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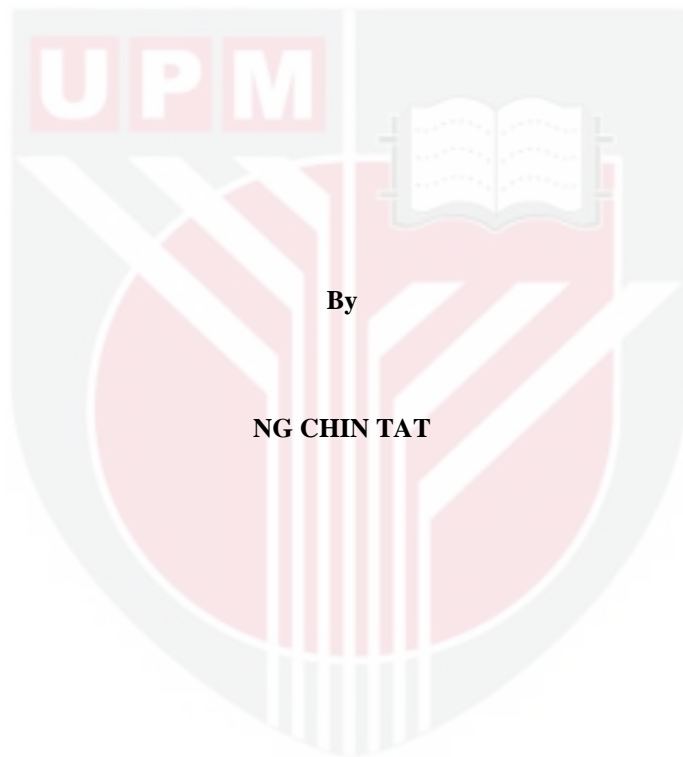
***EFFECTS OF SMALL MOLECULE INHIBITORS TARGETING PI3K, EGFR,
IGF-1R, MTOR, SMAD3 AND MEK IN TUBE FORMATION AND 3-D
SPHEROID ASSAY***

NG CHIN TAT

FPSK(m) 2014 51



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SPHEROID ASSAY**



By

NG CHIN TAT

**Thesis Submitted to the School of Graduate Studies, Universiti Putra Malaysia, in
Fulfilment of the Requirements for the Degree of Master of Science**

November 2014

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Abstract of thesis presented to the Senate of Universiti Putra Malaysia in fulfilment of the requirement for the degree of Master of Science

EFFECTS OF SMALL MOLECULE INHIBITORS TARGETING PI3K, EGFR, IGF-1R, MTOR, SMAD3 AND MEK IN TUBE FORMATION AND 3-D SPHEROID ASSAY

By

NG CHIN TAT

November 2014

Chairman: Professor Seow Heng Fong, PhD

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Angiogenesis is the process where new blood capillaries are formed from the preexisting blood vessels. The MEK/ERK and PI3K/Akt signaling pathways are involved in the processes that drive cancer progression such as cell motility, metastasis and angiogenesis. The cellular events in angiogenesis which include endothelial cell migration, invasion and differentiation contribute significant role to cancer metastasis. Inhibition of angiogenesis via disruption of signaling pathways appears to be a rational therapeutic strategy. Hence, there is need to investigate the role of PI3K-AKT and MEK-ERK signaling in microvascular endothelial cells. In this study, we investigated the role of PI3K/AKT and MEK/ERK pathway in two microvascular endothelial cell lines, namely, HMEC-1 (SV40-immortalized) and TIME (telomerase-immortalized). The specific objectives of the study were (i) to investigate the effect of blocking PI3K/AKT and MEK/ERK pathways on tube formation by HMEC-1 and TIME by using small molecules inhibitors targeting phosphoinositide 3-kinase (PI3K), epidermal growth factor receptor (EGFR), insulin-like growth factor I receptor (IGF-1R), mammalian target of rapamycin (mTOR), mothers against decapentaplegic homolog 3 (SMAD3), and mitogen-activated protein kinase kinase (MEK), (ii) to investigate the effect of blocking PI3K/AKT and MEK/ERK pathways on cell invasion by HMEC-1 and TIME in a 3-D spheroid invasion model by using small molecule inhibitors targeting PI3K, IGF-1R, EGFR, mTOR, Smad3, and MEK. and (iii) to investigate the effect of small molecule inhibitors targeting PI3K, EGFR, IGF-1R, mTOR, Smad3, and MEK on phosphorylation status of AKT and ERK by HMEC-1 and TIME. *In vitro* angiogenesis was examined using tube formation whereas the invasion properties were assessed using three-dimensional (3D) spheroid *in vitro* invasion assays. PD0325901 and NVP-AEW541 were able to inhibit tube formation by TIME cells in a dose-dependent manner but had no effect on HMEC-1. Western Blot showed MEK inhibitor PD0325901 and IGF-1R inhibitor NVP-AEW541 suppressed phosphorylation of ERK and AKT, respectively, in HMEC-1 and TIME cells. NVP-BKM 120 inhibited tube formation and suppressed phosphorylation of AKT in both cell lines in a dose-dependent manner. TIME spheroids

treated with inhibitors (NVP-AEW541, NVP-BKM120 and PD0325901) showed a significant reduction in invasion and a similar trend were observed in suppression of tube formation. However, treatment with PD0325901 showed inhibition in HMEC-1 spheroids invasion whereas there is no suppression in tube formation. HMEC-1 spheroids treated with inhibitors (NVP-BKM120, NVP-BKM120 combination with PD0325901) showed a significant reduction in invasion and a similar trend were observed in suppression of tube formation. This result suggested the different results obtained in response to inhibitors might be due to the different models chosen. In conclusion, our results indicated that tube formation of TIME cell was inhibited when MEK-ERK pathway and/or PI3K-AKT pathway was blocked. In contrast, the angiogenic activity of HMEC-1 cell was inhibited via blockade of PI3K-AKT pathway but not MEK-ERK pathway.



