

University of Groningen



Serum paraoxonase-1 activity is inversely related to free thyroxine in euthyroid subjects

van Tienhoven-Wind, Lynnda J. N.; Gruppen, Eke G.; James, Richard W.; Bakker, Stephan J. L.; Gans, Rijk O. B.; Dullaart, Robin P. F.

Published in: European Journal of Clinical Investigation

DOI: 10.1111/eci.12860

IMPORTANT NOTE: You are advised to consult the publisher's version (publisher's PDF) if you wish to cite from it. Please check the document version below.

Document Version Final author's version (accepted by publisher, after peer review)

Publication date: 2018

Link to publication in University of Groningen/UMCG research database

Citation for published version (APA): van Tienhoven-Wind, L. J. N., Gruppen, E. G., James, R. W., Bakker, S. J. L., Gans, R. O. B., & Dullaart, R. P. F. (2018). Serum paraoxonase-1 activity is inversely related to free thyroxine in euthyroid subjects: The PREVEND Cohort Study. European Journal of Clinical Investigation, 48(1), [12860]. https://doi.org/10.1111/eci.12860

Copyright Other than for strictly personal use, it is not permitted to download or to forward/distribute the text or part of it without the consent of the author(s) and/or copyright holder(s), unless the work is under an open content license (like Creative Commons).

If you believe that this document breaches copyright please contact us providing details, and we will remove access to the work immediately and investigate your claim.

Downloaded from the University of Groningen/UMCG research database (Pure): http://www.rug.nl/research/portal. For technical reasons the number of authors shown on this cover page is limited to 10 maximum.

DR. LYNNDA J VAN TIENHOVEN (Orcid ID : 0000-0001-6471-7244) DR. ROBIN P.F. DULLAART (Orcid ID : 0000-0003-4520-1239)

Article type : Original Paper

Serum paraoxonase-1 activity is inversely related to free thyroxine in euthyroid subjects: The PREVEND Cohort Study

Lynnda J.N. van Tienhoven-Wind MD¹, Eke G. Gruppen MsC¹, Richard W. James PhD², Stephan J.L. Bakker MD PhD¹, Rijk O.B. Gans MD PhD¹ and Robin P.F. Dullaart MD PhD¹

¹Department of Internal Medicine, University Medical Center Groningen and University of Groningen, The Netherlands

²Department of Internal Medicine, Faculty of Medicine, University of Geneva, Geneva, Switzerland

Corresponding author/reprints to: L.J.N. van Tienhoven-Wind, University Medical Center Groningen,

Department of Nephrology (AA53), PO Box 30.001 9700 RB Groningen The Netherlands

Telephone: +31 50 361 61 61, Fax: +31 50 361 93 10

E-mail: L.J.N.van.Tienhoven-Wind@umcg.nl

Running title: PON-1 activity in relation to free T₄

Key words: free thyroxine; high density lipoproteins; paraoxonase-1; thyroid function; metabolic

syndrome

This article has been accepted for publication and undergone full peer review but has not been through the copyediting, typesetting, pagination and proofreading process, which may lead to differences between this version and the Version of Record. Please cite this article as doi: 10.1111/eci.12860

Abstract

Background: Low-normal thyroid function within the euthyroid range has been suggested to enhance atherosclerosis susceptibility. Paraoxonase-1 (PON-1), may protect against atherosclerotic cardiovascular disease development by attenuating oxidative stress. We evaluated relationships of PON-1 with TSH, free T_4 , free T_3 , lipids and apolipoprotein (apo)A-I in euthyroid subjects, and assessed whether such relationships are modified in the context of the metabolic syndrome (MetS). **Materials and Methods**: Serum PON-1 activity (arylesterase activity), TSH, free T_4 , free T_3 , lipids and apoA-I were measured in 2206 euthyroid subjects (aged 28 to75 years; 1138 men (age 49 ± 13 years) and 1068 women (age 46 ± 12 years), recruited from the general population (PREVEND cohort).

Results: In age- and sex-adjusted analysis, PON-1 activity (divided into tertiles) was positively related to TSH (β =-0.045, *P*=0.036) and inversely to free T₄ (β =-0.042, *P*=0.050), but not to free T₃ (β =-0.027, *P*=0.20). PON-1 activity was positively related to total cholesterol, non-HDL cholesterol and triglycerides, as well as to HDL cholesterol and apoA-I (*P*<0.01 to <0.001). The inverse relationship of PON-1 activity with free T₄ remained present after adjustment for lipids and other potential confounders (β =-0.066, *P*=0.002), but the positive relationship with TSH lost significance (β =0.034, *P*=0.11). The inverse relationship of PON-1 activity with free T₄ was not different in subjects with vs. without MetS (*P*=0.94), nor modified by the presence of its individual components (*P*≥0.22 for each).

Conclusions: Serum PON-1 activity is inversely associated with free T_4 in euthyroid subjects, suggesting that low-normal thyroid function may affect PON-1 regulation.

Introduction

Low-normal thyroid function, as indicated by a higher thyroid stimulating hormone (TSH) or lower thyroid hormone levels within the euthyroid reference range, may contribute to the development of atherosclerotic cardiovascular disease (CVD) [1-4]. The mechanisms responsible for the association of (subclinical) atherosclerosis with low-normal thyroid function are still incompletely understood. Low-normal thyroid function is associated with a modest increase in plasma total cholesterol, low density lipoprotein (LDL) cholesterol and triglycerides [4-7]. Low-normal thyroid function may also attenuate high density lipoproteins (HDL) function, such as its ability to protect against oxidative stress [4,8].

Accumulating evidence supports the hypothesis that systemic oxidative stress, as at least in part reflected by enhanced oxidative modification of LDL, may contribute to the development of atherosclerosis [9-11]. In this context, it is relevant that LDL oxidation *in vitro* is exaggerated in both hypothyroidism and hyperthyroidism [12,13]. Moreover, increased circulating oxidized LDL levels have been demonstrated in euthyroid subjects with higher TSH levels [14]. Paraoxonase-1 (PON-1) is a HDL-associated hydrolytic enzyme with important anti-oxidative properties [15]. PON-1 hydrolyzes lipid peroxides, thereby preventing their accumulation in LDL particles [15,16]. Studies in rodent models and humans have suggested that the anti-atherogenic effects of the HDL fraction are to a considerable extent attributable to PON-1 activity [16]. PON-1 activity has been shown to be impaired in patients with metabolic syndrome (MetS), type 2 diabetes mellitus (T2DM) and hypercholesterolemia [15,17-19]. Furthermore, lower serum PON-1 activity may predict increased risk of coronary events [20-21] , although the association of PON-1 activity with increased CVD risk was not independent of HDL cholesterol [22].

