



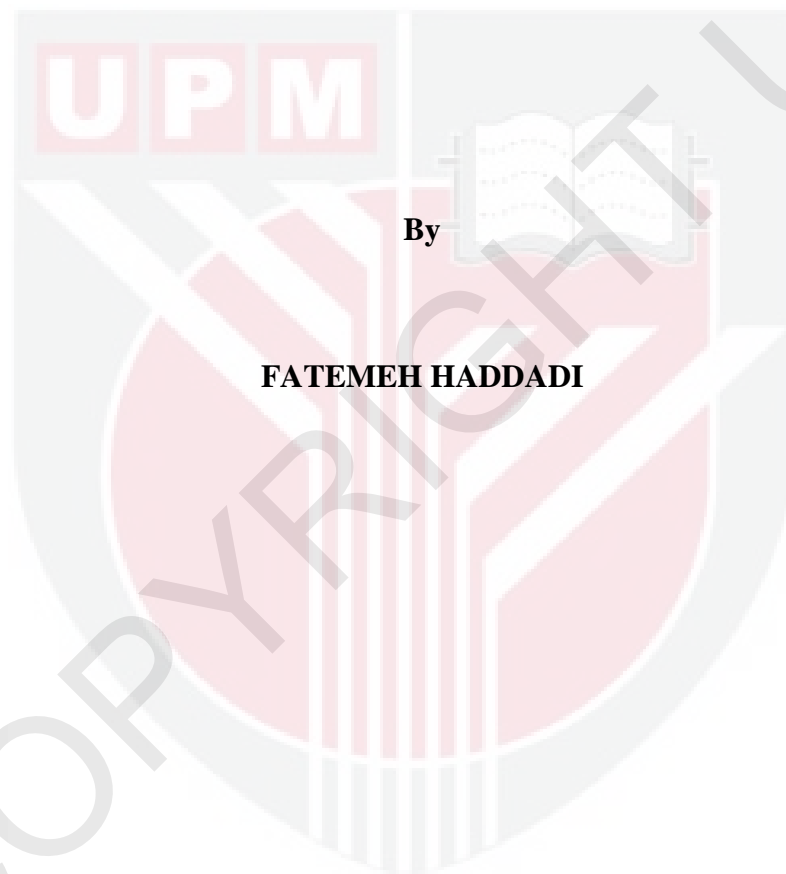
UNIVERSITI PUTRA MALAYSIA

***DEVELOPMENT OF HEAT TOLERANT STRAWBERRY (FRAGARIA X
ANANASSA DUCH., cv. CAMAROSA) PLANTS THROUGH
AGROBACTERIUM-MEDIATED TRANSFORMATION***

FATEMEH HADDADI

FP 2013 56

DEVELOPMENT OF HEAT TOLERANT STRAWBERRY (*FRAGARIA X ANANASSA* DUCH., cv. CAMAROSA) PLANTS THROUGH *AGROBACTERIUM*-MEDIATED TRANSFORMATION



By

FATEMEH HADDADI

© Thesis Submitted to the School of Graduate Studies, Universiti Putra Malaysia,
in Fulfilment of the Requirement for the Degree of Doctor of Philosophy

December 2012

In the name of ALLAH

Specially Dedicated to

All love

Specially

Hassein

My parents Halib and Parvin

And my brothers Farshad and Farsad

For their loving support

Abstract of thesis presented to the Senate of Universiti Putra Malaysia in fulfilment of the requirement for the degree of Doctor of Philosophy

DEVELOPMENT OF HEAT TOLERANT STRAWBERRY (*FRAGARIA X ANANASSA* DUCH., cv. CAMAROSA) PLANTS THROUGH *AGROBACTERIUM*-MEDIATED TRANSFORMATION

BY

FATEMEH HADDADI

December 2012

Chairman: Associate Professor Maheran Abd Aziz, PhD

Faculty : Agriculture

In Malaysia strawberry production is limited to the highland areas. Hence, based on the limitation of production areas the application of genetic engineering to produce a heat tolerant strawberry may facilitate the cultivation of the crop on the lowland areas. An *Agrobacterium*-mediated transformation method was applied to introduce the *AtHSP101* cDNA into strawberry cv. Camarosa. The *AtHSP101* cDNA under the control of CaMV35S promoter was cloned into the multiple cloning sites of pGreen0049. The construct was named pGFHSP and successfully introduced into *Agrobacterium* LBA4404. *In vitro* regeneration system from strawberry leaves used in the transformation was optimized by the application of different thidiazuron (TDZ) concentrations in MS medium. TDZ at 16 μ M showed the highest percentage (100 %) of shoot formation and the highest mean number of shoots (24) produced per explant. Effect of different antibiotics (timentin, cefotaxime, carbenicillin and ampicillin) on shoot regeneration of strawberry leaf explants showed the best shoot regeneration in the presence of 300 mg/l timentin and 150 mg/l cefotaxime. Determination of the minimum inhibitory concentration (MIC) of kanamycin was

carried out using leaf explant for effective screening of strawberry putative transformants. Kanamycin at 50 mg/l, with shoot regeneration percentage of 0 % and mean number of 0 shoot per explant was selected as the MIC. Assessment of different factors affecting *Agrobacterium* mediated-transformation of strawberry with the *AtHSP101* showed the highest efficiency of putative transformant production (83 %) in treatment with no preculture, bacterial OD₆₀₀ of 0.6 and the addition of 150 mg/l cefotaxime in the pre-selection and selection media. The presence of the *AtHSP101* in the plant genome was verified by luciferase reporter gene assay. Nested PCR amplification of genomic DNA isolated from each putative transformed plantlet showed the expected 520 and 689 bp products of *AtHSP101* and CaMV35S promoter primers, respectively. Southern blot analysis of the transgenic strawberry showed the presence of one, two and three copies of the transgene in the plant genome. Analysis of the *AtHSP101* expression at the mRNA level by RT-PCR showed the expected band of 520 bp corresponding to the *AtHSP101* cDNA. Protein dot blot and western blot analysis of the transgenic lines with one copy number of the transgene indicated the positive interaction of protein with the antibodies confirming the expression of *AtHSP101* at the protein level. DNA dot-blot analysis of the transgenic strawberry lines derived from runners and having one copy number of the transgene showed the presence of a hybridization sequence homologous to the *AtHSP101*. Greenhouse evaluation of the transgenic strawberry lines exhibited robust growth and performance compared to the non-transgenic control plant. Study on the effect of two temperatures, 20 and 30 °C showed greater growth and productivity in the transgenic lines compared to the control plants. Analysis of transgenic strawberry plants exposed to heat shock, gradual heat and drought stresses showed significant differences in their morphological characters and *AtHSP101*

protein level, measured by ELISA, compared to the control plants. The results of this study indicated that the expression of the *AtHSP101* may have an application to protect the transgenic strawberry plant under high temperature and drought conditions.



Abstrak tesis dikemukakan kepada Senat Universiti Putra Malaysia sebagai memenuhi keperluan untuk ijazah doktor falsafah

PEMBANGUNAN STRAWBERI (*FRAGARIA X ANANASSA* DUCH., cv. CAMAROSA) TUMBUHAN YANG TOLERAN PEMBAWAAN HABA MELALUI TRANSFORMASI BERPERANTARAAN *AGROBACTERIUM*

Oleh

FATEMEH HADDADI

Disember 2012

Pengerusi: Profesor Madya Maheran Abd Aziz, PhD

Fakulti : Pertanian

Di Malaysia pengeluaran strawberi adalah terhad di kawasan tanah tinggi. Oleh itu, berdasarkan kawasan pengeluaran yang terhad aplikasi kejuruteraan genetik untuk penghasilan strawberi toleran haba akan memudahkan penanaman tanaman tersebut di kawasan tanah rendah. Satu kaedah transformasi berperantaraan *Agrobacterium* telah diguna untuk memasukkan gen *AtHSP101* ke dalam strawberi cv. Camarosa. cDNA *AtHSP101* di bawah kawalan promoter CaMV35S diklonkan ke tapak pengklonan pelbagai pGreen0049. Vektor yang dibentuk dinamakan sebagai pGFHSP dan berjaya dipindahkan ke dalam *Agrobacterium* LBA4404. Sistem regenerasi *in vitro* daripada daun strawberi untuk digunakan dalam transformasi telah dioptimumkan melalui aplikasi pelbagai kepekatan berbeza TDZ dalam medium MS. TDZ pada kepekatan 16 μ M menunjukkan peratusan tertinggi pembentukan pucuk (100 %) dan purata bilangan pucuk tertinggi (24) dihasilkan per eksplan. Kesan antibiotik berbeza iaitu timentin, sifotaksim, karbenisilin dan ampisilin terhadap regenerasi eksplan pucuk daun strawberi menunjukkan regenerasi pucuk terbaik dengan kehadiran 300 mg/l timentin dan 150 mg/l sifotaksim.

Penentuan kepekatan perencatan minimum (*MIC*) kanamisin telah dijalankan menggunakan eksplan daun untuk saringan berkesan transforman strawberi putatif. Kanamisin pada kepekatan 50 mg/l dengan peratusan regenerasi pucuk 0 % dan min bilangan 0 pucuk per eksplan telah dipilih sebagai *MIC*. Penilaian faktor berbeza yang mempengaruhi transformasi strawberi berperantaraaan *Agrobacterium* dengan *AtHSP101* menunjukkan kecekapan tertinggi penghasilan transforman putatif (83 %) pada rawatan tanpa pra-kultur, OD_{600} bakteria 0.6 dan penambahan 150 mg/l sifotaksim pada medium pra-pemilihan dan pemilihan. Kehadiran *AtHSP101* dalam genom tumbuhan telah disahkan melalui asai gen pelapor lusiferase. Amplifikasi DNA genom yang dipencil dari daun muda setiap anak pokok transforman putatif melalui teknik '*Nested PCR*' menunjukkan produk 520 dan 689 bp masing-masing yang dijangka bagi primer *AtHSP101* dan promoter CaMV35S. Analisis *Southern blot* bagi strawberi transgenik menunjukkan kehadiran satu, dua dan tiga salinan transgen dalam genom tumbuhan. Analisis pengekspresan *AtHSP101* pada tahap mRNA yang dijalankan menggunakan teknik *RT-PCR* menunjukkan jalur 520 bp yang dijangka bersamaan dengan gen *AtHSP101*. Analisis *Protein dot blot* dan *Western blot* bagi baris transgenik dengan satu salinan transgen menunjukkan interaksi positif di antara protein dengan antibodi yang sekaligus mengesahkan pengekspresan gen *AtHSP101* pada tahap protein. Analisis *DNA dot blot* bagi baris strawberi transgenik dengan satu salinan transgen, yang berasal dari rayapan, menunjukkan kehadiran jujukan hibridisasi homologus *AtHSP101*. Penilaian di dalam rumah hijau bagi baris strawberi transgenik mempamerkan prestasi dan pertumbuhan yang memberangsangkan berbanding dengan tumbuhan bukan transgenik sebagai kawalan. Kajian kesan dua suhu, 20 and 30 °C menunjukkan pertumbuhan dan produktiviti lebih tinggi bagi baris transgenik berbanding

tumbuhan kawalan. Analisis tumbuhan transgenik strawberi yang didedahkan kepada kejutan haba, haba secara beransur-ansur dan tekanan kemarau menunjukkan perbezaan yang signifikan bagi ciri morfologi dan tahap protein *A α HSP101* berbanding dengan tumbuhan kawalan. Keputusan kajian ini menunjukkan bahawa pengekspresan *A α HSP101* erkemungkinan mempunyai aplikasi untuk melindungi tumbuhan strawberi transgenik di bawah suhu yang tinggi dan keadaan kemarau.



