). Briefly, the coleoptiles were homogenized in ice-cold buffer (50 mM MES/TRIS pH 7.6, 1 mM EDTA, 2 mM DTT, 1 mM PMSF, 0.1% PVPP, 10 μ M cantharidin, 1 mM Na₃VO₄). The homogenate was filtered through four layers of Miracloth and centrifuged at 6000 g for 10 min at 4 °C. The supernatant was supplied with 1 mM MgATP and pelleted with powdered (NH₄)₂SO₄ at a final concentration of 70%. Samples were centrifuged at 10 000 g for 10 min at 4 °C and the supernatant was discarded. Pelleted protein was dissolved in 35 ml of a buffer containing 20 mM HEPES/KOH pH 7.5, 1 mM PMSF, 1 μ M microcystin LR (Sigma), and dialysed against 20 mM HEPES/KOH pH 7.5. Dialysed protein was centrifuged at 32 000 g for 20 min at 4 °C, and the supernatant was used for 14-3-3 affinity chromatography. Soluble proteins (2 g) were filtered through a 0.45 μ m filter and pumped through the 14-3-3CHis column at a flow rate of 1 ml min⁻¹. To remove unspecifically

bound proteins, the column was washed with 100 ml of running buffer with 100 mM imidazole. Elution was performed in two steps: 100–200 mM and 200–600 mM of imidazole. Proteins from the second elution were collected and precipitated with 10 vols of cold acetone (–20 °C overnight).

14-3-3 overlay assay

Conditions for SDS–PAGE, the amount of protein, and blotting conditions were the same as those described for western blotting (see above). Blocking was performed with 5% BSA (Sigma) overnight at room temperature with constant agitation. 14-3-3Cbio (3 µg ml⁻¹) was incubated in TBS-T with 0.1% BSA (Sigma) with the membrane for 3 h at room temperature with constant agitation. To distinguish between specific and non-specific binding, excess 14-3-3Chis (3 mg ml⁻¹) was applied together with 14-3-3Cbio. After the membrane was washed three times with TBS-T buffer and avidin–HRP conjugate (Pierce), a concentration of 10 µg ml⁻¹ was incubated for 1 h. After washing three times with TBS-T, the signal was developed with an ECL kit (Amersham) under the same conditions as for western blotting.

[γ-³²P]ATP tonoplast protein labelling

A 20 µg aliquot of tonoplast membranes was incubated in labelling buffer (20 mM TRIS/MES pH 6.5, 250 mM sorbitol, 1 mM DTT, 2 mM Mg²⁺, 30 µM Ca²⁺, 15 mM NaF, 0.2 mg ml⁻¹ leupeptin, 10 µM cantharidin, 5 µM microcystin LR) with 0.23 µM [γ-³²P]ATP for 10, 30, and 60 min at 3 °C for the kinetic assay or a 10 min incubation with 0.23 µM [γ-³²P]ATP together with 0, 0.01, 0.1, 1, 10, 100, and 1000 µM of unlabelled ATP. The reaction was stopped by boiling for 3 min with SDS sample buffer. Sample proteins were separated by 12% SDS–PAGE. The gel was washed three times for 20 min each with acidic methanol (40% methanol, 10% acetic acid), followed by overnight incubation in TCA-PPi buffer (5% TCA, 1.5% Na-pyrophosphate) with constant agitation, and twice for 20 min each with water. After washing, the gel was stained by colloidal Coomassie, sealed in plastic, placed in a cassette of a phosphorimager, and incubated for 24 h.

In-gel digestion and MALDI TOF analysis of peptides

Protein bands were cut out from the colloidal Coomassie-stained gel. Trypsin in-gel digestion and peptide extraction were performed according to the protocol of Li *et al.* (2004). Samples of extracted peptides (0.5 µl) were mixed with 0.5 µl of α-cyano-4-hydroxycinnamic acid (5 mg ml⁻¹) and spotted on a target plate of an Applied Biosystems 4700 Proteomics Analyzer. A database search was performed using GPS Explorer V.3.0 against the Swiss-Prot green plant database with 100 ppm peptide mass tolerance, and with tandem mass spectrometry (MS/MS) tolerance of 0.2 Da. Common contaminants such as human keratin and trypsin peaks were excluded from the analysis.

Results

Blue light affects the P-ATPase

To examine the effectiveness of the blue light treatment, an investigation was carried out to determine whether blue light activates the plasma membrane (PM) P-type ATPase in etiolated coleoptiles as it does in guard cells (Goh *et al.*, 1995; Kinoshita and Shimazaki, 1999). As a marker for ATPase activation, the specific binding of FC to membranes isolated from blue light-treated coleoptiles was

measured. As high affinity FC binding sites have been widely used as a marker for plant PMs (Babakov *et al.*, 2000), it was decided to measure the binding directly on the microsomal fraction without further PM purification. Figure 1 shows a typical response of the plasma membrane FC receptor to blue light treatment (10 min 50 µE m⁻² s⁻¹) in the background of red light (100 µE m⁻² s⁻¹). After this treatment, the specific binding of FC increased about five times compared with coleoptiles treated only with red light. The increase in FC binding indicates that the number of 14-3-3–P-ATPase complexes increased and indirectly shows the activation of the P-ATPase in this experiment.

The V-ATPase activates ATP hydrolysis under blue light

To check whether the blue light treatment activates V-ATPase as it does with P-ATPase, tonoplast vesicles were purified from coleoptiles. Figure 2 presents the data of the V-ATP hydrolysis in the tonoplast fraction isolated from coleoptiles treated with blue light. The total tonoplast ATPase activity from control (red light as a background) coleoptiles was 360 nmol Pi mg⁻¹ protein min⁻¹. The specific V-ATPase activity was calculated by subtracting background hydrolysis (the same measurement in the presence 50 nM bafilomycin A1, a specific V-ATPase inhibitor) from the total ATPase activity (Fig. 2). This shows that the specific activity in the tonoplast was 300 nmol Pi mg⁻¹ protein min⁻¹, i.e. 85% of the total activity.

