

University of Groningen

Individualized versus standard FSH dosing in women starting IVF/ICSI

van Tilborg, Theodora C.; Torrance, Helen L.; Oudshoorn, Simone C.; Eijkemans, Marinus J. C.; Koks, Carolien A. M.; Verhoeve, Harold R.; Nap, Annemiek W.; Scheffer, Gabrielle J.; Manger, A. Petra; Schoot, Benedictus C.

Published in:
Human Reproduction

DOI:
[10.1093/humrep/dex318](https://doi.org/10.1093/humrep/dex318)

IMPORTANT NOTE: You are advised to consult the publisher's version (publisher's PDF) if you wish to cite from it. Please check the document version below.

Document Version
Publisher's PDF, also known as Version of record

Publication date:
2017

[Link to publication in University of Groningen/UMCG research database](#)

Citation for published version (APA):

van Tilborg, T. C., Torrance, H. L., Oudshoorn, S. C., Eijkemans, M. J. C., Koks, C. M., Verhoeve, H. R., ... OPTIMIST study group (2017). Individualized versus standard FSH dosing in women starting IVF/ICSI: An RCT. Part 1: The predicted poor responder. *Human Reproduction*, 32(12), 2496-2505. DOI: 10.1093/humrep/dex318

Copyright

Other than for strictly personal use, it is not permitted to download or to forward/distribute the text or part of it without the consent of the author(s) and/or copyright holder(s), unless the work is under an open content license (like Creative Commons).

Take-down policy

If you believe that this document breaches copyright please contact us providing details, and we will remove access to the work immediately and investigate your claim.

Downloaded from the University of Groningen/UMCG research database (Pure): <http://www.rug.nl/research/portal>. For technical reasons the number of authors shown on this cover page is limited to 10 maximum.

Individualized versus standard FSH dosing in women starting IVF/ICSI: an RCT. Part I: The predicted poor responder

Theodora C. van Tilborg^{1,*}, Helen L. Torrance¹,
Simone C. Oudshoorn¹, Marinus J.C. Eijkemans², Carolien A.M. Koks³,
Harold R. Verhoeve⁴, Annemiek W. Nap⁵, Gabrielle J. Scheffer⁶,
A. Petra Manger⁷, Benedictus C. Schoot^{8,9}, Alexander V. Sluijmer¹⁰,
Arie Verhoeff¹¹, Henk Groen¹², Joop S.E. Laven¹³, Ben Willem J. Mol^{14,15},
and Frank J.M. Broekmans¹, on behalf of the OPTIMIST study group[†]

¹Department of Reproductive Medicine and Gynaecology, University Medical Centre Utrecht, Utrecht University, PO Box 85500, 3508 GA Utrecht, The Netherlands ²Julius Centre for Health Sciences and Primary Care, University Medical Centre Utrecht, Utrecht University, PO Box 85500, 3508 GA Utrecht, The Netherlands ³Department of Obstetrics and Gynaecology, Maxima Medical Centre Veldhoven, PO Box 7777, 5500 MB Veldhoven, The Netherlands ⁴Department of Obstetrics and Gynaecology, Onze Lieve Vrouwe Gasthuis Oost, PO Box 95500, 1190 HM Amsterdam, The Netherlands ⁵Department of Reproductive Medicine, Rijnstate Hospital, PO Box 9555, 6800 TA Arnhem, The Netherlands ⁶Department of Obstetrics and Gynaecology, Gelre Hospital, PO Box 9014, 7300 DS Apeldoorn, The Netherlands ⁷Department of Obstetrics and Gynaecology, Diaconessenhuis, PO Box 80250, 3508 TG Utrecht, The Netherlands ⁸Department of Obstetrics and Gynaecology, Catharina Ziekenhuis, PO Box 1350, 5602 ZA Eindhoven, The Netherlands ⁹Department of Obstetrics and Gynaecology, University Hospital Gent, 9000 Gent, Belgium ¹⁰Department of Gynaecology, Wilhelmina Hospital, PO Box 30001, 9400 RA Assen, The Netherlands ¹¹Department of Gynaecology, Maasstad Hospital, PO Box 9100, 3007 AC Rotterdam, The Netherlands ¹²Department of Epidemiology, University of Groningen, University Medical Centre Groningen, PO Box 30.001, 9700 RB Groningen, The Netherlands ¹³Department of Obstetrics and Gynaecology, Erasmus Medical Centre, Postbus 2040, 3000 CA Rotterdam, The Netherlands ¹⁴The Robinson Research Institute, School of Paediatrics and Reproductive Health, University of Adelaide, SA 5006 Adelaide, Australia ¹⁵The South Australian Health and Medical Research Unit, PO Box 11060, SA 5001 Adelaide, Australia

*Correspondence address. Department of Reproductive Medicine and Gynaecology, University Medical Centre Utrecht, Utrecht University, PO Box 85500, 3508 GA Utrecht, The Netherlands. E-mail: ctilborg@umcutrecht.nl

Submitted on January 27, 2017; resubmitted on September 20, 2017; accepted on October 13, 2017

STUDY QUESTION: Does an increased FSH dose result in higher cumulative live birth rates in women with a predicted poor ovarian response, apparent from a low antral follicle count (AFC), scheduled for IVF or ICSI?

SUMMARY ANSWER: In women with a predicted poor ovarian response (AFC < 11) undergoing IVF/ICSI, an increased FSH dose (225/450 IU/day) does not improve cumulative live birth rates as compared to a standard dose (150 IU/day).

WHAT IS KNOWN ALREADY: In women scheduled for IVF/ICSI, an ovarian reserve test (ORT) can predict ovarian response to stimulation. The FSH starting dose is often adjusted based on the ORT from the belief that it will improve live birth rates. However, the existing RCTs on this topic, most of which show no benefit, are underpowered.

STUDY DESIGN, SIZE, DURATION: Between May 2011 and May 2014, we performed an open-label multicentre RCT in women with an AFC < 11 (Dutch Trial Register NTR2657). The primary outcome was ongoing pregnancy achieved within 18 months after randomization

[†]OPTIMIST study group: Theodora C. van Tilborg, Simone C. Oudshoorn, Marinus J.C. Eijkemans, Monique H. Mochtar, Carolien A.M. Koks, Ron J.T. van Golde, Harold R. Verhoeve, Annemiek W. Nap, Gabrielle J. Scheffer, A. Petra Manger, Annemiek Hoek, Benedictus C. Schoot, G. Jur E. Oosterhuis, Walter K.H. Kuchenbecker, Kathrin Fleischer, Jan Peter de Bruin, Alexander V. Sluijmer, Jaap Friederich, Arie Verhoeff, Marcel H.A. van Hooft, Evert J.P. van Santbrink, Egbert A. Brinkhuis, Jesper M.J. Smeenk, Janet Kwee, Cory H. de Koning, Henk Groen, Madelon van Wely, Cornelis B. Lambalk, Joop S.E. Laven, Ben Willem J. Mol, Frank J.M. Broekmans, Helen L. Torrance.

and resulting in a live birth. We needed 300 women to assess whether an increased dose strategy would increase the cumulative live birth rate from 25 to 40% (two-sided alpha-error 0.05, power 80%).

PARTICIPANTS/MATERIALS, SETTING, METHODS: Women with an AFC ≤ 7 were randomized to an FSH dose of 450 IU/day or 150 IU/day, and women with an AFC 8–10 were randomized to 225 IU or 150 IU/day. In the standard group, dose adjustment was allowed in subsequent cycles based on pre-specified criteria. Both effectiveness and cost-effectiveness of the strategies were evaluated from an intention-to-treat perspective.