The effect of thyroid dysfunction on serum PON-1 activity has only been determined in a limited number of studies [23-27]. Remarkably, PON-1 activity was found to be impaired in both hypothyroidism and hyperthyroidism [23]. In addition, PON-1 activity was decreased in (subclinical) hypothyroidism in some [24, 25], but not in other studies [26-28]. No data are currently available concerning the association of serum PON-1 activity in the context of variations in thyroid hormone levels within the euthyroid range.

Against this background, we performed the present study to evaluate the relationships of serum PON-1 activity with thyroid function in euthyroid subjects. In view of decreased PON-1 activity in MetS [15] and potential alterations in thyroid hormones in MetS [4], we also determined the extent to which such a relationship is modified by the presence of MetS and its individual components.

Subjects and Methods

Subjects

Reporting of the study conforms to STROBE (STrengthening the Reporting of OBservational studies in Epidemiology) statement along with references to STROBE statement and the broader EQUATOR guidelines [29].

The study population consisted of a random subset of participants of the PREVEND (Prevention of Renal and Vascular End Stage Disease) cohort, aged 28-75 years, living in the city of Groningen, The Netherlands. Participants were predominantly of Caucasian origin (94.2%). The protocol of this study has been described in detail elsewhere [30,31]. The local medical ethical committee approved the study; all participants gave written informed consent. For the current analysis, we excluded subjects not being euthyroid, subjects using thyroid hormones, anti-thyroid drugs, amiodarone and lithium carbonate. Euthyroidism was defined as TSH, free T_4 and free T_3 levels each within the respective reference range as provided by the manufacturer (see Laboratory Analyses). We additionally excluded subjects with positive anti-thyroid peroxidase autoantibodies (cut-off value: see Laboratory Analyses). Information on self-reported medication use was combined with information from a pharmacy-dispensing registry, which has complete information on drug of >95% of subjects in the PREVEND study. Applying these selection criteria 2206 subjects were eligible for the current analyses. The presence of a self-reported history of myocardial infarction, percutaneous coronary intervention, coronary artery bypass surgery, stroke or the diagnosis of narrowing of one or both carotid arteries was defined as CVD. Type 2 diabetes mellitus (T2DM) was defined as a fasting serum glucose concentration >7.0 mmol/L, a nonfasting plasma glucose concentration >11.1 mmol/L, a self-report of a physician diagnosis, or the use of glucose-lowering drugs. In order to categorize subjects with the metabolic syndrome (MetS) 3 or more of the following criteria were required: waist circumference > 102 cm for men and > 88 cm for women, hypertension (blood pressure \geq 130/85 mmHg or use of anti-hypertensive drugs), fasting plasma triglycerides \geq

1.70 mmol/L, fasting glucose \geq 5.6 mmol/L (or use of glucose lowering drugs), and HDL cholesterol < 1.03 mmol/L for men and < 1.29 mmol/L for women applying NCEP ATPIII criteria [32]. Patient characteristics including age, sex, alcohol use, smoking status, body mass index (BMI), waist circumference, systolic and diastolic blood pressure were obtained. The participants were instructed to let venous blood samples being drawn after an overnight fast for measurement of, glucose, total cholesterol, high density lipoprotein (HDL) cholesterol, triglycerides, TSH, free T₄ and free T₃ and PON-1 activity. Urinary albumin excretion (UAE) was documented as the mean of two 24-hour urine collections. Body mass index (BMI) was defined as weight (kg) by height (m) squared. Waist circumference (WC) was measured on bare skin between the 10th rib and iliac crest. Alcohol consumption was recorded with one drink being assumed to contain 10 grams of alcohol. Smoking was categorized into current, former and never. Estimated glomerular filtration rate (eGFR) was calculated with the use of the combined creatinine-cystatin C-based Chronic Kidney Disease Epidemiology Collaboration equation [33].

Laboratory analyses

Heparinized plasma samples were stored at -80°C until analyses. Sera were stored at -80°C until analyses. Serum TSH (Architect; Abbott Laboratories, Abbott Park, IL, USA; reference range 0.35 - 4.94 mU/L), free T₄ (AxSYM; Abbott Laboratories, Abbott Park, IL, USA; reference range 9.14 - 23.81 pmol/L) and free T₃ (AxSYM; Abbott Laboratories, Abbott Park, IL, USA; reference range; 2.23 - 5.35 pmol/L) were measured by microparticle enzyme immunoassays. Anti-thyroid peroxidase autoantibodies were determined using commercially available automated enzyme linked immunoassays (Abbott Laboratories, Abbott Park, IL, USA; kit number 5F57). Anti-thyroid peroxidase autoantibodies were considered positive using a cut-off value as indicated by the supplier ($\geq 12 \text{ kU/L}$).

Serum PON-1 enzymatic activity was measured as its arylesterase activity, i.e. as the rate of hydrolysis of phenyl acetate into phenol, as described [34]. The inter-assay CV was 8%. Arylesterase activity, measured with this assay, is positively correlated with PON-1 enzymatic activity toward paraoxon as well as with PON-1 mass [35].

Total serum cholesterol and plasma glucose were measured using Kodak Ektachem dry chemistry (Eastman Kodak, Rochester, NY, USA). Serum triglycerides were measured enzymatically. HDL cholesterol was measured with a homogeneous method (direct HDL, AEROSET system; Abbott Laboratories, Abbott Park, IL, USA; no. 7D67). Non-HDL cholesterol was calculated as the difference between total cholesterol and HDL cholesterol. Serum apoA-I was determined by nephelometry applying commercially available reagents for Dade Behring nephelometer systems (BN II; Dade Behring, Marburg, Germany; apoA-I test kit, code no. OUED).

Serum creatinine was measured by an enzymatic method on a RocheModular analyzer (Roche Diagnostics, Mannheim, Germany). Serum cystatin C was measured by Gentian Cystatin C Immunoassay (Gentian AS, Moss, Norway) on a Modular analyzer (Roche Diagnostics). Urinary albumin concentration was measured by nephelometry with a threshold of 2.3 mg/l (Dade Behring Diagnostic, Marburg, Germany).

