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# Iron metabolism as a therapeutic target for drug development and delivery

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**Abstract**

Iron fulfils a central role in many essential biochemical processes in human physiology, which makes proper processing of iron crucial. Although iron metabolism is subject to relatively strict physiological control, in recent years numerous disorders, such as cancer and neurodegenerative diseases, have been linked to deregulated iron homeostasis. Because of its involvement in the pathogenesis of these diseases, iron metabolism constitutes a promising and largely unexploited therapeutic target for the development of new pharmacological treatments. Several iron metabolism-targeted therapies are already under clinical evaluation for haematological disorders, and these and newly developed therapeutic agents will likely have substantial benefit in the clinical management of iron metabolism-associated diseases, for which few efficacious treatments are often available.

Iron is, after aluminium, the second most abundant metal on earth, and is an essential element for all forms of life on earth. Numerous enzymes involved in DNA replication, repair and translation rely on iron, often in the form of iron-sulphur (Fe-S) clusters, for proper functioning in animals, plants and fungi, as well as in organisms from the two prokaryotic domains of life, Bacteria and Archea<sup>1</sup>. The biological activity of iron lies, to a large extent, in its efficient electron transferring properties, enabling it to accept or donate electrons while switching between its ferrous bivalent (Fe(II), Fe<sup>2+</sup>), ferric trivalent (Fe(III), Fe<sup>3+</sup>) and its ferryl tetravalent (Fe(IV), Fe<sup>4+</sup>) states, thereby functioning as a catalysing cofactor in various biochemical reactions<sup>2</sup>. In vertebrates, the second main role of iron involves the oxygen-binding characteristic of porphyrin-complexed iron, better known as haem, which is crucial for the oxygen-carrying capacity of haemoglobin and myoglobin.

Taking into account these vital functions of iron in human physiology, it is clear that systemic or cellular disorders in iron metabolism may have serious consequences. At the systemic level, haem incorporated in haemoglobin (Hb) and myoglobin accounts for more than half of the approximately 4 grams of iron present in the human body, and by far the largest share of the total iron turnover is for haem production<sup>3</sup>. Consequently, an insufficient iron supply, unmet demand for iron, or substantial loss of iron will lead to a shortage of Hb, resulting in iron-deficiency anaemia<sup>4</sup>. Conversely, patients with red blood cell disorders such as  $\beta$ -thalassemia suffer from anaemia that is associated with malformed red blood cells that have a reduced life span due to dysfunctional  $\beta$ -globin expression and reduced Hb production<sup>5</sup>. In an attempt to compensate the chronic anaemia, these individuals produce large numbers of erythroid progenitors. This high erythroid activity is accompanied by a greatly increased iron demand, which promotes iron absorption and, in turn, causes serious comorbidity resulting from iron overloading.

At the cellular level, the presence of intracellular iron has a strong impact on the cellular redox status, contributing to oxidative stress in individual cells. Reactive oxygen species (ROS), such as superoxide (O<sub>2</sub><sup>-</sup>) and hydrogen peroxide (H<sub>2</sub>O<sub>2</sub>), which are formed by a single and double univalent reduction of molecular

oxygen ( $O_2$ ), respectively, are known to catalyse specific cellular redox reactions and are therefore involved in a number of signalling pathways. However, further reduction of relatively harmless  $H_2O_2$  results in the formation of hydroxyl radicals ( $OH\bullet$ ) that are highly reactive, causing nonspecific oxidation and damage to nucleic acids, lipids and proteins<sup>6</sup>. Iron, as well as other metals, catalyses the formation of  $OH\bullet$  from other ROS by Fenton chemistry<sup>7</sup>, which involves the oxidation of Fe(II) (to Fe(III)) and electron transfer to  $H_2O_2$ . The presence of superoxide further assists this process by promoting the reduction of Fe(III) to form Fe(II) (and  $O_2$ ) to complete the catalytic electron transport cycle of iron known as the Haber–Weiss reaction<sup>8</sup>.

As a consequence of its well-established roles in iron-deficiency anaemia and iron-loading anaemia, iron metabolism has historically remained within the scope of haematological pathologies. However, over the past decade, a range of ageing-related, non-haematological disorders has been associated with deregulated iron homeostasis as well. In this Review, we discuss iron metabolism as a target for the development of new therapeutics or drug delivery strategies in these diseases. We provide a systematic overview of the iron regulatory pathways and its key players, as well as the major pathophysiologies associated with dysfunctional iron homeostasis, and then review some of the most promising iron metabolism-targeted therapeutics thus developed, which could provide new therapeutic options for these often difficult to treat disorders.

## **Physiology of iron metabolism**

### *Systemic iron regulation – the hepcidin–ferroportin axis*

Hepcidin is a peptide comprising 25 amino acids that is encoded by the *HAMP* gene and named for its high expression in the liver<sup>9</sup>. Hepcidin was originally thought to be a peptide with moderate antimicrobial activity<sup>9,10</sup>, but it was soon recognized to be the master regulator of systemic iron metabolism<sup>11</sup>. Hepcidin regulates the systemic flux of iron by modulating the levels of ferroportin on the cell surface, the only known cellular exporter of unbound iron in vertebrates<sup>12</sup>. By directly binding to the extracellular domain of ferroportin, hepcidin induces endocytosis and degradation of the transmembrane protein, thereby preventing iron egress from the cell<sup>13</sup>. High levels of ferroportin are found in enterocytes in the duodenum (to transport absorbed iron), in hepatocytes (to transport stored iron), and in macrophages (to transport recycled iron), which together control systemic iron levels<sup>14–16</sup>. By reducing surface ferroportin, the expression of hepcidin limits the absorption, remobilization and recycling of iron, thereby reducing iron plasma levels (Figure 1).

With hepcidin as the central orchestrator, all major regulatory pathways of systemic iron homeostasis converge on *HAMP* transcription, mostly via SMAD signalling (Figure 2). As a result, alteration or genetic mutation of proteins that act within these pathways have been associated with a variety of disorders, including primary and secondary forms of iron overload, and chronic anaemia. Consequently, these proteins, together with ferroportin, are some of the most interesting therapeutic targets that could be exploited for a variety of disorders (Table 1).

### *Cellular iron regulation*

Cellular iron trafficking can be separated into three processes: intake, utilization and efflux (Figure 3, Table 2). There are four general pathways by which individual cells internalize iron. The most important pathway is centred around transferrin (Tf), an abundant plasma protein that acts a natural chelating agent with two high-affinity binding sites for ferric iron<sup>17</sup>. Tf fulfils several crucial physiological roles that include the solubilisation of relatively insoluble Fe(III), the prevention of iron-mediated redox toxicity, and the systemic trafficking of

iron. Although under normal circumstances less than 0.1% of total body iron is Tf-bound, the high turnover of Tf ensures that each day 20-30 mg of iron is delivered to the bone marrow for erythropoiesis. Cellular internalization of transferrin-bound iron typically occurs after docking with the membrane-bound transferrin receptor 1 (TfR1), which has nanomolar affinity for iron-bound Tf, but not for apotransferrin (apoTf) (i.e. iron-free Tf) at physiological pH<sup>18</sup>. The Tf-TfR1 complex is internalized via clathrin-mediated endocytosis, and Fe(III) is liberated from Tf as a result of a drop in pH to 5.5. Subsequently, TfR1 and apoTf, which remain associated at this low pH, are recycled back to the cell surface, where the pH is physiological and apoTf is thus released<sup>19</sup>. In addition to the TfR-mediated uptake of transferrin, cells may also utilize an alternative, low-affinity route for transferrin internalization<sup>20</sup>.

The second route by which cells can acquire iron is through the uptake of non-transferrin bound iron (NTBI), which can be regarded as 'free iron'. Currently, three cellular membrane transporters have been shown to be involved in the uptake of NTBI: divalent metal-ion transporter 1 (DMT1)<sup>21</sup>, Zrt- and Irt-like protein 14 (ZIP14)<sup>22</sup>, and, most recently, ZIP8<sup>23</sup>.

In 1997, DMT1 was identified as a key mammalian metal-ion transporter that is responsible for the intestinal iron absorption of various cationic bivalent metals, such as Fe(II), but also Cd(II), Co(II), Cu(II), Pb(II), Mn(II), Ni(II) and Zn(II)<sup>21</sup>.

Around the same time, the inheritable microcytic, hypochromic anaemia found in *mk/mk* mice and Belgrade (b) rats, which was already known to be associated with an impaired intestinal uptake and erythroid utilization of iron, was shown to originate from homozygous mutations in the gene encoding DMT1, *Slc11a2*<sup>24,25</sup>. The critical role of DMT1 in the absorption of dietary non-haem iron has recently been confirmed using intestinal DMT1 knockout mice<sup>26</sup>, and in humans, homozygous or compound heterozygous mutations in the *SLA11A2* gene have been associated with microcytic anaemia<sup>27</sup>. However, in contrast to the rodent models, patients with DMT1 mutations are overloaded with iron, rather than deficient. This effect is likely caused by a residual capacity for iron absorption, fulfilled either by the DMT1 mutant or by an alternative (haem) absorption pathway that is further stimulated by a regulatory feedback mechanism driven by the microcytic anaemia. The microcytic hypochromic anaemia in these

patients, and in the *mk/mk* mice and Belgrade (b) rats, is therefore primarily related to the other main role of DMT1: the transport of Fe(II) through the endosomal membrane. As demonstrated in *mk/mk* mice, DMT1 co-localizes with TfR1 in the endosome to handle the cellular import of iron that is liberated from the Tf-TfR1 complex. DMT1 dysfunction particularly affects the developing red blood cells, as they require a substantial supply of iron and are heavily dependent on Tf-TfR1-mediated import<sup>28</sup>.

More recently, ZIP14 has been shown to fulfil an important role in the tissue accumulation of iron by NTBI. In genetic models of iron overloading in mice, ZIP14 deficiency reduced iron content in the liver and pancreas and provided some level of protection against hereditary hemochromatosis<sup>29</sup>. Each of the iron transporters only transport soluble Fe(II), and therefore require enzymes with ferredoxin activity to reduce Fe(III) prior to internalization. Family members of cytochrome b561, the best known of which is Dcytb, which is expressed in the duodenum<sup>30</sup>, as well as STEAP, which is found in erythroid progenitor cells<sup>31</sup>, have demonstrated such ferredoxin activity<sup>32</sup>. Recently, prion proteins present on the surface of neuronal cells have been shown to possess iron-reducing capacity as well, thereby assisting cellular uptake of NTBI in the brain<sup>33,34</sup>.

The third mechanism for iron internalization is the uptake of porphyrin-bound iron in the form of Hb and haem. The scavenger receptor CD163 is almost exclusively expressed by macrophages and is specialized to take in Hb in complex with the glycoprotein haptoglobin<sup>35,36</sup>. Similarly, CD91 orchestrates the uptake of haem that is in complex with the glycoprotein hemopexin<sup>37</sup>, and although CD91 is considered a promiscuous receptor recognising other ligands, it is highly expressed by CD163-positive macrophages and may therefore be an essential route for the clearance of iron that was incorporated into Hb and other haem-containing proteins. HRG1<sup>38</sup> and FLVCR2<sup>39</sup> are also haem-importing proteins, and although their role in cellular iron trafficking needs to be further elucidated, it has been shown that HRG1 in particular is involved in the transport of haem across the lysosomal membrane upon internalization and degradation of erythrocytes and haemoglobin by macrophages<sup>40</sup>.



The fourth general pathway for iron internalization consists of the uptake of the iron-storage protein ferritin by receptors that include Scara5<sup>41</sup>, TfR1<sup>42</sup>, and a currently uncharacterized human ferritin receptor that could be related to TIM-2, a well-characterised ferritin receptor in mice<sup>43,44</sup>.

Once inside the cell, iron can be utilized and stored. Mitoferrin 1 and 2 are specialized transporters that move iron into the mitochondria, where it assists with cellular respiration and is used for the synthesis of Fe-S clusters and haem<sup>45</sup>. Intracellular iron can be 'stored' in two forms: stably incorporated into ferritin, an iron-sequestering protein that can contain up to 4,500 iron atoms<sup>46</sup>, or as active unbound species, known as the labile iron pool (LIP). Because an excess of LIP may cause oxidative stress-related toxicity, iron is generally stored as cytosolic ferritin, with the liver functioning as the primary organ for systemic iron storage. The LIP is largely responsible for the regulation of cellular iron homeostasis through the iron regulatory protein (IRP)/iron-responsive element system, which post-transcriptionally modulates the translation mRNAs of several proteins that are involved in cellular iron trafficking, including DMT1, TfR1, ferritin and Fpn<sup>47,48</sup>.

The export of iron from cells is almost exclusively managed by ferroportin (Table 1). As a result, hepcidin regulates systemic iron uptake and mobilization by inducing the degradation of ferroportin as discussed above. The multi-copper oxidases ceruloplasmin (in serum and astrocytes), hephaestin (in the intestine, placenta, heart, brain and pancreatic  $\beta$ -cells), and zyklopen (in placenta), assist the export of iron by ferroportin by oxidizing ferrous iron to ferric iron, thereby allowing it to be loaded onto transferrin<sup>49</sup>. There are, however, other possible mechanisms for cellular export of complexed iron. These include the secretion of iron-containing ferritin by hepatocytes<sup>50</sup>, macrophages and kidney proximal tubule cells<sup>51</sup>, which is likely mediated by autophagy of cytoplasmic ferritin and subsequent release via the secretory-lysosomal pathway<sup>51,52</sup>. Another pathway is the secretion of haem by Feline leukaemia virus, subgroup C, receptor 1b (FLVCR1b), which on the one hand may function as an 'emergency valve' to prevent haem overload, especially in erythroid cells, and on the other hand as a

potential route for macrophages to directly supply haem to developing erythroblasts<sup>53</sup>.