MAIN RESULTS AND THE ROLE OF CHANCE: In total, 511 women were randomized, 234 with an AFC ≤ 7 and 277 with an AFC 8–10. The cumulative live birth rate for increased versus standard dosing was 42.4% (106/250) versus 44.8% (117/261), respectively [relative risk (RR): 0.95 (95%CI, 0.78–1.15), $P = 0.58$]. As an increased dose strategy was more expensive [delta costs/woman: €1099 (95%CI, 562–1591)], standard FSH dosing was the dominant strategy in our economic analysis.

LIMITATIONS, REASONS FOR CAUTION: Despite our training programme, the AFC might have suffered from inter-observer variation. As this open study permitted small dose adjustments between cycles, potential selective cancelling of cycles in women treated with 150 IU could have influenced the cumulative results. However, since first cycle live birth rates point in the same direction we consider it unlikely that the open design masked a potential benefit for the individualized strategy.

WIDER IMPLICATIONS OF THE FINDINGS: Since an increased dose in women scheduled for IVF/ICSI with a predicted poor response (AFC < 11) does not improve live birth rates and is more expensive, we recommend using a standard dose of 150 IU/day in these women.

STUDY FUNDING/COMPETING INTEREST(S): This study was funded by The Netherlands Organisation for Health Research and Development (ZonMw number 171102020). T.C.T., H.L.T. and S.C.O. received an unrestricted personal grant from Merck BV. H.R.V. receives monetary compensation as a member on an external advisory board for Ferring pharmaceutical BV. B.W.J.M. is supported by a NHMRC Practitioner Fellowship (GNT1082548) and reports consultancy for OvsEva, Merck and Guerbet. F.J.M.B. receives monetary compensation as a member of the external advisory board for Ferring pharmaceuticals BV (the Netherlands) and Merck Serono (the Netherlands) for consultancy work for Gedeon Richter (Belgium) and Roche Diagnostics on automated AMH assay development (Switzerland) and for a research cooperation with Ansh Labs (USA). All other authors have nothing to declare.

TRIAL REGISTRATION NUMBER: Registered at the ICMJE-recognized Dutch Trial Registry (www.trialregister.nl). Registration number NTR2657.

TRIAL REGISTRATION DATE: 20 December 2010.

DATE OF FIRST PATIENT'S ENROLMENT: 12 May 2011.

Key words: Antral follicle count / FSH / RCT / live birth / poor ovarian response / cost-effectiveness / IVF / ICSI / ovarian reserve

Introduction

The objective of controlled ovarian stimulation (COS) in women undergoing IVF or ICSI is to obtain multiple oocytes. The ovarian response to COS is an expression of the quantitative ovarian reserve (De Boer *et al.*, 2002) and can differ significantly between women of the same age, as women have a wide variation in the timing of decline of their ovarian reserve (te Velde and Pearson, 2002). Approximately 20% of all women undergoing IVF/ICSI show a poor ovarian response (defined as ≤ 4 oocytes retrieved) to COS (Broer *et al.*, 2013a), which leads to higher drop-out rates (Verberg *et al.*, 2008) and results in lower live birth rates as compared to women with a normal response (van der Gaast *et al.*, 2006; Sunkara *et al.*, 2011; Drakopoulos *et al.*, 2015). An increase in the proportion of normal responders by optimizing COS is thought to improve the live birth rate. Assessment of the ovarian reserve is potentially useful for the prediction of poor response, and both the antral follicle count (AFC) and serum anti-Müllerian Hormone (AMH) are accurate predictors of ovarian response, even if used as single tests (Broer *et al.*, 2013a).

The majority of previously published RCTs on ovarian reserve test (ORT) based dosing in predicted poor responders have shown that an increased gonadotrophin dose does not increase pregnancy rates (van Tilborg *et al.*, 2016). In contrast, one study revealed a decrease in the

proportion of poor ovarian response after individualized dosing and also reported higher ongoing pregnancy rates (Popovic-Todorovic *et al.*, 2003). Nonetheless, as most of the previously published RCTs were underpowered to assess the outcomes pregnancy or live birth and suffered from methodological weaknesses (Harrison *et al.*, 2001; Popovic-Todorovic *et al.*, 2003; Klinkert *et al.*, 2005; Berkanoglu and Ozgur, 2010; Jayaprakasan *et al.*, 2010; Lefebvre *et al.*, 2015; Olivennes *et al.*, 2015), it is still unclear whether women with a predicted poor response who are undergoing IVF/ICSI benefit from a higher gonadotrophin dose. As multiple treatment cycles reflect clinical practice most accurately and live birth is the endpoint that matters most to the patient (Legro *et al.*, 2014; Scholten *et al.*, 2016), we aimed to assess whether an increased gonadotrophin dose in women scheduled for IVF/ICSI with a predicted poor response based on a low AFC resulted in higher cumulative live birth rates.

Materials and Methods

Study design and population

This open-label, multicentre RCT in predicted poor responders in which we compared an increased FSH dose with standard dosing was part of a

Dutch prospective cohort study (the OPTIMIST study, registration number Dutch Trial Register NTR2657; www.trialregister.nl). This paper reports data from the predicted poor responders only (AFC < 11). Women with an AFC > 15 were included in the predicted hyper responder RCT, of which the results are reported separately (Oudshoorn et al., 2017). Ethical approval was obtained from the Institutional Review Board of the University Medical Centre Utrecht (MEC 10–273) and the study was approved by the board of directors of all participating sites. The study protocol was published previously (van Tilborg et al., 2012).

All infertile women starting their first IVF or ICSI treatment, or the first IVF or ICSI treatment after a live birth, who were younger than 44 years of age, had a regular menstrual cycle (on average 25–35 days) and a normal transvaginal ultrasound were eligible for inclusion in the OPTIMIST study. Women with polycystic ovary syndrome (PCOS) (PCOS Consensus Workshop Group, 2004), endocrine or metabolic abnormalities or starting oocyte donation could not be included. Eligible women were recruited by their physicians or by dedicated research nurses prior to the start of their IVF/ICSI treatment, and before the ORT was performed. Written informed consent was obtained from all participants.

Study procedures

Eligible women scheduled for IVF/ICSI were counselled by physicians or research nurses for inclusion in the cohort study, with possible randomization if the AFC was < 11 or > 15. After providing informed consent, women had their AFC measured. The AFC was determined by counting follicles sized between 2 and 10 mm, and performed following previously published recommendations (Broekmans et al., 2010). At the start of the study, physicians of participating sites were trained in AFC through workshops. Furthermore, instructions on how to measure AFC, including video images, were sent to all participating sites.

Women with an AFC < 11 were included in the RCT reported here. Women were randomly assigned using a web-based randomization program to an increased or standard FSH dose using a 1:1 ratio and variable block-sizes (maximum 6). Randomization was stratified per AFC group and centre. Women assigned to the higher dose group received 450 IU/day FSH if they had an AFC ≤ 7, and 225 IU/day FSH if their AFC was 8–10. In the standard dose group, women received 150 IU/day FSH in both AFC groups. Centres were only allowed to use one type of GnRH analogue protocol as co-medication (GnRH antagonist or a long GnRH agonist schedule). So, stratification per centre also includes stratification per GnRH analogue treatment. The clinicians, participants and investigators were not masked for the assigned FSH dose. A fixed dose strategy was used. In the standard group, a maximum dose adjustment of 50 IU/day was allowed between cycles if women had a poor response (i.e. cycle cancellation owing to insufficient growth: < 2 follicles > 12 mm or < 3 follicles ≥ 17 mm; or < 5 oocytes at retrieval) or hyper response (i.e. cycle cancellation owing to excessive response: > 20 follicles > 12 mm and estradiol levels exceeding 11.700 pmol/l (= 3187.08 ng/l) or > 30 follicles > 12 mm; or > 15 oocytes at retrieval).