2.4 Statistical analyses

Data analysis was performed using IBM SPSS software (version 23.0, SPSS Inc. Chicago, IL, USA). Normally distributed data are given as mean \pm SD and non-parametrically distributed data are presented as median (interquartile range). Categorical variables are given as percentages. Differences in PON-1 activity between men and women were determined by Mann-Whitney U-test. Clinical and laboratory characteristics of the study population are presented according to sex-stratified tertiles of PON-1 activity. Differences in proportions of dichotomous variables across tertiles of PON-1 activity were determined by multinomial χ -square tests. Multivariable linear regression analyses, adjusted for age and sex, were used to test for linear trends between tertiles of PON-1 activity. Age- and sex-adjusted multivariable linear regression analyses were also used to determine the extent to which PON-1 activity (as continuous variable) was related to thyroid function parameters (TSH, free T₄, free T₃) taking clinical and laboratory covariates into account. PON-1 activity, TSH, triglycerides and UAE were natural logarithm (log_e) transformed in order to achieve approximately normal distributions. Interaction terms were calculated as the product term of TSH or free T₄ with sex or the presence of MetS or its component of interest. To account for outliers the individual TSH or free T₄

values were centered to the mean by subtracting the group mean value from individual values [36,37]. Interaction terms were considered statistically significant at *P*-values <0.10, as recommended by Selvin [38]. Otherwise, two-sided *P*-values <0.05 were considered significant.

Results

Mean age of the 2206 participants was 48 ± 13 years. 1138 participants (52.6 %) were men (age 49 ± 13 years) and 1068 (47.4 %) were women (age 46 ± 12 years). Serum PON-1 activity amounted to 56.1 (46.1 - 68.1) U/L in the whole population, and was 60.0 (46.7 - 70.5) U/L in women vs. 54.9 (45.8 - 66.0) U/L in men (P=0.002). Clinical and laboratory characteristics of the study population are, therefore, shown according to sex-stratified tertiles of PON-1 activity (Table 1). One hundred two participants (4.6 %) reported a previous cardiovascular event, 65 subjects (2.9 %) had T2DM and 418 (19%) subjects fulfilled the criteria for MetS. A history of CVD (P=0.002) and the presence of MetS (P=0.013) was more prevalent in subjects categorized in the lowest tertile of PON-1 activity, but diabetes status did not significantly vary according to the PON-1 categories (P=0.096) (Table 1). Oral glucose lowering drugs were used by 36 subjects, lipid modifying drugs (mainly statins) by 113 participants and antihypertensives by 274 subjects. Oral contraceptives were used by 283 women. The use of oral glucose lowering drugs (P < 0.002, antihypertensives (P < 0.001) was more prevalent in the subjects belonging to the lowest tertile of PON-1 activity, whereas the use of oral contraceptives was more prevalent in the women belonging to the highest tertile of PON-1 activity (P<0.001) (data not shown). The use of lipid lowering drugs did not vary across tertiles of PON-I activity (P=0.17). Accordingly, serum PON-1 activity was lower in subjects using oral glucose lowering drugs (45.9 (39.5-56.8 U/L) vs. 56.2 (46.3-68.4) U/L, P<0.001), in subjects using antihypertensives (52.6 (41.8-64.7 U/L) vs. 56.7 (46.8-68.8 U/L), P<0.001) and in women using oral contraceptives (62.9 (52.2-76.7 U/L) vs. 55.2 (45.7-66.7 U/L), P<0.001), but was not different in subjects using lipid lowering drugs compared to those who did not (56.5 (46.6-64.2 U/L vs. 56.1 (46.1-67.5) U/L, P=0.10). Serum PON-1 activity was inversely related to age. In age- and sexadjusted analysis, PON-1 activity was positively related to systolic and diastolic blood pressure (Table 1). PON-1 activity was unrelated to BMI, waist circumference, glucose, eGFR and UAE, and did not

vary significantly according to smoking status and alcohol consumption. Additionally, PON-1 activity was positively related to total cholesterol, non-HDL cholesterol, triglycerides, as well as to HDL cholesterol and apoA-I (Table 1). Of note, in age- and sex-adjusted analysis, PON-1 activity was positively related to TSH and inversely to free T₄, but not to free T₃ (Table 1). Fig.1 shows TSH, free T₄ and free T₃ levels according to sex-stratified tertiles of PON-1activity. There were no interactions of sex with TSH, free T₄ or free T₃ on PON-1 activity (*P*=0.52 to *P*=0.64; data not shown).

We then tested whether the relationships of PON-1 activity (as continuous variable) with TSH and free T₄ remained present after adjustment for relevant clinical and laboratory covariates (Table 2). In age- and sex-adjusted multivariable linear regression analysis including free T_4 and TSH together PON-1 activity was positively associated with TSH and inversely with free T₄ (Table 2, model 1). In analysis with free T_4 , free T_3 and TSH together, there was no significant independent association of PON-1 activity with free T_3 (β =-0.029, P=0.18; data not shown). The inverse relationship of PON-1 activity with free T₄ remained present after additional adjustment for non-HDL cholesterol, HDL cholesterol and triglycerides, although the positive relationship of PON-1 activity with TSH lost significance (Table 2, model 2). Likewise, PON-1 activity was inversely related to free T₄ in an alternative model which included apoA-I instead of HDL cholesterol (β = -0.055, P=0.01; data not shown). An inverse relationship of PON-1 activity with free T₄ was also found after additional adjustment for systolic and diastolic blood pressure, UAE, eGFR, alcohol consumption, smoking, a previous history of cardiovascular disease and diabetes status (Table 2, model 3), and finally after further adjustment for oral glucose lowering drugs, lipid lowering medication, antihypertensives and oral contraceptives (Table 2, model 4). The inverse relationship of PON-1 activity with free T₄ was not different in subjects with vs. without MetS (P=0.94), nor modified by the presence of its individual components (low HDL cholesterol: P=0.58); elevated triglycerides: P=0.96); enlarged waist circumference: P=0.57); elevated blood pressure: P=0.31; elevated glucose: P=0.22).

