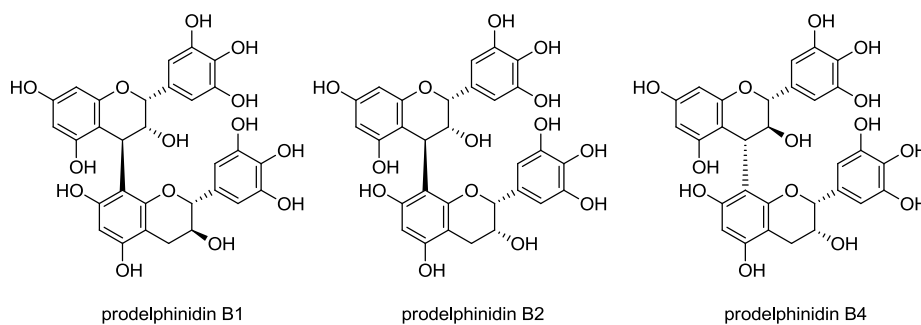


## Graphical Abstract

### Syntheses of prodelphinidin B1, B2 and B4 and their antitumor activities against human PC-3 prostate cancer cell lines

Wataru Fujii, Kazuya Toda, Kiriko Matsumoto, Koichiro Kawaguchi, Sei-ichi Kawahara, Yasunao Hattori, Hiroshi Fujii and Hidefumi Makabe



# Syntheses of prodelphinidin B1, B2 and B4 and their antitumor activities against human PC-3 prostate cancer cell lines

Wataru Fujii,<sup>a</sup> Kazuya Toda,<sup>b</sup> Kiriko Matsumoto,<sup>b</sup> Koichiro Kawaguchi,<sup>b</sup> Sei-ichi Kawahara,<sup>c</sup> Yasunao Hattori,<sup>d</sup> Hiroshi Fujii\*,<sup>b</sup> and Hidefumi Makabe\*<sup>a</sup>

<sup>a</sup>Graduate School of Agriculture, Sciences of Functional Foods, Shinshu University,

8304 Minami-minowa Kami-ina, Nagano, 399-4598, Japan

<sup>b</sup>Department of Bioscience and Biotechnology, Faculty of Agriculture, Shinshu University, 8304 Minami-minowa Kami-ina, Nagano, 399-4598, Japan

<sup>c</sup>St. Cousair Co., Ltd., 1260 Imogawa, Kami-minochi, Nagano, 389-1201, Japan

<sup>d</sup>Department of Medicinal Chemistry, Kyoto Pharmaceutical University, Yamashina-ku, Kyoto 607-8412, Japan

\*Corresponding author. Tel. +81 265 77 1626; fax +81 265 77 1626; e-mail: hfujii@shinshu-u.ac.jp

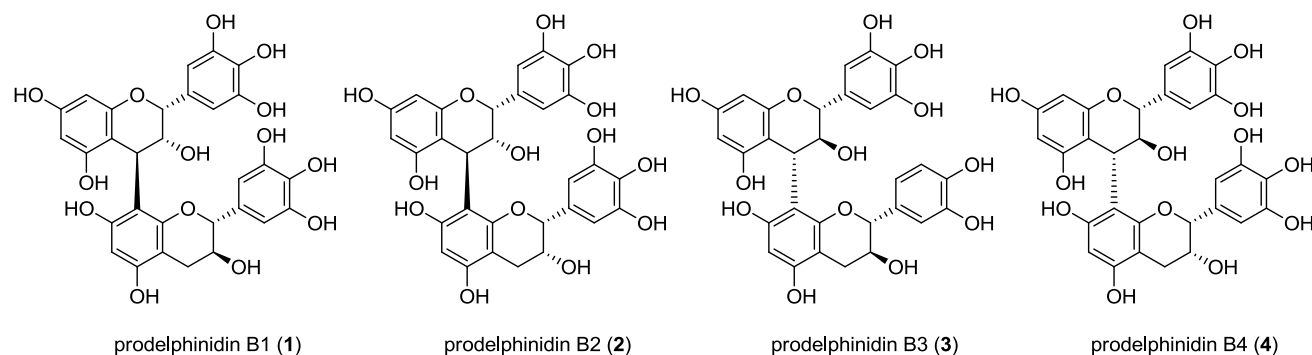
\*Corresponding author. Tel. +81 265 77 1630; fax +81 265 77 1700; e-mail: makabeh@shinshu-u.ac.jp

**Abstract:** Total synthesis of prodelphinidin B1, B2, and B4 have been accomplished. The key step is Lewis acid-mediated equimolar condensations between an epigallocatechin and/or a galliccatechin nucleophile and an epigallocatechin and/or a galliccatechin electrophile. The antitumor effects of synthetic prodelphinidin B1-B4 against human PC-3 prostate cancer cell lines have been investigated. These compounds showed significant antitumor effects. Their activity seemed to be little bit stronger than EGCG and prodelphinidin B3, known antitumor agent.