## **Iron and the pathophysiology of disease**

### *Iron overload, iron deficiency and anaemia of inflammation and chronic disease*

Iron overload is a major clinical challenge in management of hereditary hemochromatosis and haemoglobinopathy-related anaemia, also known as iron-loading anaemia. Primary iron overload, such as occurs in patients with hereditary hemochromatosis, is directly caused by disordered iron metabolism that is associated with genetic mutations in regulatory proteins. By contrast, secondary iron overload in patients with iron-loading anaemia is not intrinsically related to iron metabolism, but rather results from the anaemia that stimulates erythropoiesis and iron absorption. Moreover, the iron overload in severely anaemic patients is further aggravated by regular blood transfusions to that aim to improve haemoglobin levels. Tissues commonly affected by iron overload include the liver, spleen, heart, pancreas, and pituitary gland, and excess iron often eventually leads to complications such as liver failure, myocardial disease, diabetes mellitus and other metabolic disorders<sup>54</sup>.

Iron deficiency occurs when the systemic demand for iron surpasses its supply. As erythropoiesis is, in terms of iron requirements, the most important physiological process, iron deficiency will typically present as iron deficiency anaemia. Increasing or supplementing the dietary iron intake corrects iron deficiency in most cases, although in a rare inherited form of anaemia known as iron-refractory iron deficiency anaemia (IRIDA), mutations in *TMPRSS6* lead to inappropriately high hepcidin expression and restricted intestinal iron absorption, so intravenous iron administration is often required<sup>55</sup>. A high hepcidin level is also one of the hallmarks in anaemia of inflammation and chronic disease (AI/ACD). Under conditions of chronic inflammation, such as inflammatory diseases and cancer, increased expression of IL-6 and structurally related cytokines lead to activation of the IL-6 receptor pathway, resulting in STAT3-mediated hepcidin expression and, consequently, limited iron absorption and availability<sup>56</sup>. Additionally, activin B, a member of the TGF- $\beta$  superfamily that is expressed in the liver under inflammatory conditions, was shown to induce *HAMP* transcription via the SMAD1/5/8 signalling pathway<sup>57</sup>. Although the expression of hepcidin can be considered a useful defence mechanism against infections, as it limits the availability of iron to invading pathogens, the

presence of AI/ACD in patients with chronic diseases is a common and substantial clinical problem that can negatively affect the recovery and quality of life in these patients<sup>58</sup>.

#### *Iron and cellular redox status*

The mitochondrial generation of ATP is highly dependent on the formation and presence of ROS, and mitochondria are a major source of cellular ROS.

Mitochondrial iron homeostasis is therefore of crucial importance in the control of cellular redox and potential oxidative damage. For example, in Friedreich's ataxia, a genetic disorder affecting the expression of the mitochondrial iron-chaperone frataxin, cardiomyopathy is a major complication that has been associated to mitochondrial iron overload. Additionally, complications secondary to hereditary hemochromatosis, including liver and cardiac toxicity, have been related to ROS-mediated damage and mitochondrial pathology<sup>59</sup>.

Ferroptosis is a recently described form of controlled non-apoptotic cell death that heavily depends on iron and ROS<sup>60</sup>. Ferroptosis has been associated with a diverse range of pathologies including neurological disorders<sup>61,62</sup>, ischaemia-reperfusion injury<sup>63,64</sup>, and cancer<sup>65,66</sup>. Although the ferroptotic pathway and its function still have several unknown features, peroxidation of lipids by cytosolic ferrous iron plays a central role. Ferroptosis can be induced by blocking the system X<sub>c</sub><sup>-</sup> cystine/glutamate antiporter, which limits glutathione production and thus reduces oxidative protection by glutathione peroxidase 4<sup>67</sup>, while chelation of iron protects cells from ferroptosis<sup>60</sup>. The interplay between mitochondrial iron, ROS and ferroptosis has recently been reviewed in depth<sup>68-70</sup>, and will undoubtedly uncover key therapeutic targets that can be exploited to design novel therapies for diseases associated with abnormal ferroptotic activity.

#### *Iron and immunity*

The relationship between iron and the immune system is well established. As part of the innate immune system, macrophages are highly efficient phagocytes that fulfil a central role in the first-line defence against microorganisms. In parallel, they are responsible for iron recycling from red blood cells, and are thereby solely accountable for >95% of the iron supply needed for steady-state

erythropoiesis. In addition, they have an iron-independent growth-supporting function in erythroid development<sup>71,72</sup>. Although these macrophage activities appear to be quite dissimilar, a connection between iron status and macrophage phenotype has been firmly demonstrated. First of all, multiple reports have shown that high intracellular iron levels stimulate the secretion of inflammatory cytokines such as TNF- $\alpha$  by macrophages<sup>73,74</sup>; the clinical significance of this has been illustrated by several preclinical studies. For example, intravenous administration of ferumoxytol iron oxide nanoparticles (also known as Feraheme; AMAG Pharmaceuticals), an FDA-approved for iron replacement therapy in chronic kidney disease (see also Box 1), significantly reduced tumour growth and suppressed liver metastasis in tumour-bearing mice, and was associated with a proinflammatory phenotype in tumour-associated macrophages<sup>75</sup>. Similarly, iron-overloaded macrophages in chronic venous ulcers have a strong proinflammatory phenotype, producing of TNF- $\alpha$  and ROS, thereby delaying wound healing and supporting further tissue damage<sup>76</sup>. Finally, unbound haem and iron can induce a switch in the phenotype of macrophages from growth-supporting to inflammatory, which can be prevented by iron chelation or scavenging of haem by hemopexin<sup>77</sup>. Because various complications in patients with iron-loading anaemias have a chronic inflammatory component, such as liver fibrosis, leg ulcers, pulmonary hypertension and musculoskeletal inflammation, it is tempting to hypothesize that iron-mediated inflammatory activation of macrophages has a role in the pathogenesis of these complications<sup>5</sup>. Iron homeostasis influences adaptive immunity as well. For example, a form of immunodeficiency characterized by impaired development and function of lymphocytes has been attributed to mutations in the gene encoding TfR1, *TFRC*, which affects internalization of the surface receptor<sup>78</sup>. Although iron deficiency is the underlying cause of this disorder, illustrated by the reversal of defective lymphocyte proliferation by Fe(III) supplementation, interestingly, patients are only mildly anaemic.

Another connection between iron and immunity is related to the defence against invading pathogens. Microorganisms, as for all other forms of life, are highly dependent on the presence of iron to support growth and proliferation<sup>79</sup>. As a consequence, to limit infectivity of invading microorganisms, host organisms

have developed strategies to restrict iron supply to pathogens. NTBI is considered an important source of iron for microbes, which some of them sequester by releasing iron-binding proteins known as siderophores. Capture of these siderophores, as well as NTBI, by macrophage-produced lipocalin-2 is therefore an effective defensive response to infection<sup>80,81</sup>. Another host mechanism to limit NTBI availability under inflammatory conditions is to block cellular iron egress by reducing ferroportin expression<sup>82</sup>. This inflammation-induced reduction of ferroportin is mainly driven by upregulation of hepcidin transcription via IL-6R-activated JAK/STAT3-signalling (Figure 2, Table 1), and via TGF- $\beta$ 1–TGF- $\beta$ 1R-mediated activation of SMAD signalling, which is distinct from BMP6-induced SMAD signalling<sup>83</sup>. Alternatively, ferroportin reduction is also controlled by hepcidin-independent mechanisms, such as signalling via Toll-like receptors 2 and 6, allowing for a rapid systemic response under inflammatory conditions<sup>84</sup>.

### *Iron and cancer*

Changes in cellular redox status have a considerable role in malignant transformation and metastasis of cancer cells<sup>85</sup>, and consequently, many researchers have proposed and investigated the potential relationship between malignancies and iron<sup>86,87</sup>. Although several studies have indeed shown a correlation between the incidence and mortality of cancer and systemic iron levels, as measured by transferrin saturation or serum ferritin, this correlation is generally not strong, has shown conflicting outcomes, and it is often confounded by the presence of other pathology<sup>88,89</sup>. There is, however, clear evidence that deregulated iron homeostasis on a local — that is, microenvironmental and/or cellular — level is associated with tumour progression, as illustrated by changes in expression of iron-related genes in cancer cells (Figure 4), the levels of which are inversely correlated with patient survival<sup>90–92</sup>. In the context of tumour progression, one can envision that proliferating cancer cells, which require substantial amounts of iron to support their growth, will attempt to increase their iron intake, for example by increasing the expression of TfR1<sup>93–95</sup> and STEAP3<sup>96,97</sup>; to modulate iron storage capacity by producing ferritin<sup>94,96</sup>; and to limit their iron export by downregulating ferroportin<sup>91,98–100</sup>. However, the

additional intracellular oxidative stress that is accompanied by the increased presence of labile iron may also make them more sensitive to ferroptosis<sup>65,67</sup> and hinder tumour metastasis<sup>85</sup>, illustrating the complex relationship between iron, ROS and cancer cell survival. Furthermore, although a connection between iron and cancer growth has been demonstrated in several forms of cancers, with breast cancer<sup>90,91,99–101</sup>, prostate cancer<sup>98,102,103</sup> and glioblastoma as established examples<sup>94,104</sup>, it seems reasonable that many, but not all, cancer types will demonstrate a similar iron dependency.

### *Iron and neurodegenerative diseases*

Iron homeostasis in the central nervous system (CNS) differs from that in other tissues<sup>105</sup>. Endothelial cells of the blood–brain barrier modulate iron influx into the CNS by importing iron from the systemic circulation via TfR1 and DMT1, and subsequently releasing iron via ferroportin at the basal side into the brain interstitium, forming a control mechanism reminiscent of the absorption of iron in the duodenum<sup>106,107</sup>. The presence of an additional layer of homeostatic regulation suggests that the brain may be spared from excessive iron accumulation in iron-overloaded patients, which is supported by the fact that large iron deposits in the brain are not typically found in patients with hemochromatosis<sup>108</sup>. Nevertheless, genetic mutations associated with an inherited form of hemochromatosis are overrepresented in patients with Parkinson's disease (PD), parkinsonism and amyotrophic lateral sclerosis (ALS)<sup>109,110</sup>, indicating that cellular, rather than systemic, disturbances in brain iron homeostasis may have a role in the pathogenesis in these diseases.

Although progressive neurological disorders have been linked to disturbances in other metals such as copper (a hallmark of Wilson's Disease, but also found in Alzheimer's Disease (AD)<sup>111</sup>) and zinc (e.g. in PD<sup>112</sup>), many studies have implicated cellular iron overload and iron-induced oxidative stress in the development of PD, ALS, AD, Huntington's disease, multiple sclerosis (MS) and several other neurodegenerative diseases (Figure 4)<sup>113,114</sup>. As an interesting example, prion protein (PrP<sup>C</sup>) has been shown to promote the reduction of Fe(III) to Fe(II), enabling its uptake into cells by ZIP14 and DMT1<sup>33,34</sup>. However, the conversion of PrP<sup>C</sup> to its isoform PrP<sup>Sc</sup>, which is implicated in neurological

prion diseases such as Creutzfeldt–Jakob disease, is promoted by unbound iron<sup>115</sup>, and PrP<sup>Sc</sup> can bind to ferritin to form an insoluble complex, thereby stimulating the cellular intake of iron and promoting the formation of additional PrP<sup>Sc</sup><sup>116</sup>.

Astrocytes, glial cells in the CNS that have a function in the control of nutrient supply and brain remodelling, are thought to fulfil, in concert with endothelial cells of the blood–brain barrier, a critical role in regulating iron homeostasis in the brain<sup>106,117</sup>. Astrocytes produce hepcidin in response to inflammatory signalling and intracellular iron<sup>118</sup>, suggesting a control mechanism via ferroportin, similar to the regulation of systemic iron levels by hepatocytes. This regulatory feedback loop allows for local control over iron trafficking within the CNS, although the abundance of hepcidin in the brain interstitium suggests that at least some fraction of it is of systemic (likely hepatic) origin<sup>119</sup>. Low levels of ferroportin have been found in brains of patients with AD<sup>120</sup>, as well as in rat models of PD<sup>121</sup> and ALS<sup>122</sup>. In a mouse model of MS, reduced expression of ferroportin and intracellular iron overload were specifically found in macrophage-like microglia, but not in astrocytes<sup>123</sup>. Moreover, iron-overloaded microglia can be found in active MS lesions and have a pro-inflammatory phenotype that inhibits CNS repair, thus contributing to disease progression<sup>124,125</sup>. Neurological dysfunctions are also found in patients with aceruloplasminaemia: these individuals lack functional ceruloplasmin, a protein that facilitates iron removal from tissues by oxidising exported Fe(II) to Fe(III), thereby promoting its association with transferrin. This genetic disease is characterized by a loss of neurons, particularly in the basal ganglia, and by astrocytes with an attenuated iron efflux capacity and iron-overload<sup>126</sup>. Finally, in a clinical prospective study, higher ferritin levels in the cerebrospinal fluid were found to be correlated with a lower cognitive function and brain structural changes in patients with AD, but this correlation seemed to also be present in individuals that were considered cognitively normal<sup>127</sup>. Interestingly, genetic mutations that affect the ferritin light chain, which probably makes the iron-storage protein ‘leaky’, lead to a heterogeneous neurological clinical presentation, known as neuroferritinopathy<sup>114,128</sup>. This raises the question of whether dysfunctional ferritin or malfunctions in its cellular processing may



have some involvement in the pathogenesis of other neurodegenerative diseases<sup>129</sup>.