At the same time, blue light-treated plants showed a 2.5-fold to 3-fold increase in bafilomycin A1-sensitive ATPase activity, 950 nmol Pi mg⁻¹ protein min⁻¹. The background

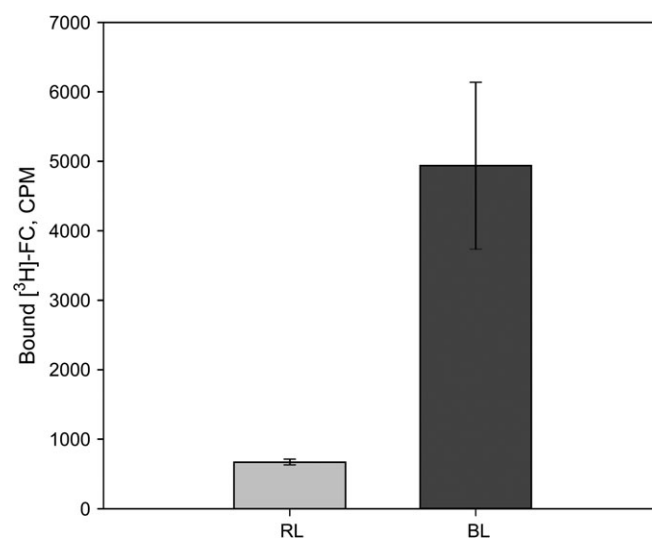


Fig. 1. Specific binding of radioactively labelled fusicoccin ([³H]FC) to microsomal membranes isolated from 4-d-old etiolated barley coleoptiles treated with red light (control, RL) and with combination of red and blue light (BL) (see Materials and methods). The average of four independent experiments (±SE) is shown.

activity in the presence of bafilomycin remained unchanged after the blue light treatment. This indicates that blue light activates only the hydrolytic activity of the tonoplast V-ATPase.

The amount of 14-3-3 bound to tonoplast vesicles increases after blue light treatment

In stomatal guard cells, blue light induces an increase in binding of 14-3-3 to the plasma membrane (Kinoshita and Shimazaki, 1999). To determine whether 14-3-3 proteins are involved in the blue light response of the plant vacuole as well, a western analysis of the tonoplast fraction isolated from coleoptiles was performed using 14-3-3 common and isoform-specific antibodies. The total amount of 14-3-3 on the tonoplast vesicles was estimated with K-19 antibodies (Santa Cruz), which can detect all five barley 14-3-3 isoforms (data not shown). Figure 3 shows representative results of a semi-quantitative western. First, quantification of the band intensity of three independent protein isolations shows that 14-3-3 is not a minor protein in the tonoplast fraction: the amount of 14-3-3 is $\sim 1.9 \pm 0.2 \mu\text{g mg}^{-1}$ of the total tonoplast protein (average \pm SE). Secondly, after the blue light treatment, the total amount of 14-3-3 associated with tonoplasts increases 1.8 times ($3.4 \pm 0.3 \mu\text{g mg}^{-1}$ of total tonoplast protein). The increase in the amount correlates with the increase in ATP hydrolysis by V-ATPase (Fig. 2).

To find out which of the 14-3-3 isoforms is involved in the blue light response, a western analysis was performed using isoform-specific antibodies raised against a synthetic peptide derived from the C-terminal part of each protein (Van den Wijngaard *et al.*, 2005). Figure 4 shows the results of a representative western blot. From three independent experiments, the optical density of the 14-3-3 bands was scanned and averaged, and the values of the blue light-treated samples were normalized to those of the respective controls (red light). These ratios are shown below the bands (Fig. 4) and only the 14-3-3A isoform shows a significant increase in density (3.9 times). In the same set of experiments and in the same fashion, the amount of the E-subunit of the V-ATPase was also measured with an antibody raised against the E-subunit (kind gift of Dr KJ Dietz; Golldack and Dietz, 2001). As shown in Fig. 4, no significant changes in the amount of E-subunit were observed. Sometimes, an extra band with an apparent molecular mass of 35 kDa was detected in the blue light treatment by the E-subunit antibodies; this may be due to a post-translational modification since it is known that phosphorylated proteins will migrate more slowly in an SDS-polyacrylamide gel (Lu *et al.*, 1994).

14-3-3 co-purifies with the V1-ATPase on the gel filtration column

Previously, it was shown that the structural and functional analogue of the V-ATPase, the F-ATPase, isolated from

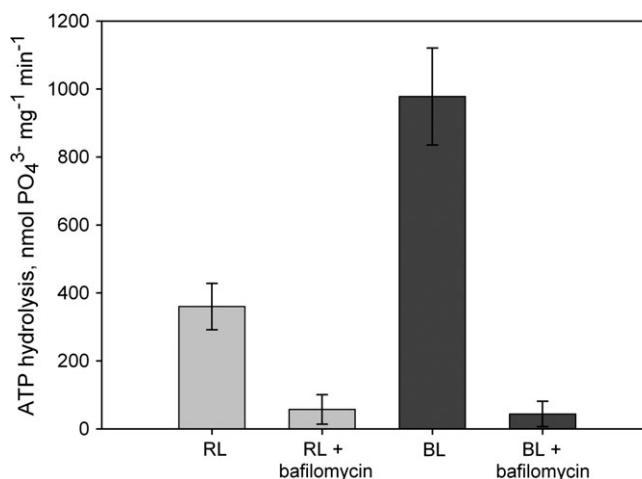


Fig. 2. ATP hydrolysis of tonoplast vesicles isolated from etiolated barley coleoptiles. After 10 min of light treatment, coleoptiles were homogenized and tonoplast membranes were isolated. The average ATPase activity of three independent experiments (\pm SE) of blue light (BL)-treated, and control (RL) plants is shown. Application of 50 nM bafilomycin A1, a specific V-ATPase inhibitor, shows a background activity, which is not related to V-ATPase.