ACKNOWLEDGEMENTS

First and foremost, I would like to express my deepest endless thanks to God for granting me the capability to complete this research successfully and best regards from God to the last prophet, Mohammad and his family.

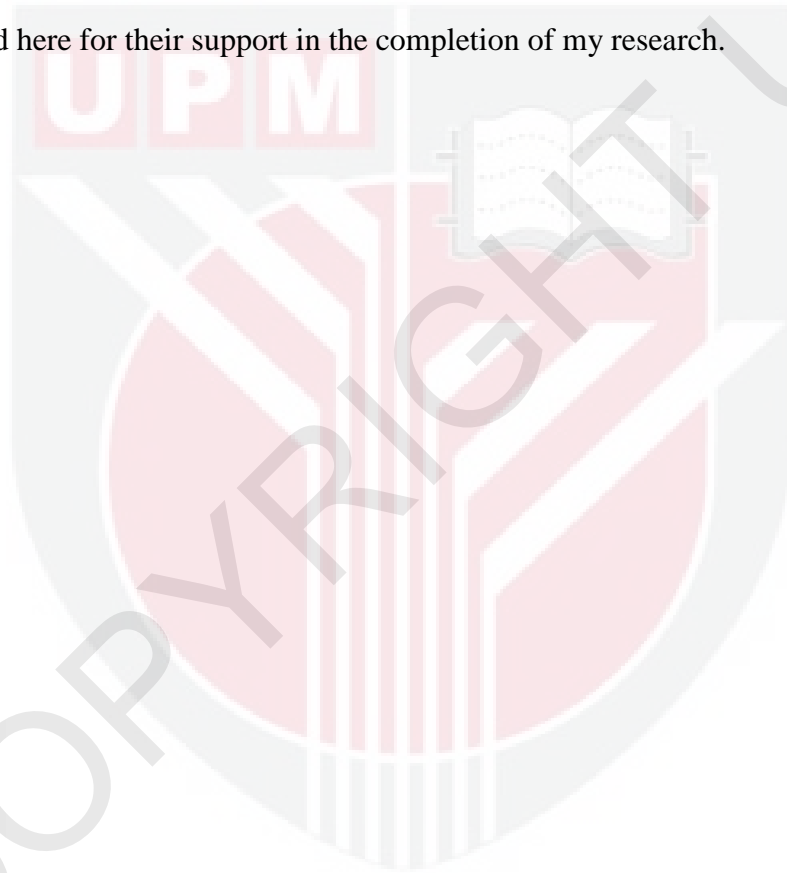
I would like to express my sincere gratitude to my supervisor Associate Professor Dr. Maheran Abdul Aziz for her generous help, patience, invaluable advice, the trust, the insightful discussion and constant support throughout the completion of the thesis.

I am also indebted and very grateful to my supervisory committee members, Assoc. Prof. Datin Dr. Siti Nor Akmar Abdullah, Prof. Dr. Tan Soon Guan and Assoc. Prof. Mohamad Ali Malbobi for their valuable advices and extended long-term support during the preparation and completion of this thesis. I am greatly thankful to Assoc. Prof. Dr. Halimi Mohd Saud, Head of Agriculture Technology Department, Assoc. Prof. Dr. Mihdzar Abdul Kadir and Mr. Azmi Abdul Rashid for their kind support, cooperation and permission to work in their laboratory.

It is a pleasure to thank all of my lab mates, staff and friends, Syed Ali Ravanfar, Ahmad Sharifkhani, Reza Azadi, Arash Rafat, Nagmeh Najat, Samira Samarfard, Moulod Koshan, Mansoreh Sharifi, Morvarid Akhavan, Shahida, Fahmi, Eng, Rohayni, Rozila and Siti Radziah, for providing a friendly atmosphere and making my life enjoyable in the laboratory. I wish to thank Mr. Mahmood Danaei for his great assistance, suggestions and knowledgeable comments and the data analysis of my thesis.

Finally, I will forever be thankful to my beloved parents and my brothers for their endless love, prayers, patience, understanding, spiritual and grateful support in all aspects of my life. I wish to thank my lovely husband Hossein for all his support and encouragement and sharing sleepless nights working together, for without him the thesis would not have come to a successful completion.

Last but certainly not least, I owe my deepest gratitude to those who are not mentioned here for their support in the completion of my research.



APPROVAL

I certify that a Thesis Examination Committee has met on 26 December 2012 to conduct the final examination of Fatemeh Haddadi on her thesis entitled "DEVELOPMENT OF HEAT TOLERANT STRAWBERRY (*FRAGARIA X ANANASSA* DUCH., cv. CAMAROSA) PLANTS THROUGH *AGROBACTERIUM*-MEDIATED TRANSFORMATION" in accordance with the Universities and University Colleges Act 1971 and the Constitution of the Universiti Putra Malaysia [P.U.(A) 106] 15 March 1998. The Committee recommends that the student be awarded the degree of Doctor of Philosophy.

Members of the Examination Committee were as follows:

Mihdzar Abd Kadir

Associate Professor
Faculty of Agriculture
Universiti Putra Malaysia
(Chairman)

Halimi Mohm Saud

Associate Professor
Faculty of Agriculture
Universiti Putra Malaysia
(Internal examiner)

Ho Chai Ling

Associate Professor
Faculty of Biotechnology and Bimolecular Science
Universiti Putra Malaysia
(Internal examiner)

Jose A. Mercado

Associate Professor
Department of Plant Biology
University of Malaga
Spain
(External Examiner)

SEOW HENG FONG, PhD

Professor and Deputy Dean
School of Graduate Studies
Universiti Putra Malaysia

Date:

This thesis was submitted to the Senate of Universiti Putra Malaysia and has been accepted as fulfilment of the requirement for the degree of Doctor of Philosophy. The members of the Supervisory Committee were as follows:

Maheran Abdul Aziz, PhD

Associate Professor
Faculty of Agriculture
Universiti Putra Malaysia
(Chairman)

Siti Nor Akmar Abdullah, PhD

Associate Professor
Faculty of Agriculture
Universiti Putra Malaysia
(Member)

Tan Soon Guan, PhD

Professor
Faculty of Biotechnology and Biomolecular Sciences
Universiti Putra Malaysia
(Member)

Mohammad Ali Malboobi, PhD

Associate Professor
Department of Plant Biotechnology
National Institute for Genetic Engineering and Biotechnology (NIGEB)
(Member)

BUJANG BIN KIM HUAT, PhD

Professor and Dean
School of Graduate Studies
Universiti Putra Malaysia

Date

DECLARATION

I declare that the thesis is my original work except for quotations and citations which have been duly acknowledged. I also declare that it has not been previously, and is not concurrently, submitted for any other degree at Universiti Putra Malaysia or at any other institution.



F. Haddadi

FATEMEH HADDADI

Date: 26 December 2012

TABLE OF CONTENT

	Page
DEDICATION	ii
ABSTRACT	iii
ABSTRAKT	vi
ACKNOWLEDGEMENTS	ix
APPROVAL	xi
DECLARATION	xiii
LIST OF TABLE	xvii
LIST OF FIGURES	xviii
LIST OF ABBREVIATIONS	xxi
1 INTRODUCTION	1
2 LITERATURE REVIEW	4
2.1 Strawberry	4
2.2 Botany and biology	4
2.3 <i>In vitro</i> culture	6
2.4 Heat shock proteins	7
2.5 Abiotic stress	10
2.6 Transformation	11
2.7 <i>Agrobacterium</i> -mediated transformation of strawberry	13
2.8 Antibiotics	14
2.9 Features of vectors	16
2.10 Confirmation methods of genetic transformation	19
2.10.1 Southern blot analysis	19
2.10.2 Western blot	20
2.10.3 Enzyme-Linked Immunosorbent Assay	21
3 OPTIMIZATION OF <i>AGROBACTERIUM</i>-MEDIATED TRANSFORMATION OF STRAWBERRY cv. CAMAROSA WITH <i>A_tHSP101</i>	24
3.1 Introduction	24
3.2 Materials and methods	27
3.2.1 Location of study	27
3.2.2 Basic medium	27
3.2.3 Plant material and culture condition	27
3.2.4 Statistical analysis	27