Ovarian stimulation was performed using recombinant-FSH or urinary-FSH. Final oocyte maturation was achieved by administration of HCG (10 000 IU Pregnyl® or 6500 IU Ovitrelle®, according to local protocol). Oocyte retrieval was carried out 36 h after HCG administration. Embryo transfer took place on Day 3–5 after oocyte retrieval, depending on local protocol. Luteal phase supplementation consisted of vaginal progesterone. Frozen/thaw treatment cycles were conducted according to local protocol.

All treatment cycles performed within 18 months after randomization were recorded. After the first fresh treatment cycle, women had to use their cryopreserved embryos before continuing to the next fresh treatment cycle. Further details of the OPTIMIST study procedures have been published elsewhere (van Tilborg et al., 2012).

Outcome measures

The primary outcome of this study was ongoing pregnancy achieved within 18 months after randomization leading to live birth, which was defined as the delivery of a live foetus after 24 weeks of gestation. All ongoing pregnancies achieved within 18 months of follow-up, irrespective of the conception mode, were taken into account. Secondary outcomes included number of cycle cancellations, reason for cycle cancellations (i.e. insufficient follicle growth, excessive response or other reason (e.g. personal reason)), number of ovum retrievals performed, number of obtained oocytes, poor response (i.e. cycle cancellation owing to insufficient growth or < 5 oocytes at retrieval), hyper response (i.e. cycle cancellation owing to excessive response or > 15 oocytes at retrieval), number of embryos available for transfer, number of embryo transfers, number of cryopreserved embryos, number of treatment cycles started, ovarian hyperstimulation syndrome (OHSS; i.e. mild, moderate and severe (for definitions see Oudshoorn et al. 2017)), biochemical pregnancy (defined as a positive pregnancy test), miscarriage (defined as the absence of a foetal heartbeat at 7 or 10 weeks of gestation) and clinical pregnancy (defined as the presence of a gestational sac). Ongoing pregnancy was defined as the detection of a foetal heartbeat on ultrasound at a gestational age of at least 10 weeks, and time to ongoing pregnancy leading to live birth as the time from randomization to an ongoing pregnancy that led to a live birth. Multiple pregnancy was defined as an ongoing pregnancy with at least two foetuses.

For the cost-effectiveness analysis, data on direct medical costs were collected. All outcomes were pre-specified in the study protocol and statistical analysis plan. We chose to adapt our protocol definitions for clinical and ongoing pregnancy in order to follow the recent consensus of ESHRE (Kolte et al., 2015).

Sample size calculation

We expected that an increased FSH dose would improve the cumulative live birth rate by 15% (from 25 to 40%) in women included in the OPTIMIST study as compared to treatment by the standard approach (Popovic-Todorovic et al., 2003). In order to be able to detect this difference, at least 300 women were needed in RCT1 (80% power, alpha-error 5%). Derived from previous data (Broer et al., 2013a, 2013b), we assumed that 20% of the eligible women would be classified as predicted poor responders and another 20% as predicted hyper responders. This meant that in total 1500 women were planned to be included in the cohort of the OPTIMIST study (van Tilborg et al. 2017).

Statistical analysis

All analyses were performed according to the intention-to-treat principle. Continuous data are presented as mean (SD) and categorical data as numbers (%). Descriptive analyses were performed with independent samples t-test and chi-square tests for continuous and categorical data, respectively. In the cumulative analyses, clustering of multiple treatment cycles per woman had to be taken into account. We used three generalized estimating equations models: normal response and identity link for continuous variables, binominal response and a log link for binominal variables, and multinomial response and cumlogit link for categorical variables, assuming independent working correlations in all models.

For the pregnancy and live birth outcomes relative risks (RR) and 95% CI were calculated. Time to ongoing pregnancy leading to live birth was graphically depicted by cumulative incidence curves. The log-rank test was used to measure whether significant differences existed in the cumulative incidence curves. Women who did not reach the primary outcome (live birth) and for whom the follow-up could not be completed because of loss of contact (despite repeated attempts) were considered as 'not having an ongoing pregnancy' at 18 months after randomization. Following our statistical analysis plan, pre-specified additional analyses were performed: the primary outcome and some

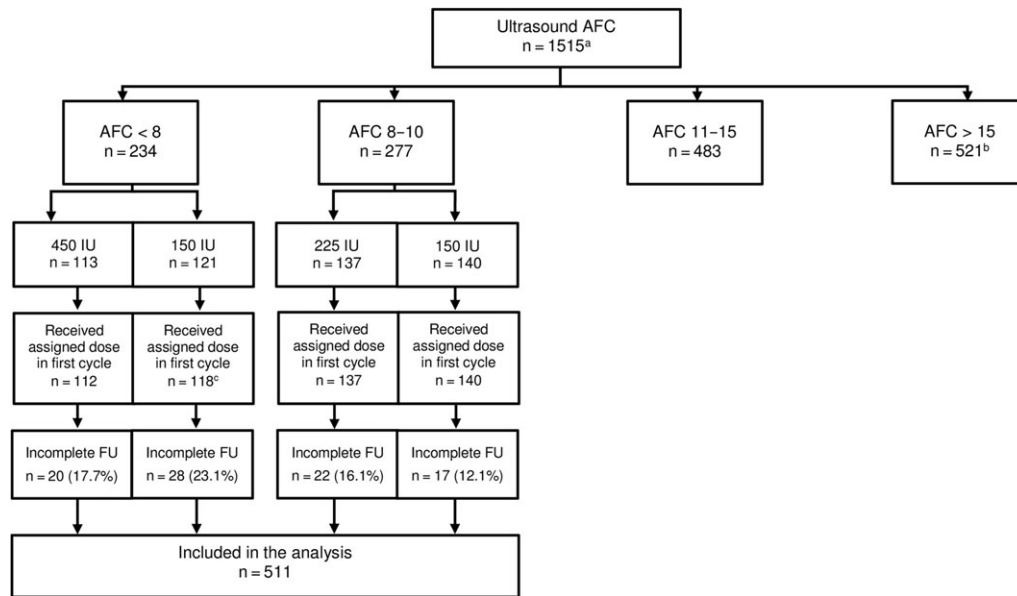


Figure 1 Flowchart of total study cohort and the included predicted poor responders. AFC, antral follicle count; FU, follow-up. ^aThe overall strategy analysis is reported in [van Tilborg et al. 2017](#); ^bThe predicted hyper responder trial is reported in [Oudshoorn et al. 2017](#). ^c $n = 1$ never started ovarian stimulation, $n = 1$ never started treatment.

of the secondary outcomes within the AFC subgroups (AFC ≤ 7 and AFC 8–10), first cycle treatment results and a per-protocol analysis of the primary outcome by excluding women with protocol violations related to their first cycle FSH starting dose, by excluding treatment cycles from a woman from their first protocol violation regarding dose-adjustments within and between treatment cycles onwards, and by excluding pregnancies that had a spontaneous or ‘other’ mode of conception. All comparisons were tested two-sided. P-values below 0.05 were considered to indicate statistical significance. Data were analysed in SPSS (version 21.0, IBM Corp Armonk, NY, USA) and R (version 3.1.3, R Foundation for Statistical Computing, Vienna, Austria).