### *Iron and atherosclerosis*

In atherosclerotic plaques, iron is present in much higher concentrations than it is in healthy tissues (Figure 4)<sup>130</sup>. The function of these iron deposits in atherosclerosis, and whether iron accumulation has a causative role or is a mere consequence of the pathogenic process, for example due to haemolysis, is still under debate. It seems reasonable that iron could be directly involved by catalysing the formation of oxidized low-density lipoprotein (LDL) in the arterial wall<sup>131</sup>, which is considered one of the hallmarks of atherosclerosis. However, in iron-overloaded patients with type 1 hemochromatosis the incidence of atherosclerosis is actually lower, rather than higher, than it is in the rest of the population<sup>132</sup>. Interestingly, the iron status of macrophages, which fulfil a key role in the development and progression of atherosclerotic plaques<sup>133,134</sup>, could explain this paradox, as macrophages of patients with this form of hemochromatosis contain less iron than do those of healthy individuals. Conversely, patients with  $\beta$ -thalassemia, a disorder that is accompanied by iron-loading especially of macrophages, indeed do have a higher risk of developing atherosclerosis, confirming the so-called 'iron hypothesis' in atherosclerosis<sup>135,136</sup>. A relationship between macrophages, iron and atherosclerosis could partially be owing to iron's direct role in the oxidation of LDL by macrophages, as well as a phenotypic change in macrophages due to iron sequestration as described above. Nevertheless, various experimental studies that have investigated intracellular iron accumulation in macrophages as a driving mechanism for atherosclerosis have produced conflicting results, suggesting that iron takes part in a complex network involving several underlying mechanisms<sup>137-141</sup>.

### *Iron and other ageing-associated diseases*

Besides cancer, neurodegenerative disorders and atherosclerosis, various other pathologies have been linked to iron homeostasis. In diseases such as type 2 diabetes mellitus<sup>142</sup>, osteoporosis<sup>143</sup>, osteoarthritis<sup>144</sup>, macular degeneration<sup>145</sup>,

and liver fibrosis<sup>146</sup>, high levels of iron have been associated with disease severity, suggesting a possible role in pathogenesis or disease progression. Importantly, many of these disorders are considered chronic ageing-associated diseases of multifactorial aetiology, and are substantial causes of mortality and morbidity in the ageing Western society. This stresses the need for a thorough exploration of the role of iron dyshomeostasis in these diseases, as well the utility of targeting iron metabolism as a therapeutic approach or as a strategy to deliver pharmaceutical agents.

## **Iron metabolism as a therapeutic target**

### *Therapeutics targeting systemic iron availability*

The use of iron-chelating agents is a straightforward approach for limiting and redistributing systemic iron availability, and is therefore frequently used in the clinical management of primary or secondary iron overload. The currently available chelators are deferoxamine (Desferal, Novartis), deferiprone (Ferriprox, ApoPharma), and deferasirox (Exjade, Novartis). More than 50 years ago, desferoxamine (DFO) was the first chelator that showed clinical promise<sup>147</sup>. Iron chelation therapy was then rapidly demonstrated to be an effective strategy for mobilizing and removing iron via faecal and urinary excretion, illustrated by a marked decrease in mortality and iron-related complications in patients with thalassemia major<sup>148</sup>. Despite its success, DFO has poor bioavailability and requires parenteral administration that is often painful, and which should be executed slowly (up to 10h infusion) and regularly (5-7 times a week), making patient compliance a major limitation. In the following decades, the oral iron chelators deferiprone (DFP)<sup>149</sup> and deferasirox (DFX)<sup>150</sup> were developed and approved for the treatment of iron overload, improving iron chelation therapy patient satisfaction as compared to treatment with DFO<sup>151</sup>. Both DFO and DFX are currently recommended as first-line chelation therapy in iron overload, but the toxicity profile of these compounds, which include hypersensitivity reactions, liver dysfunction, renal dysfunction and neuronal hearing loss<sup>152</sup>, warrant further developments in iron chelation therapies. For example, in preclinical models of iron-loading anaemia, the administration of unmodified Tf, which can be considered the physiological iron chelator, has effectively redistributed iron to developing erythroid cells<sup>153</sup>.

In addition to iron overload disorders, iron chelation has also been extensively investigated as an adjuvant therapy in other disorders. In the 1990s, clinical trials of iron chelators in patients with neuroblastoma showed antitumor efficacy<sup>154,155</sup>, which encouraged further exploration of iron chelation therapy in cancer. Several novel chelating agents have been explored, of which one is currently under clinical development (DpC, Oncochel Therapeutics)<sup>156</sup>. In parallel, iron chelation has also shown clinical potential in neurodegenerative disorders, including multiple sclerosis<sup>157</sup>, Alzheimer's<sup>158</sup> and Parkinson's

disease<sup>159</sup>, thereby confirming the involvement of iron in the pathogenesis of these disorders.

Since the discovery of hepcidin in the beginning of this millennium, a substantial number of agents stimulating or blocking its expression or activity have been investigated (Table 3). Several short hepcidin derivatives have been designed as hepcidin-mimicking agents with an improved pharmacokinetic profiles, and are currently under development by Merganser Biotech (M012, phase 1)<sup>160</sup>, La Jolla Pharmaceutical Company (LJPC-401, phase 1, see Further information), and Protagonist Therapeutics (PTG-300, preclinical, see Further information).

Interestingly, preclinical studies have not only demonstrated beneficial activity of hepcidin derivatives in several models for red blood cell and iron overload disorders, such as  $\beta$ -thalassemia, polycythaemia vera, and hemochromatosis<sup>161,162</sup>, but also as an antimicrobial agent, as it limits the availability of iron to certain types of microorganisms<sup>82</sup>. Conversely, therapeutics that bind to and inactivate hepcidin have been developed and studied for the treatment of anaemia of inflammation and chronic disease. Such hepcidin-binding agents include humanized monoclonal antibodies (LY2787106, discontinued after phase 1 in 2015, Eli Lilly<sup>163</sup>; and 12B9m, Amgen Inc<sup>164</sup>), an anticalin (PRS-080, Pieris Pharmaceutical)<sup>165</sup> and a L-RNA spiegelmer (Lexaptepid pegol (NOX-H94), NOXXON Pharma)<sup>166,167</sup>.

A similar ferroportin-reducing effect can be achieved with agents that target ferroportin directly, blocking the binding of hepcidin and preventing internalization and degradation of the iron exporter protein. The structure of ferroportin has not yet been resolved, but the solved crystal structure of a bacterial homologue and modelling of human ferroportin strongly suggest that the transporter comprises 12 transmembrane domains with a central pore<sup>168,169</sup>, sharing structural features with the major facilitator superfamily (MFS) of transporters. The cys326 residue that is essential for the binding of hepcidin is likely located inside the central pore, which could indicate that binding of hepcidin could structurally block ferroportin function before internalization has taken place<sup>169,170</sup>. Fursultiamine, a small molecule also known as thiamine tetrahydrofurfuryl disulfide that targets the cys326 residue of ferroportin, indeed reduced hepcidin-mediated internalization of ferroportin and stimulated

cellular export of iron, however its function *in vivo* is limited due to rapid conversion of fursultiamine into thiamine<sup>171</sup>. An antibody against ferroportin (LY2928057, Eli Lilly), which has been explored clinically, has had more success *in vivo*<sup>172</sup>. However, the largest challenge of any strategy directly targeting hepcidin or its receptor, ferroportin, is an underlying homeostatic feedback mechanism that may cause a rebound in hepcidin expression, thereby likely abolishing a large part of any long-term therapeutic effect<sup>164</sup>. This feedback loop may explain why the development of LY2928057 seems to have been discontinued upon completion of a phase 2 trial in 2015 (NCT01991483). Due to the homeostatic control over hepcidin expression, approaches that interfere with the underlying regulatory pathways may therefore be more useful, although none has thus progressed past phase 2 of pharmaceutical development. Several strategies have inhibited hepcidin expression by modulating the BMP6/SMAD pathway. Capturing BMP6 to prevent its binding to the BMP6 receptor (BMP6R) has been investigated using an humanized antibody (LY3113593, Eli Lilly, lost through attrition in 2016 (see Further information, NCT02604160), a fusion protein of hemojuvelin (HJV) and the Fc domain of IgG (FMX-8, phase 2 trials have been terminated in 2016 due an inability to recruit patients meeting eligibility criteria, FerruMax Pharmaceuticals<sup>173</sup>), or non-anticoagulant heparin-derivatives that bind BMP6 in preclinical studies<sup>174</sup>. A second strategy aims to reduce BMP6-induced hepcidin transcription downstream of the BMP6–BMP6R interaction, by blocking SMAD phosphorylation using dorsomorphin<sup>175</sup> or its derivatives<sup>176</sup>, or by siRNA-induced degradation of mRNA encoding for Tfr2 (ALN-HPN, discontinued for unknown reasons, Alnylam Pharmaceuticals Inc)<sup>177</sup>. A hepcidin agonistic strategy comprises inhibition of matriptase 2 (MT-2)-induced cleavage of HJV. HJV is a coreceptor in the BMP6R complex that is required for iron sensing and hepcidin induction<sup>178</sup>, and MT-2 cleaves HJV, thereby suppressing BMP/SMAD-induced hepcidin expression<sup>179,180</sup>. MT-2 is a type II transmembrane serine protease that is structurally similar to matriptase, an endothelial enzyme associated with tumour development. Although a crystal structure of MT-2 is not yet available, modelling of the active domain has identified a number of residues that can be targeted, which allows the

development of small-molecular inhibitors against MT-2<sup>181,182</sup>. Serum-stabilized antisense DNA (IONIS-TMPRSS6-LRx, Ionis Pharmaceuticals<sup>183,184</sup>) or liposomal siRNA (ALN-TMP, Alnylam Pharmaceuticals Inc<sup>185</sup>), block MT-2 mRNA translation, and have shown preclinical efficacy in animal models of  $\beta$ -thalassemia and iron overload. In addition, several physiological protease inhibitors have demonstrated specific MT-2-inhibiting activity<sup>186</sup>, and some of them have a Kunitz domain that could be an important lead for further development of hepcidin-inducing peptides<sup>187</sup>.

The IL6-JAK1/2-STAT3 pathway is a second regulatory pathway that induces hepcidin expression. The isoflavone genistein promotes phosphorylation of STAT3, in addition to SMAD4, and thereby induces hepcidin expression in zebrafish, but its activity in a preclinical or clinical setting needs to be evaluated<sup>188</sup>. Conversely, inhibition of JAK by kinase inhibitors effectively reduces hepcidin expression *in vitro* (1,2,3,4,5,6-hexabromocyclohexane<sup>189</sup>) and *in vivo* (AG 490<sup>190</sup>), and similar effects have been obtained by disrupting STAT3 dimerization using a STAT3-binding peptide<sup>191</sup>.

#### *Therapeutics targeting cellular iron trafficking*

For many diseases disordered iron metabolism typically occurs only on a cellular level, whereas systemic iron homeostasis seems unaffected. Therapies that target cellular iron uptake, storage or efflux, may therefore be particularly promising. Several compounds target cellular uptake of iron, and DMT1 in particular has been a frequent target in preclinical studies. Structural modelling of human DMT1<sup>192,193</sup>, as well as the crystal structure of ScaDMT, a prokaryotic DMT1 homologue found in *Staphylococcus capitis*<sup>194</sup>, suggest that DMT1 is a 12 transmembrane protein of 2-fold symmetry and contains a hydrophobic core that coordinates the coupled transfer of a bivalent metal ion and a proton into the cytoplasm.

Several small molecule inhibitors of DMT1-mediated iron transport have been investigated. Ferristatin restricts cellular intake of free iron *in vitro*, but its efficacy *in vivo* has not yet been evaluated<sup>195</sup>. Although the inhibitory effect of ferristatin was at first attributed to degradation of TfR1<sup>196</sup>, subsequent work demonstrated that ferristatin directly inhibited activity of DMT1<sup>195</sup>. Similarly,

ferristatin II is a structurally unrelated compound that has shown potent TfR1-inhibiting properties, mediated via DMT1 internalization, *in vitro*, as well as *in vivo*<sup>197,198</sup>. Interestingly, ferristatin II also induces hepcidin expression via JAK/STAT3 signalling *in vivo*<sup>199</sup>. However, the metabolism of ferristatin II, also known as direct black 38 or chlorazol black, includes the formation of benzidine, which is classified by the WHO as Group 1 carcinogen (carcinogenic to humans)<sup>200</sup>, thereby limiting its translation for therapeutic purposes. The selenium-containing drug ebselen, which is known for its antioxidant activity that resembles glutathione peroxidase, strongly inhibits DMT1-mediated uptake of iron (IC<sub>50</sub> ≈ 0.2 μM)<sup>201</sup>, so could have therapeutic potential in treating iron-related oxidative stress in a range of disorders, as has been shown preclinically for iron-induced cardiomyopathy<sup>202</sup>. Comparably potent DMT1 inhibition was achieved with pyrrolidine dithiobarbamate (PDTC, IC<sub>50</sub> ≈ 1.5 μM) and Δ<sup>9</sup>-tetrahydrocannabinol (THC), which also have antioxidant activity<sup>201</sup>. Recent screening studies have identified a series of pyrazole derivatives and benzylisothiureas as potent inhibitors of DMT1 both *in vitro* (IC<sub>50</sub> < 1 μM) and *in vivo* (30-85% inhibition, 50 mg/kg, taken orally)<sup>203-205</sup>. A particularly interesting, but not yet explored strategy to limit iron overload may be the oral administration of a non-bioavailable DMT1 inhibitor, i.e. a compound that is not systemically absorbed and therefore likely has no systemic off-target effects, but would inhibit absorption of iron from the intestine. Although the capacity of the intestine to absorb haem-bound iron<sup>206</sup> and nanosized particulate iron (such as the ferritin core)<sup>207</sup>, which bypass the DMT1 absorption pathway via unknown mechanisms, clearly forms a potential limitation of this approach, combining intestinal DMT1 inhibition with dietary restrictions could be a relatively safe therapeutic option in the treatment of primary or secondary iron overload. The central role of ZIP14 in transport of NTBI into tissues that are typically affected by iron overload, especially the liver and pancreas<sup>29</sup>, makes the inhibition of ZIP14 an appealing strategy for the clinical management of iron overload. Similar to other members of the ZIP transporter family, ZIP14 is predicted to comprise eight transmembrane domains, which are all involved in the transport of bivalent metals<sup>208</sup>. Whether ZIP14 is a sufficiently druggable target still needs to be evaluated.