Secondary analyses were performed after exclusion of subjects with a previous history of CVD and T2DM (n=2051, Table 3, model 1), as well as after exclusion of subjects using oral glucose lowering drugs, lipid lowering drugs, antihypertensives and oral contraceptives (n=1596, Table 3,

model 2). In both analyses, PON-I activity remained inversely associated with free T₄ (β = -0.047 to - 0.049, *P*=0.035 to 0.049) taking account of age, sex, non-HDL cholesterol, HDL cholesterol, triglycerides, systolic and diastolic blood pressure, alcohol consumption and smoking status (data not shown). In these analyses PON-1 activity was also positively related to HDL cholesterol (β = 0.236 to 0.217, *P*<0.001).

Discussion

In this large population-based study among strictly euthyroid subjects, we have shown to our knowledge for the first time, that serum PON-1 activity is positively related to TSH and inversely to free T_4 in age- and sex-adjusted analysis. In multivariable logistic regression analysis in which we included TSH, free T_4 and free T_3 together and adjusted for lipoproteins and other potentially important covariates, the inverse association of PON-1 activity with free T_4 remained present. The inverse relationship of PON-1 activity with free T_4 was not different between subjects with and without MetS nor modified by the presence of its individual components. Our current results are, therefore, in agreement with the hypothesis that variations in thyroid function within the euthyroid range may affect serum PON-1 activity.

In the interpretation of the results it is relevant that serum PON-I activity was assayed with phenyl acetate as substrate. Arylesterase activity, as measured with this type of assay, is widely used in large scale studies, and has the advantage of an approximately normal distribution, making it suitable for multivariable modeling [22]. Moreover, PON-I activity towards phenyl acetate is less variable between subjects compared to its activity towards paraoxon [overviewed in 18]. As expected [22,39-41], PON-1 activity was positively related to HDL cholesterol and apoA-I. Its correlation with non-HDL cholesterol and triglycerides is probably explained by an association of PON-1 with very low density lipoproteins which are able to act as a vector for its cellular secretion [42]. Such relations of PON-1 activity with circulating lipoproteins together with the effect of low-normal thyroid function to increase plasma cholesterol and triglycerides [4,5,43] underscore the necessity to adjust for (apo) lipoprotein levels when evaluating the relationship of PON-1 activity with variation in thyroid

function in euthyroid subjects. In the present study we only included euthyroid subjects using strict criteria, i.e. TSH, free T_4 and free T_3 each being within their respective reference range, as done in other reports [44,45]. Moreover, we excluded subjects with positive anti-thyroid peroxidase autoantibodies to avoid possible confounding of latent thyroid autoimmunity on inflammatory and oxidative stress as much as possible [25].

Inconsistent effects of thyroid function status on PON-1 activity have been reported so far [24-28]. The inverse relation of PON-1 activity with free T₄ as shown in the current study suggests that low-normal thyroid function could contribute to higher PON-1 activity, although it should be emphasized that this relationship was modest. The mechanisms responsible for this relationship are not yet known. It is unclear whether thyroid hormones are able to affect PON-1 gene expression. PON-1 is down regulated by interleukin-1 (IL-1) and tumor necrosis factor- α (TNF- α) [46]. Hypothyroidism may increase IL-1 and TNF- α [47], whereas higher levels of TNF- α are also found in subjects with low-normal thyroid function [48]. In addition, oxidized lipids are recognized to inhibit PON-1 activity [49-51]. In this context, it is relevant that low density lipoprotein (LDL) oxidation *in vitro* is exaggerated in both hypothyroidism and hyperthyroidism [12,13]. Moreover, increased levels of oxidized LDL have been demonstrated in euthyroid subjects with high normal TSH levels [14]. Taken together, these data [14,46-48] make it unlikely that a higher PON-1 activity in relation to low normal thyroid function is to explained by thyroid hormone- mediated effects on IL-1 and TNF- α or on (systemic) oxidative stress.

Given the inverse though modest relation of PON-1 activity with free T_4 , it seems plausible that other mechanisms than effects of PON-1 on oxidative stress defense could contribute to the previously reported enhanced oxidative stress in the context of low normal thyroid function [14]. It also seems unlikely that changes in PON-1 activity play a major role in an attenuated ability of HDL to protect against LDL oxidation *in vitro* in subjects with low-normal thyroid function [8], a read-out of HDL functionality which is closely related to PON-1 activity [52]. In this regard, is relevant that other factors affecting oxidative stress such as superoxide dismutase [53] and circulating bilirubin levels [44] are also affected by thyroid function. Of note, it has been demonstrated recently that the

inverse relationship of bilirubin with free T4 is stronger in more insulin resistant individuals [44], and that the relationship of PON-1 activity with its activator, apoE, is impaired in subjects with MetS [41]. For this reason we also set out to determine whether the relationship of PON-1 activity was modified in the context of MetS. We found that the inverse relationship of PON-1 activity with free T_4 was not modified by the presence of MetS nor by its individual components.

The regulation of PON-1 is dependent on many genetic and environmental factors. Regarding environmental factors, several animal and human studies have shown that dietary lipids can influence PON-1 activity [54-56]. Furthermore it has been reported that physically active subjects have higher PON-1 activity [57]. A limitation of the present study is that detailed information on nutrient intake and data with respect to physical activity were not available. Statins may also increase PON-1 activity [58], although this has not been unequivocally reported [59]. In the current report, PON-1 activity was not affected by the use of lipid lowering drugs. However, PON-1 activity was inversely associated with the use of glucose lowering medication in analysis in which we also adjusted for the presence of T2DM. We explain this finding by assuming that the use of glucose lowering drugs preferentially labels diabetic patients with more severe hyperglycemia, requiring medical drug treatment. Further, PON-1 activity was elevated in women who used oral contraceptives. Although little information is available on this issue, it seems consistent with some other data suggesting that PON-1 activity is higher in women taking oral contraceptives, and may increase in response to ethinyl oestradiol and cyproterone acetate combination [60,61].

Several other methodological aspects and limitations of our study need to be considered. We performed a cross-sectional study, so that conclusions regarding cause-effect relationships cannot be drawn with certainty. However, we are not aware of any data underscoring a physiological role of PON-1 itself in thyroid hormone regulation. In addition owing to the observational nature of our study, residual confounding due to unmeasured confounders cannot be entirely ruled out. We performed secondary analyses after exclusion of subjects with a history of CVD and T2DM, and the use of glucose lowering, lipid lowering, antihypertensive medication and oral contraceptives. Reassuringly, these analyses showed the same inverse relationship of PON-1 activity with free T₄. Of

further note, PON-1 activity was assayed in sera that were stored for a prolonged period. However, loss of PON-1 enzymatic activity are minimal if samples stored frozen at -70 C^{0} [62].