Abstrak tesis yang dikemukakan kepada Senat Universiti Putra Malaysia sebagai memenuhi keperluan untuk ijazah Sarjana Sains

KESAN-KESAN PERENCAT MOLEKUL KECIL MENSASARKAN PI3K, EGFR, IGF-1R, MTOR, SMAD3, DAN MEK PADA FORMASI TUBE DAN 3-D SFEROID ASSAL

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Angiogenesis merupakan satu proses di mana pembedakan kapilari darah baru daripada saluran darah yang wujud. Lintasan isyarat MER/ERK dan PI3K/AKT yang terlibat dalam proses yang mendorong progresi kanser seperti motiliti sel, metastasis, dan angiogenesis. Peristiwa-peristiwa sel-sel dalam angiogenesis termasuk migrasi, invasi dan pembezaan sel endothelial menyumbangkan perkembangan kepada metastasis kanser. Perencatan angiogenesis melalui gangguan laluan isyarat akan menjadi satu strategi terapeutik yang rasional. Oleh itu, pensiasatan peranan pada lintasan isyarat PI3K-AKT dan MEK-ERK dalam sel endothelial mikrovesikel adalah diperlukan. Objektif-objektif khusus kajian ini adalah (i) untuk mengkaji kesan menyekat laluan PI3K / AKT dan laluan MEK / ERK pada formasi tiub oleh HMEC-1 dan TIME dengan menggunakan perencat-*rtgtpcv* "o qngmwn" mgekn" o gpucuctmcp" ÷ r j q r j q k p q u k v k f g" 3-mkpcugø" *RK5M+." ÷ grkfgt o cn" i tqyv j" hcevqt" tgegrvqtø" *GIHT+." ÷ kpuwnkp-like growth hcevqt" K" tgegrvqtø" *KIH-3T+." ÷ o c o o cnkcp" vct igv" qh" tcrco {ekpø" *o VQT+." ÷ o qv jgtu" c ic k p u v" f g e c r g p v c r n g i k e" j q o q n q i" 5ø" *UOCF5+." fcp" ÷ o k v q i g p-activated protein kinase mkpcugø" *OGM+0" *kk+ wpvwm" o g p i m c l k" mgucp" o g p { g m c v" n c n w c p" RK5M" 1" CMV" fcp" n c n w c p" MEK / ERK pada invasi sel oleh HMEC-1 dan TIME dalam tiga dimensi (3D) sferoid assal dengan menggunakan perencat-perencat molekul kecil mensasarkan PI3K, IGF-1R, EGFR, mTOR, SMAD3, dan MEK dan (iii) untuk mengkaji kesan perencat-perencat molekul kecil mensasarkan PI3K, EGFR, IGF-1R, mTOR, SMAD3, and MEK terhadap status pemfosforilan AKT dan ERK pada HMEC-1 dan TIME. Angiogenesis *in vitro* diperiksa dengan menggunakan formasi tiub manakala sifat invasi dikaji dengan menggunakan tiga dimensi (3D) sferoid assal invasi *in-vitro*. PD0325901 dan NVP-AEW541 boleh merencatkan formasi tiub oleh sel-sel TIME dengan cara yang bergantung kepada dos tetapi tidak mempunyai kesan terhadap HMEC-1. Pemendapan Western menunjukkan PD0325901 perencat kepada MEK dan NVP-AEW541 perencat kepada IGF-1R menghalang pemfosforilan ERK dan AKT dalam sel-sel HMEC-1 dan TIME masing-masing. Perencat NVP-BKM120 menghalang pembentukan tiub dan pemfosforilan bagi AKT dalam kedua-dua jenis titisan sel dengan cara yang

bergantung kepada dos. Rawatan sferoid TIME dengan perencat-perencat (NVP-AEW541, NVP-BKM120 and PD0325901) menunjukkan pengurangan yang ketara dalam invasi dan trend yang sama diperhatikan dalam halangan bagi formasi tiub. Walau bagaimanapun, rawatan dengan PD0325901 menunjukkan perencatan dalam invasi terhadap sferoid HMEC-1 manakala tidak ada perencatan dalam tiub formasi. Rawatan sferoid TIME dengan perencat-perencat (NVP-BKM120, NVP-BKM120 kombinasi dengan PD0325901) menunjukkan pengurangan yang ketara dalam invasi dan trend yang sama diperhatikan dalam halangan bagi formasi tiub. Keputusan ini mencadangkan keputusan yang berbeza diperolehi dalam gerak balas terhadap perencat-perencat yang mungkin disebabkan oleh model kajian yang berlainan dipilih. Kesimpilannya, keputusan kami menunjukkan bahawa angiogenesis bagi sel-sel TIME telah direncatkan apabila laluan MER-ERK dan/atau laluan PI3K-AKT telah disekatkan. Sebaliknya, aktiviti angiogenik bagi sel HMEC-1 telah direncatkan melalui sekatan laluan PI3K-AKT tetapi bukan laluan MEK-ERK.