Since the economic evaluation was performed from a healthcare perspective, we focused on direct medical costs. The effectiveness outcome was the proportion of couples with an ongoing pregnancy achieved within 18 months after randomization and leading to live birth. In order to test the robustness of the results of our cost-effectiveness analysis, bootstrap resampling with 5000 resamples was performed. Based on the bootstrapped results, a Cost-effectiveness Acceptability Curve was derived. Further details regarding the cost-effectiveness analysis are described in [van Tilborg et al. \(van Tilborg et al. 2017\)](#).

Results

Between 12 May 2011 and 5 May 2014, we included 1515 women in the OPTIMIST cohort of whom 511/1515 (33.7%) had an AFC < 11 and were classified as a predicted poor responder (Fig. 1). In total, 250/511 (48.9%) women were allocated to an increased and 261/511 (51.1%) women to a standard FSH dose.

A similar proportion of women allocated to the increased and standard FSH dose did not reach the primary outcome and could not be contacted 18 months after randomization [44/250 (17.6%) versus 45/261 (17.2%), respectively; $P = 0.89$], with a comparable mean duration of follow-up [295.2 (121.2) versus 270.1 days (154.6), respectively; $P = 0.40$].

Table 1 Baseline characteristics of predicted poor responder women in a study comparing an increased versus a standard FSH dose for IVF/ICSI.

Baseline characteristics	AFC 0–10	
	450/225 IU ($n = 250$)	150 IU ($n = 261$)
Age (years)	35.6 (4.2)	35.8 (4.4)
Duration of infertility (years)	2.7 (1.8)	2.5 (1.7)
Smoking	44/238 (18.5%)	48/252 (19.0%)
BMI - kg/m ²	24.1 (4.2)	24.0 (4.3)
Primary infertility	138/248 (55.6%)	143 (54.8%)
Cause of infertility ^a		
Unexplained	96 (38.4%)	124 (47.5%)
Male factor	123 (49.2%)	114 (43.7%)
Endometriosis	10 (4.0%)	9 (3.4%)
Tubal factor	30 (12.0%)	25 (9.6%)
AFC (median, IQR)	8.0 (3.0)	8.0 (3.0)
AMH (ng/ml) (median, IQR)	1.00 (0.96)	0.88 (0.96)
GnRH co-treatment		
GnRH agonist	202 (80.8%)	208/260 (80.0%)
GnRH antagonist	48 (19.2%)	52/260 (20.0%)

Data are presented as mean (SD) or number (%) unless otherwise reported. AFC, antral follicle count; AMH, anti-Müllerian hormone; IQR, interquartile range.

^aIncluding patients with > 1 cause of infertility.

Baseline characteristics did not differ between groups (Table I). Our primary outcome, ongoing pregnancy achieved within 18 months of follow-up and leading to live birth, occurred in 106/250 (42.4%) women in the higher versus 117/261 (44.8%) women in the standard dose group [RR 0.95 (95%CI, 0.78–1.15), $P = 0.58$; Table II]. Time to ongoing pregnancy leading to live birth was comparable between groups [mean 212 (SD 133) versus 197 (SD 135) days for the increased versus standard dose group, $P = 0.40$; Table II and Figure 2]. Cumulative treatment results are shown in Table III. In the standard group more poor response and less hyper response occurred, with no difference in the proportion of normal ovarian responses as compared to an increased dose.

First cycle results

First cycle treatment results are summarized in Table IV. Despite a lower number of oocytes and a lower fresh embryo transfer rate in the standard dose group, first cycle live birth rates were not significantly different [17.6% versus 20.0%, RR 0.88 (95% CI, 0.61–1.26), $P = 0.49$].

Pre-specified additional analyses

In women with an AFC 8–10, an increased FSH dose led to a higher number of oocytes and a higher proportion of hyper responses but without improving cumulative live birth rates as compared to a standard dose [live birth rates 47.4% versus 47.1%, respectively; RR 1.01 (95%CI, 0.78–1.29), $P = 0.96$, see Supplementary Table SI, Supplementary Figure I]. The first treatment cycle showed similar results, except for a higher proportion of poor responses in the standard dosing group (Supplementary Table SII).

For women with an AFC 0–7, cumulative results revealed less poor response and more normal- and hyper responses in women allocated to the increased dose strategy as compared to the standard strategy. Despite more fresh embryo transfers in women allocated to the increased dose, no significant effect on live birth rates was found [36.3% versus 42.1%, respectively, RR 0.86 (95%CI, 0.62–1.19), $P = 0.36$; Supplementary Table SI, Supplementary Figure S2]. With respect to the first cycle results, women in the standard group more often showed a poor response, were more likely to be cancelled in case of insufficient follicle growth and had less fresh embryo transfers (Supplementary Table SII). These results however did not lead to significant differences in first cycle live birth rates between treatment strategies (Supplementary Table SII).

In the per protocol analysis, 73/246 (29.7%) women that were allocated to the increased FSH dose had an ongoing pregnancy resulting in live birth compared to 82/250 (32.8%) women that were allocated to the standard FSH dose [RR 0.90 (95%CI, 0.70–1.18), $P = 0.45$].

Health economic evaluation

An increased FSH dose is more expensive [€6397 versus €5298, mean difference costs per woman €1099 (95%CI, 562–1591)] while it does not increase live birth rates (Supplementary Table SIII). Therefore, standard dosing has to be considered the dominant strategy in predicted poor responders (Supplementary Figure S3 and S4).

Discussion

This study shows that in women undergoing IVF/ICSI with a predicted poor response (defined as AFC < 11) increased FSH dosing does not improve live birth rates, while it increases treatment costs as

Table II Pregnancy data per woman.

Pregnancy data per woman ^a	AFC 0–10			
	450/225 IU (n = 250)	150 IU (n = 261)	RR (95%CI)	P-value
Ongoing pregnancy resulting in live birth within 18 months of FU	106 (42.4%)	117 (44.8%)	0.95 (0.78–1.15)	0.58
First cycle (250 versus 260)	44 (17.6%)	52 (20.0%)	0.88 (0.61–1.26)	0.49
Second cycle (165 versus 176)	24 (14.5%)	32 (18.2%)	0.80 (0.49–1.30)	0.37
Third cycle (79 versus 87)	16 (20.3%)	14 (16.1%)	1.26 (0.66–2.41)	0.49
Fourth cycle (18 versus 27)	4 (22.2%)	1 (3.7%)	5.99 (0.73–50.0)	0.05
Fifth and sixth cycle	0	0		
Ongoing pregnancy	108 (43.2%) ^d	119 (45.6%) ^e	0.95 (0.78–1.15)	0.29
Multiple pregnancy ^b	5 (2.0%)	8 (3.1%)	0.65 (0.22–1.97)	0.45
Conception mode (% of live birth)				0.29
Spontaneous	18 (17.0%)	14 (12.0%)		
IVF/ICSI fresh ^c	74 (69.8%)	85 (72.6%)		
IVF/ICSI frozen	14 (13.2%)	15 (12.8%)		
Unknown	0	3 (2.6%)		
Time to ongoing pregnancy leading to live birth, days (mean, SD)	212 (133)	197 (135)		0.40

FU, follow-up; RR, relative risk. P-value are calculated by using a Chi-square test or an independent samples t-test.

^aShown with a maximum of one event per woman.

^bBased on the number of multiple ongoing pregnancies.

^cIncluding live births from escape IUI treatments.

^dOne woman with an immature birth and one woman with a foetal demise.

^eOne woman with an immature birth and one woman with a foetal demise.