In conclusion, this large population-based cohort study demonstrates for the first time that serum PON-1 activity is inversely associated with free T_4 in euthyroid subjects. It is conceivable that low-normal thyroid function may influence PON-1 regulation.

Author Contributions: Conception and design of the study: LTW, EG, RJ, SB, RG and RD. Data collection and analysis: LTW, EG and RD. Interpretation of data: LTW, EG and RD. Drafting the manuscript: LTW, EB, and RD. All authors have revised and approved the submitted manuscript.

Grant support: The Dutch Kidney Foundation supported the infrastructure of the PREVEND program from 1997 to 2003 (Grant E.033). The Groningen University Medical Center supported the infrastructure from 2003 to 2006. Dade Behring, Ausam, Roche and Abbott, financed laboratory equipment and reagents by which various laboratory determinations could be performed. Richard W. James conducted the assays for paraoxonase-1 activity in PREVEND. The Dutch Heart Foundation supported studies on lipid metabolism (2001-005).

Acknowledgments: The authors wish to acknowledge the services of the PREVEND Study, contributing research centers, participating general practitioners and pharmacists and all the study participants.

Disclosure Statement: The authors have nothing to disclose.

References

- Taylor PN, Razvi S, Pearce SH, Dayan CM. Clinical review: A review of the clinical consequences of variation in thyroid function within the reference range. J Clin Endocrinol Metab 2013;98:3562-3571.
- Dullaart RP, de Vries R, Roozendaal C, Kobold AC, Sluiter WJ. Carotid artery intima media thickness is inversely related to serum free thyroxine in euthyroid subjects. Clin Endocrinol (Oxf) 2007;67:668-673.
- Zhang Y, Kim BK, Chang Y, Ryu S, Cho J, Lee WY, et al. Thyroid hormones and coronary artery calcification in euthyroid men and women. Arterioscler Thromb Vasc Biol 2014;34:2128-2134.
- van Tienhoven-Wind, Dullaart RP. Low-normal thyroid function and novel cardiometabolic biomarkers. Nutrients 2015;16:7:1352-1377.
- Roos A, Bakker SJ, Links TP, Gans RO, Wolffenbuttel BH. Thyroid function is associated with components of the metabolic syndrome in euthyroid subjects. J Clin Endocrinol Metab 2007;92: 491–496.
- Kim BJ, Kim TY, Koh JM, Kim HK, Park JY, Lee KU et al. Relationship between serum free T4 (FT4) levels and metabolic syndrome (MS) and its components in healthy euthyroid subjects. Clin Endocrinol 2009;70:152–160.
- Wang F, Tan Y, Wang C, Zhang X, Zhao Y, Song X, et al. Thyroid-stimulating hormone levels within the reference range are associated with serum lipid profiles independent of thyroid hormones. J Clin Endocrinol Metab 2012;97:2724–2731.
- Triolo M, de Boer JF, Annema W, Kwakernaak AJ, Tietge UJ, Dullaart RP. Low normal free T4 confers decreased high-density lipoprotein antioxidative functionality in the context of hyperglycaemia. Clin Endocrinol (Oxf) 2013;79:416-423.
- Shih DM, Gu L, Xia YR, Navab M, Li WF, Hama S, et al. Mice lacking serum paraoxonase are susceptible to organophosphate toxicity and atherosclerosis. Nature 1998;394:284–287.
- Tward A, Xia YR, Wang XP, Shi YS, Park C, Castellani LW, et al. Decreased atherosclerotic lesion formation in human serum paraoxonase transgenic mice. Circulation 2002;106:484–490.

- 11. Soran H, Younis NN, Charlton-Menys V, Durrington P. Variation in paraoxonase-1 activity and atherosclerosis. Curr Opin Lipidol 2009;20:265–274.
- Sundaram V, Hanna AN, Koneru L, Newman HA, Falko JM. Both hypothyroidism and hyperthyroidism enhance low density lipoprotein oxidation. J Clin Endocrinol Metab 1997;82:3421-3424.
- Costantini F, Pierdomenico SD, De Cesare D, De Remigis P, Bucciarelli T, Bittolo-Bon G, et al. Effect of thyroid function on LDL oxidation. Arterioscler Thromb Vasc Biol 1998;18:732-737.
- Ittermann T, Baumeister SE, Völzke H, Wasner C, Schminke U, Wallaschofski H, et al. Are serum TSH levels associated with oxidized low-density lipoprotein? Results from the Study of Health in Pomerania. Clinical Endocrinology (Oxford) 2012;76:526-532.
- 15. Deakin SP, James RW. Genetic and environmental factors modulating serum concentrations and activities of the antioxidant enzyme paraoxonase-1. Clin Sci (Lond) 2004;107:435–447.
- 16. Karabina SA, Lehner AN, Parthasarathy S, Santanam N. Oxidative inactivation of paraoxonaseimplications in diabetes mellitus and atherosclerosis. Biochim Biophys Acta 2005;1725:213-221.
- Mackness MI, Harty D, Bhatnagar D, Winocour PH, Arrol S, Ishola M, etal. Serum paraoxonase activity in familial hypercholesterolaemia and insulin-dependent diabetes mellitus. Atherosclerosis 1991;86:193–199.
- Dullaart RP, de Vries R, Sluiter WJ, Voorbij HA. High plasma C-reactive protein (CRP) is related to low paraoxonase-I (PON-I) activity independently of high leptin and low adiponectin in type 2 diabetes mellitus. Clin Endocrinol 2009;70:221–226.
- Fülöp P, Harangi M, Seres I, Paragh G. Paraoxonase-1 and adipokines: Potential links between obesity and atherosclerosis. Chem Biol Interact 2016;25 259(Pt B):388-393.
- 20. Bhattacharyya T, Nicholls SJ, Topol EJ, Zhang R, Yang X, Schmitt D, et al. Relationship of paraoxonase 1 (PON1) gene polymorphisms and functional activity with systemic oxidative stress and cardiovascular risk. JAMA 2008;299:1265e76.