**Key words:** polyphenols, synthesis, natural product, anticancer agents

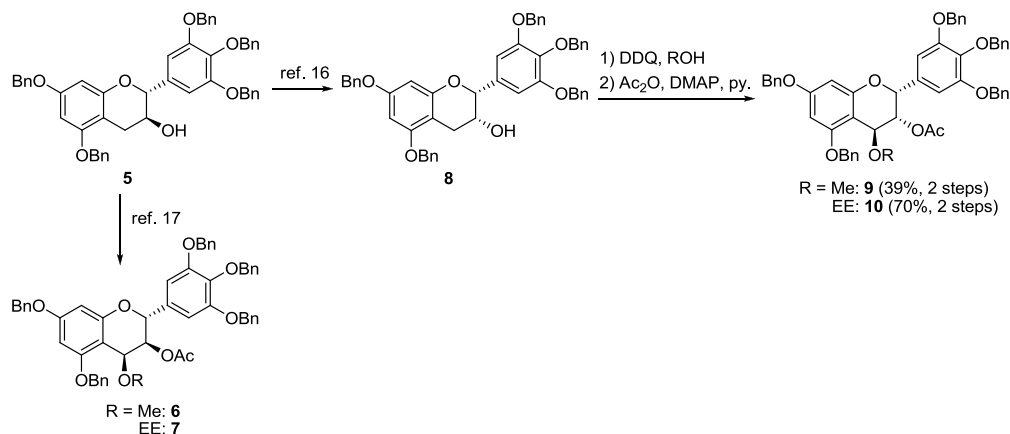
Prodelphinidins are paid much attention due to their significant biological activities. For example, prodelphinidin B2 3-*O*-gallate and 3,3'-di-*O*-gallate inhibit proliferation of A549 cancer cells,<sup>1,2</sup> and prodelphinidin B4 3'-*O*-gallate inhibits COX-2 and iNOS.<sup>3</sup> As to the prodelphinidins B1, B2 and B4, a number of isolation form plants have been reported; prodelphinidin B1 (**1**) from *Citstus incanus*<sup>4</sup> and *Lotus pedunculatus*<sup>5</sup>, and prodelphinidin B2 (**2**) from *Lotus pedunculatus*,<sup>5</sup> and prodelphinidin B4 (**4**) from *Ribes nigrum*,<sup>6</sup> *Vicia faba*,<sup>7</sup> and *Stryphnodendron adstringens*.<sup>8</sup> Because purification and identification of prodelphinidins from plants are very difficult, the

mechanism for their biological activities remains unknown. Thus syntheses of prodelphinidins are quite important to obtain pure materials for evaluating their biological activities. The many examples of the syntheses of procyanidins were reported in this decade including our syntheses,<sup>9-14</sup> however, synthetic studies on prodelphinidins are quite limited due to difficulty in obtaining (-)-gallocatechin or (+)-epigallocatechin as synthetic starting materials.<sup>15</sup> Although (-)-gallocatechin or (+)-epigallocatechin is commercially available, both of compounds are very expensive. Thus it was necessary to prepare enough amount of (-)-gallocatechin or (+)-epigallocatechin derivatives according to the reported procedure.<sup>16</sup> Until now only an example of total synthesis of prodelphinidin B3 (**3**) and C2 has been reported by us using equimolar coupling between nucleophilic and electrophilic partners using Lewis acid.<sup>17</sup> Herein we demonstrate equimolar condensation of (-)-gallocatechin and/or (+)-gallocatechin nucleophile with a gallocatechin and/or epigallocatechin derived electrophile and the first total syntheses of prodelphinidin B1 (**1**), B2 (**2**) and B4 (**4**) (Figure 1).