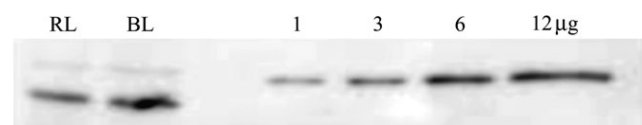


Fig. 3. Amount of 14-3-3 on the tonoplast membrane. Tonoplast proteins (5 μg) from control (red light, RL) and blue light (BL)-treated plants were separated by 12% SDS-PAGE and probed with common anti-14-3-3 antibodies. As a sample with a known concentration of the 14-3-3s (1, 3, 6, and 12 μg), the isoform of the recombinant 14-3-3CH3 was used.

barley chloroplasts or mitochondria, can be co-purified with 14-3-3 (Bunney *et al.*, 2001). To check whether 14-3-3 interacts and co-migrates with the V-ATPase holoenzyme, the V1 head structure was dissociated from the whole enzyme and run on a size exclusion column. As shown in Fig. 5, in fractions collected between the void column volume and salt peak, both 14-3-3 and the E-subunit of the V-ATPase were detected on western blot. The molecular mass of the V1-domain of the V-ATPase is 440 kDa, and column calibration showed that this corresponds to fraction number 4.

The A-subunit of the V-ATPase specifically interacts with 14-3-3 in an overlay assay

Next, the question of which subunit of the V-ATPase multisubunit complex interacts with 14-3-3 proteins was addressed. To do this, a so-called far-western or overlay assay with 14-3-3 was performed. In the assay, the tonoplast fraction and one of the 14-3-3 isoforms labelled with biotin (14-3-3Cbio) were used to detect 14-3-3-interacting partners. Bands were visualized with avidin-HRP conjugate

14-3-3A		14-3-3B		14-3-3C		14-3-3D		14-3-3E		V-ATPase E	
RL	BL	RL	BL	RL	BL	RL	BL	RL	BL	RL	BL
1.0	3.9±0.7	1.0	1.3±0.2	1.0	1.1±0.2	1.0	0.8±0.3	1.0	0.7±0.4	1.0	0.9±0.1

Fig. 4. 14-3-3 isoforms in the tonoplast isolated from control (red light, RL) and blue light (BL)-treated plants. A representative western blot quantification of the 14-3-3 band density and the E-subunit of the V-ATPase from three independent experiments is shown. Intriguingly, sometimes an extra 35 kDa band recognized by antibodies against the E-subunit appeared after the blue light treatment.

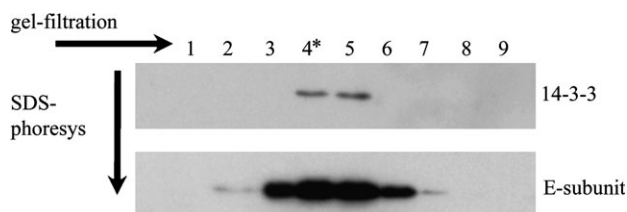


Fig. 5. 14-3-3 and the V1-ATPase co-purify on size exclusion chromatography. The 'head' structure of the V-ATPase was dissociated from tonoplast vesicles with high salt, and after that the sample was separated on the gel filtration column (Superdex-200). Western blotting of total 14-3-3 proteins and the E-subunit in the fractions was performed. Calibration of the column with the F_1 -moiety of the F_0F_1 -ATPase showed that 440 kDa proteins (the V1-ATPase molecular mass) eluted from the column in fraction number 4 together with 14-3-3.

using enhanced chemiluminescence (ECL). To show the specificity of the interaction, a control experiment was done where the 14-3-3Cbio overlay buffer was supplemented with a 1000-fold excess of unlabelled 14-3-3C (histidine-tagged, but this tag is not detected with avidin-HRP). Figure 6B shows that 14-3-3Cbio specifically recognizes one band at 68 ± 2 kDa of the tonoplast membrane fraction. This band is not seen in the control experiment with excess 14-3-3His (Fig. 6C). This band was excised from the gel, digested with trypsin, and the peptide mass fingerprint of the protein was determined by matrix-assisted laser desorption ionization tandem time of flight mass spectrometry (MALDI TOF/TOF-MS; model 4700 ABI). Figure 7 shows that this proved to be the A-subunit of the V-ATPase. No statistically significant difference between the 14-3-3 signal intensity in the control (red light) and the blue light-treated sample of the tonoplast fractions (Fig. 6) was seen, which may be due to the fact that the overlay with the 14-3-3C is not quantitative enough.

The V1-ATPase interacts with 14-3-3 on a 14-3-3 affinity column

To identify possible targets for 14-3-3 interaction after blue light treatment of etiolated barley shoots, 14-3-3 affinity chromatography of the 70% ammonium sulphate-precipitated protein fraction was performed. The protein fraction was applied to a 14-3-3 affinity column; non-specifically bound proteins interacting with the gel matrix were washed away with an imidazole gradient from 100 mM

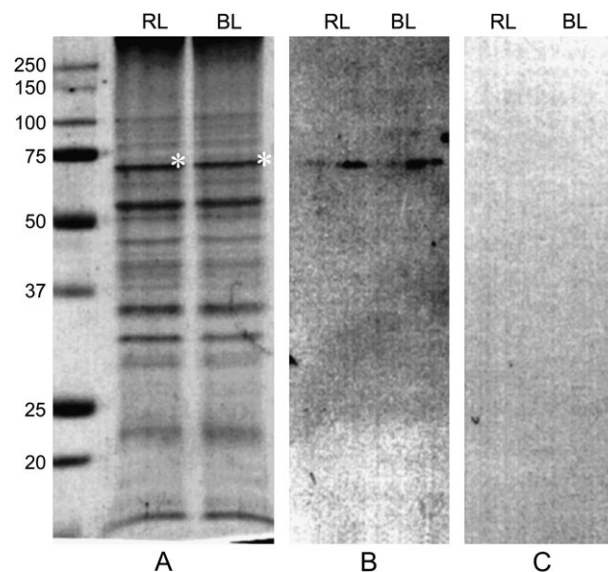


Fig. 6. Overlay of tonoplast proteins with 14-3-3bio. To find out which subunit of the V-ATPase interacts with 14-3-3, the so-called 'overlay' technique (far-western) was performed. Biotin-labelled 14-3-3 was used as a probe to find interacting 14-3-3 partners with the V-ATPase. To prove the specificity of the interaction, a 1000-fold excess of unlabelled 14-3-3 was used. (A) Coomassie-stained gel of the tonoplast fraction. (B) 14-3-3bio ($3 \mu\text{g ml}^{-1}$) was incubated for 3 h with tonoplast proteins on the blot. (C) 14-3-3bio ($3 \mu\text{g ml}^{-1}$) mixed with 14-3-3His (3 mg ml^{-1}), used as a competitor for specific binding. The interacting 69 kDa band (labelled with an asterisk) was shown to be the A-subunit of the V-ATPase with MS mass fingerprinting (see Fig. 7).