- 21. van Himbergen TM, van der YT, Voorbij HA, van Tits LJ, Stalenhoef AF, Peeters PH, et al. Paraoxonase (PON1) and the risk for coronary heart disease and myocardial infarction in a general population of Dutch women. Atherosclerosis 2008;199:408-414.
- 22. Kunutsor SK, Bakker SJL, James RW, Dullaart RPF. Serum paraoxonase-1 activity and risk of incident cardiovascular disease: The PREVEND study and meta-analysis of prospective population studies. Atherosclerosis 2016;245:143–154.
- 23. Azizi F, Raiszadeh F, Solati M, Etemadi A, Rahmani M, Arabi M. Serum paraoxonase 1 activity is decreased in thyroid dysfunction. J Endocrinol Invest 2003;26:703–709.
- Cebeci E, Alibaz-Oner F, Usta M, Yurdakul S, Erguney M. Evaluation of oxidative stress, the activities of paraoxonase and arylesterase in patients with subclinical hypothyroidism. J Investig Med 2012;60:23-28.
- 25. Ates I, Altay M, Yilmaz FM, Topcuoglu C, Yilmaz N, Berker D, et al. The impact of levothyroxine sodium treatment on oxidative stress in Hashimoto's thyroiditis. Eur J Endocrinol 2016;174:727-734.
- 26. Sigal GA, Medeiros-Neto G, Vinagre JC, Diament J, Maranhão RC. Lipid metabolism in subclinical hypothyroidism: plasma kinetics of triglyceride-rich lipoproteins and lipid transfers to high-density lipoprotein before and after levothyroxine treatment. Thyroid 2011;21:347-353.
- 27. Milionis HJ, Tambaki AP, Kanioglou CN, Elisaf MS, Tselepis AD, Tsatsoulis A. Thyroid substitution therapy induces high-density lipoprotein-associated platelet-activating factoracetylhydrolase in patients with subclinical hypothyroidism: a potential antiatherogenic effect. Thyroid 2005;15:455-460.
- 28. Kebapcilar L, Comlekci A, Tuncel P, Solak A, Secil M, Gencel O, et al. Effect of levothyroxine replacement therapy on paraoxonase-1 and carotid intima-media thickness in subclinical hypothyroidism. Med Sci Monit 2010;16:CR41-47.
- 29. Simera I, Moher D, Hoey J, Schulz KF, Altman DG. A catalogue of reporting guidelines for health research. Eur J Clin Invest 2010;40:35–53.

- Halbesma N, Brantsma AH, Bakker SJ, Jansen DF, Stolk RP, De Zeeuw D, et al. Gender differences in predictors of the decline of renal function in the general population. Kidney Int 2008;74:505-512.
- 31. Corsetti JP, Gansevoort RT, Bakker SJ, Sparks CE, Vart P, Dullaart RP. Apolipoprotein B attenuates albuminuria-associated cardiovascular disease in prevention of renal and vascular endstage disease (PREVEND) participants. J Am Soc Nephrol 2014;25:2906-2915.
- 32. Grundy SM, Cleeman JI, Daniels SR, Donato KA, Eckel RH, Franklin BA, et al. Diagnosis and management of the metabolic syndrome: an American Heart Association/National Heart, Lung, and Blood Institute Scientific Statement. Circulation 2005;112:2735–2752.
- 33. Inker LA, Schmid CH, Tighiouart H, Eckfeldt JH, Feldman HI, Greene T, et al; CKD-EPI Investigators. Estimating glomerular filtration rate from serum creatinine and cystatin C. N Engl J Med 2012;367:20-29.
- Richter RJ, Jarvik GP, Furlong CE. Paraoxonase 1 (PON1) status and substrate hydrolysis. Toxicology and applied pharmacology 2009;235:1-9.
- 35. van Himbergen TM, Roest M, de Graaf J, Jansen EH, Hattori H, Kastelein JJ, et al. Indications that paraoxonase-1 contributes to plasma high density lipoprotein levels in familial hypercholesterolemia. Journal of lipid research 2005;46:445-451.
- 36. Shieh G. Clarifying the role of mean centring in multicollinearity of interaction effects. Br J Math Stat Psychol 2011;64:462-477.
- Kraemer HC, Blasey CM. Centring in regression analyses: A strategy to prevent errors in statistical inference. Int J Methods Psychiatr Res 2004;13:141-151.
- 38. Selvin S. Statistical analysis of epidemiological data. New York: Oxford University Press. 1996.
- 39. Deakin SP, James RW. Genetic and environmental factorsmodulating serumconcentrations and activities of the antioxidant enzyme paraoxonase-1. Clin Sci (Lond) 2004;107:435–447.
- Blatter Garin MC, Moren X, James RW. Paraoxonase-1 and serum concentrations of HDLcholesterol and apoA-I. J Lipid Res 2006;47:515–520.

- Dullaart RPF, Kwakernaak AJ, Dallinga-Thie GM. The positive relationship of serum paraoxonase-1 activity with apolipoprotein E is abrogated in metabolic syndrome. Atherosclerosis 2013;230:6–11.
- 42. Deakin S, Moren X, James RW. Very low density lipoproteins provide a vector for secretion of paraoxonase-1 from cells. Atherosclerosis 2005;179:17-25.
- 43. van Tienhoven-Wind L, Dullaart RP. Low normal thyroid function as a determinant of increased large very low density lipoprotein particles. Clin Biochem 2015;48:489-494.
- 44. Deetman PE, Bakker SJ, Kwakernaak AJ, Navis G, Dullaart RP; PREVEND Study Group. The relationship of the anti-oxidant bilirubin with free thyroxine is modified by insulin resistance in euthyroid subjects. PLoS One 2014;9:e90886.
- 45. van den Berg EH, van Tienhoven-Wind LJ, Amini M, Schreuder TC, Faber KN, Blokzijl H, et al. Higher free triiodothyronine is associated with non-alcoholic fatty liver disease in euthyroid subjects: the Lifelines Cohort Study. Metabolism 2017;67:62-71.
- 46. Kumon Y, Nakauchi Y, Suehiro T, Shiinoki T, Tanimoto N, Inoue M, et al. Proinflammatory cytokines but not acute phase serum amyloid A or C-reactive protein, down regulate paraoxonase 1 (PON1) expression by Hep G2. Amyloid 2002:9:160-164.
- Weetman AP. Cellular immune responses in autoimmune thyroid disease. Clin Endocrinol (Oxf) 2004;61:405-413.
- 48. van Tienhoven-Wind LJ, Dullaart RP. Tumor Necrosis Factor-α is Inversely Related to Free Thyroxine in Euthyroid Subjects Without Diabetes. Horm Metab Res 2017;49:95-102.
- Sundaram V, Hanna AN, Koneru L, Newman HA, Falko JM. Both hypothyroidism and hyperthyroidism enhance low density lipoprotein oxidation. J Clin Endocrinol Metab 1997;82:3421-3424.
- Mackness MI, Arrol S, Abbott C, Durrington PN. Protection of low-density lipoprotein against oxidative modification by high-density lipoprotein associated paraoxonase. Atherosclerosis 1993;104:129-135.