**Figure 1.** The structures of prodelphinidin B1 (**1**)-B4 (**4**).

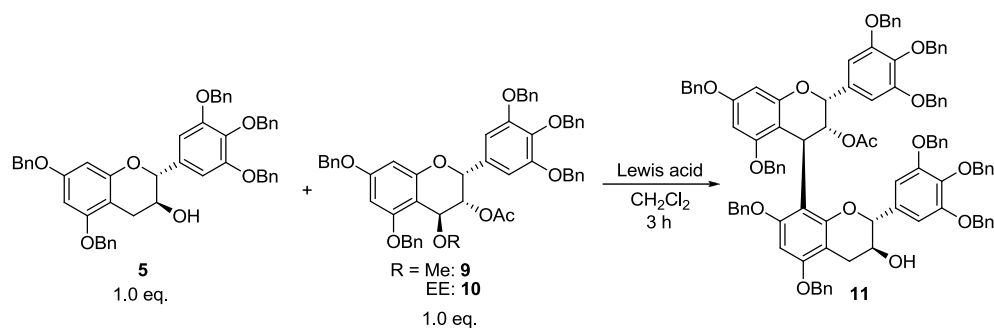
The gallocatechin-derived nucleophile **5** was constructed as Chan and co-workers reported.<sup>16</sup> Gallocatechin-derived electrophiles **6** and **7** were prepared as we reported earlier.<sup>17</sup> The epigallocatechin-derived nucleophile **8** was prepared according to the Chan and co-workers' method.<sup>16</sup> DDQ oxidation of **8** in the presence of methanol or ethoxyethanol followed by acetylation gave epigallocatechin-derived electrophiles **9** and **10**, respectively (Scheme 1).



**Scheme 1.** Synthesis of gallocatechin and/or epigallocatechin nucleophiles and electrophiles.

First, we examined the condition of equimolar condensation of gallo catechin nucleophile **5** with epigallo catechin electrophile **9** or **10** to construct prodelphinidin B1 (**1**) skeleton. We chose Yb(OTf)<sub>3</sub> as a Lewis acid for condensation because equimolar coupling worked well in the case of procyaninidin dimers.<sup>10d,10h</sup> We also examined silver Lewis acids because Ferreira and co-workers reported that using AgBF<sub>4</sub> as the thiophilic Lewis acid offered advantages to control the level of oligomeration in the synthesis of procyaninidin B1-B4.<sup>18</sup> As shown in Table 1, 4-(2''-ethoxyethoxy) derivative **10** afforded condensed product **11** in 66% yield when Yb(OTf)<sub>3</sub> was used as Lewis acid. On the other hand, the reaction using methoxy derivative **9** gave **11** in moderate to poor yield. We found that the choice of leaving group at the C4 position and Lewis acid was important for equimolar condensation (Table 1).<sup>19</sup>

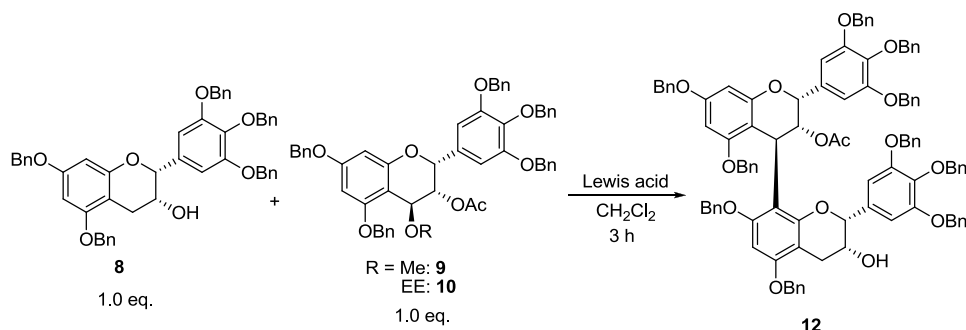
**Table 1.** Equimolar condensation of gallo catechin nucleophile **5** with epigallo catechin electrophile **9** or **10**.<sup>a</sup>



entry	electrophile	Lewis acid	yield (%)
1	<b>9</b>	Yb(OTf) <sub>3</sub>	7
2	<b>9</b>	AgOTf	54
3	<b>9</b>	AgBF <sub>4</sub>	33
4	<b>10</b>	Yb(OTf) <sub>3</sub>	66

<sup>a</sup> All reactions were carried out at room temperature.