3.2.5	Effect of different concentrations of TDZ on shoot regeneration of strawberry leaves	28
3.2.6	Effect of different antibiotics on shoot regeneration from strawberry leaves	28
3.2.7	Construction of recombinant plasmid harbouring the <i>A/HSP101</i> by dual plasmid system	29
3.2.8	<i>Agrobacterium</i> –mediated transformation of strawberry	34
3.3	Results	40
3.3.1	Effect of different concentrations of TDZ on shoot regeneration of strawberry leaves	40
3.3.2	Effect of different antibiotics on shoot regeneration from strawberry leaves	42
3.3.3	Construction of recombinant plasmid harbouring the <i>A/HSP101</i> by dual plasmid system	45
3.3.4	<i>Agrobacterium</i> –mediated transformation of strawberry	50
3.3.5	Optimization of different factors affecting transformation of strawberry	54
3.3.6	Evaluation of the antibiotic ability on bacteria elimination from leaf explant of putative transformants	57
3.4	Discussion	59
4	CONFIRMATION OF STABLE INTEGRATION AND EXPRESSION OF THE <i>A/HSP101</i> IN STRAWBERRY cv. CAMAROSA PLANT GENOME	68
4.1	Introduction	68
4.2	Materials and methods	70
4.2.1	Location of study	70
4.2.2	Plant materials	70
4.2.3	Luciferase reporter assay	70
4.2.4	Nested PCR analysis of putative transformed strawberry plantlets	71
4.2.5	Southern blot analysis	72
4.2.6	Reverse transcription PCR analysis of the transgenic strawberry plants	75
4.2.7	Protein dot blot analysis	76
4.2.8	Western blot analysis	77
4.3	Results	79
4.3.1	Luciferase reporter assay	79
4.3.2	Nested PCR	80
4.3.3	Southern blot	82

4.3.4	RT-PCR	84
4.3.5	Protein dot blot	85
4.3.6	Western blot	86
4.4	Discussion	87
5	ANALYSIS OF THE EFFECTS OF THE <i>AtHSP101</i> EXPRESSION IN TRANSGENIC STRAWBERRY EXPOSED TO ABIOTIC STRESS	91
5.1	Introduction	91
5.2	Materials and methods	93
5.2.1	Location of study	93
5.2.2	Plant materials	93
5.2.3	DNA dot blot analysis of plants derived from runners of the transgenic strawberry lines	93
5.2.4	Greenhouse evaluation of the transgenic strawberry lines	94
5.2.5	Effect of two different temperatures on morphological parameters of the transgenic strawberry	94
5.2.6	Effect of heat stress on the transgenic strawberry	95
5.2.7	Effect of drought stress on the transgenic strawberry	96
5.2.8	Chlorophyll measurement	96
5.2.9	ELISA assay	97
5.2.10	Statistical analysis	98
5.3	Results	100
5.3.1	DNA dot blot analysis of plants derived from runners of the transgenic strawberry	100
5.3.2	Greenhouse evaluation of the transgenic strawberry lines	100
5.3.3	Effect of two different temperatures on morphological parameters and the <i>AtHSP101</i> expression of the transgenic strawberry	105
5.3.4	Effect of heat stress on <i>AtHSP101</i> expression of the transgenic strawberry lines	109
5.3.5	Effect of drought stress on <i>AtHSP101</i> expression of the transgenic strawberry	114
5.4	Discussion	117
6	GENERAL DISCUSSION, CONCLUSION AND RECOMMENDATION FOR FUTURE	123
	REFERENCES	132
	APPENDICES	149
	BIODATA OF STUDENT	164

LIST OF TABLE

Table	Page
2.1 Commonly used binary and super-binary vectors (Komari <i>et al.</i> , 2006)	17
2.2 Different features of reporter genes (Luehrsen and Walbot, 1993).	18
2.3 Comparison of direct and indirect ELISA detection methods. Source: http://www.piercenet.com .	22
3.1 Primers designed for PCR analysis of the pGFHSP	32
4.1 Effect of kanamycin on shoot regeneration percentage of strawberry cv. Camarosa leaf explants after five weeks of culture.	51
4.2 Effect of kanamycin on percentage of shoot regeneration and mean number of shoots derived from leaf explant of strawberry after five weeks of culture.	53
4.3 Effect of preculture medium, bacterial OD ₆₀₀ and antibiotics on mean number and percentage of putative transformant production after eight weeks of culture.	55
5.1 Primers designed for nested PCR analysis of genomic DNA of putative transformants	72
5.2 RLU of putative kanamycin resistant strawberry plantlets	79
5.3 Percentage of transformation efficiency obtained based on PCR positive results	81

LIST OF FIGURES

Figure	Page
2.1 The chemical reaction catalysed by firefly luciferase (Luehrsen and Walbot, 1993).	19
2.2 Schematic conjugation of ELISA assay common formats.	21
2.3 Sandwich ELISA; A: Direct and B: Indirect (Crowther, 2009)	23
3.1 Effect of different concentrations of TDZ on: A, mean number of shoots produced per leaf explant; and B, percentage of shoot formation of strawberry cv. Camarosa.	41
3.2 Leaf regeneration of strawberry cv. Camarosa after five weeks of culture on MS medium containing: A, 4 μ M TDZ; B, 8 μ M TDZ; C, 12 μ M TDZ; D, 24 μ M TDZ; E, 32 μ M TDZ and F, 16 μ M TDZ.	42
3.3 Effect of different concentrations of timentin, cefotaxime, carbenicillin and ampicillin on: A, mean number of shoots produced per leaf explant and B, percentage of shoot regeneration from the leaf explants of strawberry after five weeks of culture.	44
3.4 Effect of different concentrations of antibiotics on shoot regeneration of strawberry leaf explant after five weeks of culture.	45
3.5 PCR amplification of <i>AtHSP101</i> cDNA under the control of CaMV35S promoter from pCAMHSP vector.	46
3.6 Double digestion with <i>EcoRI</i> and <i>PstI</i> restriction enzymes after purification using PCR purification kit.	47
3.7 Ligation of double digested pGreen0049 and <i>AtHSP101</i> cDNA under the control of CaMV35S promoter.	47
3.8 Cloned products in <i>E. coli</i> were grown on LB agar plate in the presence of X-gal.	48
3.9 Double digestion of pGFHSP using <i>EcoRI</i> and <i>PstI</i> restriction enzymes.	49

3.10	Amplified PCR product using the forward and reverse primers shown in Table 3.1.	49
3.11	Schematic representation of constructed pGFHSP, pGreen0049 harbouring the <i>AtHSP101</i> under the control of CaMV35S promoter and pSoup vector.	50
3.12	Minimum lethal dosage of kanamycin on shoot regeneration from strawberry cv. Camarosa leaf explant after five weeks of culture..	52
3.13	Minimum lethal dose of kanamycin on shoot regeneration from strawberry shoot tips after five weeks of culture..	53
3.14	Regeneration response of strawberry leaf explant with respect to week of culture after inoculation with <i>Agrobacterium</i>	56
3.15	Acclimatization of strawberry putative plantlets after 14 weeks of culture	57
3.16	Evaluation of <i>Agrobacterium</i> contamination of leaf derived from putative kanamycin resistant strawberry plantlets.	58
4.1	Luciferase activity of control and putative trasformed plants..	80
4.2	Amplification of PCR product in independent lines of putatively transformed strawberry harbouring <i>AtHSP101</i> cDNA.	81
4.3	Southern blot analysis of four independent PCR-positive lines of transgenic strawberry.	83
4.4	Acclimatized transgenic strawberry plants confirmed by Southern blot analysis.	84
4.5	RT-PCR product of strawberry harbouring the <i>AtHSP101</i> .	85
4.6	Dot blot analysis of <i>AtHSP101</i> protein, C: Non-transformed control; Transgenic lines LS ₁ and LS ₂ of strawberry with one copy number.	86
4.7	Western blot analysis of the transgenic strawberry plants.	86
5.1	DNA dot blot analysis of plants derived from the transgenic lines LS ₁ and LS ₂ and control..	100
5.2	Greenhouse evaluation of the transgenic strawberry lines LS ₁ and LS ₂ and control.	103
5.3	Greenhouse eveluation of the transgenic strawberry lines LS ₁ and LS ₂ and control.	104

5.4 Greenhouse evaluation of the transgenic strawberry lines LS ₁ and LS ₂ and control after seven months.	104
5.5 Four month-old transgenic strawberry lines LS ₁ and LS ₂ and the control plant	105
5.6 Fruits of the transgenic lines LS ₁ and LS ₂ and control (Bar=1 cm)	105
5.7 Effect of two different temperatures on A: Mean number of leaves; B: Mean height of plant (cm); C: Mean number of flower; D: Mean number of fruit and E: Total chlorophyll content in transgenic lines LS ₁ and LS ₂ and the control plant of strawberry tow months after transfer.	108
5.8 Effect of temperature on flower of strawberry cv. Camarosa formed at 30 °C.	109
5.9 Effect of heat stress on total chlorophyll content of the transgenic lines LS ₁ and LS ₂ and control plant of strawberry.	112
5.10 ELISA assay of the transgenic lines and control plant of strawberry exposed to abiotic stress.	113
5.11 Effect of heat stress on <i>AtHSP101</i> protein content of the transgenic lines LS ₁ and LS ₂ and control plant of strawberry; A: Heat shock stress; B: Gradual heat stress..	113
5.12 Effect of heat stress on the transgenic lines LS ₁ and LS ₂ and control plant of strawberry cv. Camarosa one month after stress, A: Heat shock stress; B: Gradual heat stress.	114
5.13 Effect of drought stress on: A, Total chlorophyll content; and B, <i>AtHSP101</i> protein content of the transgenic lines LS ₁ and LS ₂ and control plants of strawberry.	116
5.14 Effect of drought stress on the transgenic lines LS ₁ and LS ₂ and control plants of strawberry cv. Camarosa two weeks after stress.	116

LIST OF ABBREVIATIONS

<i>A. tumefaciens</i>	<i>Agrobacterium tumefaciens</i>
ANOVA	analysis of variance
BA	N6-benzyladenine
BAP	6-benzylaminopurine
bp	base pair
BSA	bovine serum albumin
CaMV	cauliflower mosaic virus
cDNA	complementary DNA
CRD	randomized complete design
DMRT	duncan multiple range test
<i>E.coli</i>	<i>Escherichia coli</i>
<i>et al.</i>	et alia
ELISA	enzyme-linked immune sorbent assay
GH	gradual heat
GFP	green fluorescent protein
GUS	β -glucuronidase
HS	heat shock
HSP	heat shock protein
IAA	indole-3-acetic acid
IBA	indole-3-butyric acid
OD	optical density
PCR	polymerase chain reaction
PBS	phosphate buffered saline

RLU	relative luminescence unit
RT-PCR	reverse transcription polymerase chain reaction
PGR	plant growth regulator
RNase	ribonuclease
rpm	revolutions per minute
SDS	sodium dodecyl sulphate
sp.	species
TAE	tris-acetate/EDTA electrophoresis buffer
TDZ	thidiazuron
T _m	temperature
w/v	weight to volume
X-gal	5-bromo-4-chloro-3-indolyl-D-galactopyranoside
2iP	6-(α,α -dimethylallylamino)-purine
2,4-D	2,4-dichlorophenoxyacetic acid

CHAPTER 1

INTRODUCTION

There is a worldwide growing demand for strawberry due to its richness in vitamins, polyphenolics and phytochemicals, pleasant aroma and flavor, and fresh appearance. Strawberry constitutes an important part of greenhouse production, and many studies on strawberry have been conducted. Complicated genetic background contributed by high heterozygosity and polyploidy of strawberry impedes improvement of this crop through traditional breeding methods. Manipulation at gene level paved the way for further amelioration of strawberry improvement although the octoploid genome of strawberry poses difficulties for genetic and molecular studies. Studies on gene function in strawberry are few (Folta *et al.*, 2006). Furthermore concomitant application of genetic manipulation and classical breeding methods could accelerate cultivar developmental programs.