The hypoxia-inducible factor (HIF) transcription factors, which are composed of a heterodimerized  $\alpha$ - and  $\beta$ -subunit, may be another very attractive target for interfering with iron metabolism<sup>209</sup>. HIFs modulate, by binding to a hypoxia response element (HRE) in the promoter of a target gene, the expression of a wide range of proteins that are involved in cellular oxygen status, glucose metabolism, angiogenesis and erythropoiesis. In parallel, the promoters of several genes encoding key iron trafficking proteins, namely DMT1<sup>210,211</sup>, DCytB<sup>211</sup>, TfR1<sup>212</sup>, and FPN<sup>213</sup>, especially those with roles in intestinal cells, also contain such HREs and have been shown to be susceptible to HIF regulation. As a consequence, inhibitors of HIFs, for example those that bind within the C-terminal PAS domains of HIF-2 $\alpha$  to prevent HIF-2 heterodimerization<sup>214</sup>, may form an appealing strategy for limiting iron absorption and iron overload<sup>215</sup>. The oxygen-sensitive prolyl-4-hydroxylases (PHDs) are physiological intracellular inhibitors that regulate the post-translational expression of HIFs by promoting the hydroxylation of two proline residues in its  $\alpha$ -subunit, labelling it for recognition by the VHL-E3-ubiquitin ligase complex and subsequent degradation<sup>216,217</sup>. PHD2, which seems to be most dominant of the three PHDs with respect to HIF regulation, is a homotrimeric protein that contains a coordinated Fe(II) and a hydrophobic pocket at its active site, and requires 2-oxoglutarate (2-OG) as a co-substrate<sup>218</sup>. By using 2-OG analogues as competitive substrates to pharmacologically inhibit PHDs, it is possible to stabilize HIF expression, which is a very promising strategy for promoting erythropoiesis in patients with anaemia<sup>219</sup>. Although the direct effect of HIF stabilization on the expression of erythropoietin is a key characteristic of this strategy, PHD inhibition also improves duodenal iron adsorption and utilization, as illustrated by an improved mean corpuscular volume (MCV) and mean corpuscular haemoglobin (MCH) of erythrocytes in a rat model for anaemia of inflammation and chronic disease<sup>220</sup>. Moreover, increased HIF-1 expression by inhibition of PHD2 improved neuronal survival in models of PD, likely due to an increased expression of the lysosomal proton pump ATP13A2, which reduced the cytosolic iron pool by improving lysosomal iron storage conditions<sup>221</sup>. A substantial number of pharmaceutical companies, as well as one tobacco company, are actively pursuing the development of such inhibitors for the



treatment of anaemia in chronic kidney disease and end-stage renal disease. Promising candidates that have already entered phase 3 clinical trials include FG-4592 (Roxadustat, Fibrogen/Astellas Pharma/AstraZeneca<sup>222</sup>), GSK1278863 (Daprodustat, GlaxoSmithKline<sup>223</sup>), and AKB-6548 (Vadadustat, Akebia Therapeutics<sup>224</sup>), and several others are currently being evaluated in a phase 2 (BAY85-3934, Molidustat, Bayer Pharma AG<sup>225</sup>; JTZ-951, Japan Tobacco<sup>226</sup>), phase 1 (DS-1093, Daiichi Sankyo (discontinued in 2016)); ZYAN1, Zydus Cadila<sup>227</sup>), or preclinical (JNJ-42905343, Johnson & Johnson<sup>220</sup>) setting.

#### *Drug targeting strategies exploiting iron metabolism*

Iron and iron-based materials have been widely researched for drug targeting and diagnostic applications, and iron oxide nanoparticles have especially received attention due to their superparamagnetic properties that can be used for therapeutic and diagnostic purposes (Box 1). The metabolism of iron can be utilized for the delivery of therapeutics to specific sites in the body by targeting iron-related proteins that are upregulated in certain tissues and pathological processes (Figure 5). Targeting TfR1 using drugs and drug delivery systems functionalized with transferrin, antibodies, antibody-derivatives, or TfR-targeted peptides, is a commonly employed and arguably the best-known strategy, as illustrated by its usage as a prototypical system for mechanistic studies of active targeting to cancer cells<sup>228</sup>. An impressive amount of different TfR-targeted delivery systems have been designed and evaluated for cancer therapy by exploiting the increased expression of TfR in some tumours, and this strategy has shown preclinical benefit when using a low-dose regimen<sup>229</sup>. At least three TfR-targeted systems are currently under clinical development (Table 3)(MBP-426, Mebiopharm; SGT-53, SGT-94, SynerGene Therapeutics)<sup>230,231</sup>.

An elegant strategy that has been evaluated in preclinical cancer models is the application of transferrin-artemisinin conjugates as cytotoxic agents<sup>232</sup>. Artemisinin, discovered and developed by Tu Youyou, for which she was awarded the 2015 Nobel Prize in physiology and medicine, is a drug with antimalarial activity that is isolated from the herb *Artemisia annua* and contains an intramolecular peroxide bridge. Fe(II) is involved in the activation of artemisinin by cleavage of this endoperoxide bridge, resulting in the formation of

reactive carbon-centred radicals and subsequent cytotoxicity<sup>233</sup>. By using transferrin as a targeting moiety, both iron and the inactive artemisinin can be delivered to TfR-overexpressing cancer cells; upon internalization of the complex the released Fe(III) is oxidized to Fe(II), which in turn reduces and activates artemisinin into its cytotoxic form<sup>232</sup>. However, the effect of such artemisinin-constructs on other tissues with high TfR1 expression, notably the bone marrow, has not yet been reported. A second notable application of TfR-targeting therapeutics is the transport of drugs and drug delivery systems across the blood–brain barrier. The endothelial cells of the blood–brain barrier transcytose TfR-transferrin complexes to the brain parenchyma to fulfil the iron requirements of the CNS. Exploiting this mechanism, transferrin and antibodies that target TfR can be used as a Trojan horse to deliver drug carriers and macromolecules to the brain, but future evaluation should demonstrate its practical potential in a clinical setting<sup>234,235</sup>.

Ferritin is a commonly used biomolecule for drug targeting, as it can efficiently encapsulate large amounts of drugs because of its ‘nanocage’ structure<sup>236</sup>.

Ferritin can function as a ligand as well to target cells that highly express ferritin-internalizing receptors, such as Scara5 and TfR1<sup>237</sup>. Moreover, ferritin can be disassembled and reassembled by changing the pH to low or physiological, respectively, which facilitates drug loading and endows the protein with pH-triggered drug release properties, enabling discharge specifically upon uptake and lysosomal processing by the target cell<sup>238</sup>.

A relatively specific approach to target a subset of macrophages is the use of haemoglobin clearance mechanisms. The haemoglobin-scavenger receptor, CD163, is exclusively expressed by macrophages and monocytes that recycle iron<sup>35</sup>, and it can also be found in solid tumours<sup>239</sup> and inflamed tissues<sup>240</sup>, for example, as these tissues have increased iron requirements. Consequently, a liposomal targeting strategy exploiting tissue-specific expression of CD163 using an anti-CD163 antibody has demonstrated effective macrophage targeting *in vivo* as compared to untargeted nanocarriers<sup>241</sup>.

The value of haem as a targeting ligand, which uses differences in cellular affinity for haem to specifically target subsets of cells, has not yet been fully explored. As *in vitro* studies have shown that PLGA nanoparticles functionalized with

haematin, the oxidized Fe(III) form of haem, localize in tumour cells<sup>242</sup>, and haem-targeting improves the uptake of oligonucleotides in hepatocytes<sup>243</sup>, further evaluation of haem-targeting is certainly warranted.

#### *Merits and challenges of targeting iron metabolism as therapeutic strategy*

The majority of therapeutics that target iron metabolism has been developed in the context of haematological diseases and iron homeostasis disorders, such as iron deficiency and overload. However, considering the rapidly growing evidence implicating iron in a variety of other (often ageing-related) disorders, such as neurodegenerative diseases and atherosclerosis, further exploration of these therapeutics agents and their targets in other pathologies is certainly warranted. Although unbalanced iron trafficking should not be considered the sole cause, the chronic and inflammatory character of the underlying pathologic processes in iron metabolism-related diseases suggest that an iron-induced change in redox state within the affected tissues may form the missing link in the pathogenesis of several multifactorial disorders<sup>86,105,244</sup>. Importantly, apart from directly modulating the function of cells responsible for the condition, such as neurons and tumour cells, iron also impacts the inflammatory activity of local immune cells, especially macrophages<sup>134,245,246</sup> (See also Box 1). Consequently, interfering with iron metabolism may be a promising therapeutic strategy in these disorders for which there are currently few effective treatments available. A clear additional advantage of such an approach is that it typically addresses pathologic pathways that are complementary to those targeted by other therapeutic approaches, allowing for additive or possibly even synergistic effects when used in a combination strategy.

However, the broad involvement of iron metabolism in many, rather diverse physiological and pathological processes potentially makes its use as a therapeutic target a double-edged sword. Although iron-targeted therapeutic interventions may effectively change the iron status in diseased tissues to treat a disorder, they also have an inherent risk of perturbing systemic iron homeostasis that may lead to substantial side effects, such as erythropoietic

disorders. Using a targeted strategy that allows the delivery of therapeutics to specific tissues could limit such off-target activity.

In addition, most iron metabolism-related disorders are associated with cellular iron accumulation, which, due to the late onset of symptoms, is most likely a chronic process that takes several years to establish. Interfering with this process once iron-induced cellular damage has already occurred may not have a direct therapeutic effect, and prevention may be more efficacious in this context. Nevertheless, limiting further deterioration and modulating the inflammatory state within the affected tissue could certainly make iron metabolism-targeted therapies an effective adjuvant treatment<sup>124,247</sup>.

### **Conclusion and outlook**

Over the past decade, a multitude of iron metabolism-targeting agents have been developed and clinically evaluated in diseases with an obvious iron-related background, such as anaemia, iron overload and iron-loading anaemia. However, alterations in iron metabolism have demonstrated involvement in a wide range of disorders, such as cancer, neurodegeneration and atherosclerosis, but also type 2 diabetes, osteoporosis, osteoarthritis, retinal disorders and liver fibrosis. The chronic inflammatory and ageing-related character of many of these conditions suggests that a slowly changing intracellular iron level, causing a shift in cellular redox status that eventually may result in cell damage, is an underlying mechanism that could be exploited for the development of new therapeutic approaches. Further investigation is therefore required to explore current and newly developed therapies that target iron metabolism in a much broader context than haematological pathologies alone. Modulation of macrophage activity by targeting their role in iron recycling may be a particularly interesting approach. Similar to iron metabolism, the role of an altered macrophage phenotype has been increasingly implicated in various disorders<sup>248</sup>, and both concepts seem to be interconnected<sup>245</sup>. Targeted approaches — for example those using nanosized drug delivery systems that preferentially accumulate in phagocytic cells, to specifically interfere with iron trafficking in macrophages — would thereby enable therapeutic modulation of macrophage activity in cancer and chronic inflammatory disorders<sup>75,249,250</sup>. To

achieve this, the cellular mechanisms governing the relationship between macrophage phenotype and its iron load need to be further elucidated. For example, recent work has demonstrated that internalization of unbound haem induces an inflammatory phenotype in macrophages, whereas using haem in complex with hemopexin, which is scavenged by CD91, such a phenotypic switch was not observed<sup>77</sup>. This observation indicates that the pathway of internalization, rather than the absolute load of iron, has a dominant role in the influence of iron on the phenotype of macrophages, thereby providing valuable clues for the identification of new strategies for macrophage-modulating therapies.

Overall, owing to the substantial advances in our understanding of the role of iron in a range of diseases, it has become clear that targeting iron metabolism is a compelling new therapeutic strategy for these disorders. Although further research is still required to systematically explore the value of targeting the various iron-related proteins in individual pathologies, these efforts will almost certainly prove to be valuable in the quest for novel efficacious therapies for diseases associated with disturbed iron metabolism.