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This thesis was submitted to the Senate of Universiti Putra Malaysia and has been accepted as fulfilment of the requirement for the degree of Master of Science. The members of the Supervisory Committee were as follows:

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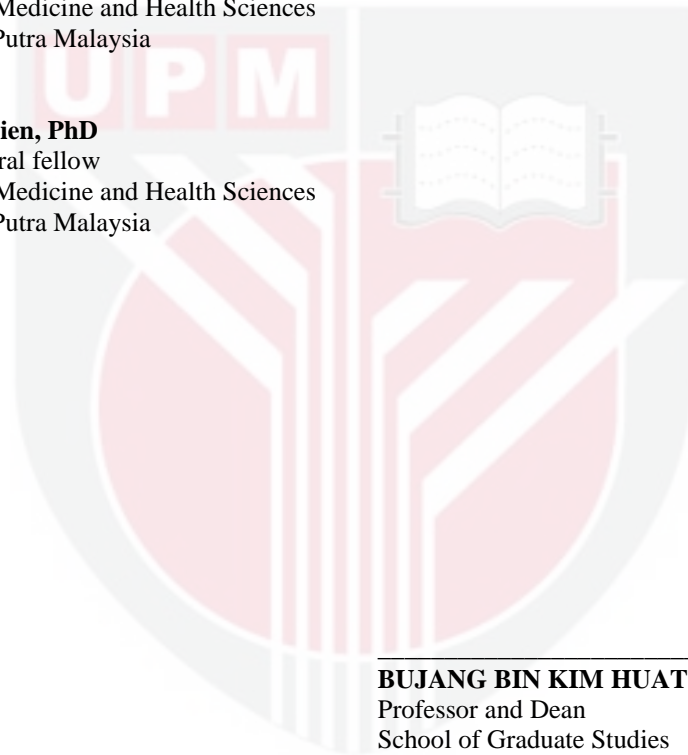
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LIST OF ABBREVIATIONS

%	Percent
μ	Micro
AKT	v-akt murine thymoma viral oncogene homolog
ATP	Adenosine triphosphate
bFGF	Basic fibroblast growth factor
BSA	Bovine serum albumin
CR	Conserved region
DAPI	6,8-diamidine-4-phenylindole
DMSO	Dimethyl sulfoxide
DTT	Dithiothreitol
ECM	Extracellular matrix
EGF	Epidermal growth factor
EGFR	Epidermal growth factor receptor
eNOS	Endothelial nitric oxide synthase
ERK	Extracellular signal-regulated kinase
FBS	Fetal bovine serum
FGFR	Fibroblast growth factor receptor
FITC	Fluorescein isothiocyanate
GDP	Guanosine diphosphate
Grb2	Growth factor receptor-bound protein 2
GPCR	G-protein-coupled receptor
GSK	Glycogen synthase kinase
GTP	Guanosine triphosphate
HER2	Human epidermal growth factor receptor 2
HIF-3	Hypoxia-inducible factor-1 alpha
H-Ras	v-Ha-ras Harvey rat sarcoma viral oncogene homolog
IC ₅₀	Half maximal inhibitory concentration
IGF-1	Insulin-like growth factor 1
IRS1/2	Insulin receptor substrate 1 or 2
JNK	c-Jun N-terminal kinase
MAPK	Mitogen-activated protein kinase
MAPKK	Mitogen-activated protein kinase kinase
MAPKKK	Mitogen-activated protein kinase kinase kinase
MEK1/2	MAPK/ERK kinase 1 or 2
mTOR	Mammalian target of rapamycin
mTORC2	Rapamycin-insensitive mTOR complex
MTT	3-(4,5-dimethylthiazol-2-yl)-2,5-diphenyl-2H-tetrazolium bromide
NF- NB	Nuclear factor- NB
NIH	National Institutes of Health
PBS	Phosphate buffered saline
PDGFR	Platelet-derived growth factor receptor
PDK1	3-Phosphoinositide-dependent kinase-1
PH domain	Pleckstrin homology domain
PI	Propidium iodide
PI(3,4)P ₂	Phosphatidylinositol 3,4-bisphosphate
PI(3,4,5)P ₃	Phosphatidylinositol 3,4,5-trisphosphate
PI(4,5)P ₂	Phosphatidylinositol 4,5-bisphosphate
PI3K	Phosphatidylinositol 3-kinase

PIK3CA	Phosphatidylinositol 3-kinase catalytic subunit
PIP ₂	Phosphatidylinositol 4,5-bisphosphate
PIP ₃	Phosphatidylinositol 3,4,5-trisphosphate
PKB	Protein kinase B
PTC	Papillary thyroid carcinoma
PTEN	Phosphatase and tensin homolog
Raf	v-raf-1 murine leukemia viral oncogene homolog
RPMI 1640	Roswell Park Memorial Institute 1640
RTKs	Receptor tyrosine kinases
SCLC	Small cell lung carcinoma
SD	Standard deviation
SDS	Sodium dodecyl sulfate
Ser	Serine
SH2	Src homology 2
SHIP	SH2 domain-containing inositol phosphatase
Src	v-src sarcoma (Schmidt-Ruppin A-2) viral oncogene homolog
TGF-	Transforming growth factor beta
Thr380	Threonine380
TSC1/2	Tuberous sclerosis complex 1/2
Tyr	Tyrosine
VEGFR	Vascular endothelial growth factor receptor

CHAPTER 1

INTRODUCTION

1.1 Background of Study

Angiogenesis is a process in which new blood vessels are formed (Mikirova, Casciari, & Riordan, 2010) and is characterized by a number of cellular events including endothelial cell migration, invasion and differentiation into capillaries. Oxygen and nutrients are delivered via blood vessels to every part of the body, and also nourish malignant tumors (Carmeliet & Jain, 2011). Tumor angiogenesis is a process based upon a sequence of interplay between endothelial cells and tumor cells. It plays an important role for local tumor progression and development of distant metastasis (Khodarev, 2003).

Active form of PI3K is an oncogene, and amplifications and mutations of PI3K are commonly found in many kinds of human cancers (Okumura *et al.*, 2012). In endothelial cells, the PI3K/AKT pathway is also involved in angiogenesis through its regulation of nitric oxide (NO) signaling (Karar & Maity, 2011) and endothelial nitric oxide synthase (eNOS) which are important in VEGF-induced angiogenesis and vascular permeability (Fukumura *et al.*, 2001). RAS/RAF/MEK/ERK pathway is deregulated in approximately one third of all human cancers (Neuzillet *et al.*, 2014) and is involved in the promotion of tumor growth, invasion, angiogenesis, and metastasis (Emuss & Marais, 2008).