to 200 mM, and finally 14-3-3 protein together with interacting proteins were stripped from the column in a 200–600 mM imidazole gradient. The eluted proteins were identified by MALDI TOF/TOF MS (Fig. 8) and compiled into the list of proteins presented in Table 1. Besides a number of other interesting proteins, the core subunits of the V1-ATPase catalytic moiety were clearly identified, i.e. both the A- and the B-subunit. This corroborates the outcome of the gel filtration and overlay experiment and provides further evidence for the conclusion that the tonoplast V-ATPase in coleoptiles interacts with members of the 14-3-3 family.

The A-subunit can be phosphorylated in vitro

Although there are exceptions to the rule, the paradigm of 14-3-3 interaction states that a 14-3-3 target protein

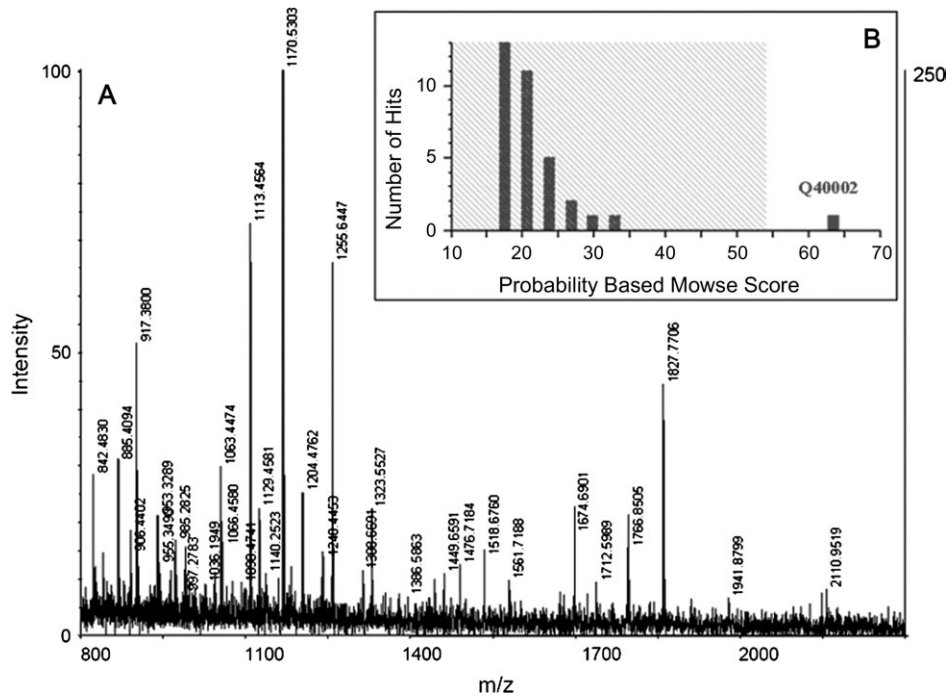


Fig. 7. Mass fingerprint identification of the protein from the SDS-PAGE 69 kDa band. (A) The MALDI-TOF spectrum of the trypsin-treated 69 kDa band. (B) Probability-based Mowse score graph. The spectrum was analysed with Mascot Peptide Mass Fingerprint (<http://www.matrixscience.com>) against the Swiss-Prot green plant database with 100 ppm mass peptide tolerance. Only one protein with a significant score of 64 (probability $P < 0.05$; an eight peptide match; 16% of protein coverage) was identified as the catalytic A-subunit of the V-ATPase (Q40002, EC 3.6.3.14).

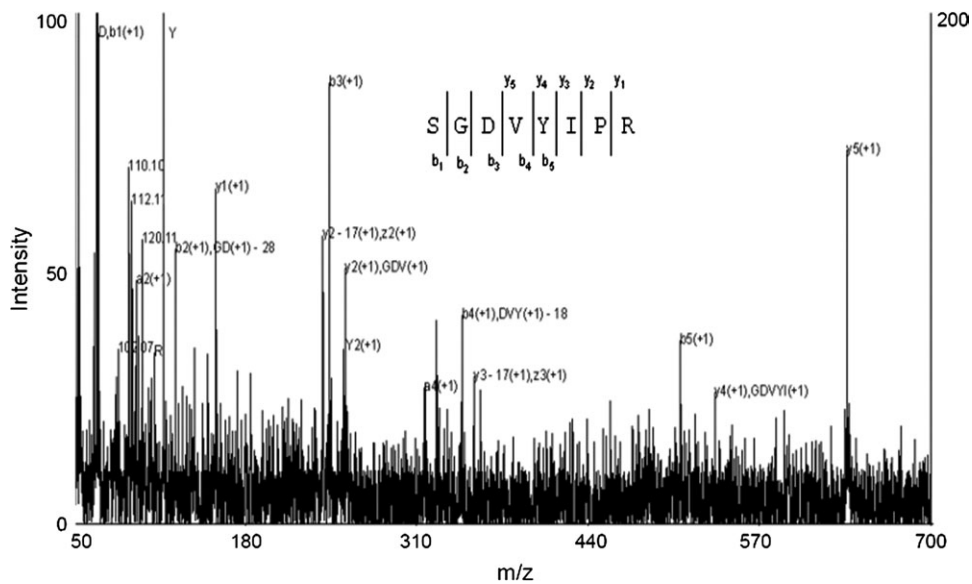


Fig. 8. Fragmentation spectrum of the peptide SGDVIYIPR of the V-ATPase A-subunit Q41775 (EC 3.6.3.14). The peptide was identified in the eluate after 14-3-3 affinity chromatography. The analysis was performed with 0.1 Da mass tolerance. A Mascot search against the Swiss-Prot green plant database gave total an ion score of 37 with 100% confidence interval for this peptide.

contains a conserved 14-3-3 binding motif, that will be phosphorylated on a serine or threonine residue for high-affinity interaction (Van Heusden, 2005). The A-subunit of the barley V-ATPase potentially can bind 14-3-3 because it has putative 14-3-3 binding motifs that are conserved in

many homologous proteins (RGVS⁸³VP and RLVS⁵⁴⁴QK; numbering according to alignment with the full-length A-subunit from *Hordeum*; accession no. Q40002). The latter peptide was shown to be phosphorylated in the maize homologue (Liu *et al.*, 2004). To address the question of

Table 1. List of proteins eluted from the 14-3-3 affinity column and identified with the Swiss-Prot green plant database

The search of the proteins was performed with 100 ppm precursor peptides mass tolerance and 0.2 Da MS/MS tolerance.