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## **Competing financial interests**

S.R. has restricted stocks in Merganser Biotech. S.R. is a consultant for Novartis Pharmaceuticals, Bayer Healthcare, Merganser Biotech and Keryx Pharmaceuticals. S.R. is a member of scientific advisory board of Merganser Biotech and Ionis Pharmaceuticals. T.L. is a member of scientific advisory board of Cristal Therapeutics B.V..

## **Further information**

Protagonist's pipeline. <http://www.protagonist-inc.com/pipeline-main.php>

La Jolla Pharmaceutical LJPC-401 Phase 1 Results and Development Update.

<http://lajollapharmaceutical.com/wp-content/uploads/2016/09/LJPC-401-Phase-1-Results-and-Development-Update-9.7.16.pdf>

Eli Lilly's clinical pipeline.

[https://www.lilly.com/\\_assets/SiteCollectionDocuments/Pipeline/Clinical-Development-Pipeline/index.html](https://www.lilly.com/_assets/SiteCollectionDocuments/Pipeline/Clinical-Development-Pipeline/index.html)

## **Glossary terms**

### *Microcytic, hypochromic anaemia*

Low systemic Hb levels associated with a reduced Hb concentration inside each red blood cell, making them small (microcytic) and pale (hypochromic). This is most commonly caused by iron deficiency, but is also found in patients with red blood cell disorders such as thalassemia.

### *Hemochromatosis*

Hemochromatosis is a family of hereditary diseases that are characterised by genetic mutations affecting an iron metabolism-regulating protein and resulting in excessive iron absorption. Since there is no physiological mechanism for iron excretion, this will eventually lead to iron overload.

### *Anticalin*

Technology developed by Pieris Pharmaceuticals that modifies lipocalins, which are natural extracellular proteins that bind and transport a variety of ligands, to make therapeutics that target a specific molecule.

### *Spiegelmer*

Technology developed by NOXXON Pharma that uses L-RNA aptamers as therapeutic agents that bind a specific molecule. L-RNA, the stereoisomeric form of physiological D-RNA, is not sensitive to nuclease activity and therefore has improved biological stability.

### *Kunitz domain*

A Kunitz domain, or Kunitz-type protease inhibitor domain, is the active domain of various protease inhibitors. Their relatively short peptide sequence of around 60 amino acids makes them an interesting lead for the development new biopharmaceutical protease inhibitors.

### Box 1 – Iron oxide nanoparticles

Over the past few years, an enormous variety of iron oxide nanoparticles have been developed for diagnostic and therapeutic applications, although few have reached the clinic and several have been withdrawn for various reasons<sup>251</sup>. Also known as superparamagnetic iron oxide nanoparticles (SPIONs), these sterically stabilised particles typically contain magnetite ( $\text{Fe}_3\text{O}_4$ ) and maghemite ( $\gamma\text{Fe}_2\text{O}_3$ ), and the resulting magnetic properties make them particularly interesting as magnetic resonance imaging-contrast agents. Due to their nanosized dimensions, iron oxide nanoparticles primarily localize in phagocytic cells, mostly macrophages<sup>252</sup>, and in the liver, spleen and lymphatic tissues, so iron oxide nanoparticles can be utilized for imaging these tissues. For example, Feraheme (ferumoxytol, AMAG Pharmaceuticals), which is clinically approved for iron replacement therapy in chronic kidney disease, has been investigated as marker for the presence of macrophages in solid tumours<sup>253</sup>, and has recently been shown to modulate the activity of these macrophages towards a proinflammatory, anti-tumoural phenotype<sup>75</sup>. There are several additional promising applications of SPIONs, such as using their magnetic properties and localisation tumours to create local hyperthermia by applying an alternating magnetic field (NanoTherm, MagForce AG), or using them as drug carriers that release their payload as a result of magnetically induced heating<sup>254</sup>. While SPIONs are generally considered non-toxic, it is important to realise that there is no physiological route of iron excretion from the human body, other than a passive intestinal loss that is typically limited to 2 mg iron per day. Therefore, intravenously administered iron oxide, which would be in the range of 400 mg iron (Feraheme, NCT01336803), for example, will add to the total iron pool and may, in parallel, cause an proinflammatory phenotypic switch in the macrophages clearing these nanoparticles<sup>75</sup>. Further evaluation regarding the long-term tolerability of SPIONs as routinely used imaging agents may be therefore warranted.



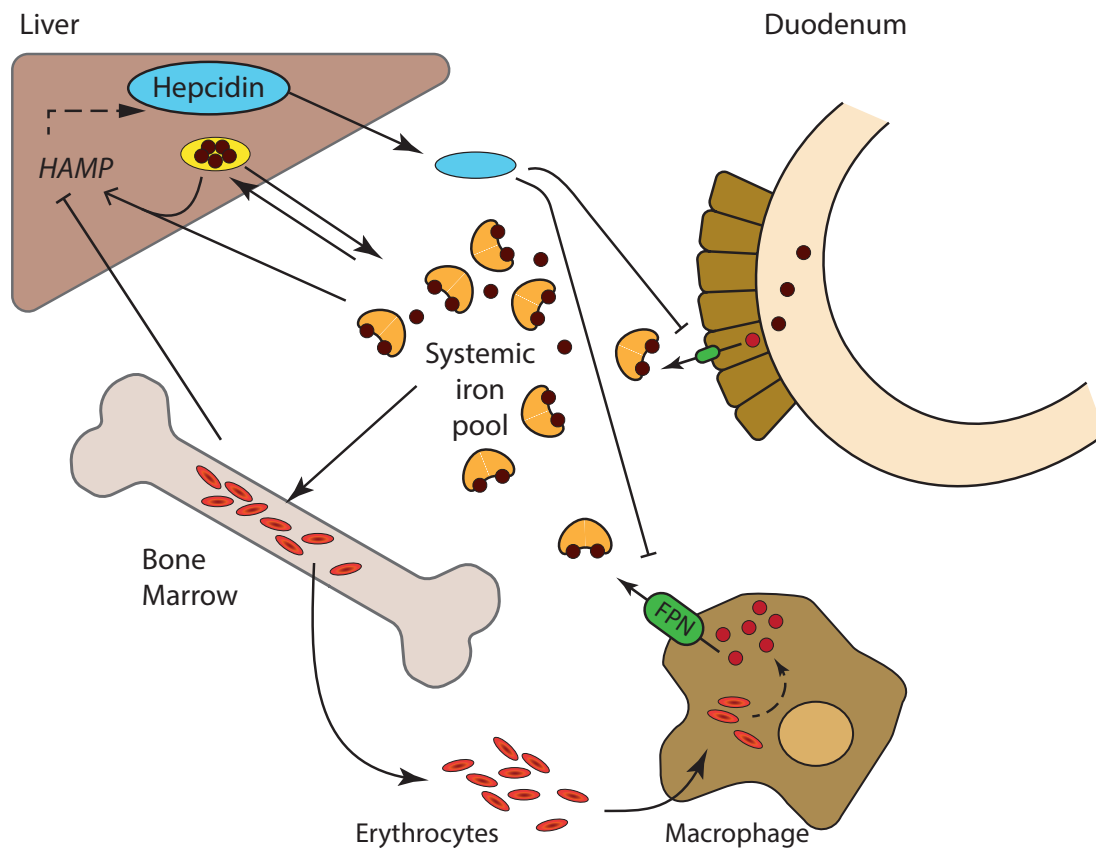


Figure 1 – **Systemic iron metabolism.** Dietary iron is absorbed by duodenal enterocytes and added to the systemic iron pool upon export via ferroportin (FPN, green). The systemic iron pool, which under normal steady-state conditions is mainly bound to transferrin (orange), is primarily utilized for erythropoiesis. Excess systemic iron is stored as ferritin (yellow) in the liver, and to a lesser extent in other tissues. The iron utilized in senescent erythrocytes is recycled by macrophages and released, via ferroportin, back into the systemic iron pool. Ferroportin expression, and consequently systemic iron availability, is inhibited by hepcidin (blue), a peptide primarily produced by the liver. In turn, transcription of the gene encoding hepcidin, *HAMP*, is stimulated by the presence of iron and inhibited by erythropoietic demand.

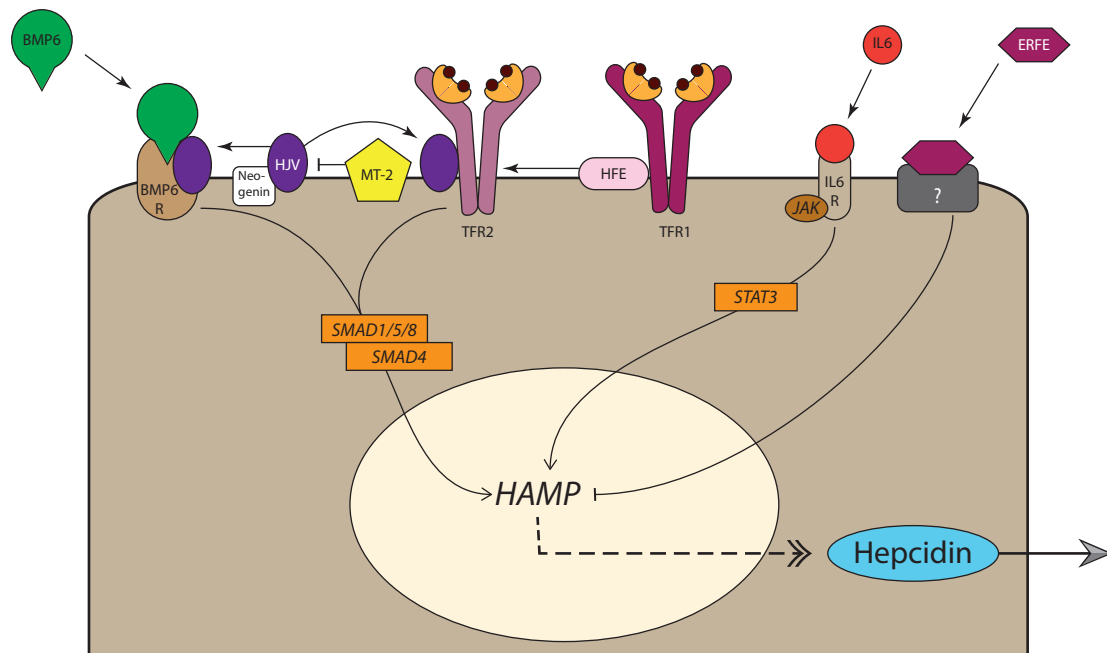


Figure 2 – **Regulation of hepcidin expression in hepatocytes.** Bone morphogenetic protein 6 (BMP6) is produced by non-parenchymal cells in the liver in response to iron stores and flux. Binding of BMP6 to the BMP receptor complex on hepatocytes, in conjunction with hemojuvelin (HJV) results in downstream signalling via SMAD1/5/8 and SMAD4, which in turn stimulates *HAMP* transcription and expression of hepcidin. HJV is stabilized by neogenin, and cleaved by matriptase 2 (MT-2), allowing for fine-tuning of BMP6 signalling. SMAD signalling is additionally regulated by transferrin receptor 2 (TfR2), upon association with human hemochromatosis protein (HFE) and HJV. HFE also interacts with TfR1, where it competes with holotransferrin. Consequently, when iron supply is high, HFE is displaced from TfR1 allowing it to associate with TfR2, thereby increasing hepcidin expression via SMAD signalling. Under inflammatory conditions, interleukin 6 (IL-6) and related cytokines bind IL-6 receptor, resulting in JAK1/2-STAT3 activation and hepcidin expression to restrict iron availability. Finally, developing erythroid cells confer their iron requirements via secretion of erythroferrone (ERFE), which inhibits hepcidin expression, via a currently unknown receptor and signalling pathway, to stimulate iron uptake under circumstances of high erythropoietic demand.