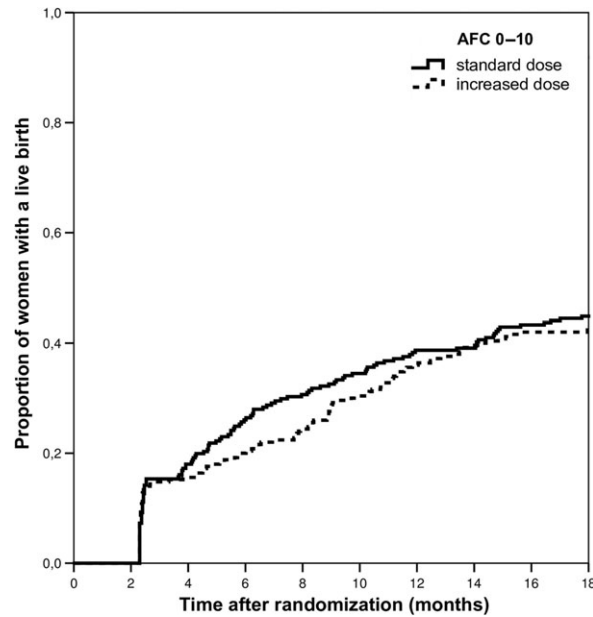


Figure 2 Cumulative live birth incidence in predicted poor responders. Log-rank = 0.511.

Time after randomization (months)	0	2	4	6	8	10	12	14	16	18
Number at risk (%); increased dose	250	250 (100)	211 (84.4)	200 (80.0)	189 (75.6)	174 (69.6)	160 (64.0)	151 (60.4)	145 (58.0)	144 (57.6)
Number at risk (%); standard dose	261	261 (100)	111 (79.3)	193 (73.9)	181 (69.3)	171 (65.5)	160 (61.3)	159 (60.9)	148 (56.7)	144 (55.2)

compared to a standard dose regimen. Subgroup analyses in women with an AFC 0–7 or AFC 8–10 further strengthen the overall result as increased dosing is not beneficial with respect to live birth rates in either of these groups.

It is difficult to directly compare our results with previous findings as none of the available RCTs evaluated the effect of an increased FSH dose on cumulative live birth rates nor assessed the cost-effectiveness of such a strategy. Nonetheless, our results are in line with several previous RCTs on increased FSH dosing in predicted poor responders (Harrison *et al.*, 2001; Klinkert *et al.*, 2005; Berkkanoglu and Ozgur, 2010; Lefebvre *et al.*, 2015). These RCTs, however, suffered from methodological weaknesses which were summarized in a recent systematic review on individualized dosing (van Tilborg *et al.*, 2016). Most studies on poor responders included a heterogeneous group of women with both predicted and proven poor responders. Such a heterogeneous study population makes it more difficult to assess the true effect of an intervention as it is likely that expected and unexpected poor responders have a different prognosis (Klinkert *et al.*, 2004). In contrast to our findings, one small study by Popovic-Todorovic *et al.* (2003) reported that individualized FSH dosing based on a multi-factor algorithm resulted in a higher ongoing pregnancy rate after the first IVF/ICSI treatment cycle as compared to a standard dose of 150 IU/day. This study mainly included women with a good prognosis and was not powered to find a difference in ongoing pregnancy rates.

A recently published RCT showed that mild ovarian stimulation consisting of a GnRH antagonist protocol with a daily FSH dose of 150 IU led to similar single cycle ongoing pregnancy rates in predicted or

actual poor responders as compared to treatment with a GnRH agonist protocol and 450 IU/day HMGs (Youssef *et al.*, 2017). Although this study compared two totally different stimulation protocols instead of just two different FSH dosages, it confirms that high dosages of gonadotrophins will not improve IVF outcomes in predicted poor responders (Land *et al.*, 1996).

For women with an AFC 0–7, the cumulative incidence curves for live birth indicated that women in the increased dose group had a 10% lower cumulative live birth rate after 8 months of randomization, despite a lower cancellation rate, as compared to the standard dose group (Supplementary Figure 2). At 18 months of follow-up, a possible clinically relevant (though statistically non-significant) difference of 6% remained in the advantage of the standard FSH dose group (see Supplementary Table I). A retrospective study analysing over 600 000 fresh COS cycles showed that an average daily FSH dose >300 IU resulted in a statistically decreased live birth rate irrespective of the number of oocytes retrieved or a woman's age, even in women without a diagnosis of a diminished ovarian reserve (Baker *et al.*, 2015). As an explanation for this observation, one might hypothesize that a high exogenous FSH dose induces chromosomal abnormalities in embryos derived from IVF/ICSI (Munne *et al.*, 1997; Katz-jaffe *et al.*, 2005; Baart *et al.*, 2006). Another explanation might be found in the disturbed endometrial receptivity in women treated with a high FSH dose, as the pregnancy results from fresh cycles were lower but no difference was found in pregnancy rates from cryo/thaw cycles (Munch *et al.*, 2017).

Table III Cumulative treatment cycle results.

Results 18 months follow-up	AFC 0–10		P-value
	450/225 IU (n = 250)	150 IU (n = 260)	
Number of fresh cycles per woman	2.1 (0.9)	2.1 (1.1)	0.29
Total number of fresh cycles started	513	558	
First cycle	250 (100%)	260/261 (99.6%)	
Second cycle	165 (66.0%)	176 (67.7%)	
Third cycle	79 (31.6%)	87 (33.3%)	
Fourth cycle	18 (7.2%)	27 (10.3%)	
Fifth cycle	1 (0.4%)	6 (2.3%)	
Sixth cycle	0	2 (0.8%)	
Cancellation classification			<0.001 ^c
No cancellation	464 (90.4%)	444/557 (79.7%)	<0.001 ^{c,d}
Insufficient growth	36 (7.0%)	99/557 (17.8%)	<0.001 ^{c,d}
Excessive response	3 (0.6%)	1/557 (0.2%)	0.34 ^{c,d}
Other	10 (1.9%)	13/557 (2.3%)	0.67 ^{c,d}
Number of oocytes ^a	7.6 (5.0)	6.4 (4.3)	0.002 ^c
Poor response	178/510 (34.9%)	275/556 (49.5%)	<0.001 ^c
Hyper response	44/510 (8.6%)	19/556 (3.4%)	0.003 ^c
Number of fresh embryos for transfer ^a	3.6 (3.1)	3.1 (2.7)	0.04 ^c
Number of fresh embryo transfers	399 (77.8%)	385/557 (69.1%)	0.005 ^c
Number of fresh embryos/transfer	1.3 (0.5)	1.3 (0.5)	0.60 ^c
Number of frozen embryos ^b	3.0 (2.2)	2.6 (1.7)	0.11 ^c
OHSS overall	6/509 (1.2%)	10/555 (1.8%)	0.45 ^c
Number of cryo cycles with transfer/woman	0.7 (1.2)	0.6 (1.1)	0.49
Number of frozen embryos/transfer	1.1 (0.4)	1.1 (0.3)	0.42 ^c

Data are presented as mean (SD) or number (%). OHSS, ovarian hyperstimulation syndrome.

^aBased on the women who had an ovum pick-up.

^bBased on the women who had frozen embryos.

^cP-value calculated by using a generalized estimating equation model.

^dPost hoc testing, P-value is considered to be significant when <0.013 (Bonferroni correction (0.05/4)).

One of the main strengths of this multicentre RCT is the fact that the study was powered to report live birth rates, which is the outcome of interest for infertile couples (Fauser et al., 2005; Legro et al., 2014). Furthermore, not just single cycle live birth rates, but results from consecutive cycles performed within an 18-month period of follow-up were evaluated thereby mimicking daily practice (Scholten et al., 2016). Other strengths include the low proportion of loss to follow-up and the performance of an intention-to-treat analysis, both leading to an increased reliability of our outcomes (Higgins and Altman, 2011). Besides, this is the first RCT that performed a cost-effectiveness analysis, which provides essential information before implementation of study results might be considered.