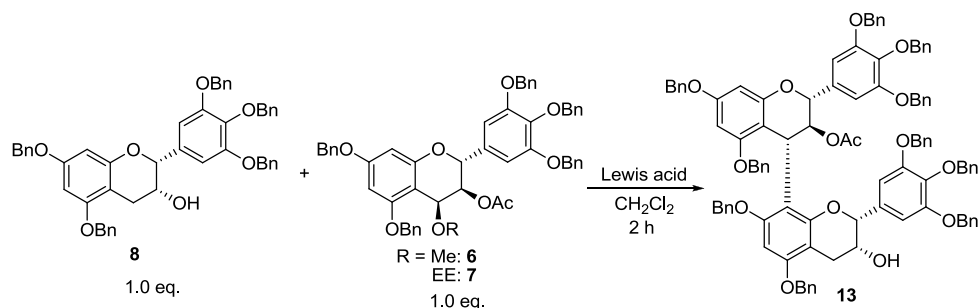
Next, we investigated the condition of equimolar condensation of epigallo catechin nucleophile **8** with epigallo catechin electrophile **9** or **10** to construct prodelphinidin B2 (**2**) skeleton. As shown in Table 2, methoxy derivative **9** afforded condensed product **12** in good yield when AgBF<sub>4</sub> was used as Lewis acid. Yb(OTf)<sub>3</sub> also gave **12** in good yield. In this case, we found that the methoxy group at the C-4 position was important for Lewis acid mediated condensation (Table 2).

**Table 2.** Equimolar condensation of epigallocatechin nucleophile **8** with epigallocatechin electrophile **9** or **10**.<sup>a</sup>

entry	electrophile	Lewis acid	yield (%)
1	<b>9</b>	Yb(OTf) <sub>3</sub>	70
2	<b>9</b>	AgOTf	53
3	<b>9</b>	AgBF <sub>4</sub>	76
4	<b>10</b>	Yb(OTf) <sub>3</sub>	22

<sup>a</sup> All reactions were carried out at room temperature.

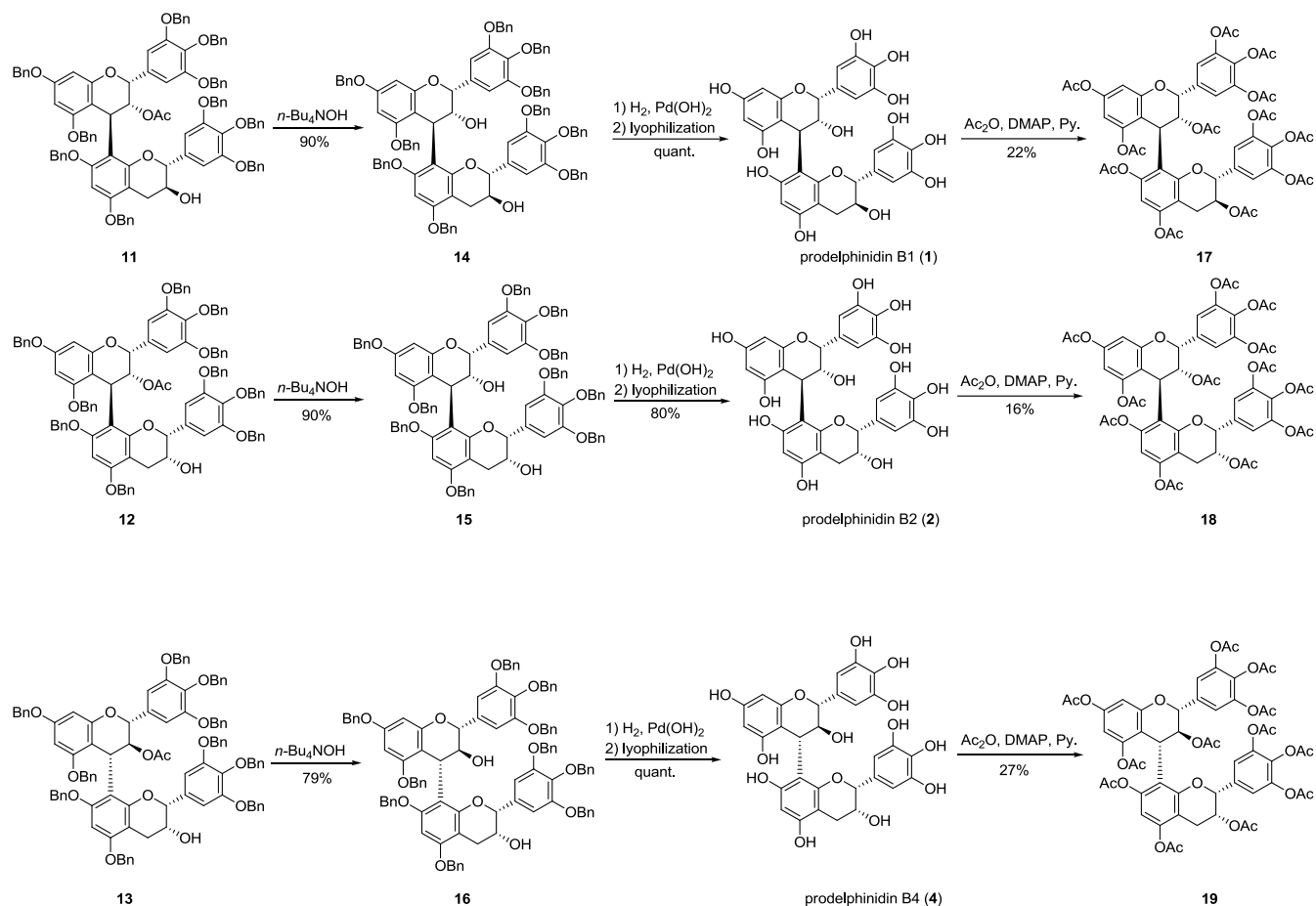
We further investigated the condition of equimolar condensation of epigallocatechin nucleophile **8** with gallo catechin electrophile **6** or **7** to construct prodelphinidin B4 (**4**) skeleton. As shown in Table 3, 4-(2''-ethoxyethoxy) derivative **7** afforded condensed product **13** in 78% yield when Yb(OTf)<sub>3</sub> was used as Lewis acid. On the other hand, the reaction using methoxy derivative **6** gave **13** in moderate to poor yield. (Table 3).