Plants are generally exposed to various stress factors and environmental conditions which affect their productivity and growth. Within the context of environmental changes, global warming or climate change as one of the abiotic stress factors has the foremost global impact on yield of crops. The optimal growth temperature for temperate species varies from 11.5 to 26 °C, whereas for tropical/subtropical species is between 23 and 32 °C (Lee *et al.*, 2007). Increase in temperature, 5–10 °C above ambient, causes damage of proteins and enzymes involved in metabolism (Porter, 2005). These changes may take a few minutes to a few hours as a result of the heat shock (Cho *et al.*, 2012; Yan *et al.*, 2011). Biological stress like elevated temperatures result in expression of heat shock proteins (HSPs) that influence

survival of plants under stress conditions (Park *et al.*, 2012). HSPs are highly conserved and commonly expressed constitutively by facilitating the synthesis and folding of proteins. HSP101, a member of the HSP families, plays a critical role in acquiring thermotolerance.

Strawberry as a temperate crop is grown in the highland regions of Malaysia such as in Cameron Highlands at an altitude of 6,001ft (1,829 m) above sea-level. Cameron Highlands have the optimum weather and condition for strawberry cultivation with temperatures ranging between 13.7 °C and 23.6 °C and a mean value of 17.4 °C. Temperatures higher than 30 °C affect growth, runner production, flowering and fruiting of strawberries resulting in production of lower quality strawberries (Li *et al.*, 2010). Increment of temperature even by 1 °C was found to decrease strawberry firmness (Pyrotis *et al.*, 2012). Besides, limited availability of areas for cultivation other problems encountered in strawberry production in Cameron highlands are the sloppy lands, land degradation and distance.

Pressed by these concerns, an attempt was undertaken in this study to produce strawberry plants tolerant to high temperature stress via genetic engineering using *Arabidopsis thaliana* HSP101 (*AtHSP101*) cDNA which may enable the crop to be grown at the lowland. By far the most exploited method of strawberry gene transformation is *Agrobacterium*-mediated transformation (Qin *et al.*, 2011; Mezzetti, 2009). Indeed strawberry cv. Camarosa is the most common cultivar grown in Malaysia. Hence optimization and development of *Agrobacterium*-mediated transformation of strawberry cv. Camarosa using the *AtHSP101* for heat tolerance were conducted in this study.

Therefore the main aims of this study were:

- 1- To determine the effect of different concentrations of TDZ and antibiotics on *in vitro* shoot regeneration from strawberry leaves
- 2- To construct pGreen0049 binary vector harbouring *AtHSP101* cDNA, to optimize factors affecting *Agrobacterium*-mediated transformation of strawberry with the cDNA, and to verify the presence and expression of the gene in the plant genome
- 3- To evaluate the performance of the transgenic plants in the transgenic greenhouse and to study the effect of abiotic stresses on the expression of the *AtHSP101* in the transgenic strawberry lines

REFERENCES

- Abdul-Baki, A. A. and Stommel, J. R. (1995). Pollen viability and fruit set of tomato genotypes under optimum and high-temperature regimes. *HortScience*, 30(1), 115-117.
- Adam, Z., Adamska, I., Nakabayashi, K., Ostersetzter, O., Haussuhl, K., Manuell, A., Zheng, B., Vallon, O., Rodermel, S. R. and Shinozaki, K. (2001). Chloroplast and mitochondrial proteases in *Arabidopsis*. A proposed nomenclature. *Plant Physiology*, 125(4), 1912-1918.
- Agarwal, M., Katiyar-Agarwal, S., Sahi, C., Gallie, D. R. and Grover, A. (2001). *Arabidopsis thaliana* Hsp100 proteins: kith and kin. *Cell Stress and Chaperones*, 6(3), 219.
- Al-Whaibi, M. H. (2011). Plant heat-shock proteins: A mini review. *Journal of King Saud University-Science*, 23(2), 139-150.
- Allen, G. C., Hall Jr, G., Michalowski, S., Newman, W., Spiker, S., Weissinger, A. K. and Thompson, W. F. (1996). High-level transgene expression in plant cells: effects of a strong scaffold attachment region from tobacco. *The Plant Cell Online*, 8(5), 899-913.
- Alsheikh, M., Suso, H. P., Robson, M., Battey, N. and Wetten, A. (2002). Appropriate choice of antibiotic and *Agrobacterium* strain improves transformation of antibiotic-sensitive *Fragaria vesca* and *F. v. semperflorens*. *Plant Cell Reports*, 20(12), 1173-1180.
- Amil-Ruiz, F., Blanco-Portales, R., Munoz-Blanco, J. and Caballero, J. L. (2011). The strawberry plant defense mechanism: A molecular review. *Plant and Cell Physiology*, 52(11), 1873-1903.
- An, G., Ebert, P., Mitra, A. and Ha, S. (1988). Binary vectors. In *Plant Molecular Biology Manual* (Gelvin, S. B. and Schilperoort, R. A., eds) pp. A3 1-19, Kluwer Academic.
- Angadi, S., Cutforth, H., Miller, P., McConkey, B., Entz, M., Brandt, S. and Volkmar, K. (2000). Response of three Brassica species to high temperature stress during reproductive growth. *Canadian Journal of Plant Science*, 80(4), 693-701.
- Aoshima, Y. (2005). Efficient embryogenesis in the callus of tea (*Camellia sinensis*) enhanced by the osmotic stress or antibiotics treatment. *Plant Biotechnology*, 22(4), 277-280.
- Asao, H., Nishizawa, Y., Arai, S., Sato, T., Hirai, M., Yoshida, K., Shinmyo, A. and Hibi, T. (1997). Enhanced resistance against a fungal pathogen *Sphaerotheca humuli* in transgenic strawberry expressing a rice chitinase gene. *Plant Biotechnology*, 14(3), 145-149.

- Bachelier, C., Graham, J., Machray, G., Du Manoir, J., Roucou, J., McNicol, R. and Davies, H. (1996). Integration of an invertase gene to control sucrose metabolism in strawberry cultivars. *Acta Horticulturae*, 439, 161-164.
- Barcelo, M., El-Mansouri, I., Mercado, J. A., Quesada, M. A. and Pliego Alfaro, F. (1998). Regeneration and transformation via *Agrobacterium tumefaciens* of the strawberry cultivar Chandler. *Plant Cell, Tissue and Organ Culture*, 54(1), 29-36.
- Bhat, S. and Srinivasan, S. (2002). Molecular and genetic analyses of transgenic plants: Considerations and approaches. *Plant Science*, 163(4), 673-681.
- Biswas, M., Dutt, M., Roy, U., Islam, R. and Hossain, M. (2009). Development and evaluation of *in vitro* somaclonal variation in strawberry for improved horticultural traits. *Scientia Horticulturae*, 122(3), 409-416.
- Borrelli, G., Di Fonzo, N. and Lupotto, E. (1992). Effect of cefotaxime on callus culture and plant regeneration in durum wheat. *Journal of Plant Physiology*, 140(3), 372-374.
- Boston, R. S., Viitanen, P. V. and Vierling, E. (1996). Molecular chaperones and protein folding in plants. *Plant Molecular Biology*, 32(1), 191-222.
- Břiza, J., Pavingerová, D., Přikrylová, P., Gazdova, J., Vlasák, J. and Niedermeierová, H. (2008). Use of phosphomannose isomerase-based selection system for *Agrobacterium*-mediated transformation of tomato and potato. *Biologia Plantarum*, 52(3), 453-461.
- Brown, M. A., Zhu, L., Schmidt, C. and Tucker, P. W. (2007). Hsp90-from signal transduction to cell transformation. *Biochemical and Biophysical Research Communications*, 363(2), 241-246.
- Brown, T. (1993). Analysis of DNA sequences by blotting and hybridization. *Current Protocols in Molecular Biology (Suppl 45)*. New York: Wiley. p, 2, 1-2.9.
- Brown, T. (2001). Dot and slot blotting of DNA. *Current Protocols in Molecular Biology*, 21, 2.9.15–12.19.20.
- Buchanan, B. B., Gruissem, W. and Jones, R. L. (2000). *Biochemistry and Molecular Biology of Plants* (Vol. 40): American Society of Plant Physiologists.
- Burke, J. J. (2009). Enhancement of Reproductive Heat Tolerance in Plants. United State Patent US 2011/0055977A0055971.
- Chalavi, V., Tabaeizadeh, Z. and Thibodeau, P. (2003). Enhanced resistance to *Verticillium dahliae* in transgenic strawberry plants expressing a *Lycopersicon chilense* chitinase gene. *Journal of the American Society for Horticultural Science*, 128(5), 747-753.