Some limitations of the study need to be addressed. A potential weakness concerns the multiple AFC observers since inter-observer variability is known to be present (Scheffer et al., 2002; Broekmans et al., 2010). To minimize inter-observer variation, observers were trained by arranging AFC workshops and offering instructions for the AFC procedure prior to the start of the study (Broekmans et al., 2010). The remaining variability reflects daily practice variation and could also be seen as a strength of the study as this increases the

generalizability of the results. Furthermore, based on the baseline characteristics from the OPTIMIST trial as a whole, we can conclude that the AFC is an adequate response predictor because age and AMH distributions, other factors associated with response prediction, show expected patterns over the AFC subgroups (van Tilborg et al. 2017). Live birth rates in predicted poor responders, normal responders (data not shown) and hyper responders (Oudshoorn et al., 2017) further confirm the assumption that AFC is an adequate tool. Nonetheless, some concern regarding the AFC operator accuracy may be raised as OHSS events occurred in predicted poor responders randomized to 150 IU/day. These OHSS events were spread over many centres and most of these women had less than 15 oocytes at oocyte retrieval (data not shown). As the AFC has a certain inter- and intra-observer variation we also studied the data from women classified as predicted poor responders based on AMH and treated with a standard dose. A similar proportion of women classified as predicted poor responder based on AMH (AMH ≤ 0.769 ng/ml and AMH 0.770–1.330 ng/ml) developed OHSS (data not shown). So, it is likely that using AMH will not prevent misclassification with respect to response prediction. These results are in line with previous studies

Table IV First treatment cycle results.

First cycle results	AFC 0–10		P-value
	450/225 IU (n = 250)	150 IU (n = 260)	
Total FSH used IU	3705 (1616)	1871 (542)	<0.001
Cancellation classification			<0.001
No cancellation	231 (92.4%)	199 (76.5%)	<0.001 ^c
Insufficient growth	16 (6.4%)	58 (22.3%)	<0.001 ^c
Excessive response	1 (0.4%)	0	0.31 ^c
Other	2 (0.8%)	3 (1.2%)	0.69 ^c
Number of oocytes ^a	7.9 (5.3)	6.5 (4.2)	0.003
Poor response	86 (34.4%)	134 (51.5%)	<0.001
Hyper response	26 (10.4%)	7 (2.7%)	<0.001
Number of embryos for transfer ^a	3.8 (3.5)	3.2 (2.6)	0.04
Number of fresh embryo transfers	195 (78.0%)	175 (67.3%)	0.007
Number of frozen embryos ^b	3.3 (2.5)	2.7 (1.7)	0.09
OHSS	5/249 (2.0%)	7 (2.7%)	0.61
Number of cryo cycles with transfer/woman	0.4 (0.8)	0.4 (0.8)	0.85
Number of cryo embryos per transfer	1.1 (0.3)	1.2 (0.4)	0.36 ^d
Live birth (fresh only)	37 (14.8%)	41 (15.8%)	0.76
Live birth (fresh and cryo)	44 (17.6%)	52 (20.0%)	0.49

Data are presented as mean (SD) or number (%).

^aBased on the women who had an ovum pick-up.

^bBased on the women who had frozen embryos.

^cPost hoc testing, P-value is considered to be significant when <0.013 (Bonferroni correction (0.05/4)).

^dP-value calculated by using a generalized estimating equation model.

that show similar test accuracy (receiver operating characteristic) for AMH and AFC for the prediction of both poor and excessive response (Broer *et al.*, 2013a, 2013b). Unfortunately, test accuracy will never be perfect, as there will always be some misclassification in response prediction (i.e. false positives and false negatives). Nevertheless, recent reports suggest that AMH is a better ovarian response predictor, and could therefore be a better tool for dose individualization (Iliodromiti *et al.*, 2015; Nelson *et al.*, 2015a) but evidence directly comparing AMH- with AFC-based dosing is scarce (Lan *et al.*, 2013). AMH is likely to be a more stable biomarker, but challenges remain with different assays and different laboratories (Broer *et al.*, 2014; Nelson *et al.*, 2015b). An additional analysis by using AMH cut-offs for response prediction and subsequent individualized dosing on the overall OPTIMIST data revealed comparable treatment and live birth results as compared to AFC-based dosing (van Tilborg *et al.*, 2017). Another limitation might be the permitted between-cycle dose adjustments in the standard dose strategy. Although, criteria for those adjustments were pre-specified in the protocol it might have introduced performance bias due to selective cancellation in the standard dose group since this was an open-label trial. In the first cycle results of the AFC 0–7 subgroup, we found more cancellations in the standard group for not fulfilling the HCG criterion. Nonetheless, despite fewer cancellations in the increased dose group, no benefit on first cycle live birth rates was found (see Supplementary Table II). So, if this selective cancelling in the standard dose group would have negatively influenced the live birth rate, it is hypothetically possible that standard dosing would have been superior to increased dosing in women with an AFC 0–7.

The choice to allow two different types of GnRH analogues could be considered a flaw in the study design. In total, 80% of the women were treated by GnRH agonist co-treatment with a similar proportion between both strategy groups. Although, there is no consensus about what is the best protocol for predicted poor responders, a long GnRH agonist protocol has been claimed to be co-treatment of choice for predicted poor responders, especially in older women (Sbracia *et al.*, 2005, 2009). However, based on more recently published papers, it is unlikely that the inclusion of both types of GnRH analogue co-treatment influenced the overall live birth rate (Al-Inany *et al.*, 2016; Toftager *et al.*, 2017; Youssef *et al.*, 2017).

As our study was designed prior to the publication of the Bologna criteria, the poor response definition used in this study does not fully match the Bologna criteria (Ferraretti *et al.*, 2011). Recently, the POSEIDON group published a new classification for 'low prognosis' patients based on either age, ovarian reserve parameters prior to ovarian stimulation, with or without the number of oocytes retrieved (Humaidan *et al.*, 2016; Poseidon Group *et al.*, 2016). In contrast to our study design, their groups all include a combination of age and ORT. Further OPTIMIST data analyses are planned to evaluate whether the efficacy of individualized dosing based on AFC is influenced by age. Furthermore, the OPTIMIST data provide the opportunity to evaluate whether unexpected poor responders benefit from a dose-adjustment in a subsequent treatment cycle as it is unclear from current literature what policy should be recommended for these women. Unexpected poor responders seem to have other biological characteristics and prognosis and may form a

different entity than the predicted poor responder (Klinkert et al., 2004).

As this RCT indicates that increased FSH dosing should not be used in predicted poor responders, the question remains whether it is (cost-)effective to perform an ORT in all women starting IVF/ICSI treatment with the objective to personalize the FSH strategy in certain subgroups of women. In line with this RCT, the overall OPTIMIST study shows that standard dosing (150 IU/day), so without performing an AFC, is the dominant strategy (van Tilborg et al., 2017). The results for reduced FSH dosing in predicted hyper responders are described by Oudshoorn et al. (2017), in which no benefit on cumulative live birth rates was reported, although an advantage regarding safety could not be ruled out.

In conclusion, in women with a predicted poor response starting their IVF/ICSI treatment, increased FSH dosing based on the AFC does not improve live birth rates while it increases costs. Therefore, physicians should accept that prescribing above standard FSH doses in predicted poor responders is not effective.

Supplementary data

Supplementary data are available at *Human Reproduction* online.

Acknowledgement

We thank the women who participated in this study, the staff of the participating hospitals, and the office members of the Dutch Consortium for their contribution.