**Table 3.** Equimolar condensation of epigallocatechin nucleophile **8** with gallo catechin electrophile **6** or **7**.<sup>a</sup>

entry	electrophile	Lewis acid	yield (%)
1	<b>6</b>	Yb(OTf) <sub>3</sub>	48
2	<b>6</b>	AgOTf	49
3	<b>6</b>	AgBF <sub>4</sub>	62
4	<b>7</b>	Yb(OTf) <sub>3</sub>	78

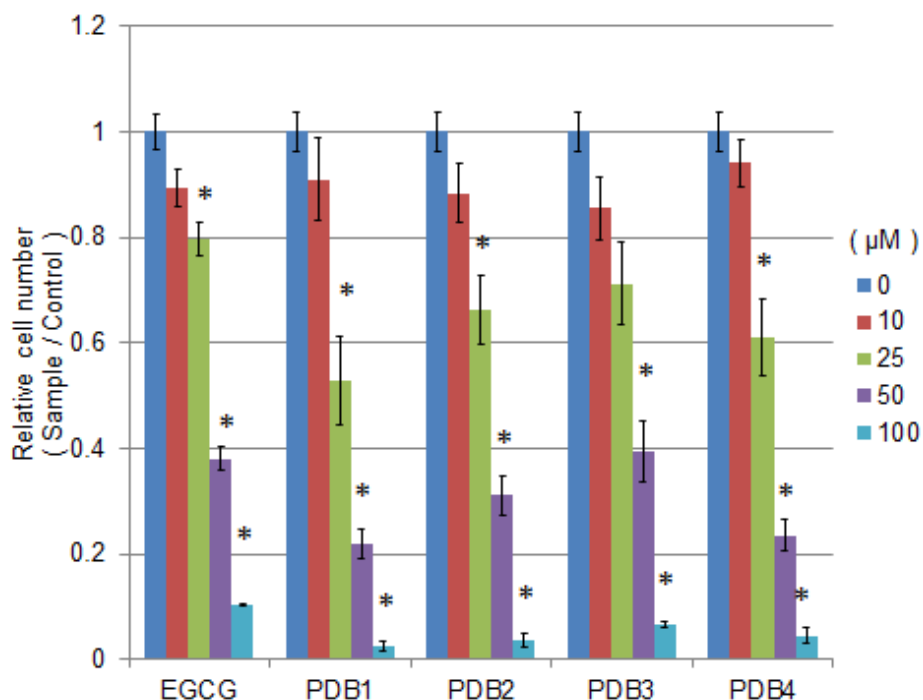
<sup>a</sup> All reactions were carried out at room temperature.