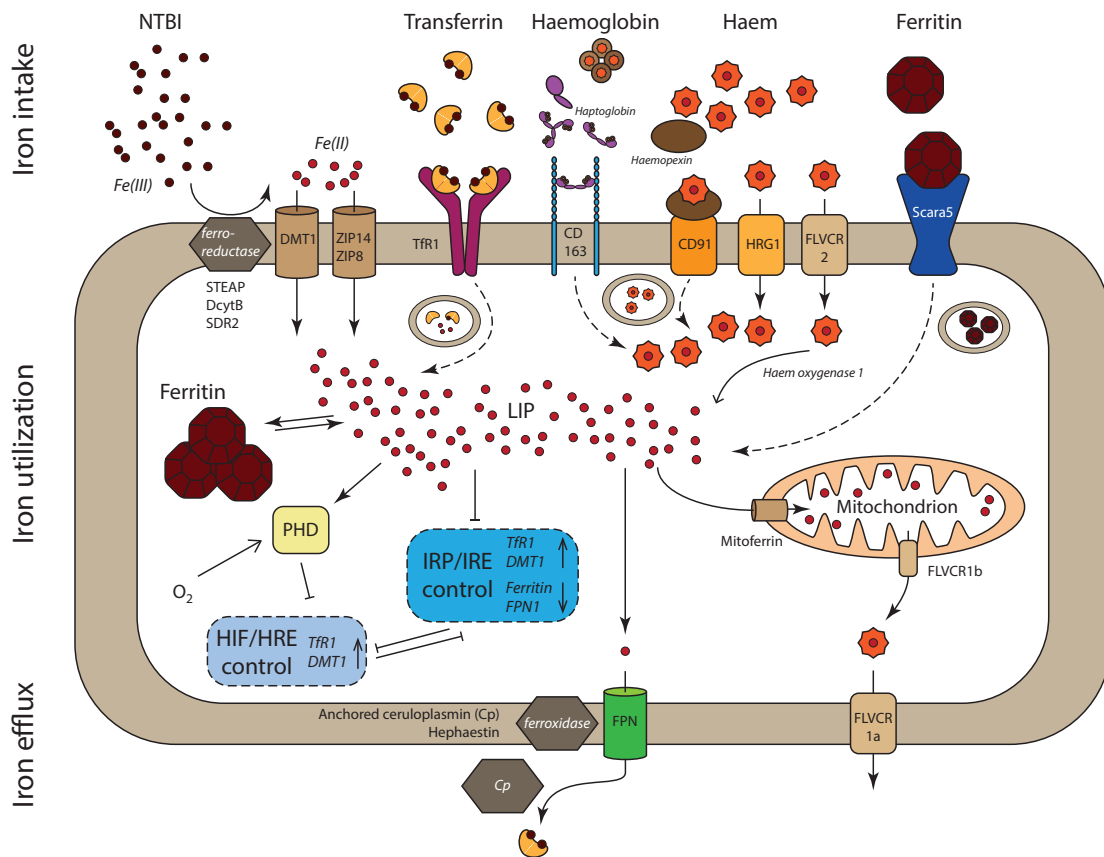


Figure 3 – **Cellular iron trafficking pathways.** *Iron intake.* Non-transferrin bound iron (NTBI) can be transported directly (solid arrow) into the cell by DMT1, ZIP14 and ZIP8, after reduction of Fe(III) to Fe(II) by ferrireductases STEAP, DcytB or SDR2. Transferrin-bound iron is taken up via binding to Tfr1 and endocytosis (dashed arrow) of the receptor, release and reduction of Fe(III) and transport into the cytoplasm via DMT1. Haemoglobin associates with haptoglobin to allows endocytosis by CD163, upon which haemoglobin is degraded, and haem transported into the cytosol. Systemic haem is scavenged by complexation with haemopexin and endocytosis via CD91. Haem can be directly transported, also from the endosome, into the cytosol by HRG1 and FLVCR2, where it is processed by haem oxygenase 1 and iron released as Fe(II). Scara5 internalizes ferritin that consists of light chains, after which iron is liberated and transported to the cytosol. Ferritin composed of heavy chains can be endocytosed by Tfr1 (not depicted here).

*Iron utilization.* The labile iron pool (LIP) comprises cytosolic Fe(II), which can be stored in cytoplasmic ferritin or utilized for biochemical processes. Most of these processes take place in the mitochondria, for which iron is supplied via

mitoferrin, while FLVCR1b allows export of mitochondrially produced haem. The LIP controls the expression of TfR1, DMT1, ferritin and ferroportin via the posttranscriptional IRP/IRE regulatory system. Similarly, the transcriptional HIF/HRE regulatory system controls several intracellular processes, including the transcription of TfR1 and DMT1. HIF2 $\alpha$  translation is inhibited by IRP1, while, conversely, IRP1 transcription is inhibited by HIF. HIF expression is regulated by PHD, which stimulates the degradation of HIFs under conditions of high oxygen.

*Iron egress.* Fe(II) is exported by ferroportin, followed by oxidation to Fe(III) by the ferroxidases hephaestin or ceruloplasmin, and subsequent binding to transferrin. Intracellular haem can be directly exported via FLVCR1a.

*Not all pathways are present in all cells.*

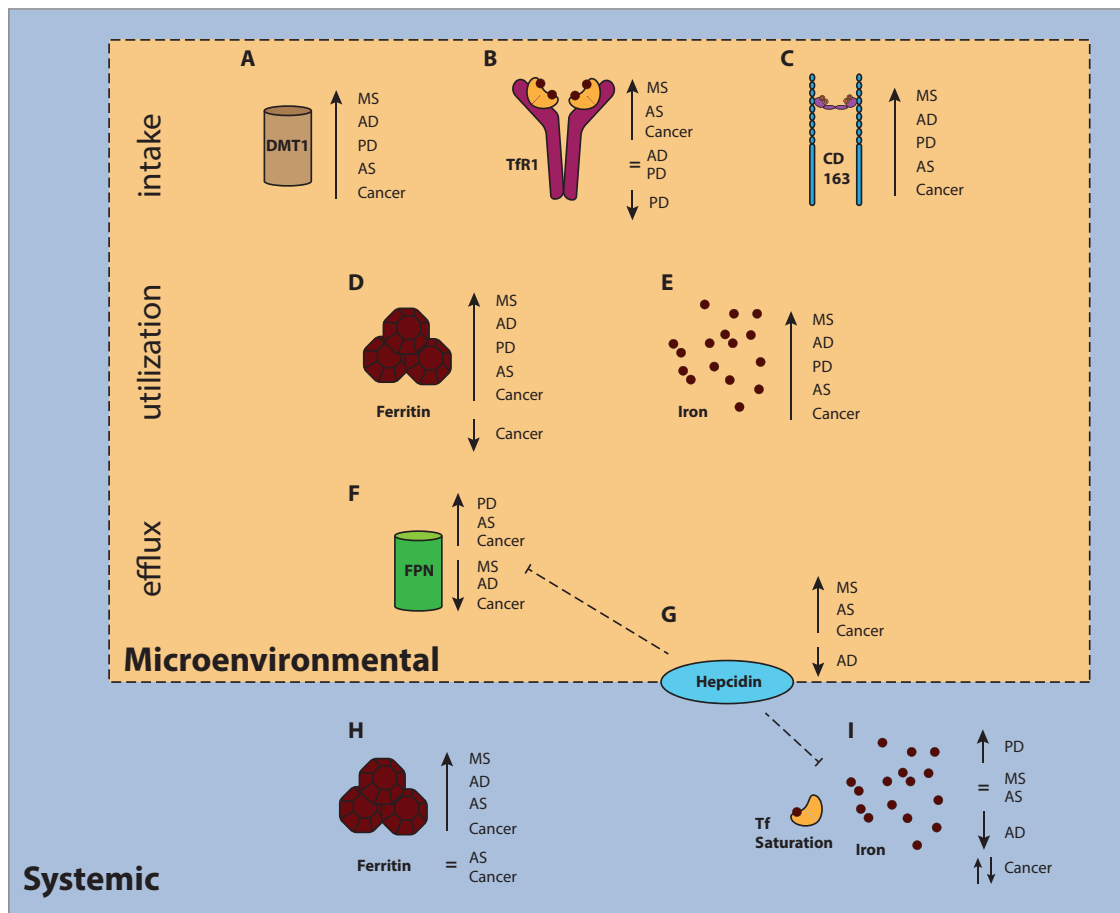
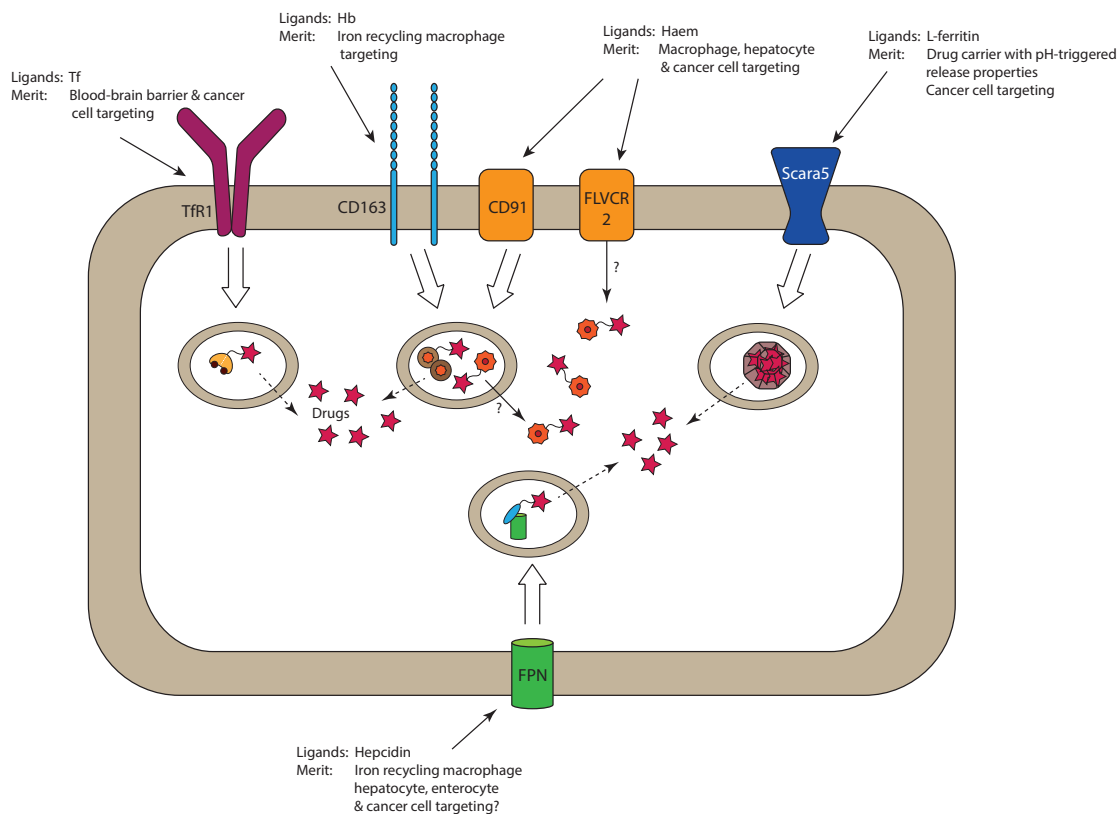


Figure 4 – **Up- and downregulation of several key players in non-haematological pathologies.** Schematic overview illustrating the microenvironmental and systemic changes in iron and iron metabolism-related proteins in selected pathologies in patients: multiple sclerosis (MS, sclerotic plaques), Alzheimer’s disease (AD, frontal cerebral cortex), Parkinson’s disease (PD, basal ganglia), atherosclerosis (AS, sclerotic plaques), and cancer (tumor tissue).

**A.** Upregulation of DMT1 has been observed in MS (preclinical)<sup>123</sup>, AD (preclinical)<sup>255</sup>, PD<sup>256</sup>, and cancer (colorectal)<sup>257</sup>. **B.** TfR1 upregulation has been found in MS<sup>258</sup>, atherosclerosis<sup>259</sup>, and cancer (colorectal<sup>257</sup>, breast<sup>95,101</sup>, glioblastoma<sup>94</sup>; normal TfR1 levels have been observed in AD<sup>260,261</sup>, while for PD normal levels<sup>262</sup>, and low levels have been reported<sup>263,264</sup>. **C.** Increased levels of the macrophage-associated scavenger receptor CD163 have been found in affected tissues of patients with MS<sup>265</sup>, AD<sup>266</sup>, PD<sup>266</sup>, AS<sup>267</sup>, and cancer (breast<sup>268</sup>, prostate<sup>269</sup>, glioblastoma<sup>270</sup>. **D.** Tissue iron stored as ferritin has found to be increased in MS<sup>271</sup>, AD<sup>272</sup>, PD<sup>264</sup>, AS<sup>259,273</sup>, and for cancer both high levels

(glioblastoma)<sup>94</sup> and low levels have been reported (breast)<sup>101</sup>. **E.** Similarly, microenvironmental iron deposits have been observed in MS<sup>271</sup>, AD<sup>271,272,274</sup>, PD<sup>256,264,274</sup>, AS<sup>273</sup>, and cancer (colorectal)<sup>257</sup>. **F.** Upregulation of FPN has been demonstrated in tissues of patients with PD<sup>264</sup>, AS<sup>273</sup>, and cancer (colorectal<sup>257</sup>, breast<sup>101</sup>), while low levels of FPN have been observed in MS (preclinical)<sup>123</sup>, AD<sup>120</sup> and also in breast<sup>91</sup> and prostate<sup>98</sup> cancer. **G.** Hepcidin levels are not in all diseases consistent with (an inversed) FPN expression. High hepcidin concentrations have been found in MS (preclinical, tissue)<sup>123</sup>, AS (serum)<sup>275</sup>, breast cancer (tissue<sup>91,101</sup> and serum<sup>276</sup>) and prostate cancer (tissue)<sup>98</sup>, whereas a low level has been observed in the tissues of patients with AD<sup>120</sup>. **H.** Increased systemic ferritin concentrations have found in the cerebrospinal fluid (CSF) of patients with MS<sup>277</sup>, AD<sup>127</sup>, and glioblastoma<sup>278</sup>, as well as in the serum of patients with AS<sup>279</sup> and breast cancer<sup>276</sup>, although the levels of serum ferritin have also been found to be normal in patients with AS<sup>275</sup> and non-skin cancers<sup>280</sup>. **I.** Systemic iron levels measured by iron concentration and transferrin saturation have been quite inconsistent: higher than average levels have been found in a meta analysis in PD<sup>281</sup>, normal levels have been found in MS<sup>282</sup> and AS<sup>275</sup>, and low levels have been observed in AD<sup>283,284</sup>. With regard to cancer, the association with systemic iron levels is quite unclear: women with high levels of serum iron were at higher risk for developing non-skin cancer, while men were at lower risk<sup>280</sup>.