- Chandler, V. L. and Vaucheret, H. (2001). Gene activation and gene silencing. *Plant Physiology*, 125(1), 145-148.
- Chen, C., Zheng, Q., Xiang, X., Soneji, J. R., Huang, S., Choi, Y. A., Rao, M. N. and Gmitter Jr, F. G. (2007). Development of pGreen-derived GFP binary vectors for citrus transformation. *HortScience*, 42(1), 7-10.
- Cheng, M., Lowe, B. A., Spencer, T. M., Ye, X. and Armstrong, C. L. (2004). Factors influencing *Agrobacterium*-mediated transformation of monocotyledonous species. *In Vitro Cellular and Developmental Biology-Plant*, 40(1), 31-45.
- Chnappagoudar, S. B. (2007). *Studies on vitro regeneration and genetic transformation in chilli (Capsicum annuumL.)*. Dharwad University of Agricultural Science, Dharwad.
- Cho, M. A., Moon, C. Y., Liu, J. R. and Choi, P. S. (2008). *Agrobacterium*-mediated transformation in *Citrullus lanatus*. *Biologia Plantarum*, 52(2), 365-369.
- Cho, S. C., Chao, Y. Y. and Kao, C. H. (2012). Calcium deficiency increases Cd toxicity and Ca is required for heat-shock induced Cd tolerance in rice seedlings. *Journal of Plant Physiology*, 169(9), 892-898.
- Corden de Mesa, C., Jiménez-Bermúdez, S., Pliego-Alfaro, F., Quesada, M. A. and Mercado, J. A. (2000). *Agrobacterium* cells as microprojectile coating: a novel approach to enhance stable transformation rates in strawberry. *Functional Plant Biology*, 27(12), 1093-1100.
- Costa, M., Nogueira, F., Figueira, M., Otoni, W., Brommonschenkel, S. and Cecon, P. (2000). Influence of the antibiotic timentin on plant regeneration of tomato (*Lycopersicon esculentum* Mill.) cultivars. *Plant Cell Reports*, 19(3), 327-332.
- Crowther, J. (2009). *Methods in Molecular Biology. The ELISA guidebook. 2. painos: New York: Humana Press.*
- da Silva Pinto, M., De Carvalho, J. E., Lajolo, F. M., Genovese, M. I. and Shetty, K. (2010). Evaluation of antiproliferative, anti-type 2 diabetes, and antihypertension potentials of ellagitannins from strawberries (*Fragaria × ananassa* Duch.) using in vitro models. *Journal of Medicinal Food*, 13(5), 1027-1035.
- Danilova, S. and Dolgikh, Y. I. (2004). The stimulatory effect of the antibiotic cefotaxime on plant regeneration in maize tissue culture. *Russian Journal of Plant Physiology*, 51(4), 559-562.
- Debnath, S. C. (2005). Strawberry sepal: another explant for thidiazuron-induced adventitious shoot regeneration. *In Vitro Cellular and Developmental Biology-Plant*, 41(5), 671-676.

- Demirevska-Kepova, K., Holzer, R., Simova-Stoilova, L. and Feller, U. (2005). Heat stress effects on ribulose-1, 5-bisphosphate carboxylase/oxygenase, Rubisco binding protein and Rubisco activase in wheat leaves. *Biologia Plantarum*, 49(4), 521-525.
- Dhir, S., Hinchee, M. A. W., Layton, J. G. and Oakes, J. V. (2001). Methods for strawberry transformation using *Agrobacterium tumefaciens*. United State Patent US 6274, 6791 B6271.
- du Plessis, H. J., Brand, R. J., Glyn-Woods, C. and Goedhart, M. A. (1997). Efficient genetic transformation of strawberry (*Fragaria x ananassa* Duch.) cultivar Selekt. *Acta Horticulturae*, 447, 289-294.
- Eapen, S. and George, L. (1990). Influence of phytohormones, carbohydrates, aminoacids, growth supplements and antibiotics on somatic embryogenesis and plant differentiation in finger millet. *Plant Cell, Tissue and Organ Culture*, 22(2), 87-93.
- El-Mansouri, I., Mercado, J. A., Valpuesta, V., López-Aranda, J. M., Pliego-Alfaro, F. and Quesada, M. A. (1996). Shoot regeneration and *Agrobacterium*-mediated transformation of *Fragaria vesca* L. *Plant Cell Reports*, 15(8), 642-646.
- Estopa, M., Marfa, V., Mele, E. and Messeguer, J. (2001). Study of different antibiotic combinations for use in the elimination of *Agrobacterium* with kanamycin selection in carnation. *Plant Cell, Tissue and Organ Culture*, 65(3), 211-220.
- Fahmy, A. H., El-Shafy, Y. H., El-Shihy, O. M. and Madkour, M. (2006). Highly efficient regeneration system via somatic embryogenesis from immature embryos of Egyptian wheat cultivars (*Triticum aestivum* L.) using different growth regulators. *World Journal of Agricultural Sciences*, 2(3), 282-289.
- FAOSTAT (2011). www.fao.org/waicent/portal/statistics_en.asp.
- Ferreira, S., Hjern, K., Larsen, M., Wingsle, G., Larsen, P., Fey, S., Roepstorff, P. and Pais, M. S. (2006). Proteome profiling of *Populus euphratica* Oliv. upon heat stress. *Annals of Botany*, 98(2), 361-377.
- Finstad, K. and Martin, R. R. (1994). Transformation of strawberry for virus resistance. *Acta Horticulturae*, 385, 86-90.
- Firsov, A. and Dolgov, S. (1999). Agrobacterial transformation and transfer of the antifreeze protein gene of winter flounder to the strawberry. *Acta Horticulturae*, 484, 581-586.
- Fleury, D., Jefferies, S., Kuchel, H. and Langridge, P. (2010). Genetic and genomic tools to improve drought tolerance in wheat. *Journal of Experimental Botany*, 61(12), 3211-3222.

- Folta, K. M. and Dhingra, A. (2006). Invited review: Transformation of strawberry: The basis for translational genomics in Rosaceae. *In Vitro Cellular and Developmental Biology -Plant*, 42(6), 482-490.
- Folta, K. M., Dhingra, A., Howard, L., Stewart, P. J. and Chandler, C. K. (2006). Characterization of LF9, an octoploid strawberry genotype selected for rapid regeneration and transformation. *Planta*, 224(5), 1058-1067.
- Gamborg, O. L., Miller, R. A. and Ojima, K. (1968). Nutrient requirements of suspension cultures of soybean root cells. *Experimental Cell Research*, 50(1), 151-158.
- Gantait, S., Mandal, N. and Nandy, S. (2011). Advances in micro propagation of selected aromatic plants: A review on vanilla and strawberry. *American Journal of Biochemistry and Molecular Biology*, 1(1), 1-19.
- Giampieri, F., Tulipani, S., Alvarez-Suarez, J. M., Quiles, J. L., Mezzetti, B. and Battino, M. (2012). The strawberry: composition, nutritional quality, and impact on human health. *Nutrition*, 28(1), 9-19.
- Glover, J. R. and Lindquist, S. (1998). Hsp104, Hsp70, and Hsp40: A novel chaperone system that rescues previously aggregated proteins. *Cell*, 94(1), 73-82.
- Goloubinoff, P., Mogk, A., Zvi, A. P. B., Tomoyasu, T. and Bukau, B. (1999). Sequential mechanism of solubilization and refolding of stable protein aggregates by a bichaperone network. *Proceedings of the National Academy of Sciences*, 96(24), 13732.
- Goto, F., Yoshihara, T. and Saiki, H. (2000). Iron accumulation and enhanced growth in transgenic lettuce plants expressing the iron-binding protein ferritin. *TAG Theoretical and Applied Genetics*, 100(5), 658-664.
- Graham, J., McNicol, R. and Greig, K. (1995). Towards genetic based insect resistance in strawberry using the cowpea trypsin inhibitor gene. *Annals of Applied Biology*, 127(1), 163-173.
- Gravel, P. (2002). Identification of glycoproteins on nitrocellulose membranes using lectin blotting. *The Protein Protocols Handbook*, 779-793.
- Grewal, D., Gill, R. and Gosal, S. S. (2006). Influence of antibiotic cefotaxime on somatic embryogenesis and plant regeneration in indica rice. *Biotechnology Journal*, 1(10), 1158-1162.
- Gruchala, A., Korbin, M. and Żurawicz, E. (2004). Conditions of transformation and regeneration of Induka' and Elista' strawberry plants. *Plant Cell, Tissue and Organ Culture*, 79(2), 153-160.
- Gulen, H. and Eris, A. (2003). Some physiological changes in strawberry (*Fragaria x ananassa* 'Camarosa') plants under heat stress. *Journal of Horticultural Science and Biotechnology*, 78(6), 894-898.

- Gurley, W. B. (2000). HSP101: a key component for the acquisition of thermotolerance in plants. *Plant Cell*, 12(4), 457-460.
- Haddadi, F., Aziz, M. A., Saleh, G., Rashid, A. A. and Kamaladini, H. (2010). Micropropagation of Strawberry cv. Camarosa: Prolific shoot regeneration from *in vitro* shoot tips using Thidiazuron with N6-benzylamino-purine. *HortScience*, 45(3), 453-456.
- Hartl, F. U. (1996). Molecular chaperones in cellular protein folding. *Nature*, 381, 571-580.
- Hartl, F. U. (2011). Chaperone-assisted protein folding: the path to discovery from a personal perspective. *Nature Medicine*, 17(10), 1206-1210.
- Hartl, F. U. and Hayer-Hartl, M. (2002). Molecular chaperones in the cytosol: from nascent chain to folded protein. *Science*, 295(5561), 1852-1858.
- Harwood, W., Ross, S., Bulley, S., Travella, S., Busch, B., Harden, J. and Snape, J. (2002). Use of the firefly luciferase gene in a barley (*Hordeum vulgare*) transformation system. *Plant cell reports*, 21(4), 320-326.
- Haymes, K. and Davis, T. (1998). *Agrobacterium*-mediated transformation of Alpine 'Fragaria vesca', and transmission of transgenes to R1 progeny. *Plant Cell Reports*, 17(4), 279-283.
- He, P., Ma, Y., Dai, H., Li, L., Liu, Y., Li, H., Zhao, G. and Zhang, Z. (2012). Characterization of the hormone and stress-induced expression of FaRE1 retrotransposon promoter in strawberry. *Journal of Plant Biology*, 55(1), 1-7.
- Heikkila, J. J. (2010). Heat shock protein gene expression and function in amphibian model systems. *Comparative Biochemistry and Physiology-Part A: Molecular and Integrative Physiology*, 156(1), 19-33.
- Hellens, R. P., Edwards, E. A., Leyland, N. R., Bean, S. and Mullineaux, P. M. (2000). pGreen: a versatile and flexible binary Ti vector for *Agrobacterium*-mediated plant transformation. *Plant Molecular Biology*, 42(6), 819-832.
- Hellman, E. and Travis, J. (1988). Growth inhibition of strawberry at high temperatures. *Advances in Strawberry Production*, 7, 36-38.
- Himes, S. R. and Shannon, M. F. (2000). Assays for transcriptional activity based on the luciferase reporter gene. *Methods in Molecular Biology-Clifton then Totowa*, 130, 165-174.
- Ho, M. W. (2000). Hazards of transgenic plants containing the cauliflower mosaic viral promoter: Authors' reply to critiques of "The Cauliflower Mosaic Viral Promoter-a Recipe for Disaster?". *Microbial Ecology in Health and Disease*, 12(1), 6-11.
- Hoekema, A., Hirsch, P., Hooykaas, P. and Schilperoort, R. (1983). A binary plant vector strategy based on separation of vir- and T-region of the *Agrobacterium tumefaciens* Ti-plasmid. *Nature*, 303, 179-180.