The condensed products **11-13** were transformed into diols **14-16** using *n*-Bu<sub>4</sub>NOH.<sup>13c</sup> Finally deprotection of the benzyl ethers of **14-16** and subsequent lyophilization afforded prodelphinidin B1 (**1**), B2 (**2**), and B4 (**4**) in good yield.<sup>20</sup> The <sup>1</sup>H and <sup>13</sup>C NMR spectral data of peracetate of **1** (**17**),<sup>5</sup> **2** (**18**),<sup>5</sup> and **4** (**19**)<sup>7</sup> were in good agreement with the reported values (Scheme 2).



**Scheme 2.** Synthesis of prodelpinidin B1 (**1**), B2 (**2**), and B4 (**4**) and their peracetate **17-19**.

Because prodelpinidin B1 (**1**), B2 (**2**), and B4 (**4**) were obtained in sufficient quantities, we investigated antitumor activities against PC-3 prostate cancer cell lines. Results were obtained by cell count measurement. Epigallocatechin gallate (EGCG) and prodelpinidin B3 (**3**) were used as positive controls. As shown in Figure 2, EGCG, prodelpinidin B1 (**1**), B2 (**2**), and B4 (**4**) exhibited significant cytotoxic activities with  $\text{IC}_{50}$  values below 50  $\mu\text{M}$ . At higher concentration ( $>50 \mu\text{M}$ ), prodelpinidin B1 (**1**), B2 (**2**), and B4 (**4**) which have two pyrogallol moieties seemed to be stronger activity than that of prodelpinidin B3 (**3**) which has one pyrogallol moiety. The additional pyrogallol moieties might enhance the cytotoxic effects. Making a comparison of prodelpinidin B1 (**1**), B2 (**2**), and B4 (**4**) with EGCG, the activities of **1**, **2** and **4** seemed to be a little bit stronger than that of EGCG at higher concentration ( $>25 \mu\text{M}$ ). Recently we have examined the cytotoxic effects on PC-3 prostate cancer cell lines of procyanidin gallates and found that esterified pyrogallol moiety showed weaker activity than prodelpinidin B3 (**3**).<sup>13</sup> EGCG has two pyrogallol moieties but one of them is esterified one. This might be a reason of weaker activity of EGCG than prodelpinidins B1, B2, and B4 (Figure 2).



**Figure 2.** Effects of various concentrations of test compounds on cell proliferation.

After treatment of cells with EGCG, prodelphinidin B1 (**1**, PDB1), prodelphinidin B2 (**2**, PDB2), prodelphinidin B3 (**3**, PDB3),<sup>17</sup> and prodelphinidin B4 (**4**, PDB4) for 48 h, the cell proliferation was determined by cell count as described in experimental section. The values were represented as the rate of inhibition of cell proliferation by the treated sample compared to the untreated control (vehicle). Values are means  $\pm$  S.Ds. for three independent experiments. Asterisks indicated a significant difference between the control- and test-compound-treated cells, as analyzed by Student's test ( $p < 0.001$ ).

The first total syntheses of prodelphinidin B1 (**1**), B2 (**2**) and B4 (**4**) have been achieved via Lewis acid-mediated equimolar condensation of a gallicocatechin and/or epigallocatechin nucleophile with gallicocatechin and/or epigallocatechin electrophiles. In addition to demonstrating the total synthesis, we examined their antitumor activities against PC-3 prostate cancer cells. Prodelphinidin B1 (**1**), B2 (**2**), and B4 (**4**) showed significant cytotoxic activity with  $IC_{50}$  values below 50  $\mu$ M. The potencies of prodelphinidins **1**, **2**, and **4** seemed to be a little bit stronger than those of EGCG and prodelphinidin B3 (**3**).

### Acknowledgements

This work was supported in part by Grant-in-Aid for Scientific Research from the Ministry of Education, Science, Culture, Sports, and Technology of Japan (22570112 to H. F.), by Grant from Uehara Memorial Foundation (to H. F.) and Shinshu Foundation for Promotion of Agricultural and Forest Science (to H. M.). We also thank Prof. Dr. Toshiyuki Kan for providing EGCG.