Authors' roles

M.J.C.E., H.L.T., F.J.M.B. and B.W.J.M. designed the trial. T.C.T., H.L.T., F.J.M.B. and B.W.J.M. coordinated the trial. T.C.T., S.C.O., C.A.M.K., H.R.V., A.W.N., G.J.S., A.P.M., B.C.S., A.S., A.V., J.S.E.L., F.J.M.B. and all other members from the OPTIMIST study group collected data. M.J.C.E. and H.G. provided input on the statistical analyses. T.C.T. and M.J.C.E. performed the analyses. T.C.T. drafted the manuscript. T.C.T., M.J.C.E., H.L.T., F.J.M.B. and B.W.J.M. interpreted the data. All authors revised the report, and approved the final submitted version.

Funding

This study was funded by The Netherlands Organisation for Health Research and Development (ZonMW number 171102020).

Conflict of interest

T.C.T., H.L.T. and S.C.O. received an unrestricted personal grant from Merck BV. H.R.V. receives monetary compensation as a member on an external advisory board for Ferring pharmaceutical BV. B.W.J.M. is supported by a NHMRC Practitioner Fellowship (GNT1082548) and reports consultancy for OvsEva, Merck and Guerbet. F.J.M.B. receives monetary compensation as a member of the external advisory board for Ferring pharmaceuticals BV (the Netherlands) and Merck Serono (the Netherlands) for consultancy work for Gedeon Richter (Belgium) and Roche Diagnostics on automated AMH assay

development (Switzerland) and for a research cooperation with Ansh Labs (USA). All other authors have nothing to declare.

References

- Al-Inany HG, Youssef MA, Ayeleke RO, Brown J, Lam WS, Broekmans FJ. Gonadotrophin-releasing hormone antagonists for assisted reproductive technology (Review). *Cochrane Database of Syst Rev* 2016. Art. No.: CD001750.
- Baart EB, Martini E, van den Berg I, Macklon NS, Galjaard RJH, Fauser BCJM, van Opstal D. Preimplantation genetic screening reveals a high incidence of aneuploidy and mosaicism in embryos from young women undergoing IVF. *Hum Reprod* 2006;**21**:223–233.
- Baker VL, Brown MB, Luke B, Smith GW, Ireland JJ. Gonadotropin dose is negatively correlated with live birth rate: analysis of more than 650,000 assisted reproductive technology cycles. *Fertil Steril* 2015;**104**:1145–1152.
- Berkkanoglu M, Ozgur K. What is the optimum maximal gonadotropin dosage used in microdose flare-up cycles in poor responders? *Fertil Steril* 2010;**94**:662–665.
- de Boer EJ, den Tonkelaar I, te Velde ER, Burger CW, Klip H, van Leeuwen FE. A low number of retrieved oocytes at in vitro fertilization treatment is predictive of early menopause. *Fertil Steril* 2002;**77**:978–985.
- Broekmans FJM, Ziegler D De, Howles CM, Gougeon A, Trew G, Olivennes F. The antral follicle count: practical recommendations for better standardization. *Fertil Steril* 2010;**94**:1044–1051.
- Broer SL, van Disseldorp J, Broeze KA, Dolleman M, Opmeer BC, Bossuyt P, Eijkemans MJC, Mol BWJ, Broekmans FJM, Anderson R et al. Added value of ovarian reserve testing on patient characteristics in the prediction of ovarian response and ongoing pregnancy: An individual patient data approach. *Hum Reprod Update* 2013a;**19**:26–36.
- Broer SL, Dólleman M, van Disseldorp J, Broeze KA, Opmeer BC, Bossuyt PMM, Eijkemans MJC, Mol BWJ, Broekmans FJM. Prediction of an excessive response in in vitro fertilization from patient characteristics and ovarian reserve tests and comparison in subgroups: an individual patient data meta-analysis. *Fertil Steril* 2013b;**100**:420–429.
- Broer SL, Broekmans FJM, Laven JSE, Fauser BCJM. Anti-Müllerian hormone: Ovarian reserve testing and its potential clinical implications. *Hum Reprod Update* 2014;**20**:2688–2701.
- Drakopoulos P, Blockeel C, Stoop D, Camus M, De Vos M, Tournaye H, Polyzos NP. Conventional ovarian stimulation and single embryo transfer for IVF/ICSI. How many oocytes do we need to maximize cumulative live birth rates after utilization of all fresh and frozen embryos? *Hum Reprod* 2015;**31**:370–376.
- Fauser BCJM, Devroey P, Macklon NS. Multiple birth resulting from ovarian stimulation for subfertility treatment. *Lancet* 2005;**365**:1807–1816.
- Ferraretti AP, La Marca A, Fauser BC, Tarlatzis B, Nargund G, Gianaroli L, ESHRE working group on Poor Ovarian Response Definition. ESHRE consensus on the definition of 'poor response' to ovarian stimulation for in vitro fertilization: the Bologna criteria. *Hum Reprod* 2011;**26**:1616–1624.
- Harrison RF, Jacob S, Spillane H, Mallon E, Hennelly B. A prospective randomized clinical trial of differing starter doses of recombinant follicle-stimulating hormone (folitropin- beta) for first time in vitro fertilization and intracytoplasmic sperm injection treatment cycles. *Fertil Steril* 2001;**75**:23–31.
- Higgins JPT, Altman DG (Editors). Chapter 8: Assessing risk of bias in included studies. *Cochrane Handb Syst Rev Interv* Version 5.10 (updated March 2011) 2011; Date last accessed November 12th 2016.
- Humaidan P, Alviggi C, Fischer R, Esteves SC. The novel POSEIDON stratification of 'Low prognosis patients in Assisted Reproductive Technology' and its proposed marker of successful outcome. *F1000 Res* 2016;**5**:2911.
- Iliodromiti S, Anderson RA, Nelson SM. Technical and performance characteristics of anti-Müllerian hormone and antral follicle count as biomarkers of ovarian response. *Hum Reprod Update* 2015;**21**:698–710.