- 51. Aviram M, Rosenblat M, Billecke S, Erogul J, Sorenson R, Bisgaier CL, et al. Human serum paraoxonase (PON 1) is inactivated by oxidized low density lipoprotein and preserved by antioxidants. Free Radic Biol Med 1999;26:892-904.
- 52. Kappelle PJ, de Boer JF, Perton FG, Annema W, de Vries R, Dullaart RP, et al. Increased LCAT activity and hyperglycaemia decrease the antioxidative functionality of HDL. Eur J Clin Invest 2012;42:487-495.
- 53. Baskol G, Atmaca H, Tanriverdi F, Baskol M, Kocer D, Bayram F. Oxidative stress and enzymatic antioxidant status in patients with hypothyroidism before and after treatment. Exp Clin Endocrinol Diabetes 2007;115:522-526.
- Kudchodkar BJ, Lacko AG, Dory L, Fungwe TV. Dietary fat modulates serum paraoxonase 1 activity in rats. J. Nutr 2000; 130:2427-33.
- 55. Sutherland WH, Walker RJ, de Jong SA, van Rij AM, Philips V, Walker HL. Reduced postprandial serum paraoxonase activity after a meal rich in used cooking fat. Arterioscler Thromb Vasc Biol 1999; 19:1340-7.
- 56. Kim DS, Maden SK, Burt AA, Ranchalis JE, Furlong CE, Parvik GP. Dietary fatty acid intake is associated with paraoxonase 1 activity in a cohort-based analysis of 1,548 subjects. Lipids in health and Disease 2013; 112:183.
- 57. Senti M, Tomás M, Anglada R, Elosua R, Marrugat J, Covas MI, et al. Interrelationship of smoking, paraoxonase activity, and leisure time physical activity: a population-based study. Eur J Intern Med. 2003 May;14(3):178-184.
- 58. Tomás M, Sentí M, García-Faria F, Vila J, Torrents A, Covas M, Marrugat J. Effect of simvastatin therapy on paraoxonase activity and related lipoproteins in familial hypercholesterolemic patients. Arterioscler Thromb Vasc Biol. 2000 Sep;20(9):2113-9.
- 59. Dullaart RPF, de Vries R, Voorbij HAM, Sluiter WJ, van Tol A. Serum paraoxonase-I activity is unaffected by short-term administration of simvastatin, bezafibrate and their combination in type 2 diabetes mellitus. Eur J Clin Invest 2009; 39(3):200-203.

- 60. Kowalska K, Ściskalska M, Bizoń A, Śliwińska-Mossoń M, Milnerowicz H. Influence of oral contraceptives on lipid profile and paraoxonase and commonly hepatic enzymes activities. Clin Lab Anal. 2017 Mar 9. doi: 10.1002/jcla.22194. [Epub ahead of print].
- 61. Carlioglu A, Kaygusuz I, Karakurt F, Gumus II, Uysal A, Kasapoglu B, Armutcu F, Uysal S, Keskin EA, Koca C. The platelet activating factor acetyl hydrolase, oxidized low-density lipoprotein, paraoxonase 1 and arylesterase levels in treated and untreated patients with polycystic ovary syndrome. Arch Gynecol Obstet. 2014 Nov;290(5):929-35.
- 62. Huen K, Richter R, Furlong C, Eskenazi B, Holland N. Validation of PON1 enzyme activity assays for longitudinal studies. Clin Chim Acta 2009;402:67-74.

Figure legend:

Figure 1. TSH, and free T_4 and free T_3 levels according to sex-stratified tertiles of serum paraoxonase-1 (PON-1) activity . *P*-values for linear trend (adjusted for age and sex): TSH: *P*=0.036, free T_4 : *P*=0.050 and free T_3 : *P*=0.20. Data are given in means and standard errors. TSH is logarithmically transformed.

 Table 1. Clinical and laboratory characteristics in 2206 subjects according to sex-stratified tertiles of paraoxonase-1 (PON-1) activity.

	Sex stratified ter				
	1	2	3		
	Men	Men	Men		
	19.7-48.8	48.9-62.2	62.2-130.7		
	Women	Women	Women		
	17.0-50.2	50.2-65.1	65.2-119.3	β	<i>P</i> -value
Participants, n	735	736	735		
Men, n (%)	379 (51.6)	380 (51.6)	379 (51.6)		
Women, n (%)	356 (48.4)	356 (48.4)	356 (48.4)		
Age (years)	49.8 ± 13.2	48.1 ± 12.6	45.7 ± 11.4	-0.132	< 0.001
BMI (kg/m ²)	25.9 ± 4.3	26.1 ± 4.6	25.4 ± 3.9	-0.013	0.65
Waist circumference (cm)	87.8 ± 13.3	88.1 ± 13.5	86.3 ± 12.7	0.000	0.98
Systolic blood pressure (mmHg)	129 ± 20	129 ± 21	127 ± 18	0.038	0.037
Diastolic blood pressure (mmHg)	74 ± 9	74 ± 10	74 ± 10	0.053	0.004
Alcohol					0.43
< 10 gram per day (%)	538 (73.6)	521 (71.2)	518 (70.8)		
\geq 10 gram per day (%)	193 (26.4)	211 (28.8)	214 (29.2)		