## Supplementary data

Supplementary data associated with this article can be found, in the online version, at <http://dx.doi>

## References and notes

1. Kuo, P. L.; Hsu Y. L.; Lin T. C.; Lin, C. C. *Food and Chem. Toxic.* **2005**, *43*, 315.
2. Kuo, P. L.; Hsu Y. L.; Lin T. C.; Lin, C. C. *Eur. J. Phem.* **2004**, *501*, 41.
3. Hou, D.-X.; Luo, D.; Tanigawa, S.; Hashimoto, F.; Uto, T.; Masuzaki, S.; Fujii, M.; Sakata, Y. *Biochem. Pharm.* **2007**, *74*, 742.
4. Danne, A.; Peterreit, F.; Nahrstedt, A. *Phytochemistry* **1993**, *34*, 1129.
5. Foo, L. Y.; Lu, Y.; McNabb, W. C.; Waghor, G.; Ulyatt, M. J. *Phytochemistry* **1997**, *45*, 1689.
6. Tits, M.; Angenot, L.; Poukens, P.; Warin, R. Dierckxsens, Y. *Phytochemistry* **1992**, *31*, 971.
7. Helsper, J. P. F. G.; Kolodziej, H.; Hoogendijk J. M.; Van Norel, A. *Phytochemistry* **1993**, *34*, 1255.
8. De Mello, J. P.; Peterreit, F.; Nahrstedt, A. *Phytochemistry* **1996**, *41*, 807.
9. Recent review of synthesis of procyanidins: (a) Ferreira, D.; Coleman, C. M. *Planta Med.* **2011**, *77*, 1071; (b) Oyama, K. -i.; Yoshida, K.; Kondo, T. *Curr. Org. Chem.* **2011**, *15*, 2567; (c) Ohmori, K.; Suzuki, K. *Curr. Org. Chem.* **2012**, *16*, 566.
10. Recent syntheses of procyanidin dimers: (a) Nakajima, N.; Horikawa, K.; Takekawa, N.; Hamada, M.; Kishimoto, T. *Heterocycles* **2012**, *84*, 349; (b) Katoh, M.; Oizumi, Y.; Mohri, Y.; Hirota, M.; Makabe, H. *Lett. Org. Chem.* **2012**, *9*, 233; (c) Alharthy, R. D.; Hayes, C. J. *Tetrahedron Lett.* **2010**, *51*, 1193; (d) Oizumi, Y.; Mohri, Y.; Hirota, M.; Makabe, H. *J. Org. Chem.* **2010**, *75*, 4884; (e) Mohri, Y.; Sagehashi, M.; Yamada, T.; Hattori, Y.; Morimura, K.; Hamauzu, Y.; Kamo, T.; Hirota, M.; Makabe, H. *Heterocycles* **2009**, *79*, 549; (f) Oyama, K. -i.; Kuwano, M.; Ito, M.; Yoshida, K.; Kondo, T. *Tetrahedron Lett.* **2008**, *49*, 3176; (g) Viton, F.; Landreau, C.; Rustidge, D.; Robert, F.; Williamson, G.; Barron, G. *Eur. J. Org. Chem.* **2008**, 6069. (h) Mohri, Y.; Sagehashi, M.; Yamada, T.; Hattori, Y.; Morimura, K.; Kamo, T.; Hirota, M.; Makabe, H. *Tetrahedron Lett.* **2007**, *48*, 5891; (i) Tarascou, I.; Barathieu, K.; André, Y.; Pianet, I.; Dufourc, E. J.; Fouquet, E. *Eur. J. Org. Chem.* **2006**, 5367; (j) Saito, A.; Nakajima, N.; Matsuura, N.; Tanaka, A.; Ubukata, M. *Heterocycles* **2004**, *62*, 479; (k) Saito, A.; Nakajima, N.; Tanaka, A.; Ubukata, M. *Heterocycles* **2003**, *61*, 287; (l) Saito, A.; Nakajima, N.; Tanaka, A.; Ubukata, M. *Tetraherdron* **2002**, *58*, 7829. (m) Tückmantel, W.; Kozikowski, A. P.; Romanczyk, Jr. L. J. *J. Am. Chem. Soc.* **1999**, *121*, 12073.
11. Recent syntheses of procyanidin dimers with gallates: (a) Suda, M.; Katoh, M.; Toda, K.; Matsumoto, K.; Kawaguchi, K.; Kawahara, S. -i.; Hattori, Y.; Fujii, H.; Makabe, H. *Bioorg. Med. Chem. Lett.* **2013**, *23*,