Recently, many studies have revealed the importance of PI3K/AKT and MEK/ERK signaling in angiogenesis. However, the role of these two signaling pathways in microvascular endothelial cells is less-well studied. In our study, two microvascular endothelial cell, namely human dermal microvascular endothelial cell (HMEC-1) and telomerase-immortalized microvascular cell (TIME) were selected. HMEC-1 is immortalized via transfection with a plasmid containing SV40A gene and has been used for a wide range of application in endothelial cell research and drug development. To represent the population of microvascular endothelial cell, TIME is also included in this study. These cell lines are immortalized and provide a convenient model for *in vitro* studies. The human endothelial cell line, HUVEC, is not immortalized and dies off after a number of passages.

In our study, we aim to investigate the role of PI3K/AKT and MEK/ERK pathway in angiogenesis by using tube formation assay and three dimensional (3D) spheroid invasion assay with TIME and HMEC-1. Our hypothesis is that, blockade of PI3K/AKT and MEK/ERK signaling pathway with small molecule inhibitors are able to suppress tube formation, phosphorylation of AKT and ERK, and inhibit cell invasion in a collagen-embedded three dimensional (3D) model.

Tube formation assay is chosen for this study due to its simplicity, rapidness, quantitative, reliability and more comprehensive than other *in vitro* assays. 3D spheroid invasion assay closely mimics invasion *in vivo* due to invasion occurring from cell clusters with well-established cell–cell interactions rather than from single cells. Therefore, it is a good model to study HMEC-1 and TIME cell invasion.

1.2 Objectives of Study

The objectives of the study were:

1. To investigate the effect of blocking PI3K/AKT and MEK/ERK pathways on tube formation by HMEC-1 and TIME by using small molecules inhibitors targeting phosphoinositide 3-kinase (PI3K), epidermal growth factor receptor (EGFR), insulin-like growth factor I receptor (IGF-1R), mammalian target of rapamycin (mTOR), mothers against decapentaplegic homolog 3 (SMAD3), and mitogen-activated protein kinase kinase (MEK).
2. To investigate the effect of blocking PI3K/AKT and MEK/ERK pathways on cell invasion by HMEC-1 and TIME in a 3-D spheroid invasion model by using small molecules inhibitors targeting PI3K, IGF-1R, EGFR, mTOR, Smad3, and MEK.
3. To investigate the effect of small molecules inhibitors targeting PI3K, EGFR, IGF-1R, mTOR, Smad3, and MEK on phosphorylation status of AKT and ERK by HMEC-1 and TIME.

REFERENCES

- Ades, E. W., Candal, F. J., Swerlick, R. a, George, V. G., Summers, S., Bosse, D. C., & Lawley, T. J. (1992). HMEC-1: establishment of an immortalized human microvascular endothelial cell line. *The Journal of Investigative Dermatology*, **99(6)**, 683–90.
- Akinleye, A., Avvaru, P., Furqan, M., Song, Y., & Liu, D. (2013). Phosphatidylinositol 3-kinase (PI3K) inhibitors as cancer therapeutics. *Journal of Hematology and Oncology*, **6(88)**, 1–17.
- Arnaoutova, I., & Kleinman, H. K. (2010). In vitro angiogenesis: endothelial cell tube formation on gelled basement membrane extract. *Nature Protocols*, **5(4)**, 628–35.
- Auerbach, R., Lewis, R., Shinnars, B., Kubai, L., & Akhtar, N. (2003). Angiogenesis assays: a critical overview. *Clinical Chemistry*, **49(1)**, 32–40.
- Baker, M., Robinson, S. D., Lechertier, T., Jones, D., Vojnovic, B., and Hodivala-Dilke, K. (2012). Use of the mouse aortic ring assay to study angiogenesis. *Nature Protocols*, **7(1)**, 89–104.
- Bates, R. C., Edwards, N. S., & Yates, J. D. (2000). Spheroids and cell survival. *Critical Reviews in Oncology/hematology*, **36**: 61–74.
- Boyden, S. (1961). The Chemotactic Effect of Mixtures of Antibody and Antigen on Polymorphonuclear Leucocytes. *JEM*, **115(3)**, 453–466.
- Brooks, S. a, Lomax-Browne, H. J., Carter, T. M., Kinch, C. E., & Hall, D. M. S. (2010). Molecular interactions in cancer cell metastasis. *Acta Histochemica*, **112(1)**, 3–25.
- Burger, M.T., Pecchi, S., Wagman, A., Ni, Z. J., Knapp, M., Hendrickson, T., Atallah, G., Pfister, K., Zhang, Y., Bartulis, S., Frazier, K., Ng, S., Smith, A., Verhagen, J., Hazneder, J., Huh, K., Iwanowocz, E., Xin, X., Menezes, D., Merritt, H., Lee, I., Wiesmann, M., Kaufman, S., Crawford, K., Chin, M., Bussiere, D., Shoemaker, K., Zaror, I., Maira, S., and Voliva, C. F. (2011) Identification of NVP-BKM120 as a Potent, Selective, Orally Bioavailable Class I PI3 Kinase Inhibitor for Treating Cancer. *ACS Med. Chem. Lett.*, 2011, **2(10)**, 774–9.
- Carmeliet, P., & Jain, R. K. (2011). Molecular mechanisms and clinical applications of angiogenesis. *Nature*, **473(7347)**, 298–307.
- Chen, W., Kuo, K., Chou, T., Chen, C., Wang, C., Wei, Y., & Wang, L. (2012). The role of cytochrome c oxidase subunit Va in non-small cell lung cancer association with migration, invasion and prediction of distant metastasis. *BMC Cancer*, **12(273)**, 1471–2407.