**Figure 5 – Proposed mechanisms for targeted drug delivery exploiting iron metabolism-associated cellular targets.** The natural ligand and the merits of its use as a delivery strategy are presented for each target. TfR1, CD163 and Scara5 allow endocytosis of drugs associated with their natural ligands. Subsequent lysosomal processing would then liberate the targeted drug, which then enters the cytosol by passive diffusion from the lysosome. This likely also occurs for haem-targeted constructs upon internalization by CD91, but possibly direct transport of the haem-linked drug into the cytosol by HRG1 and/or FLVCR2 could represent an alternative mechanism. Drug targeting to FPN, for example by using hepcidin, has not been explored until now, but forms an interesting approach for directing therapeutics to cell that overexpress FPN, such as some types of tumour cells. Tf, transferrin; Hb, haemoglobin; FPN, ferroportin

Table 1 – Key players in systemic iron metabolism.

Name	Gene	Characteristics	Function in systemic iron physiology	Dysfunctional phenotype	Refs
Bone morphogenetic protein 6 (BMP6)	<i>BMP6</i>	<ul style="list-style-type: none"> <li>• Homodimeric member of the TGF-<math>\beta</math> superfamily, expressed by non-parenchymal liver cells, cartilage, bone</li> </ul>	<ul style="list-style-type: none"> <li>• Major mediator in negative feedback loop between iron and hepcidin</li> <li>• Expressed in liver in response to intracellular iron stores or flux of iron</li> <li>• Interacts with the BMP receptor complex on hepatocytes to induce <i>HAMP</i> transcription and expression of hepcidin through the SMAD signalling pathway</li> </ul>	<ul style="list-style-type: none"> <li>• In humans, heterozygous mutations in <i>BMP6</i> result in relatively moderate, late-onset iron overload</li> <li>• Complete deficiency in mice leads to severe iron accumulation in liver, pancreas, heart and kidney</li> </ul>	173,285–288
Erythroferrone (ERFE, also known as myonectin or CTRP15)	<i>FAM132B</i>	<ul style="list-style-type: none"> <li>• Member of the TNF-<math>\alpha</math> superfamily, produced and secreted by erythroblasts</li> </ul>	<ul style="list-style-type: none"> <li>• Produced in response to erythropoietin</li> <li>• Suppresses <i>HAMP</i> transcription to increase iron supply</li> <li>• Receptor and signalling pathway in the liver are unknown</li> </ul>	<ul style="list-style-type: none"> <li>• No known mutations in humans.</li> <li>• In mice, ERFE deficiency is not associated with an abnormal phenotype, but the erythropoietic response to erythropoietin is delayed</li> </ul>	289,290
Ferroportin (FPN1, also known as SLC11A3, MTP1 or IREG1)	<i>SLC40A1</i>	<ul style="list-style-type: none"> <li>• Transmembrane protein, highly expressed in hepatocytes, enterocytes, macrophages and placental cells</li> </ul>	<ul style="list-style-type: none"> <li>• Exports Fe(II) across the cellular membrane</li> <li>• Is the only known iron exporter in vertebrates</li> </ul>	<ul style="list-style-type: none"> <li>• Human heterozygous mutations lead to autosomal dominant hemochromatosis type 4, which is associated with liver fibrosis (Kupffer cell iron overload), marginal anaemia.</li> <li>• Complete loss of function is embryonic lethal in mice.</li> </ul>	12,14–16,291–293



Hemojuvelin (HJV, also known as repulsive guidance molecule C; RGMc)	<i>HFE2</i>	<ul style="list-style-type: none"> <li>• Heterodimeric protein expressed on the cell surface of hepatocytes</li> </ul>	<ul style="list-style-type: none"> <li>• Coreceptor in the BMP receptor complex: also interacts with BMP6, possibly to assist its recruitment to the membrane</li> <li>• Required for SMAD signalling and <i>HAMP</i> transcription</li> <li>• Forms a complex with transferrin receptor 2 and HFE and functions as iron sensor</li> </ul>	<ul style="list-style-type: none"> <li>• Human homozygous or compound heterozygous mutations lead to juvenile hereditary hemochromatosis type 2A</li> <li>• Severe iron overload with similar clinical presentation as type 2B (<i>HAMP</i> mutations)</li> </ul>	178,294–296
Hepcidin (also known as LEAP-1)	<i>HAMP</i>	<ul style="list-style-type: none"> <li>• 25 amino acid secreted peptide</li> <li>• Produced primarily in liver, but also by macrophages and several other cell types</li> </ul>	<ul style="list-style-type: none"> <li>• Master regulator of iron metabolism</li> <li>• Binds to ferroportin, inducing its internalization and degradation</li> </ul>	<ul style="list-style-type: none"> <li>• Human homozygous or compound heterozygous <i>HAMP</i> mutations lead to juvenile hereditary hemochromatosis type 2B, which is associated with severe iron-overload resulting in iron-related toxicity early in life: liver fibrosis, arthropathy, cardiomyopathy, hypogonadism, and reduced glucose tolerance</li> </ul>	9–11,13,297,298
Human hemochromatosis protein (HFE, also known as HLA-H)	<i>HFE</i>	<ul style="list-style-type: none"> <li>• Monomeric MHC class I-type protein, expressed by hepatocytes, intestinal epithelial cells, macrophages, placental cells</li> </ul>	<ul style="list-style-type: none"> <li>• Competes with transferrin for binding to transferrin receptor type 1 (TfR1)</li> <li>• Associates with transferrin receptor type 2 (TfR2), in concert with HJV to induce <i>HAMP</i> transcription via SMAD signalling</li> <li>• HFE displacement from TfR1 and its subsequent association with TfR2 and HJV likely function as independent extracellular sensors</li> </ul>	<ul style="list-style-type: none"> <li>• Homozygous (majority C282Y) or compound heterozygous mutations in <i>HFE</i> cause adult hereditary hemochromatosis type 1: the most common form of hemochromatosis, associated with progressive iron overload later in life, leading to liver fibrosis and cardiomyopathy</li> </ul>	296,299–302

		for transferrin-bound iron,			
Interleukin 6 receptor (IL-6R, also known as CD126)	<i>IL6R</i>	<ul style="list-style-type: none"> <li>• Type I cytokine receptor expressed on the surface of hepatocytes and immune cells</li> <li>• Soluble IL-6R (sIL-6R) may be involved in IL-6 (trans-) signalling in all cells</li> </ul>	<ul style="list-style-type: none"> <li>• Binding of IL-6 or related cytokines to IL-6R promotes <i>HAMP</i> transcription through glycoprotein 130 (gp130) association and downstream JAK1/2-STAT3 activation.</li> <li>• Hepcidin expression during inflammation is considered one of the hallmarks of anaemia of inflammation and chronic disease</li> </ul>	<ul style="list-style-type: none"> <li>• No known mutations in humans</li> <li>• Mice deficient for IL-6 have normal serum iron levels, but are less sensitive to inflammation-induced anaemia as compared to wild-type mice</li> <li>• The iron status in IL-6R deficient mice has not yet been investigated</li> </ul>	56,246,3 03,304
Matriptase 2 (MT-2)	<i>TMPRSS6</i>	<ul style="list-style-type: none"> <li>• Type II transmembrane serine protease, expressed primarily by hepatocytes</li> </ul>	<ul style="list-style-type: none"> <li>• Inhibits hepcidin expression by limiting BMP/SMAD signalling, which, in vitro, is the result of MT-2-mediated cleavage of HJV</li> <li>• Gene expression, stability, and enzymatic activity of MT-2 is part of a feedback loop that responds to iron deficiency, hypoxia and erythropoietin, but also to chronic iron loading and BMP6</li> </ul>	<ul style="list-style-type: none"> <li>• Homozygous or compound heterozygous mutations in <i>TMPRSS6</i> result in hereditary iron-refractory iron deficiency anaemia (IRIDA), which is marked by inappropriate high levels of hepcidin</li> </ul>	55,179,1 80,305,3 06
Neogenin	<i>NEO1</i>	<ul style="list-style-type: none"> <li>• Transmembrane protein with 4 immunoglobulin-like and 6 fibronectin III-like extracellular domains</li> </ul>	<ul style="list-style-type: none"> <li>• Stabilizes HJV on the membrane by binding in a 2:2 stoichiometry via interactions between the RGD sequence on HJV and the fibronectin domain in neogenin</li> <li>• Associates with the BMP receptor complex via BMP receptor type Ia</li> </ul>	<ul style="list-style-type: none"> <li>• No known mutations in humans</li> <li>• In mice, loss of interaction of HJV with neogenin impairs BMP6-induced hepcidin expression</li> <li>• Neogenin knockout in mice is perinatal lethal</li> </ul>	294,307, 308
Transferrin	<i>TFR2</i>	<ul style="list-style-type: none"> <li>• Type II membrane</li> </ul>	<ul style="list-style-type: none"> <li>• Binds transferrin saturated with 2</li> </ul>	<ul style="list-style-type: none"> <li>• Homozygous or compound</li> </ul>	296,300,

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receptor 2 (TfR2, also known as HFE3)	protein that is relatively similar (66%) to TfR1, and is primarily expressed in the liver and erythroid cells	<p>Fe(III) ions (holotransferrin) and associates with HFE and HJV to form an extracellular transferrin-bound iron sensor and regulate hepcidin expression via the SMAD signalling pathway</p> <ul style="list-style-type: none"> <li>• TfR2 can be cleaved and released from the surface under iron deficient conditions, but the function of this soluble TfR remains unclear</li> </ul>	<p>heterozygous mutations in the <i>TFR2</i> gene underlie adult hereditary hemochromatosis type 3: the iron overload is usually more severe and presents itself earlier in life than in HFE-associated hemochromatosis type 1, but less severe than HJV-associated hemochromatosis type 2B</p> <ul style="list-style-type: none"> <li>• Similar observations have been made in knockout mice.</li> </ul>	301,309-311
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Table 2 – Key players in cellular iron homeostasis.

Name	Gene	Characteristics	Function in cellular iron physiology	Dysfunctional phenotype	Refs
<i>Iron intake</i>					
Class A scavenger receptor 5 (Scara5, also known as TESR, NET33 or SR-A5)	<i>SCARA5</i>	<ul style="list-style-type: none"> <li>• Membrane-bound member of the class A scavenger receptor family</li> <li>• Primarily expressed by fibroblasts in the respiratory tract, the urinary tract, skin, retina, testes and ovaries</li> </ul>	<ul style="list-style-type: none"> <li>• Binds and internalizes L-ferritin</li> <li>• Tissue distribution suggests a role in anti-bacterial defence, similar to other class A scavenger receptors</li> </ul>	<ul style="list-style-type: none"> <li>• Scara5-deficient mice develop normally, but later in life antinuclear autoantibodies are formed and lymphocyte infiltrations occur in various connective tissues, reminiscent of human connective tissue autoimmune diseases</li> </ul>	41,312,313
Low density lipoprotein receptor-related protein (LRP, also known as CD91)	<i>LRP1</i>	<ul style="list-style-type: none"> <li>• Member of the LDL receptor family that binds a wide array of ligands</li> <li>• Expressed by many different cells, including macrophages, hepatocytes, fibroblasts and neurons</li> </ul>	<ul style="list-style-type: none"> <li>• Scavenges and endocytoses haem in complex with haemopexin</li> <li>• Induces expression of haem oxygenase-1</li> </ul>	<ul style="list-style-type: none"> <li>• LRP1 deficiency is embryonic lethal in mice</li> <li>• In mice, macrophage-specific deletion promoted atherosclerosis, but the relationship with iron was not evaluated.</li> </ul>	37,314,315
CD163 (also known as M130, p155, haemoglobin scavenger receptor or SR-I1)	<i>CD163</i>	<ul style="list-style-type: none"> <li>• Family member of the scavenger cysteine-rich domain-containing proteins</li> <li>• Receptor expressed by monocyte-like cells, including macrophages</li> <li>• Can be shed from</li> </ul>	<ul style="list-style-type: none"> <li>• Endocytoses haemoglobin bound to haptoglobin as dimeric or multimeric complexes</li> <li>• Induces IL-10 release and haem oxygenase-1 synthesis</li> </ul>	<ul style="list-style-type: none"> <li>• In mice, CD163 deficiency does not lead to phenotypic changes</li> </ul>	35,36,316

		membrane as soluble sCD163			
Cytochrome b561, family including duodenal cytochrome b561 (Dcytb) and ferric chelate reductase 1 (also known as stromal cell-derived receptor 2; SDR2)	<i>CYBRD1</i> <i>FRRS1</i>	<ul style="list-style-type: none"> <li>• Family of transmembrane proteins, containing two haem cofactors</li> <li>• Isoforms are highly expressed in duodenum (Dcytb), liver, and kidney (SDR2), as well as in other tissues including the brain</li> </ul>	<ul style="list-style-type: none"> <li>• Enzymatically reduces Fe(III) to Fe(II) using ascorbate (vitamin C) as an electron donor</li> <li>• Enables iron transport across the cell/lysosomal membrane by DMT, ZIP14 or ZIP8</li> </ul>	<ul style="list-style-type: none"> <li>• In mice, <i>Cybrd1</i> knockout does not change steady-state serum iron levels or iron stores, but intestinal iron absorption is limited during erythropoietic stress</li> </ul>	30,317-319
Divalent metal-ion transporter 1 (DMT1, also known as DCT1 or Nramp2)	<i>SLC11A2</i>	<ul style="list-style-type: none"> <li>• 12-transmembrane protein, ubiquitously expressed, with high levels in duodenum, erythroid cells, brain and kidney</li> </ul>	<ul style="list-style-type: none"> <li>• Transports unbound extracellular bivalent metals, including Fe, into the cell</li> </ul>	<ul style="list-style-type: none"> <li>• In humans, mutations in <i>DMT1</i> have been found in patients with microcytic anaemia and iron overload</li> <li>• Complete DMT1 disruption is lethal shortly after birth in mice</li> </ul>	21,27,194,320
Feline leukaemia virus, subgroup C, receptor 2 (FLVCR2, also known as SLC49A2)	<i>FLVCR2</i>	<ul style="list-style-type: none"> <li>• Transmembrane protein highly transcribed in various tissues, including the brain, placenta, lung, liver, kidney, spleen and pancreas</li> </ul>	<ul style="list-style-type: none"> <li>• Imports haem into the cell</li> <li>• Exact function in overall iron homeostasis not yet clear</li> </ul>	<ul style="list-style-type: none"> <li>• Homozygous or compound heterozygous mutations in <i>FLVCR2</i> underlie Fowler syndrome, a syndrome that often results in prenatal lethality marked by CNS malformations, including hydrocephaly and hydranencephaly.</li> </ul>	39,321