- Holford, P. and Newbury, H. (1992). The effects of antibiotics and their breakdown products on the in vitro growth of *Antirrhinum majus*. *Plant Cell Reports*, 11(2), 93-96.
- Hong, S. W., Lee, U. and Vierling, E. (2003). *Arabidopsis* hot mutants define multiple functions required for acclimation to high temperatures. *Plant Physiology*, 132(2), 757-767.
- Houde, M., Dallaire, S., N'Dong, D. and Sarhan, F. (2004). Overexpression of the acidic dehydrin WCOR410 improves freezing tolerance in transgenic strawberry leaves. *Plant Biotechnology Journal*, 2(5), 381-387.
- Huang, B. and He, Y. (2007). Protein changes during heat stress in three Kentucky bluegrass cultivars differing in heat tolerance. *Crop Science*, 47(6), 2513-2520.
- Huetteman, C. A. and Preece, J. E. (1993). Thidiazuron: a potent cytokinin for woody plant tissue culture. *Plant Cell, Tissue and Organ Culture*, 33(2), 105-119.
- Hummer, K. E. and Hancock, J. (2009). Strawberry genomics: botanical history, cultivation, traditional breeding, and new technologies. *Genetics and Genomics of Rosaceae*, 6(7): 413-435.
- Husaini, A. M. (2010). Pre-and post-agroinfection strategies for efficient leaf disk transformation and regeneration of transgenic strawberry plants. *Plant Cell Reports*, 29(1), 97-110.
- Husaini, A. M. and Abdin, M. Z. (2008). Development of transgenic strawberry (*Fragaria x ananassa* Duch.) plants tolerant to salt stress. *Plant Science*, 174(4), 446-455.
- Husaini, A. M. and Rafiqi, A. M. (2012). Role of osmotin in strawberry improvement. *Plant Molecular Biology Reporter*, 30(5), 1055-1064.
- Ieamkhang, S. and Chatchawankanphanich, O. (2005). Augmentin as an alternative antibiotic for growth suppression of *Agrobacterium* for tomato (*Lycopersicon esculentum*) transformation. *Plant Cell, Tissue and Organ Culture*, 82(2), 213-220.
- James, D. J., Passey, A. J. and Barbara, D. J. (1990). *Agrobacterium*-mediated transformation of the cultivated strawberry (*Fragaria x ananassa* Duch.) using disarmed binary vectors. *Plant Science*, 69(1), 79-94.
- Jiménez-Bermúdez, S., Redondo-Nevado, J., Muñoz-Blanco, J., Caballero, J. L., López-Aranda, J. M., Valpuesta, V., Pliego-Alfaro, F., Quesada, M. A. and Mercado, J. A. (2002). Manipulation of strawberry fruit softening by antisense expression of a pectate lyase gene. *Plant Physiology*, 128(2), 751-759.

- Jin, W. M., Dong, J., Liu, Y., Zhang, Y. P. and Pan, Q. H. (2009). Genetically transformed strawberry (*Fragaria × ananassa* Duch.) with cold-inducible transcription factor CBF1. *Acta Horticulturae*, 842, 529-532.
- Jones, N. L. (2002). PCR. Principles, procedures, and parameters. *Methods in Molecular Biology*, 187, 37-46.
- Kadir, S., Sidhu, G. and Al-Khatib, K. (2006). Strawberry (*Fragaria xananassa* Duch.) growth and productivity as affected by temperature. *HortScience*, 41(6), 1423-1430.
- Kadir, S., Von Weihe, M. and Al-Khatib, K. (2007). Photochemical efficiency and recovery of photosystem II in grapes after exposure to sudden and gradual heat stress. *Journal of the American Society for Horticultural Science*, 132(6), 764-769.
- Karami, O., Esna-Ashari, M., Karimi Kurdistani, G. and Aghavaisi, B. (2009). *Agrobacterium*-mediated genetic transformation of plants: the role of host. *Biologia Plantarum*, 53(2), 201-212.
- Katiyar-Agarwal, S., Agarwal, M. and Grover, A. (2003). Heat-tolerant basmati rice engineered by over-expression of hsp101. *Plant Molecular Biology*, 51(5), 677-686.
- Kaur, A., Gill, M., Ruma, D. and Gosal, S. (2008). Enhanced *in vitro* shoot multiplication and elongation in sugarcane using cefotaxime. *Sugar Technology*, 10(1), 60-64.
- Keeler, S. J., Boettger, C. M., Haynes, J. G., Kuches, K. A., Johnson, M. M., Thureen, D. L., Keeler Jr, C. L. and Kitto, S. L. (2000). Acquired thermotolerance and expression of the HSP100/ClpB genes of lima bean. *Plant Physiology*, 123(3), 1121-1132.
- Komari, T., Takakura, Y., Ueki, J., Kato, N., Ishida, Y. and Hiei, Y. (2006). Binary vectors and super-binary vectors. *Methods in Molecular Biology*, 343, 15-41.
- Kost, B., Schnorf, M., Potrykus, I. and Neuhaus, G. (1995). Non-destructive detection of firefly luciferase (LUC) activity in single plant cells using a cooled, slow-scan CCD camera and an optimized assay. *The Plant Journal*, 8(1), 155-166.
- Kumakura, H. (1994). The effect of daytime, nighttime, and mean diurnal temperatures on the growth of 'Morioka-16' strawberry fruit and plants. *Journal of Japanese Society for Horticultural Science*, 62, 827-832.
- Kumar, V., Sharma, A., Narasimha Prasad, B. C., Bhaskar Gururaj, H. and Aswathanarayana Ravishankar, G. (2006). *Agrobacterium rhizogenes* mediated genetic transformation resulting in hairy root formation is enhanced by ultrasonication and acetosyringone treatment. *Electronic Journal of Biotechnology*, 9(4), 349-357.

- Kurien, B. T. and Scofield, R. H. (2005). Blotting techniques. *Encyclopedia of Analytical Science, Second Edition (Worsfold, P.J, Townshend, A., and Poole, C.F, eds.)*. Elsevier, Oxford, 425.
- Kurien, B. T. and Scofield, R. H. (2009). Introduction to protein blotting. *Methods in Molecular Biology*, 536, 9-22.
- Landi, L. and Mezzetti, B. (2006). TDZ, auxin and genotype effects on leaf organogenesis in *Fragaria*. *Plant Cell Reports*, 25(4), 281-288.
- Ledesma, N., Nakata, M. and Sugiyama, N. (2008). Effect of high temperature stress on the reproductive growth of strawberry cvs. 'Nyoho' and 'Toyonoka'. *Scientia Horticulturae*, 116(2), 186-193.
- Lee, K. S., Park, S. R. and Kim, Y. K. (2007). Effects of irradiance, temperature, and nutrients on growth dynamics of seagrasses: A review. *Journal of Experimental Marine Biology and Ecology*, 350(1-2), 144-175.
- Li, H., Li, T., Gordon, R. J., Asiedu, S. K. and Hu, K. (2010). Strawberry plant fruiting efficiency and its correlation with solar irradiance, temperature and reflectance water index variation. *Environmental and Experimental Botany*, 68(2), 165-174.
- Li, H., Zhang, Z., Huang, F., Chang, L. and Ma, Y. (2009). MicroRNA expression profiles in conventional and micropropagated strawberry (*Fragaria × ananassa* Duch.) plants. *Plant Cell Reports*, 28(6), 891-902.
- Li, J., Han, J., Chen, P., Chang, Z., He, Y., Liu, P. and Wang, Q. (2012). Cloning of a heat shock protein 90 (HSP90) gene and expression analysis in the ridgetail white prawn *Exopalaemon carinicauda*. *Fish and Shellfish Immunology*, 32(6), 1191-1197.
- Lin, J. J., Assad-Garcia, N. and Kuo, J. (1995). Plant hormone effect of antibiotics on the transformation efficiency of plant tissues by *Agrobacterium tumefaciens* cells. *Plant Science*, 109(2), 171-177.
- Liu, C. W., Chen, J. J. W., Kang, C. C., Wu, C. H. and Yiu, J. C. (2012). Transgenic lettuce (*Lactuca sativa* L.) expressing H1N1 influenza surface antigen (neuraminidase). *Scientia Horticulturae*, 139, 8-13.
- Liu, F., Guo, Y., Gu, D., Xiao, G., Chen, Z. and Chen, S. (1997). Salt tolerance of transgenic plants with BADH cDNA. *Acta Genetica Sinica*, 24(1), 54-58.
- Liu, H., Chen, H., Jing, J. and Ma, X. (2011). Cloning and characterization of the HSP90 beta gene from *Tanichthys albonubes* Lin (Cyprinidae): effect of copper and cadmium exposure. *Fish Physiology and Biochemistry*, 38(3), 1-12.
- Luehrsen, K. R. and Walbot, V. (1993). Firefly luciferase as a reporter for plant gene expression studies. *Promega Notes*, 44, 24-29.