- Desideri, M., Sanza, C. Di, Stoppacciaro, A., Ricciardi, M. R., Chiaretti, S., Tavolaro, S., Benassi, B., Bellacosa, A., Foa, R., Tafuri, A., Cognetti, F., and Anichini, A. (2009). Growth-Inhibitory and Antiangiogenic Activity of the MEK Inhibitor PD0325901 in Malignant Melanoma with or without BRAF Mutations. *Neoplasia*, **11(8)**, 720–731.
- Detmar, M. (2009). TScratch: a novel and simple software tool for automated analysis of monolayer wound healing assays. *Bio Techniques*, **46(4)**, 265–274.
- Doehn, U., Hauge, C., Frank, S. R., Jensen, C. J., Duda, K., Nielsen, V., Cohen, M. S., Johansen, J. V., Winther, B. R., Lund, L. R., Taunton, J., Hansen, S. H., and Frodin, M. (2013). RSK is a principle effector of the RAS-ERK pathway for eliciting a coordinate, pro-motile/invasive gene program and phenotype in epithelial cells. *Mol Cell*, **35(4)**, 511–522.
- Dolznic, H., Rupp, C., Puri, C., Haslinger, C., Schweifer, N., Wieser, E., Kerjaschki, D., and Garin-chesa, P. (2011). Modeling Colon Adenocarcinomas in Vitro. *AJPA*, **179(1)**, 487–501.
- Dutta, P. R., & Maity, A. (2008). Cellular responses to EGFR inhibitors and their relevance to cancer therapy. *Cancer Letters*, **254(2)**, 165–177.
- Eliceiri, B. P., Klemke, R., Strömblad, S., & Cheres, D. A. (1998). Integrin $\alpha v \beta 3$ Requirement for Sustained Mitogen-activated Protein Kinase Activity during Angiogenesis. *The Journal of Cell Biology*, **140(5)**, 1255–1264.
- Emuss, V., & Marais, R. (2008). The biology and oncology of RAF-ERK signaling. In *Cancer Drug Design and Discovery*. pp.382–402.
- Fedorenko, I. V., Gibney, G. T., & Smalley, K. S. M. (2013). NRAS mutant melanoma: biological behavior and future strategies for therapeutic management. *Oncogene*, **32(25)**, 3009–18.
- Fresno Vara, J. A., Casado, E., de Castro, J., Cejas, P., Belda-Iniesta, C., & González-Barón, M. (2004). PI3K/Akt signalling pathway and cancer. *Cancer Treatment Reviews*, **30(2)**, 193–204.
- Fukumura, D., Gohongi, T., Kadambi, a, Izumi, Y., Ang, J., Yun, C. O., Buerk, D. G., Huang, P. L., and Jain, R. K. (2001). Predominant role of endothelial nitric oxide synthase in vascular endothelial growth factor-induced angiogenesis and vascular permeability. *Proceedings of the National Academy of Sciences of the United States of America*, **98(5)**, 2604–9.
- Gabellini, C., Del Bufalo, D., & Zupi, G. (2006). Involvement of RB gene family in tumor angiogenesis. *Oncogene*, **25(38)**, 5326–32.
- Garci, C., Pearson, M. A., Marti, A., Meyer, T., Mestan, J., Zimmermann, J., Gao, J., Brueggen, J., Capraro, H., Cozens, R., Evans, D. B., Fabbro, D., Furet, P., Porta,

- D. G., Liebetanz, J., Martiny-baron, G., Ruetz, S., and Hofmann, F. (2004). In vivo antitumor activity of NVP-AEW541 — A novel , potent , and selective inhibitor of the IGF-IR kinase. *Cancer cell*, **5**: 231–239.
- Gariboldi, M. B., Ravizza, R., and Monti, E. (2010). The IGF-R1 inhibitor NVP-AEW541 disrupts a pro-survival and pro-angiogenic IGF-STAT3-HIF1 pathway in human glioblastoma cells. *Biochemical Pharmacology*, **80(4)**, 455–62.
- Giroux, S., Tremblay, M., Bernard, D., Aubry, S., Larouche, L., Huot, J., Landry, J., Jeannotte, L., and Charron, J. (1999). Embryonic death of Mek1-deficient mice reveals a role for this kinase in angiogenesis in the labyrinthine region of the placenta. *Current Biology*, **9**, 369–372.
- Go, R. S., & Owen, W. G. (2003). The rat aortic ring assay for in vitro study of angiogenesis. *Methods in Molecular Medicine*, **85**: 59–64.
- Hak, S., Reitan, N. K., Haraldseth, O., & de Lange Davies, C. (2010). Intravital microscopy in window chambers: a unique tool to study tumor angiogenesis and delivery of nanoparticles. *Angiogenesis*, **13(2)**, 113-130.
- Harisi, R., Kenessey, I., Olah, J. N., Timar, F., Babo, I., Pogany, G., Paku, S., and Jeney, A. (2009). Differential Inhibition of Single and Cluster Type Tumor Cell Migration. *Anticancer Research*, **29**: 2981–85.
- Henderson, Y. C., Chen, Y., Frederick, M. J., Lai, S. Y., & Clayman, G. L. (2010). MEK inhibitor PD0325901 significantly reduces the growth of papillary thyroid carcinoma cells in vitro and in vivo. *Molecular Cancer Therapeutics*, **9(7)**, 1968–76.
- Hiraoka, D., Okumura, E., & Kishimoto, T. (2011). Turn motif phosphorylation negatively regulates activation loop phosphorylation in Akt. *Oncogene*, **30(44)**, 4487–97.
- Hirschhaeuser, F., Menne, H., Dittfeld, C., West, J., Mueller-Klieser, W., & Kunz-Schughart, L. a. (2010). Multicellular tumor spheroids: an underestimated tool is catching up again. *Journal of Biotechnology*, **148(1)**, 3–15.
- Inoue, A., Sawata, S. Y., Taira, K., & Wadhwa, R. (2007). Loss-of-function screening by randomized intracellular RNAi identifies a potential target for metastasis. *PNAS*, **104(21)**, 8983–88.
- Jinnin, M., Ihn, H., & Tamaki, K. (2006). Characterization of SIS3 , a Novel Specific Inhibitor of Smad3 , and Its Effect on Transforming Growth Factor- beta 1- Induced Extracellular Matrix Expression. *Mol Pharmacol*, **69(2)**, 597–607.
- Jiang, B.-H., & Liu, L.-Z. (2008). PI3K/PTEN signaling in tumorigenesis and angiogenesis. *Biochimica et Biophysica Acta*, **1784(1)**, 150–8.