- 4935; (b) Sakuda, H.; Saito, A.; Mizushina, Y.; Yoshida, H.; Tanaka, A.; Nakajima, N. *Heterocycles* **2006**, *67*, 175; (c) Saito, A.; Mizushina, Y.; Ikawa, H.; Yoshida, H.; Doi, Y.; Tanaka, A.; Nakajima, N. *Bioorg. Med. Chem.* **2005**, *13*, 2759; (d) Saito, A.; Emoto, M.; Tanaka, A.; Doi, Y.; Shoji, K.; Mizushina, Y.; Ikawa, H.; Yoshida, H.; Matsuura, N.; Nakajima, N. *Tetrahedron* **2004**, *60*, 12043.
12. Recent syntheses of procyanidin trimers and related compounds: (a) Yano, T.; Ohmori, K.; Takahashi, H.; Kusumi, T.; Suzuki, K. *Org. Biomol. Chem.* **2012**, *10*, 7685; (b) Oizumi, Y.; Katoh, M.; Hattori, Y.; Toda, K.; Kawaguchi, K.; Fujii, H.; Makabe, H. *Heterocycles* **2012**, *85*, 2241; (c) Oizumi, Y.; Mohri, Y.; Hattori, Y.; Makabe, H. *Heterocycles* **2011**, *83*, 739; (d) Ohmori, K.; Ushimaru, N.; Suzuki, K. *Proc. Natl. Acad. Sci. U. S. A.* **2004**, *101*, 12002; (e) Saito, A.; Doi, Y.; Tanaka, A.; Matsuura, N.; Ubukata, M.; Nakajima, N. *Bioorg. Med. Chem.* **2004**, *12*, 4783; (f) Saito, A.; Tanaka, A.; Ubukata, M.; Nakajima, N. *Synlett* **2004**, 1069.
13. Syntheses of procyanidin oligomers: (a) Ohmori, K.; Shono, T.; Hatakoshi, Y.; Yano, T.; Suzuki, K. *Angew. Chem. Int. Ed.* **2011**, *50*, 4862; (b) Saito, A.; Mizushina, Y.; Tanaka, A.; Nakajima, N. *Tetrahedron* **2009**, *65*, 7422; (c) Kozikowski, A. P.; Tückmantel, W.; Böttcher, G.; Romanczyk, Jr. L. J. *J. Org. Chem.* **2003**, *68*, 1641.
14. Synthesis of procyanidin B6: Watanabe, G.; Ohmori, K.; Suzuki, K. *Chem. Commun.* **2013**, *49*, 5210.
15. Krohn, K.; Ahmed, I.; John, M.; Letzel, M. C.; Kuck, D. *Eur. J. Org. Chem.* **2010**, 2544.
16. Wan, S. B.; Dou, Q. P.; Chan, T. H. *Tetrahedron* **2006**, *62*, 5897.
17. Fujii, W.; Toda, K.; Kawaguchi, K.; Kawahara, S.-i.; Hattori, Y.; Fujii, H.; Makabe, H. *Tetrahedron* **2013**, *69*, 3543.
18. Steynberg, P. J.; Nel, R. J. J.; Rensberg, H. van; Bezuidenhoudt, B. C. B.; Ferreira, D. *Tetrahedron*, **1998**, *54*, 8153.
19. Representative procedure for equimolar coupling reaction using Yb(OTf)<sub>3</sub>: To a solution of nucleophile **5** (42 mg, 55 μmol) and electrophile **10** (49 mg, 55 μmol) in CH<sub>2</sub>Cl<sub>2</sub> (4.0 mL) was added Yb(OTf)<sub>3</sub> (34 mg, 55 μmol). After the resulting mixture had been stirred for 3 h at room temperature, the reaction was quenched with water. The mixture was extracted with EtOAc (10 mL×2) and the combined organic layer was washed with brine, dried over MgSO<sub>4</sub>, filtered, and concentrated. The crude product was purified with preparative TLC (hexane:AcOEt:CH<sub>2</sub>Cl<sub>2</sub> = 6:1:3) to afford **11** (57 mg, 66%) as pale yellow oil.
20. HPLC measurement condition of prodelphinidin B1 (**1**): column; InertSustain C18 250×4.6 mm Waters, eluent 0.1% HCOOH-CH<sub>3</sub>CN, flow rate: 0.5 mL/min, detection: UV 280 nm, retention time: 12.28 min., prodelphinidin B2 (**2**): 13.62 min., prodelphinidin B4 (**4**): 14.27 min.