Protein name	Accession no.	Protein mol. wt, Da	Protein pI	Peptide count	Total ion score	Total ion score CI %	14-3-3 binding motif
V-ATPase 69 kDa subunit	VATA_MAIZE	61 952	5.9	2	37	100	Yes
Putative disease resistance protein	DRL38_ARATH	101 219	5.6	2	27	99	Yes
Exopolysaccharonase clone GBGE184 precursor	PGLR1_ARATH	44 402	8.5	1	27	99	No
Malate dehydrogenase, glyoxysomal precursor	MDHG_ORYSA	37 362	8.1	1	26	98	No
Zinc finger protein	CONS_ARATH	41 960	6.5	2	23	97	Yes
Chalcone synthase	CHS1_CASGL	42 576	6.0	1	23	97	No
Ribulose bisphosphate carboxylase small chain, chloroplast precursor	RBS_AEGTA	19 504	9.1	1	21	95	Yes
ATP synthase B chain	ATPF_MARPO	21 068	7.9	1	21	95	No
ADP, ATP synthase	ADT_CHLKE	36 663	9.9	1	21	95	No
Cell division control protein 48 homologue B	CD48B_ARATH	65 868	6.6	1	21	95	Yes
Kunitz-type trypsin inhibitor beta chain	ID5B_PROJU	4310	9.2	1	19	93	Yes
Ribulose bisphosphate carboxylase/oxygenase activase A, chloroplast precursor	RCAA_HORVU	51 041	8.0	1	19	92	No
Heat shock cognate 70 kDa protein 2	HSP72_LYCES	70 663	5.1	1	19	92	Yes
Maturase K (intron maturase)	MATK_MIMAU	60 363	9.9	1	16	86	Yes
Vacuolar ATP synthase subunit B isoform 2	VATB2_ACEAT	54 680	5.15	1	11	70	No

whether the barley V-ATPase can be phosphorylated, *in vitro* labelling with $[\gamma\text{-}^{32}\text{P}]\text{ATP}$ was carried out. Integral protein kinase activity, both Ca^{2+} and calmodulin independent, is expected to be present in purified tonoplast membranes, as shown by Chen *et al.* (2002). The tonoplast fraction was mixed with $\text{Mg}\text{-}[\gamma\text{-}^{32}\text{P}]\text{ATP}$ in the presence of phosphatase inhibitors and 10 μM free Ca^{2+} (Liu *et al.*, 2004). Figure 9 shows several phosphorylated bands, with the strongest labelling shown by a 69 kDa and a 55 kDa band. The peptides of these bands were identified with MALDI TOF/TOF MS as the A- and the B-subunit of the V-ATPase, respectively (data not shown). Phosphorylation occurs very rapidly: already after 10 min the bands are visible, and further incubation with $[\gamma\text{-}^{32}\text{P}]\text{ATP}$ did not increase the intensity. At the same time, no significant difference between control and blue light-treated samples was seen. However, it is possible that *in vitro* assay light-induced phosphorylation of the subunit is rapidly saturated. A competition assay was performed to reveal possible differences in the kinase activity after blue light treatment. The same amounts of tonoplast vesicles were incubated with a fixed amount of $[\gamma\text{-}^{32}\text{P}]\text{ATP}$ and increasing amounts of unlabelled ATP. If the blue light affects the affinity of the kinase for ATP as compared with the control treatment, differences in the competition curve are to be expected. Phosphorylation of the 69 kDa band was quantified with a phosphoimager and plotted against the amount of unlabelled ATP (Fig. 10). Blue light treatment shifted the curve about two orders of magnitude, which means that 100-fold more unlabelled ATP was necessary to reduce labelling by 50% as compared with the control (red light). Therefore, blue light treatment can stimulate phosphorylation of the A-subunit affecting the activity of the kinases.

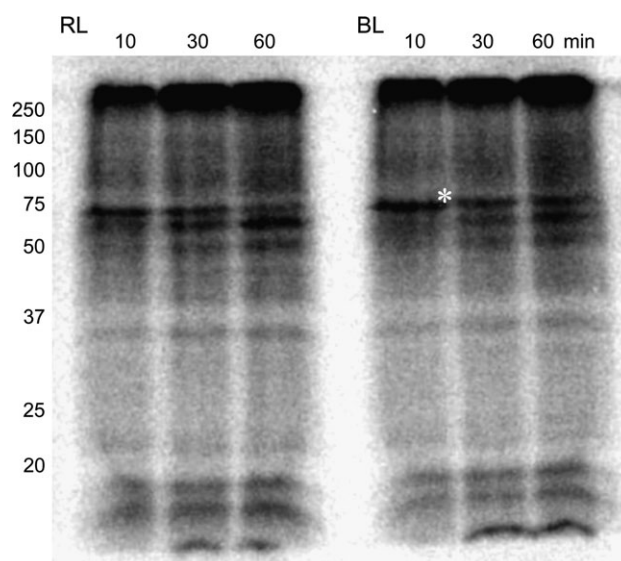


Fig. 9. *In vitro* labelling of tonoplast proteins with $[\gamma\text{-}^{32}\text{P}]\text{ATP}$. Tonoplast vesicles (20 μg per lane) from the control (red light, RL) and the blue light (BL)-treated coleoptiles were mixed with 0.23 μM $[\gamma\text{-}^{32}\text{P}]\text{ATP}$, incubated for 10, 30, and 60 min, boiled, and run on an SDS-polyacrylamide gel. The gel was exposed on a screen of a phosphoimager for 24 h. The band labelled with an asterisk was identified by mass fingerprinting as the A-subunit of the V-ATPase.