- Lunkenbein, S., Coiner, H., de Vos, C. H. R., Schaart, J. G., Boone, M. J., Krens, F. A., Schwab, W. and Salentijn, E. M. J. (2006a). Molecular characterization of a stable antisense chalcone synthase phenotype in strawberry (*Fragaria* × *ananassa*). *Journal of Agricultural and Food Chemistry*, 54(6), 2145-2153.
- Lunkenbein, S., Salentijn, E. M. J., Coiner, H. A., Boone, M. J., Krens, F. A. and Schwab, W. (2006b). Up-and down-regulation of *Fragaria* × *ananassa* O-methyltransferase: impacts on furanone and phenylpropanoid metabolism. *Journal of Experimental Botany*, 57(10), 2445-2453.
- Mahajan, R., Kaur, R., Sharma, A. and Sharma, D. R. (2001). Micropropagation of strawberry cultivar Chandler and Fern. *Crop Improvement*, 28, 19-25.
- Mahajan, S. and Tuteja, N. (2005). Cold, salinity and drought stresses: An overview. *Archives of Biochemistry and Biophysics*, 444(2), 139-158.
- Manchado, M., Salas-Leiton, E., Infante, C., Ponce, M., Asensio, E., Crespo, A., Zuasti, E. and Cañavate, J. P. (2008). Molecular characterization, gene expression and transcriptional regulation of cytosolic HSP90 genes in the flatfish *Senegalese sole* (*Solea senegalensis* Kaup). *Gene*, 416(1), 77-84.
- Marcotrigiano, M., McGlew, S. P., Hackett, G. and Chawla, B. (1996). Shoot regeneration from tissue-cultured leaves of the American cranberry (*Vaccinium macrocarpon*). *Plant Cell, Tissue and Organ Culture*, 44(3), 195-199.
- Marta, A. E., Camadro, E. L., Díaz-Ricci, J. C. and Castagnaro, A. P. (2004). Breeding barriers between the cultivated strawberry, *Fragaria* × *ananassa*, and related wild germplasm. *Euphytica*, 136(2), 139-150.
- Mathews, H., Dewey, V., Wagoner, W. and Bestwick, R. K. (1998). Molecular and cellular evidence of chimaeric tissues in primary transgenics and elimination of chimaerism through improved selection protocols. *Transgenic Research*, 7(2), 123-129.
- Mathews, H., Wagoner, W., Kellogg, J. and Bestwick, R. (1995). Genetic transformation of strawberry: stable integration of a gene to control biosynthesis of ethylene. *In Vitro Cellular and Developmental Biology-Plant*, 31(1), 36-43.
- Mauromicale, G., Ierna, A. and Marchese, M. (2006). Chlorophyll fluorescence and chlorophyll content in field-grown potato as affected by nitrogen supply, genotype, and plant age. *Photosynthetica*, 44(1), 76-82.
- Mezzetti, B. (2009). GMO strawberry: methods, risk and benefits. *Genetics and Genomics of Rosaceae*, 6(7), 487-506.
- Milioni, D. and Hatzopoulos, P. (1997). Genomic organization of hsp90 gene family in *Arabidopsis*. *Plant Molecular Biology*, 35(6), 955-961.

- Molina-Bravo, R., Arellano, C., Sosinski, B. R. and Fernandez, G. E. (2011). A protocol to assess heat tolerance in a segregating population of raspberry using chlorophyll fluorescence. *Scientia Horticulturae*, 130(3), 524-530.
- Monjardino, P., Jones, R. J. and Smith, A. G. (2005). Heat stress effects on protein accumulation of maize endosperm. *Crop Science*, 45(4), 1203-1210.
- Morgan, A., Baker, C. M., Chu, J. S. F., Lee, K., Crandall, B. A. and Jose, L. (2002). Production of herbicide tolerant strawberry through genetic engineering. *Acta Horticulturae*, 567(1), 113-115.
- Murashige, T. and Skoog, F. (1962). A revised medium for rapid growth and bio assays with tobacco tissue cultures. *Physiologia Plantarum*, 15(3), 473-497.
- Murthy, B., Singh, R. and Saxena, P. K. (1996). Induction of high-frequency somatic embryogenesis in geranium (*Pelargonium x hortorum* Bailey cv. Ringo Rose) cotyledonary cultures. *Plant Cell Reports*, 15(6), 423-426.
- Na, J., Wang, G., Xia, R. and Yang, H. (2006). Construction of anti-sense gene of *annfap* and genetic transformation of strawberry. *Scientia Agricultura Sinica*, 39, 582-586.
- Nagel, R. J., Manners, J. M. and Birch, R. G. (1992). Evaluation of an ELISA assay for rapid detection and quantification of neomycin phosphotransferase II in transgenic plants. *Plant Molecular Biology Reporter*, 10(3), 263-272.
- Nehra, N. S., Chibbar, R. N., Kartha, K. K., Datla, R. S. S., Crosby, W. L. and Stushnoff, C. (1990). Genetic transformation of strawberry by *Agrobacterium tumefaciens* using a leaf disk regeneration system. *Plant Cell Reports*, 9(6), 293-298.
- Netto, A. T., Campostrini, E., de Oliveira, J. G. and Bressan-Smith, R. E. (2005). Photosynthetic pigments, nitrogen, chlorophyll a fluorescence and SPAD-502 readings in coffee leaves. *Scientia Horticulturae*, 104(2), 199-209.
- Nolan, T., Hands, R. E. and Bustin, S. A. (2006). Quantification of mRNA using real-time RT-PCR. *Nature Protocols*, 1(3), 1559-1582.
- Nyman, M. and Wallin, A. (1992). Transient gene expression in strawberry (*Fragaria x ananassa* Duch.) protoplasts and the recovery of transgenic plants. *Plant Cell Reports*, 11(2), 105-108.
- Obembe, O., Adesoye, A. and Machuka, J. (2005). Evaluation of the effectiveness of selected antibiotics in the suppression of *Agrobacterium* from Cowpea (*Vigna unguiculata* L. Walp.) embryo explants and as potential selective agents in *Agrobacterium*-mediated transformation. *Journal of Tropical Microbiology and Biotechnology*, 1(1), 27-33.
- Olhoft, P. M., Flagel, L. E., Donovan, C. M. and Somers, D. A. (2003). Efficient soybean transformation using hygromycin B selection in the cotyledonary-node method. *Planta*, 216(5), 723-735.

- Oosumi, T., Gruszewski, H. A., Blischak, L. A., Baxter, A. J., Wadl, P. A., Shuman, J. L., Veilleux, R. E. and Shulaev, V. (2006). High-efficiency transformation of the diploid strawberry (*Fragaria vesca*) for functional genomics. *Planta*, 223(6), 1219-1230.
- Padilla, I. M. G. and Burgos, L. (2010). Aminoglycoside antibiotics: structure, functions and effects on *in vitro* plant culture and genetic transformation protocols. *Plant Cell Reports*, 29(11), 1203-1213.
- Palomer, X., Llop-Tous, I., Vendrell, M., Krens, F. A., Schaart, J. G., Boone, M. J., van der Valk, H. and Salentijn, E. M. J. (2006). Antisense down-regulation of strawberry *endo*- β -(1, 4)-glucanase genes does not prevent fruit softening during ripening. *Plant Science*, 171(5), 640-646.
- Pan, L., Zhang, Y., Wang, Y., Lv, J., Zhou, P., Zhang, Z., Fang, Y. and Jiang, S. (2011). Expression and detection of the FMDV VP1 transgene and expressed structural protein in *Arabidopsis thaliana*. *Turkish Journal of Veterinary and Animal Sciences*, 35(4), 235-242.
- Park, H. S., Jeong, W. J., Kim, E., Jung, Y., Lim, J. M., Hwang, M. S., Park, E. J., Ha, D. S. and Choi, D. W. (2012). Heat shock protein gene family of the *Porphyra seriata* and enhancement of heat stress tolerance by PsHSP70 in *Chlamydomonas*. *Marine Biotechnology*, 14(3), 332-342.
- Passey, A., Barrett, K. and James, D. (2003). Adventitious shoot regeneration from seven commercial strawberry cultivars (*Fragaria* \times *ananassa* Duch.) using a range of explant types. *Plant Cell Reports*, 21(5), 397-401.
- Pauli, S., Rothnie, H. M., Chen, G., He, X. and Hohn, T. (2004). The cauliflower mosaic virus 35S promoter extends into the transcribed region. *Journal of Virology*, 78(22), 12120-12128.
- Percival, G. C., Keary, I. P. and Noviss, K. (2008). The potential of a chlorophyll content SPAD meter to quantify nutrient stress in foliar tissue of Sycamore (*Acer pseudoplatanus*), English Oak, (*Quercus robur*), and European Beech (*Fagus sylvatica*). *Arboriculture and Urban Forestry*, 34(2), 89-100.
- Petri, C., Lopez-Noguera, S., Albuquerque, N., Egea, J. and Burgos, L. (2008). An antibiotic-based selection strategy to regenerate transformed plants from apricot leaves with high efficiency. *Plant Science*, 175(6), 777-783.
- Picard, D. (2002). Heat-shock protein 90, a chaperone for folding and regulation. *Cellular and Molecular Life Sciences*, 59(10), 1640-1648.
- Picard, D. (2006). Chaperoning steroid hormone action. *Trends in Endocrinology and Metabolism*, 17(6), 229-235.
- Plus, J., George, L., Eapen, S. and Rao, P. (1993). Enhanced plant regeneration in pearl millet (*Pennisetum americanum*) by ethylene inhibitors and cefotaxime. *Plant Cell, Tissue and Organ Culture*, 32(1), 91-96.