- Johnson, G. L., & Lapadat, R. (2002). Mitogen-Activated Protein Kinase Pathways Mediated by ERK , JNK , and p38 Protein Kinases. *Science*, **298**: 1911–34.
- Karar, J., & Maity, A. (2011). PI3K/AKT/mTOR Pathway in Angiogenesis. *Frontiers in Molecular Neuroscience*, **4**(51), 1–8.
- Khodarev, N. N. (2003). Tumour-endothelium interactions in co-culture: coordinated changes of gene expression profiles and phenotypic properties of endothelial cells. *Journal of Cell Science*, **116**(6), 1013–1022
- Kim, J. Bin, Stein, R., & O ' H a r e , M. J-dimensional in vitro tissue culture models of breast cancer-- a review. *Breast Cancer Research and Treatment*, **85**(3), 281–91.
- Kim, C. K., Choi, Y. K., Lee, H., Ha, K. S., Won, M. H., Kwon, Y. G., & Kim, Y. M. (2010). The farnesyltransferase inhibitor LB42708 suppresses vascular endothelial growth factor-induced angiogenesis by inhibiting ras-dependent mitogen-activated protein kinase and phosphatidylinositol 3-kinase/Akt signal pathways. *Mol Pharmacol*, **78**(1), 142-150.
- King, E., & Bowser, A. D. (2009). *Expanding the Targets in Adjuvant Therapy* , pp. 1–26).
- Kohn, E. C. (1992). Invasion and metastasis: Biology and Clinical Potential. *Pharmacology & Therapeutics*, **52**: 235–244.
- Kramer, N., Walzl, A., Unger, C., Rosner, M., Krupitza, G., Hengstschliger, M., & Dolznig, H. (2013). In vitro cell migration and invasion assays. *Mutation Research*, **752**(1), 10–24.
- Kumar, H. R., Zhong, X., Hoelz, D. J., Rescorla, F. J., Robert, J., Malkas, L. H., & Sandoval, J. A. (2010). Three-dimensional neuroblastoma cell culture: proteomic analysis between monolayer and multicellular tumor spheroids. *Pediatr Surg Int*, **24**(11), 1229–1234.
- Kunz-Schughart, L. a, Freyer, J. P., Hofstaedter, F., & Ebner, R. (2004). The use of 3-D cultures for high-throughput screening: the multicellular spheroid model. *Journal of Biomolecular Screening*, **9**(4), 273–85.
- Lee, S.-J., Namkoong, S., Kim, Y.-M., Kim, C.-K., Lee, H., Ha, K.-S., Chung, H.-T., Lee, S.-J., Namkoong, S., Kim, Y.-M., Kim, C.-K., Lee, H., Ha, K.-S., Chung, H.-T., Kwon, Y.-G., Kim, Y.-M. (2006). Fractalkine stimulates angiogenesis by activating the Raf-1/MEK/ERK- and PI3K/Akt/eNOS-dependent signal pathways. *American Journal of Physiology. Heart and Circulatory Physiology*, **291**(6), 2836–2846.

- Li a n g , C . , P a r k , A . Y . , & G u a n , J . (2 0 0 7) .
inexpensive method for analysis of cell migration in vitro. *Nature Protocols*, **2(2)**,
329–333.
- Lokman, N. a, Elder, A. S. F., Ricciardelli, C., & Oehler, M. K. (2012). Chick
Chorioallantoic Membrane (CAM) Assay as an In Vivo Model to Study the
Effect of Newly Identified Molecules on Ovarian Cancer Invasion and
Metastasis. *International Journal of Molecular Sciences*, **13(8)**, 9959–70.
- Ma, J., Sawai, H., Ochi, N., Matsuo, Y., Xu, D., Yasuda, A., Takahashi, H., Wakasugi,
T., and Takeyama, H. (2009). PTEN regulates angiogenesis through
PI3K/Akt/VEGF signaling pathway in human pancreatic cancer cells. *Molecular
and Cellular Biochemistry*, **331(1-2)**, 161–71.
- Maira, S.-M., Pecchi, S., Huang, A., Burger, M., Knapp, M., Sterker, D., Schnell, C.,
Guthy, D., Nagel, T., Wiesmann, M., Brachmann, S., Fritsch, Christine Dorsch,
M., Chae, P., Shoemaker, K., De Pover, A., Menezes, D., Martiny-Baron, G.,
Fabbro, D., Wilson, C.J., Schlegel, R., Hofmann, F., Garca-Echeverria, C.,
Sellers, W.R., and Voliva, C. F. (2012). Identification and characterization of
NVP-BKM120, an orally available pan-class I PI3-kinase inhibitor. *Molecular
Cancer Therapeutics*, **11(2)**, 317–28.
- Maity, P. R. D. and A. (2008). Cellular responses to EGFR inhibitors and their
relevance to cancer therapy. *Cancer Letters*, **254(2)**, 165–177.
- Maurer, G., Tarkowski, B., & Baccharini, M. (2011). Raf kinases in cancer-roles and
therapeutic opportunities. *Oncogene*, **30(32)**, 3477–88.
- Mavria, G., Vercoulen, Y., Yeo, M., Paterson, H., Karasarides, M., Marais, R., Bird,
D., and Marshall, C. J. (2006). ERK-MAPK signaling opposes Rho-kinase to
promote endothelial cell survival and sprouting during angiogenesis. *Cancer
Cell*, **9(1)**, 33–44.
- McCubrey, J. a, Steelman, L. S., Chappell, W. H., Abrams, S. L., Wong, E. W. T.,
Chang, F., Lehmann, B., Terrian, D., Milella, M., Tafuri, A., Stivala, F., Libra,
M., Basccke, J., Evangelisti, C., Martelli, A., and Franklin, R. (2007). Roles of
the Raf/MEK/ERK pathway in cell growth, malignant transformation and drug
resistance. *Biochimica et Biophysica Acta*, **1773(8)**, 1263–84.
- Mendoza-naranjo, A., Cormie, P., Serrano, A. E., Hu, R., Neill, S. O., Wang, M.,
Thrasivoulou, C., Power, K. T., White, A., Serene, T., Philips, A. R. J., and
Becker, D. L. (2012). Targeting Cx43 and N-Cadherin , Which Are Abnormally
Upregulated in Venous Leg Ulcers , Influences Migration , Adhesion and
Activation of Rho GTPases. *PLoS one*, **7(5)**, 1-16.
- Mikirova, N. a, Casciari, J. J., & Riordan, N. H. (2010). Ascorbate inhibition of
angiogenesis in aortic rings ex vivo and subcutaneous Matrigel plugs in vivo.
Journal of Angiogenesis Research, **2**: 1-6.