Smoking					0.081
	199 (25.9)	221 (20.2)	221 (20.1)		
never (%)	188 (25.8)	221 (30.2)	221 (30.1)		
former (%)	246 (33.7)	245 (33.5)	263 (35.8)		
aurrant (0/)	206 (40.5)	266 (26 2)	250 (24.1)		
current (%)	290 (40.3)	200 (30.3)	230 (34.1)		
Glucose (mmol/L)	4.6±1.3	4.6±1.1	4.5±1.1	0.007	0.74
Total cholesterol (mmol/L)	5.5 ± 1.14	5.6 ± 1.22	5.7 ± 1.12	0.12	<0.001
Non-HDL cholesterol (mmol/L)	4.2 ± 1.25	4.2 ± 1.31	4.3 ± 1.22	0.069	<0.001
HDL cholesterol (mmol/L)	1.28 ± 0.39	1.36 ± 0.40	1.42 ± 0.42	0.131	<0.001
Triglycerides (mmol/L)	1.14 (0.81-1.63)	1.12 (0.82-1.62)	1.16 (0.85-1.69)	0.057	0.005
ApoA-I (g/L)	1.35 ± 0.32	1.40 ± 0.31	1.44 ± 0.33	0.127	<0.001
ApoB (g/L)	1.04 ± 0.35	1.04 ± 0.34	1.03 ± 0.33	0.034	0.099
CVD (n, %)	46 (6.3)	38 (5.2)	18 (2.4)		0.002
MetS (n, %)	161 (21.9)	140 (19.0)	117 (15.9)		0.013
T2DM (n, %)	29 (3.9)	21 (2.9)	15 (2.0)		0.096
eGFR (ml/min/1.73m ²)	95.1 (83.2-105.9)	97.3 (86.3-107.8)	98.7 (87.5-109.6)	0.020	0.23
UAE (mg/24 hrs)	9.0 (6.1-17.2)	9.0 (6.2-16.9)	8.6 (6.1-15.7)	0.028	0.18
TSH (mU/L)	1.28 (0.94-1.80)	1.29 (0.95-1.81)	1.39 (0.99-1.87)	0.045	0.036
free T ₄ (pmol/L)	12.97 ± 1.83	12.93 ± 1.71	12.80 ± 12.8	-0.042	0.050
free T ₃ (pmol/L)	3.75 ± 0.64	3.72 ± 0.61	3.73 ± 0.62	-0.027	0.20
				1	

Data in mean \pm SD or in median (interquartile range). For continuous variables *P*-values for linear trend are adjusted for age and sex, except for age which was adjusted for sex only. Data with respect to smoking and alcohol consumption are missing in 10 (0.5%) and 11 (0.5%) of the subjects, respectively. Triglycerides, UAE and TSH are log_e transformed. For dichotomous variables *P*-values are by multinomial χ -square test. Apo, apolipoprotein; BMI, body mass index; CVD, cardiovascular disease; eGFR, estimated glomerular filtration rate, HDL, high density lipoproteins; MetS, metabolic syndrome; T2DM, type 2 diabetes mellitus; UAE, urinary albumin excretion. β : standardized regression coefficient.

Table 2. Multiple linear regression models demonstrating the independent association of free T_4 and TSH with paraoxonase-1 (PON-1) activity.

	Model 1		Model 2 M		Мос	lel 3	Model 4	
	β	P-value	β	P-value	β	P-value	β	P-value
Age (years)	-0.141	<0.001	-0.183	<0.001	-0.160	<0.001	-0.131	<0.001
Sex (men vs. women)	-0.050	0.018	-0.009	0.69	-0.005	0.85	0.029	0.276
free T ₄ (pmol/L)	-0.064	0.003	-0.067	<0.001	-0.066	0.002	-0.0642	0.002
TSH (mU/L)	0.051	0.016	0.035	0.091	0.034	0.11	0.033	0.124
non-HDL cholesterol (mmol/L)			0.095	<0.001	0.089	0.001	0.097	<0.001
HDL cholesterol (mmol/L)			0.242	< 0.001	0.230	<0.001	0.220	<0.001
Triglycerides (mmol/L)			0.116	< 0.001	0.121	<0.001	0.103	<0.001
Systolic blood pressure (mm Hg)					-0.010	0.78	-0.005	0.878
Diastolic blood pressure (mm Hg)					0.013	0.70	0.006	0.870
eGFR (ml/min/1.73m ²)					0.029	0.28	0.030	0.271
UAE (mg/24 hrs)					0.033	0.15	0.028	0.226
CVD history (yes/no)					-0.021	0.34	-0.016	0.496

Diabetes status (yes/no)			-0.031	0.15	0.039	0.222
Alcohol consumption			0.025	0.26	0.029	0.190
$(< vs. \ge 10 \text{ gram per day})$						
Smoking (never, former,			-0.049	0.024	-0.050	0.023
current)						
Glucose lowering drugs					-0.089	0.004
Lipid lowering drugs					0.007	0.766
Antihypertensives					-0.018	0.469
Oral contraceptives					0.092	<0.001

β: standardized regression coefficient. eGFR, estimated glomerular filtration rate; HDL, high density lipoproteins; UAE, urinary albumin excretion. PON-1 activity, TSH, triglycerides and UAE are log_e transformed. Alcohol consumption is categorized in per day. Smoking is categorized in never, former and current. Variables included in the models:

Model 1:age, sex, free T₄, TSH

Model 2: model 1 plus non-HDL cholesterol, HDL cholesterol and triglycerides

Model 3: model 2 plus systolic and diastolic pressure, alcohol consumption, smoking, plus

cardiovascular disease (CVD) history and diabetes status.

Model 4: model 3 plus glucose lowering drugs, lipid modifying medication, antihypertensives and oral contraceptives.

Table 3. Multivariable linear regression analyses demonstrating relationships of paraoxonase-1 (PON-1) activity with after exclusion of subjects with a history of cardiovascular disease and type 2 diabetes mellitus (n=2051; model 1) or oral glucose lowering drugs, lipid lowering drugs, oral contraceptives and antihypertensives (n=1596; model 2).

	Model 1		Model 2	
	ρ	D vialua	ρ	D vialua
	р	<i>P</i> -value	р	<i>P</i> -value
Age	-0.161	<0.001	-0.140	<0.001
Sex (men vs women)	-0.011	0.67	-0.026	0.37
TSH	0.028	0.21	0.047	0.06
free T ₄	-0.047	0.035	-0.049	0.049

β: standardized regression coefficient. All models are adjusted for age, sex, non-high density lipoprotein (HDL) cholesterol, HDL cholesterol, triglycerides, systolic blood pressure, diastolic blood pressure, alcohol consumption and smoking. PON-1 activity, TSH and triglycerides are log_e transformed.







This article is protected by copyright. All rights reserved.