- Porter, J. R. (2005). Rising temperatures are likely to reduce crop yields. *Nature*, 436(7048), 174-174.
- Prasad, P. V. V., Craufurd, P. Q., Summerfield, R. J. and Wheeler, T. R. (2000). Effects of short episodes of heat stress on flower production and fruit-set of groundnut (*Arachis hypogaea* L.). *Journal of Experimental Botany*, 51(345), 777-784.
- Pyrotis, S., Abayomi, L., Rees, D. and Orchard, J. (2012). Effect of temperature and humidity on strawberry firmness at two different sites in the Huelva region of Spain. *Acta Horticulturae*, 926, 567-570.
- Qin, Y., Shanglong, Z., Asghar, S., Lingxiao, Z., Qiaoping, Q., Kunsong, C. and Changjie, X. (2005). Regeneration mechanism of Toyonoka strawberry under different color plastic films. *Plant Science*, 168(6), 1425-1431.
- Qin, Y., Teixeira da Silva, J. A., Bi, J., Zhang, S. and Hu, G. (2011). Response of in vitro strawberry to antibiotics. *Plant Growth Regulation*, 65(1), 1-11.
- Qin, Y., Teixeira da Silva, J. A., Zhang, L. and Zhang, S. (2008). Transgenic strawberry: State of the art for improved traits. *Biotechnology Advances*, 26(3), 219-232.
- Qin, Y. and Zhang, S. (2007). Factors influencing the efficiency of *Agrobacterium*-mediated transformation in strawberry cultivar *Toyonaka*. *Journal of Nuclear Agricultural Science*, 21, 461-465.
- Queitsch, C., Hong, S. W., Vierling, E. and Lindquist, S. (2000). Heat shock protein 101 plays a crucial role in thermotolerance in *Arabidopsis*. *Plant Cell*, 12(4), 479-492.
- Quesada, M., Martin-Pizarro, C., Garcia-Gago, J., Posé, S., Santiago, N., Sesmero, R., Pliego-Alfaro, F. and Mercado, J. (2007). Transgenic strawberry: current status and future perspectives. *Transgenic Plant Journal*, 1, 280-288.
- Quirino, B. F. and Bent, A. F. (2003). Deciphering host resistance and pathogen virulence: the *Arabidopsis/Pseudomonas* interaction as a model. *Molecular Plant Pathology*, 4(6), 517-530.
- Rafat, A., Aziz, M. A., Rashid, A. A., Abdullah, S. N. A., Kamaladini, H., Sirchi, M. and Javadi, M. (2010). Optimization of *Agrobacterium tumefaciens*-mediated transformation and shoot regeneration after co-cultivation of cabbage (*Brassica oleracea* subsp. capitata) cv. KY Cross with *AtHSP101* gene. *Scientia Horticulturae*, 124(1), 1-8.
- Rao, A., Sree, K. P. and Kishor, P. B. K. (1995). Enhanced plant regeneration in grain and sweet sorghum by asparagine, proline and cefotaxime. *Plant Cell Reports*, 15(1), 72-75.
- Sabelli, P. A. and Shewry, P. R. (1995). Northern analysis and nucleic acid probes. *Methods in Molecular Biology-Clifton then Totowa*, 49, 213-228.

- Salas, M., Park, S., Srivatanakul, M. and Smith, R. (2001). Temperature influence on stable T-DNA integration in plant cells. *Plant Cell Reports*, 20(8), 701-705.
- Sambrook, J. and Russell, D. W. (2001). *Molecular cloning: a laboratory manual* (Vol. 2): CSHL press.
- Sarma, K., Evans, N. E. and Selby, C. (1995). Effect of carbenicillin and cefotaxime on somatic embryogenesis of Sitka spruce (*Picea sitchensis* (Bong.) Carr.). *Journal of Experimental Botany*, 46(11), 1779-1781.
- Schaart, J. G., Krens, F. A., Pelgrom, K. T. B., Mendes, O. and Rouwendal, G. J. A. (2004). Effective production of marker-free transgenic strawberry plants using inducible site-specific recombination and a bifunctional selectable marker gene. *Plant Biotechnology Journal*, 2(3), 233-240.
- Schestibratov, K. and Dolgov, S. (2005). Transgenic strawberry plants expressing a thaumatin II gene demonstrate enhanced resistance to *Botrytis cinerea*. *Scientia Horticulturae*, 106(2), 177-189.
- Schnurr, J. and Guerra, D. (2000). The CaMV35S promoter is sensitive to shortened photoperiod in transgenic tobacco. *Plant cell reports*, 19(3), 279-282.
- Shin, S., Kim, I. H., Kang, W., Yang, J. K. and Hah, S. S. (2010). An alternative to Western blot analysis using RNA aptamer-functionalized quantum dots. *Bioorganic and Medicinal Chemistry Letters*, 20(11), 3322-3325.
- Shou, H., Frame, B. R., Whitham, S. A. and Wang, K. (2004). Assessment of transgenic maize events produced by particle bombardment or *Agrobacterium*-mediated transformation. *Molecular Breeding*, 13(2), 201-208.
- Signore, M. and Reeder, K. A. (2012). Antibody validation by Western blotting *Molecular Profiling: Methods and Protocols, Methods in Molecular Biology* Vol. 823, pp. 139-155.
- Singh, A. and Grover, A. (2008). Genetic engineering for heat tolerance in plants. *Physiology and Molecular Biology of Plants*, 14(1-2), 155-166.
- Stam, M., Mol, J. N. M. and Kooter, J. M. (1997). Review article: The silence of genes in transgenic plants. *Annals of Botany*, 79(1), 3-12.
- Székács, A., Hegedus, G., Tóbiás, I., Pogány, M. and Barna, B. (2000). Immunoassays for plant cytokinins as tools for the assessment of environmental stress and disease resistance. *Analytica Chimica Acta*, 421(2), 135-146.
- Tang, J., Liang, S., Zhang, J., Gao, Z. and Zhang, S. (2009). pGreen-S: A clone vector bearing absence of enhanced green fluorescent protein for screening recombinants. *Analytical Biochemistry*, 388(1), 173-174.

- Teixeira da Silva, J. A. T. (2003). Filter paper type affects the morphogenic programs and buffers the phytotoxic effect of antibiotics in chrysanthemum and tobacco thin cell layers. *HortScience*, 38(7), 1403-1407.
- Teixeira da Silva, J. A. T. and Fukai, S. (2001). The impact of carbenicillin, cefotaxime and vancomycin on chrysanthemum and tobacco TCL morphogenesis and *Agrobacterium* growth. *Journal of Applied Horticulture*, 3(1), 3-12.
- Towbin, H., Staehelin, T. and Gordon, J. (1979). Electrophoretic transfer of proteins from polyacrylamide gels to nitrocellulose sheets: procedure and some applications. *Proceedings of the National Academy of Sciences*, 76(9), 4350-4354.
- Tulipani, S., Marzban, G., Herndl, A., Laimer, M., Mezzetti, B. and Battino, M. (2011). Influence of environmental and genetic factors on health-related compounds in strawberry. *Food Chemistry*, 124(3), 906-913.
- Vellicce, G. R., Ricci, J. C. D., Hernández, L. and Castagnaro, A. P. (2006). Enhanced resistance to *Botrytis cinerea* mediated by the transgenic expression of the chitinase gene ch5B in strawberry. *Transgenic Research*, 15(1), 57-68.
- Vicente, A. R., Costa, M. L., Martínez, G. A., Chaves, A. R. and Civello, P. M. (2005). Effect of heat treatments on cell wall degradation and softening in strawberry fruit. *Postharvest Biology and Technology*, 38(3), 213-222.
- Vierling, E. (1991). The roles of heat shock proteins in plants. *Annual Review of Plant Biology*, 42(1), 579-620.
- Wadl, P. A. (2006). *Improved regeneration and Agrobacterium-mediated transformation of wild strawberry (Fragaria vesca L.)*. Virginia Polytechnic Institute and State University.
- Wang, S. Y. and Camp, M. J. (2000). Temperatures after bloom affect plant growth and fruit quality of strawberry. *Scientia Horticulturae*, 85(3), 183-199.
- Wang, J., Ge, H., Peng, S., Zhang, H., Chen, P. and Xu, J. (2004a). Transformation of strawberry (*Fragaria x ananassa* Duch.) with late embryogenesis abundant protein gene. *Journal of Horticultural Science and Biotechnology*, 79(5), 735-738.
- Wang, W., Vinocur, B. and Altman, A. (2003). Plant responses to drought, salinity and extreme temperatures: towards genetic engineering for stress tolerance. *Planta*, 218(1), 1-14.
- Wang, W., Vinocur, B., Shoseyov, O. and Altman, A. (2004b). Role of plant heat-shock proteins and molecular chaperones in the abiotic stress response. *Trends in Plant Science*, 9(5), 244-252.

- Waters, E. R., Lee, G. J. and Vierling, E. (1996). Evolution, structure and function of the small heat shock proteins in plants. *Journal of Experimental Botany*, 47(3), 325-338.
- Wehr, N. and Levine, R. (2012). Quantitation of protein carbonylation by dot blot. *Analytical Biochemistry*, 423(2), 241-245.
- Weng, J. H. and Lai, M. F. (2005). Estimating heat tolerance among plant species by two chlorophyll fluorescence parameters. *Photosynthetica*, 43(3), 439-444.
- Willits, D. and Peet, M. (2001). Measurement of chlorophyll fluorescence as a heat stress indicator in tomato: laboratory and greenhouse comparisons. *Journal of the American Society for Horticultural Science*, 126(2), 188-194.
- Woolley, L., James, D. and Manning, K. (2001). Purification and properties of an endo- β -1,4-glucanase from strawberry and down-regulation of the corresponding gene, cell1. *Planta*, 214, 11-21.
- Wu, H., Sparks, C., Amoah, B. and Jones, H. (2003). Factors influencing successful *Agrobacterium*-mediated genetic transformation of wheat. *Plant Cell Reports*, 21(7), 659-668.
- Xu-Gang, L., Song-Biao, C., Zi-Xian, L., Tuan-Jie, C., Qian-Chun, Z. and Zhen, Z. (2002). Impact of copy number on transgene expression in tobacco. *Acta Botanica Sinica*, 44, 120-123.
- Yan, X., Chenyang, Z. and Bingru, H. (2011). Heat shock proteins in association with heat tolerance in grasses. *International Journal of Proteomics*, 2011, 1-11.
- Yang, J., Sun, Y., Sun, A., Yi, S., Qin, J., Li, M. and Liu, J. (2006). The involvement of chloroplast HSP100/ClpB in the acquired thermotolerance in tomato. *Plant Molecular Biology*, 62(3), 385-395.
- Zambre, M., Terryn, N., De Clercq, J., De Buck, S., Dillen, W., Van Montagu, M., Van Der Straeten, D. and Angenon, G. (2003). Light strongly promotes gene transfer from *Agrobacterium tumefaciens* to plant cells. *Planta*, 216(4), 580-586.
- Zhang, Y., Seeram, N. P., Lee, R., Feng, L. and Heber, D. (2008). Isolation and identification of strawberry phenolics with antioxidant and human cancer cell antiproliferative properties. *Journal of Agricultural and Food Chemistry*, 56(3), 670-675.
- Zhang, Z. H. and Wu, L. P. (1998). Development of genetic transformation system of the commercial strawberry cultivar "Tudla". *Journal of Agricultural Biotechnology*, 6, 200-204.
- Zhao, Y., Liu, Q. and Davis, R. E. (2004). Transgene expression in strawberries driven by a heterologous phloem-specific promoter. *Plant Cell Reports*, 23(4), 224-230.

Ziemenowicz, A. (2001). Plant selectable markers and reporter genes. *Acta Physiologiae Plantarum*, 23(3), 363-374.



© COPYRIGHT UPM