Mirzoeva, O. K., Das, D., Heiser, L. M., Bhattacharya, S., Siwak, D., Gendelman, R., Bayani, N., Wang, N., Neve, R., Guan, Y., Hu, Z., Knight, Z., Feiler, H. S., Gascard, P., Parvin, B., Spellman, P. T., Shokat, K. M., Wyrobek, A. J., Bissell, M. J., McCormick, F., Kuo, W.-L., Mills, G. B., Gray, J. W., and Korn, W. M. (2009). Basal subtype and MAPK/ERK kinase (MEK)-phosphoinositide 3-kinase feedback signaling determine susceptibility of breast cancer cells to MEK inhibition. *Cancer Research*, **69**(2), 565–72.

Mollloy, T., & van't Veer, L. J. (2008). *Current Opinion in Genetics & Development*, **18**(1), 35–41.

Montagut, C., & Settleman, J. (2009). Targeting the RAF-MEK-ERK pathway in cancer therapy. *Cancer Letters*, **283**(2), 125–34.

Murphy, D. a, Makonnen, S., Lassoued, W., Feldman, M. D., Carter, C., & Lee, W. M. F. (2006). Inhibition of tumor endothelial ERK activation, angiogenesis, and tumor growth by sorafenib (BAY43-9006). *The American Journal of Pathology*, **169**(5), 1875–85.

Nanobashvili, J., Jozkowicz, A., Neumayer, C., Fögl, a., Sporn, E., Polterauer, P., & Huk, I. (2003). Comparison of Angiogenic Potential of Human Microvascular Endothelial Cells and Human Umbilical Vein Endothelial Cells. *European Surgery*, **35**(4), 214–19.

Neuzillet, C., Tijeras-Raballand, A., de Mestier, L., Cros, J., Faivre, S., & Raymond, E. (2014). MEK in cancer and cancer therapy. *Pharmacology & Therapeutics*, **141**(2), 160–71.

Noro, R., Gemma, A., Kosaihiira, S., Kokubo, Y., Chen, M., Seike, M., Kataoka, K., Matsuda, K., Okano, T., Minegishi, Y., Yoshimura, A., and Kudoh, S. (2006). Gefitinib (IRESSA) sensitive lung cancer cell lines show phosphorylation of Akt without ligand stimulation. *BMC Cancer*, **6**: 277.

Okumura, N., Yoshida, H., Kitagishi, Y., Murakami, M., Nishimura, Y., & Matsuda, S. (2012). PI3K/AKT/PTEN Signaling as a Molecular Target in Leukemia Angiogenesis. *Advances in Hematology*, **2012**(843085), 1–6.

Ono, M., Hirata, A., & Kometani, T. (2004). Sensitivity to gefitinib (Iressa , ZD1839) in non-small cell lung cancer cell lines correlates with dependence on the epidermal growth factor (EGF) receptor / extracellular signal-regulated kinase 1 / 2 and EGF receptor / Akt pathway for proliferation. *Molecular Cancer Therapeutics*, **3**: 465–72.

Pardali, E., Goumans, M.-J., & ten Dijke, P. (2010). Signaling by members of the TGF-beta family in vascular morphogenesis and disease. *Trends in Cell Biology*, **20**(9), 556–67.