- Jayaprakasan K, Hopkisson J, Campbell B, Johnson I, Thornton J, Raine-Fenning N. A randomised controlled trial of 300 versus 225 IU recombinant FSH for ovarian stimulation in predicted normal responders by antral follicle count: fertility and assisted reproduction. *BJOG An Int J Obstet Gynaecol* 2010;**117**:853–862.
- Katz-Jaffe MG, Trounson AO, Cram DS. Chromosome 21 mosaic human preimplantation embryos predominantly arise from diploid conceptions. *Fertil Steril* 2005;**84**:634–643.
- Klinkert ER, Broekmans FJM, Looman CWN, Habbema JDF, te Velde ER. Expected poor responders on the basis of an antral follicle count do not benefit from a higher starting dose of gonadotrophins in IVF treatment: a randomized controlled trial. *Hum Reprod* 2005;**20**:611–615.
- Klinkert ER, Broekmans FJM, Looman CWN, te Velde ER. A poor response in the first in vitro fertilization cycle is not necessarily related to a poor prognosis in subsequent cycles. *Fertil Steril* 2004;**81**:1247–1253.
- Kolte AM, Bernardi LA, Christiansen OB, Quenby S, Farquharson RG, Goddijn M, Stephenson MD. Terminology for pregnancy loss prior to viability: a consensus statement from the ESHRE early pregnancy special interest group. *Hum Reprod* 2015;**30**:495–498.
- Lan VTN, Linh NK, Tuong HM, Wong PC, Howles CM. Anti-Mullerian hormone versus antral follicle count for defining the starting dose of FSH. *Reprod Biomed Online* 2013;**27**:390–399.
- Land JA, Yarmolinskaya MI, Dumoulin JC, Evers JL. High-dose human menopausal gonadotropin stimulation in poor responders does not improve in vitro fertilization outcome. *Fertil Steril* 1996;**65**:961–965.
- Lefebvre J, Antaki R, Kadach IJ, Dean NL, Sylvestre C, Bissonnette F, Benoit J, Ménard S, Lapensée L. 450 IU versus 600 IU gonadotropin for controlled ovarian stimulation in poor responders: a randomized controlled trial. *Fertil Steril* 2015;**104**:1419–1425.
- Legro RS, Wu X, Barnhart KT, Farquhar C, Fauser BCJM, Mol B. Improving the reporting of Clinical trials of infertility treatments (IMPRINT): modifying the CONSORT statement. *Hum Reprod* 2014;**29**:2075–2082.
- Munch EM, Sparks AE, Zimmerman MB, Van Voorhis BJ, Hakan Duran E. High FSH dosing is associated with reduced live birth rate in fresh but not subsequent frozen embryo transfers. *Hum Reprod* 2017;**32**:1402–1409.
- Munne S, Magli C, Adler A, Wright G, de Boer K, Mortimer D, Tucker M, Cohen J, Gianaroli L. Treatment-related chromosome abnormalities in human embryos. *Hum Reprod* 1997;**12**:780–784.
- Nelson SM, Klein BM, Arce J-C. Comparison of antimüllerian hormone levels and antral follicle count as predictor of ovarian response to controlled ovarian stimulation in good-prognosis patients at individual fertility clinics in two multicenter trials. *Fertil Steril* 2015a;**103**:923–930.
- Nelson SM, Pastuszek E, Kloss G, Malinowska I, Liss J, Lukaszuk A, Plociennik L, Lukaszuk K. Two new automated, compared with two enzyme-linked immunosorbent, antimüllerian hormone assays. *Fertil Steril* 2015b;**104**:1016–1021.
- Olivennes F, Trew G, Borini A, Broekmans F, Arriagada P, Warne DW, Howles CM. Randomized, controlled, open-label, non-inferiority study of the CONSORT algorithm for individualized dosing of follitropin alfa. *Reprod Biomed Online* 2015;**30**:248–257.
- Oudshoorn SC, van Tilborg TC, Eijkemans MJC, Oosterhuis GJE, Friederich J, van Hooff MHA, van Santbrink EJP, Brinkhuis EA, Smeenk JM, Kwee J et al. Individualized versus standard FSH dosing in predicted hyper responders starting IVF/ICSI: an RCT. Part 2: The predicted hyper responder. *Hum Reprod* 2017;**32**:2506–2514.
- PCOS Consensus Workshop Group TRE-SPCWG. Revised 2003 consensus on diagnostic criteria and long-term health risks related to polycystic ovary syndrome. *Fertil Steril* 2004;**81**:19–25.
- Popovic-Todorovic B, Loft A, Ejdrup Bredkjær H, Bangsbo S, Nielsen IK, Nyboe Andersen A. A prospective randomized clinical trial comparing an individual dose of recombinant FSH based on predictive factors versus a 'standard' dose of 150 IU/day in 'standard' patients undergoing IVF/ICSI treatment. *Hum Reprod* 2003;**18**:2275–2282.
- Poseidon Group, Alviggi C, Andersen CY, Buehler K, Conforti A, De Placido G, Esteves SC, Fischer R, Galliano D, Polyzos NP, Sunkara SK et al. A new more detailed stratification of low responders to ovarian stimulation: from a poor ovarian response to a low prognosis concept. *Fertil Steril* 2016;**105**:1452–1453.
- Sbracia M, Colabianchi J, Giallonardo A, Giannini P, Piscitelli C, Morgia F, Montigiani M, Schimberni M. Cetrorelix protocol versus gonadotropin-releasing hormone analog suppression long protocol for superovulation in intracytoplasmic sperm injection patients older than 40. *Fertil Steril* 2009;**91**:1842–1847.
- Sbracia M, Farina A, Poverini R, Morgia F, Schimberni M, Aragona C. Short versus long gonadotropin-releasing hormone analogue suppression protocols for superovulation in patients >40 years old undergoing intracytoplasmic sperm injection. *Fertil Steril* 2005;**84**:644–648.
- Scheffer GJ, Broekmans FJM, Bancsi LF, Habbema JDF, Looman CWN, te Velde ER. Quantitative transvaginal two- and three-dimensional sonography of the ovaries: reproducibility of antral follicle counts. *Ultrasound Obstet Gynecol* 2002;**20**:270–275.
- Scholten I, Braakhekke M, Limpens J, Hompes PA, van der Veen F, Mol BWJ, Gianotten J. Reporting multiple cycles in trials on medically assisted reproduction. *Reprod Biomed Online* 2016;**33**:646–651.
- Sunkara SK, Rittenberg V, Raine-Fenning N, Bhattacharya S, Zamora J, Coomarasamy A. Association between the number of eggs and live birth in IVF treatment: an analysis of 400 135 treatment cycles. *Hum Reprod* 2011;**26**:1768–1774.
- te Velde ER, Pearson PL. The variability of female reproductive aging. *Hum Reprod Update* 2002;**8**:141–154.
- Toftager M, Bogstad J, Løssl K, Prætorius L, Zedeler A, Bryndorf T, Nilas L, Pinborg A. Cumulative live birth rates after one ART cycle including all subsequent frozen-thaw cycles in 1050 women: secondary outcome of an RCT comparing GnRH-antagonist and GnRH-agonist protocols. *Hum Reprod* 2017;**32**:556–567.
- van der Gaast MH, Eijkemans MJC, van der Net JB, de Boer EJ, Burger CW, van Leeuwen FE, Fauser BCJM, Macklon NS. Optimum number of oocytes for a successful first IVF treatment cycle. *Reprod Biomed Online* 2006;**13**:476–480.
- van Tilborg TC, Broekmans FJM, Dólleman M, Eijkemans MJC, Mol BW, Laven JSE, Torrance HL. Individualized follicle-stimulating hormone dosing and in vitro fertilization outcome in agonist downregulated cycles: a systematic review. *Acta Obstet Gynecol Scand* 2016;**95**:1333–1344.
- van Tilborg TC, Eijkemans MJC, Laven JSE, Koks CAM, Bruin JP, de Scheffer GJ, van Golde RJT, Fleischer K, Hoek A, Nap AW et al. The OPTIMIST study: optimisation of cost effectiveness through individualised FSH stimulation dosages for IVF treatment. A randomised controlled trial. *BMC Womens Health* 2012;**12**:29.
- van Tilborg TC, Oudshoorn SC, Eijkemans MJC, Mochtar MH, van Golde RJT, Hoek A, Kuchenbecker WKH, Fleischer K, de Bruin JP, Groen H et al. Individualized FSH dosing based on ovarian reserve testing in women starting IVF/ICSI: a multicentre trial and cost-effectiveness analysis. *Hum Reprod* 2017;**32**:2485–2495.
- Verberg MFG, Eijkemans MJC, Heijnen EMEW, Broekmans FJ, de Klerk C, Fauser BCJM, Macklon NS. Why do couples drop-out from IVF treatment? A prospective cohort study. *Hum Reprod* 2008;**23**:2050–2055.
- Youssef MA, van Wely M, Al-Inany H, Madani T, Jahangiri N, Khodabakhshi S, Alhalabi M, Akhondi M, Ansari-pour S, Tokhmechy R et al. A mild ovarian stimulation strategy in women with poor ovarian reserve undergoing IVF: a multicenter randomized non-inferiority trial. *Hum Reprod* 2017;**32**:112–118.