Discussion

Tonoplast-associated 14-3-3 proteins

Although 14-3-3 proteins have previously been shown to down-regulate the activity of the slow vacuolar channels located on the tonoplast (Van den Wijngaard *et al.*, 2001), they have not been reported to be co-isolated with the tonoplast before. In the present work, the tonoplast was

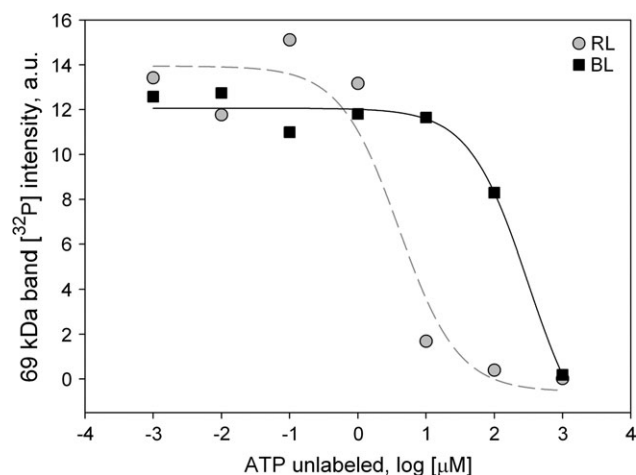


Fig. 10. *In vitro* labelling of tonoplast proteins with $[\gamma\text{-}^{32}\text{P}]\text{ATP}$: the competition assay. A 10 μg aliquot of tonoplast protein was mixed with 0.15 μM $[\gamma\text{-}^{32}\text{P}]\text{ATP}$ and 0, 0.01, 0.1, 1, 10, 100, and 1000 μM of unlabelled ATP, incubated for 10 min, separated by 12% SDS-PAGE and exposed on a phosphorimager screen for 4 d. The 69 kDa band of the tonoplast control (RL) and blue light (BL)-treated samples was quantified.

isolated in the presence of phosphatase inhibitors which increases the stability of the 14-3-3–target complexes because phosphorylation increases the affinity. Quantification of western blotting data shows that 14-3-3 is not a minor protein in the tonoplast fraction (Fig. 3). An estimation shows that 0.2% of tonoplast-associated 14-3-3s (60 kDa as a dimer) is enough to regulate up to 20% of the V-ATPase (600 kDa the holoenzyme) in the case of formation of a complex between 14-3-3 and the V-ATPase. After blue light treatment, the amount of 14-3-3 on the tonoplast vesicles increased almost 2-fold. It should be noted that not all 14-3-3 isoforms react with their targets in a similar fashion. Only one isoform, 14-3-3A, responded to blue light treatment with a significant increase in the amount of tonoplast bound (>3.5-fold). A similar increase in 14-3-3 protein binding to the plasma membrane after a blue light impulse has been shown for a plasma membrane fraction isolated from guard cells (Kinoshita and Shimazaki, 1999). Interestingly, in the case of the effect of blue light on guard cells of *Vicia faba*, isoform-specific binding of vf14-3-3a (81% homology with 14-3-3A on the protein level) to the plasma membrane has also been observed (Emi *et al.*, 2001).

14-3-3 interaction with the V-ATPase

The V-ATPase as a key enzyme has to be tightly regulated in several different ways involving different mechanisms of regulation. In its multimeric structure, the V-ATPase contains regulatory subunits that influence enzyme activity. However, there are indications that regulation can be carried out by proteins that are not conventionally included in the holoenzyme. In mammals, two proteins were discovered that activate the V-ATPase:

a 35 kDa cytosolic activator from bovine kidney (Zhang *et al.*, 1992) and a 6 kDa membrane-bound activator from bovine brain (Xie *et al.*, 1993). Thus, the V-ATPase can be regulated not only autonomously, but ‘external’ regulatory subunits might be involved.

In our view, 14-3-3 proteins are good candidates to have such an activator function after blue light treatment of barley etiolated coleoptiles. 14-3-3s have already been reported to be important regulators of both the P-ATPase (Korthout and de Boer, 1994; Marra *et al.*, 1994; Oecking *et al.*, 1994) and the F-ATPases (Bunney *et al.*, 2001). Moreover, 14-3-3 proteins bind very rapidly to the P-ATPase in response to a blue light pulse (Kinoshita and Shimazaki, 1999). Evidence has been provided for direct interaction of 14-3-3 proteins with the V1-holoenzyme complex and specific interaction with one of the subunits, the A-subunit of the V-ATPase, in the gel filtration assay (Fig. 5) and the 14-3-3 overlay assay (Fig. 6). Affinity chromatography of soluble coleoptile proteins shows that both the V1-ATPase core subunits A and B can be retrieved with 14-3-3 after blue light treatment (Table 1). Phosphorylation of 14-3-3 target proteins, needed for interaction of 14-3-3s with their partners, occurs in the case of the V-ATPase on the catalytic A-subunit of the V-ATPase (Liu *et al.*, 2004; Fig. 9). Blue light also increases the level of kinase activity, which results in higher concentrations of ATP to compete with the 68 kDa band phosphorylation (Fig. 9B).