- Rajalingam, K., Schreck, R., Rapp, U. R., & Albert, S. (2007). Ras oncogenes and their downstream targets. *Biochimica et Biophysica Acta*, **1773**(8), 1177–95.
- Ribatti, D. (2012). Cardiovascular Development: Methods and Protocols. *Methods in Molecular Biology*, **843**: 47–57.
- Rogers, M. S., Birsner, A. E., & Amato, R. J. D. (2007). The mouse cornea micropocket angiogenesis assay. *Nature protocols*, **2**(10), 2545–2550.
- Sabeh, F., Shimizu-hirota, R., & Weiss, S. J. (2009). Protease-dependent versus independent cancer cell-dimensional anisotropic movement revisited. *The Journal of Cell Biology*, **185**(1), 11–19.
- Serra, V., Scaltriti, M., Prudkin, L., Eichhorn, P. J. a, Ibrahim, Y. H., Chandarlapaty, S., Markman, B., Rodriguez, O., Guzman, M., Rodriguez, S., Gili, M., Russillo, M., Parra, J. L., Singh, S., Arribas, J., Rosen, N., and Baselga, J. (2011). PI3K inhibition results in enhanced HER signaling and acquired ERK dependency in HER2-overexpressing breast cancer. *Oncogene*, **30**(22), 2547–57.
- Sussman, M. A., Volkers, M., Fischer, K., Bailey, B., Cottage, C. T., Din, S., Gude, N., Avitabile, D., Alvarez, R., Sundararaman, B., Quijada, P., Mason, M., Konstandin, M., Malhowski, A., Cheng, Z., Khan, M., and McGregor, M. (2011). Myocardial AKT: The Omnipresent Nexus. *Physiological Reviews*, **91**: 1023–1070.
- Schlessinger, J. (2000). Cell Signaling by Receptor Tyrosine Kinases. *Cell*, **103**: 211–225.
- Sckell, A., & Leunig, M. (2009). The dorsal skinfold chamber: studying angiogenesis by intravital microscopy. *Methods Mol Biol*, **467**, 305-317.
- Shen, J., Meng, X., Schiffmann, R., Brady, R. O., and Kaneski, C. R. (2007). Establishment and characterization of Fabry disease endothelial cells with an extended lifespan. *Molecular Genetics and Metabolism*, **92**: 137–144.
- Staton, C. A., Reed, M. W., & Brown, N. J. (2009). A critical analysis of current in vitro and in vivo angiogenesis assays. *Int J Exp Pathol*, **90**(3), 195-221.
- Staton, C. A., Stribbling, S. M., Tazzyman, S., Hughes, R., Brown, N. J., and Lewis, C. E. (2004). Current methods for assaying angiogenesis in vitro and in vivo. *International Journal of Experimental Pathology*, **85**: 233–248.
- Tanno, B., Mancini, C., Vitali, R., Mancuso, M., McDowell, H. P., Dominici, C., & Raschellà G. (2006). Down-regulation of insulin-like growth factor I receptor activity by NVP-AEW541 has an antitumor effect on neuroblastoma cells in vitro and in vivo. *Clinical Cancer Research*, **12**(22), 6772–80.

- Valastyan, S., & Weinberg, R. a. (2011). Tumor metastasis: molecular insights and evolving paradigms. *Cell*, **147**(2), 275–92.
- Venetsanakos, E., Mirza, A., Fanton, C., Romanov, S. R., Tlsty, T., & McMahon, M. (2002). Induction of tubulogenesis in telomerase-immortalized human microvascular endothelial cells by glioblastoma cells. *Experimental Cell Research*, **273**(1), 21–33.
- Wang, L., Chen, Q., Li, G., & Ke, D. (2012). Ghrelin stimulates angiogenesis via GHSR1a-dependent MEK/ERK and PI3K/Akt signal pathways in rat cardiac microvascular endothelial cells. *Peptides*, **33**(1), 92–100.
- Wang, Y., Hailey, J., Williams, D., Wang, Y., Lipari, P., Malkowski, M., Wang, X., Xie, L., Li, G., Saha, D., Ling, W., L.W., Cannon-Carlson, S., Greenberg, R., Ramos, Shields, R., Presta, L., Brams, P., Bishop, W. R., and Pachter, J. a. (2005). Inhibition of insulin-like growth factor-I receptor (IGF-IR) signaling and tumor cell growth by a fully human neutralizing anti-IGF-IR antibody. *Molecular Cancer Therapeutics*, **4**(8), 1214–21.
- Wen, S., Stolarov, J., Myers, M. P., Su, J. D., Wigler, M. H., Tonks, N. K., & Durden, D. L. (2001). PTEN controls tumor-induced angiogenesis. *Proceedings of the National Academy of Sciences of the United States of America*, **98**(8), 4622–7.
- Xia, C., Meng, Q., Cao, Z., Shi, X., & Jiang, B. H. (2006). Regulation of angiogenesis and tumor growth by p110 alpha and AKT1 via VEGF expression. *J Cell Physiol*, **209**(1), 56-66.
- Yun, J., Poulogiannis, G., Brower, E. T., Klemptner, S., & Cantley, L. L. (2013). The PI3K Pathway in Colorectal Cancers. In K. M. Haigis (Ed.), *Molecular Pathogenesis of Colorectal Cancer*, pp.157–99.
- Yang, S.-H., Sharrocks, A. D., & Whitmarsh, A. J. (2003). Transcriptional regulation by the MAP kinase signaling cascades. *Gene*, **320**: 3–21.
- Yuan, T. L., & Cantley, L. C. (2008). PI3K pathway alterations in cancer: variations on a theme. *Oncogene*, **27**(41), 5497–510.
- Zeng, Q., Li, S., Chepeha, D. B., Giordano, T. J., Li, J., Zhang, H., Polverini, P. J. Nor, J. Kitajewski, J. Wang, C. Y. (2005). Crosstalk between tumor and endothelial cells promotes tumor angiogenesis by MAPK activation of Notch signaling. *Cancer Cell*, **8**(1), 13-23.
- Zhang, H., Berel, D., Wang, Y., Li, P., Bhowmick, N. a, Figlin, R. a, & Kim, H. L. (2013). A comparison of Ku0063794, a dual mTORC1 and mTORC2 inhibitor, and temsirolimus in preclinical renal cell carcinoma models. *PloS One*, **8**(1), 1-12.

Zhang, S., Cao, Z., & Tian, H. (2011). SKLB1002 , a Novel Potent Inhibitor of VEGF Receptor 2 Signaling , Inhibits Angiogenesis and Tumor Growth In Vivo. *Clinical Cancer Research*, **17(13)**, 4439–4450.