Co-ordinated regulation of the P- and V-ATPases under blue light

What might be happening at very early stages of blue light treatment at the cellular level in etiolated coleoptiles? Hager (1996) reported that in maize coleoptiles, blue light activates a photoreceptor kinase localized in the plasma membrane, which then phosphorylates the P-ATPase. This receptor kinase is probably encoded by *phot1*, since it was recently shown that Phot1 mediates the rapid inhibition of *Arabidopsis* stem growth by blue light (Folta and Spalding, 2001). Upon blue light activation, Phot1 itself interacts with 14-3-3 proteins (Kinoshita *et al.*, 2003), and subsequently rapid phosphorylation of the C-terminal autoinhibitory domain of the P-ATPase results in binding of 14-3-3 and activation. Activation of the H⁺-ATPase will hyperpolarize the membrane potential, which can result in the activation of hyperpolarization-activated K⁺ and Ca²⁺ channels. At the same time, the blue light treatment results in increased expression of inward-rectifying K⁺ channels (Fuchs *et al.*, 2003), activation of chloride channels (Noh and Spalding, 1998), and activation of voltage-dependent and Ca²⁺-permeable channels (Stoelzle *et al.*, 2003). Therefore, blue light strongly affects the machinery responsible for ion homeostasis and in this light it is not surprising that the vacuolar H⁺-ATPase is

also involved in blue light regulation. For example, blue light-induced Ca^{2+} influx in the cytoplasm (Stoelzle *et al.*, 2003) can activate a kinase located on the tonoplast (Chen *et al.*, 2002), phosphorylate the A-subunit of the V-ATPase, followed by 14-3-3 binding and activation of the whole holoenzyme. The increase in the electrochemical gradient across the tonoplast can then drive secondary active transport of other ions and solutes across the tonoplast, necessary for the proper photomorphogenic response.

Acknowledgements

We wish to thank Dr HS van Walraven for valuable discussions, and Dr KJ Dietz for the gift of antibodies against the E-subunit of the V-ATPase. We are grateful to NWO/CW (grant number 700.50.030) for financial support to OK and AH de B.

References

- Aitken A. 2006. 14-3-3 proteins: a historic overview. *Seminars in Cancer Biology* **16**, 162–172.
- Allen GJ, Chu SP, Schumacher K, *et al.* 2000. Alteration of stimulus-specific guard cell calcium oscillations and stomatal closing in *Arabidopsis det3* mutant. *Science* **289**, 2338–2342.
- Babakov AV, Chelysheva VV, Klychnikov OI, Zorinyanz SE, Trofimova MS, De Boer AH. 2000. Involvement of 14-3-3 proteins in the osmotic regulation of H^+ -ATPase in plant plasma membranes. *Planta* **211**, 446–448.
- Barkla BJ, Zingarelli L, Blumwald E, Smith J. 1995. Tonoplast Na^+/H^+ antiport activity and its energization by the vacuolar H^+ -ATPase in the halophytic plant *Mesembryanthemum crystallinum* L. *Plant Physiology* **109**, 549–556.
- Bunney TD, van Walraven HS, de Boer AH. 2001. 14-3-3 protein is a regulator of the mitochondrial and chloroplast ATP synthase. *Proceedings of the National Academy of Sciences, USA* **98**, 4249–4254.
- Chelysheva VV, Smolenskaya IN, Trofimova MC, Babakov AV, Muromtsev GS. 1999. Role of the 14-3-3 proteins in the regulation of H^+ -ATPase activity in the plasma membrane of suspension-cultured sugar beet cells under cold stress. *FEBS Letters* **456**, 22–26.
- Chen S, Chen J, Wang XC. 2002. Existence and characteristics of tonoplast-bound protein kinase in the tip cell of maize root. *Acta Botanica Sinica* **44**, 661–666.
- Dettmer J, Hong-Hermesdorf A, Stierhof YD, Schumacher K. 2006. Vacuolar H^+ -ATPase activity is required for endocytotic and secretory trafficking in *Arabidopsis*. *The Plant Cell* **18**, 715–730.
- Dietz KJ, Tavakoli N, Kluge C, Mimura T, Sharma SS, Harris GC, Chardonnens AN, Gollmack D. 2001. Significance of the V-type ATPase for the adaptation to stressful growth conditions and its regulation on the molecular and biochemical level. *Journal of Experimental Botany* **52**, 1969–1980.
- Emi T, Kinoshita T, Shimazaki K. 2001. Specific binding of v14-3-3a isoform to the plasma membrane H^+ -ATPase in response to blue light and fusicoccin in guard cells of broad bean. *Plant Physiology* **125**, 1115–1125.
- Finbow ME, Harrison MA. 1997. The vacuolar H^+ -ATPase: a universal proton pump of eukaryotes. *Biochemical Journal* **324**, 697–712.
- Folta KM, Spalding EP. 2001. Unexpected roles for cryptochrome 2 and phototropin revealed by high-resolution analysis of blue-light-mediated hypocotyl growth inhibition. *The Plant Journal* **26**, 471–478.
- Fuchs I, Philippar K, Ljung K, Sandberg G, Hedrich R. 2003. Blue light regulates an auxin-induced K^+ -channel gene in the maize coleoptile. *Proceedings of the National Academy of Sciences, USA* **100**, 11795–11800.
- Goh CH, Oku T, Shimazaki K. 1995. Properties of proton-pumping in response to blue-light and fusicoccin in guard-cell protoplasts isolated from adaxial epidermis of *Vicia* leaves. *Plant Physiology* **109**, 187–194.
- Gollmack D, Dietz KJ. 2001. Salt-induced expression of the vacuolar H^+ -ATPase in the common ice plant is developmentally controlled and tissue specific. *Plant Physiology* **125**, 1643–1654.
- Hager A. 1996. Properties of a blue-light-absorbing photoreceptor kinase localized in the plasma membrane of the coleoptile tip region. *Planta* **198**, 294–299.
- Kinoshita T, Emi T, Tominaga M, Sakamoto K, Shigenaga A, Doi M, Shimazaki K. 2003. Blue-light- and phosphorylation-dependent binding of a 14-3-3 protein to phototropins in stomatal guard cells of broad bean. *Plant Physiology* **133**, 1453–1463.
- Kinoshita T, Shimazaki K. 1999. Blue light activates the plasma membrane H^+ -ATPase by phosphorylation of the C-terminus in stomatal guard cells. *EMBO Journal* **18**, 5548–5558.
- Kinoshita T, Shimazaki K. 2002. Biochemical evidence for the requirement of 14-3-3 protein binding in activation of the guard-cell plasma membrane H^+ -ATPase by blue light. *Plant and Cell Physiology* **43**, 1359–1365.
- Korthout HA, de Boer AH. 1994. A fusicoccin binding protein belongs to the family of 14-3-3 brain protein homologs. *The Plant Cell* **6**, 1681–1692.
- Laemmli UK. 1970. Cleavage of structural proteins during the assembly of the head of bacteriophage T4. *Nature* **227**, 680–685.
- LeBel D, Poirier GG, Beaudoin AR. 1978. A convenient method for the ATPase assay. *Analytical Biochemistry* **85**, 86–89.
- Li KW, Hornshaw MP, van Der Schors RC, *et al.* 2004. Proteomics analysis of rat brain post-synaptic density. Implications of the diverse protein functional groups for the integration of synaptic physiology. *Journal of Biological Chemistry* **279**, 987–1002.
- Liu GS, Chen S, Chen J, Wang XC. 2004. Identification of the phosphorylation site of the V-ATPase subunit a in maize roots. *Acta Botanica Sinica* **46**, 428–435.
- Lu G, Sehnke PC, Ferl RJ. 1994. Phosphorylation and calcium binding properties of an *Arabidopsis* GF14 brain protein homolog. *The Plant Cell* **6**, 501–510.
- Maeshima M, Yoshida S. 1989. Purification and properties of vacuolar membrane proton-translocating inorganic pyrophosphatase from mung bean. *Journal of Biological Chemistry* **264**, 20068–20073.
- Marra M, Fullone MR, Fogliano V, Pen J, Mattei M, Masi S, Aducci P. 1994. The 30-kilodalton protein present in purified fusicoccin receptor preparations is a 14-3-3-like protein. *Plant Physiology* **106**, 1497–1501.
- Merzendorfer H, Graf R, Huss M, Harvey WR, Wiczorek H. 1997. Regulation of proton-translocating V-ATPases. *Journal of Experimental Biology* **200**, 225–235.
- Moorhead G, Douglas P, Cotelle V, *et al.* 1999. Phosphorylation-dependent interactions between enzymes of plant metabolism and 14-3-3 proteins. *The Plant Journal* **18**, 1–12.
- Muller M, Irkens-Kiesecker U, Rubinstein B, Taiz L. 1996. On the mechanism of hyperacidification in lemon. Comparison of the vacuolar H^+ -ATPase activities of fruits and epicotyls. *Journal of Biological Chemistry* **271**, 1916–1924.

- Muslin AJ, Tanner JW, Allen PM, Shaw AS.** 1996. Interaction of 14-3-3 with signaling proteins is mediated by the recognition of phosphoserine. *Cell* **84**, 889–897.
- Nelson N.** 1992. Evolution of organellar proton-ATPases. *Biochimica et Biophysica Acta* **1100**, 109–124.
- Nelson N, Taiz L.** 1989. The evolution of H⁺-ATPases. *Trends in Biochemical Sciences* **14**, 113–116.
- Noh B, Spalding EP.** 1998. Anion channels and the stimulation of anthocyanin accumulation by blue light in *Arabidopsis* seedlings. *Plant Physiology* **116**, 503–509.
- Oecking C, Eckerskorn C, Weiler EW.** 1994. The fusicoccin receptor of plants is a member of the 14-3-3 superfamily of eukaryotic regulatory proteins. *FEBS Letters* **352**, 163–166.
- Ratajczak R.** 2000. Structure, function and regulation of the plant vacuolar H⁺-translocating ATPase. *Biochimica et Biophysica Acta* **1465**, 17–36.
- Ratajczak R, Richter J, Luttge U.** 1994. Adaptation of the tonoplast V-type H⁺-ATPase of *Mesembryanthemum crystallinum* to salt stress, C-3-Cam transition and plant age. *Plant, Cell and Environment* **17**, 1101–1112.
- Schumacher K, Vafeados D, McCarthy M, Sze H, Wilkins T, Chory J.** 1999. The *Arabidopsis det3* mutant reveals a central role for the vacuolar H⁺-ATPase in plant growth and development. *Genes and Development* **13**, 3259–3270.
- Sinnige MP, Roobeek I, Bunney TD, Visser AJWG, Mol JNM, Boer AH.** 2005. Single amino acid variation in barley 14-3-3 proteins leads to functional isoform specificity in the regulation of nitrate reductase. *The Plant Journal* **44**, 1001–1009.
- Stoelzle S, Kagawa T, Wada M, Hedrich R, Dietrich P.** 2003. Blue light activates calcium-permeable channels in *Arabidopsis* mesophyll cells via the phototropin signaling pathway. *Proceedings of the National Academy of Sciences, USA* **100**, 1456–1461.
- Towbin H, Staehelin T, Gordon J.** 1979. Electrophoretic transfer of proteins from polyacrylamide gels to nitrocellulose sheets—procedure and some applications. *Proceedings of the National Academy of Sciences, USA* **76**, 4350–4354.
- Tsiantis MS, Bartholomew DM, Smith JAC.** 1996. Salt regulation of transcript levels for the c subunit of a leaf vacuolar H⁺-ATPase in the halophyte *Mesembryanthemum crystallinum*. *The Plant Journal* **9**, 729–736.
- Van den Wijngaard PW, Bunney TD, Roobeek I, Schonknecht G, de Boer AH.** 2001. Slow vacuolar channels from barley mesophyll cells are regulated by 14-3-3 proteins. *FEBS Letters* **488**, 100–104.
- Van den Wijngaard PWJ, Sinnige MP, Roobeek I, Reumer A, Schoonheim PJ, Mol JNM, Wang M, De Boer AH.** 2005. Abscisic acid and 14-3-3 proteins control K⁺ channel activity in barley embryonic root. *The Plant Journal* **41**, 43–55.
- Van Heusden GPH.** 2005. 14-3-3 proteins: regulators of numerous eukaryotic proteins. *IUBMB Life* **57**, 623–629.
- Xie XS, Crider BP, Stone DK.** 1993. Isolation of a protein activator of the clathrin-coated vesicle proton pump. *Journal of Biological Chemistry* **268**, 25063–25067.
- Yaffe MB, Rittinger K, Volinia S, Caron PR, Aitken A, Leffers H, Gamblin SJ, Smerdon SJ, Cantley LC.** 1997. The structural basis for 14-3-3: phosphopeptide binding specificity. *Cell* **91**, 961–971.
- Zhang K, Wang ZQ, Gluck S.** 1992. Identification and partial purification of a cytosolic activator of vacuolar H(+)-ATPases from mammalian kidney. *Journal of Biological Chemistry* **267**, 9701–9705.