Haem responsive gene-1 (HRG-1)	<i>SLC48A1</i>	<ul style="list-style-type: none"> <li>• Membrane transporter widely expressed in brain, kidney, heart and muscle</li> <li>• Low expression in liver, lung, placenta and intestine</li> </ul>	<ul style="list-style-type: none"> <li>• Transports haem across the endosomal membrane, primarily in macrophages to recycle haem after erythrophagocytosis.</li> </ul>	<ul style="list-style-type: none"> <li>• No known or generated mutations in mammals</li> <li>• In zebrafish, HRG-1 knockout leads to severe developmental defects, such as hydrocephalus and erythropoietic impairment</li> </ul>	38,40,322
Six-transmembrane epithelial antigen of prostate (STEAP family proteins)	<i>STEAP1-4</i>	<ul style="list-style-type: none"> <li>• Transmembrane proteins with 4 homologous members</li> <li>• Ubiquitous expression; elevated levels of STEAP1 in particular can be found in a variety of solid tumours, including prostate</li> </ul>	<ul style="list-style-type: none"> <li>• STEAP2-4 can reduce Fe(III) to Fe(II)</li> <li>• STEAP3 has been implicated as the main ferrireductase in the Tf cycle, enabling iron transport out of the endosome; STEAP2 and STEAP4 have shown a similar capacity</li> <li>• STEAP2-4 are also involved in copper reduction</li> </ul>	<ul style="list-style-type: none"> <li>• In humans, low or absent expression of functional STEAP3 leads to hypochromic anaemia, due to an insufficient iron supply for Hb synthesis.</li> <li>• STEAP3-deficient mice retain some ferrireductive activity, possibly via STEAP2 and STEAP4</li> </ul>	31,32,323-325
Transferrin (Tf)	<i>TF</i>	<ul style="list-style-type: none"> <li>• Abundant monomeric serum glycoprotein, primarily produced in the liver</li> <li>• Tf comprises two lobes, each capable of binding a Fe(III) ion with high affinity (<math>K_d \approx 10^{-20}</math> M at pH 7.4).</li> <li>• Homologous family members in mammals include lactoferrin, melanotransferrin and ovotransferrin</li> </ul>	<ul style="list-style-type: none"> <li>• Chelates and transports insoluble Fe(III) in the circulation</li> <li>• Binding to transferrin is the principal method for systemic iron distribution</li> <li>• About 3 mg of iron is transferrin-bound at any time (<math>\pm 30\%</math> Tf saturation), and 30 mg is turned over by Tf each day</li> </ul>	<ul style="list-style-type: none"> <li>• Homozygous or compound heterozygous mutations in <i>TF</i> result in the rare disorder atransferrinemia; this disease is associated with anaemia and secondary iron overload, due to an insufficient iron supply to immature erythroid cells</li> </ul>	17,18,326

Transferrin receptor 1 (TfR1, also known as CD71)	<i>TFRC</i>	<ul style="list-style-type: none"> <li>• Homodimeric type II transmembrane glycoprotein ubiquitously expressed on the surface of most cells</li> <li>• A non-membrane bound, truncated monomeric form (sTfR) can be found in the serum</li> </ul>	<ul style="list-style-type: none"> <li>• Each monomer can bind a single Tf with nanomolar affinity for biferric Tf.</li> <li>• Upon internalization of the Tf-TfR complex by clathrin-mediated endocytosis, iron is released under acidic conditions in the lysosome and the apo-Tf/TfR complex is recycled</li> <li>• TfR1 can also bind and support the internalization of h-ferritin</li> <li>• The function of sTfR is currently unknown, but it serves as a biomarker for anaemia and some malignancies</li> </ul>	<ul style="list-style-type: none"> <li>• Complete deficiency is embryonic lethal in mice</li> <li>• In humans, a homozygous missense mutation in <i>TFRC</i> that affects its internalization results in combined immunodeficiency and mild anaemia; the limited impact of this mutation on haemoglobin levels could be related to compensation by STEAP3</li> </ul>	18,19,42,78,327
Zrt- and Irt-like protein 8 (ZIP8) and ZIP14	<i>SLC39A8</i> <i>SLC39A14</i>	<ul style="list-style-type: none"> <li>• Transmembrane proteins, part of the zinc transporter family, highly expressed in liver, pancreas, brain and heart</li> </ul>	<ul style="list-style-type: none"> <li>• Takes up unbound Fe(II) (as well as zinc) into the cell</li> <li>• ZIP14 is found at high levels in liver and other tissues that accumulate iron in iron-loading disorders</li> <li>• ZIP8 is involved in the uptake of unbound iron in neurons</li> </ul>	<ul style="list-style-type: none"> <li>• No known mutations in humans</li> <li>• ZIP14-deficient mice show reduced capacity to internalize iron in the liver and pancreas, but serum iron and Hb levels are similar to wild-type mice</li> <li>• ZIP14-deficient mice have skeletal underdevelopment due to Zn deficiency.</li> </ul>	22,23,29,208,328,329
<i>Iron utilization</i>					
Ferritin (Ftn)	<i>FTH1</i> (heavy chain), <i>FTL</i> (light chain)	<ul style="list-style-type: none"> <li>• Ubiquitously expressed protein that is composed of 24 subunits and comprises a</li> </ul>	<ul style="list-style-type: none"> <li>• Stores iron: each ferritin molecule can bind up to 4,500 Fe(III) atoms</li> </ul>	<ul style="list-style-type: none"> <li>• A heterozygous mutation in <i>FTL</i> is associated with neuroferritinopathy, a</li> </ul>	46,50,51,128,330

		combination of heavy and light chains, which together form a hollow shell of around 12 nm in diameter	<ul style="list-style-type: none"> <li>• Each H-subunit comprises a ferroxidase centre that allows the oxidation of Fe<sup>2+</sup> into Fe<sup>3+</sup>, while the L-subunits promote iron mineral nucleation.</li> <li>• Extracellular (serum) ferritin, mostly consisting of L-ferritin, secreted by the liver and macrophages, serves as a biomarker for systemic iron load.</li> </ul>	<p>neurodegenerative syndrome</p> <ul style="list-style-type: none"> <li>• The only known case of complete <i>FTL</i> deficiency resulted in seizures and restless leg syndrome, likely resulting from iron deficiency due to the higher affinity of the heavy chain for iron</li> <li>• <i>Fth1</i> deficiency in mice is embryonic lethal</li> </ul>	
Haem oxygenase-1 (HO-1)	<i>HMOX1</i>	<ul style="list-style-type: none"> <li>• Highly inducible catalytic enzyme, expression can, in principle, occur in most cells</li> </ul>	<ul style="list-style-type: none"> <li>• Degrades haem into iron, carbon monoxide and biliverdin; the latter two have anti-inflammatory properties</li> <li>• Is important for iron recycling from erythrocytes by macrophages</li> <li>• Expression is largely controlled by nuclear factor-like 2 (Nrf2).</li> </ul>	<ul style="list-style-type: none"> <li>• Only one reported case of HO-1 deficiency in humans: patient with compound heterozygous mutations presented with anaemia, hemolysis, macrophage dysfunction, hyperlipidemia and atherosclerosis, and died at age 6</li> <li>• Mice lacking <i>HMOX1</i> develop iron-deficiency anaemia, as well as iron overload in liver and kidneys</li> </ul>	331-333
Hypoxia-inducible factor (HIF)	<i>HIF1A</i> (HIF1 $\alpha$ ) <i>EPAS1</i> (HIF2 $\alpha$ ) <i>HIF3A</i> (HIF3 $\alpha$ )	<ul style="list-style-type: none"> <li>• Oxygen-responsive <math>\alpha</math>-subunit that dimerizes with a <math>\beta</math>-subunit to form an intracellular heterodimeric protein</li> <li>• HIF1<math>\alpha</math> is ubiquitously expressed, HIF2<math>\alpha</math> is predominantly expressed by renal, hepatic,</li> </ul>	<ul style="list-style-type: none"> <li>• HIF2<math>\alpha</math> in particular modulates the expression of various proteins involved in iron metabolism by binding to hypoxic response elements (HREs) in gene promoters to increase iron trafficking (including <i>HAMP</i> suppression)</li> <li>• HIF2<math>\alpha</math> translation is regulated by</li> </ul>	<ul style="list-style-type: none"> <li>• In humans, loss-of-function mutations in HIF1 or HIF2 have not been described.</li> <li>• Gain-of-function mutations in HIF2 have been found in two patients with polycythaemia, and stabilization of HIF1 or HIF2 has been associated with various forms of cancer</li> </ul>	209- 213,334-336



		macrophageal, glial, cardiac and endothelial cells	iron via the presence of an iron response element (IRE) element in its mRNA	<ul style="list-style-type: none"> <li>• In mice, HIF1<math>\alpha</math>-knockout is embryonic lethal</li> <li>• HIF2<math>\alpha</math>-knockout mice are only viable in a mixed genetic background, and have severe abnormalities in numerous organs</li> </ul>	
Mitoferrin 1 and mitoferrin 2	<i>SLC25A37</i> (mitoferrin 1) and <i>SLC25A28</i> (mitoferrin 2)	<ul style="list-style-type: none"> <li>• Mitochondrial transporters that are part of the mitochondrial carrier proteins (SLC25) family</li> <li>• Mitoferrin 1 is primarily expressed in erythroid cells and mitoferrin 2 is widely expressed</li> </ul>	<ul style="list-style-type: none"> <li>• Imports iron across the mitochondrial inner membrane, supplying iron for haem and Fe-S cluster synthesis</li> </ul>	<ul style="list-style-type: none"> <li>• No known mutations in humans</li> <li>• In mice, knockout of mitoferrin 1 is embryonic lethal</li> <li>• Selective knockout of <i>Slc25a37</i> in erythroid tissues in mice leads to severe anaemia</li> </ul>	45,337
Prolyl-hydroxylase domain-containing protein (PHD, also known as HIF-PH)	<i>ELGN2</i> (PHD1) <i>ELGN1</i> (PHD2) <i>ELGN3</i> (PHD3) <i>P4HTM</i> (PHD4)	<ul style="list-style-type: none"> <li>• Ubiquitously expressed enzyme family, localized in the nucleus (PHD1 and PHD3) and the cytosol (PHD2 and PHD3)</li> <li>• PHD4 contains an endoplasmic reticulum transmembrane domain</li> </ul>	<ul style="list-style-type: none"> <li>• The oxygen-sensitive PHDs, especially PHD2, hydroxylate the <math>\alpha</math>-subunit of HIF to promote its ubiquitylation and degradation, and thereby limit iron trafficking</li> <li>• Especially PHD2 fulfils a crucial role.</li> <li>• Fe(II) is required as a cofactor</li> </ul>	<ul style="list-style-type: none"> <li>• In humans, heterozygous mutations in <i>ELGN1</i> result in loss of function, which increases levels of EPO and Hb</li> <li>• <i>Elgn1</i> knockout is embryonic lethal in mice; <i>Elgn2</i> and <i>Elgn3</i> knockout mice are phenotypically normal</li> </ul>	216,217,219, 338,339
<i>Iron efflux</i>					
Ceruloplasmin (Cp)	<i>CP</i>	<ul style="list-style-type: none"> <li>• Glycoprotein member of the multi-copper oxidase family, each protein contains 6 copper ions</li> </ul>	<ul style="list-style-type: none"> <li>• Oxidizes Fe(II) to Fe(III) via concurrent reduction of dioxygen to water</li> <li>• Enables binding of Fe(III) to</li> </ul>	<ul style="list-style-type: none"> <li>• Deficiency of ceruloplasmin due to mutations in <i>Cp</i> are associated with cognitive and neurological degeneration, type II diabetes,</li> </ul>	49,340

		<ul style="list-style-type: none"> <li>• Abundantly available in the blood; produced in the liver</li> <li>• Membrane-anchored Cp is found in astrocytes and retinal cells</li> </ul>	transferrin to facilitate cellular iron export by ferroportin	retinal degradation and hepatic iron overload	
Feline leukaemia virus, subgroup C, receptor 1 (FLVCR1a and FLVCR1b, also known as SLC49A1)	<i>FLVCR1</i>	<ul style="list-style-type: none"> <li>• Transmembrane protein isoforms with ubiquitous expression in placenta, uterus, duodenum, liver, kidney, lung, brain and bone marrow</li> </ul>	<ul style="list-style-type: none"> <li>• Exports of haem from cells (FLVCR1a), especially erythroid progenitors and macrophages</li> <li>• May function as an 'emergency valve', preventing oxidative damage in case of excess iron</li> <li>• Might directly supply haem to developing erythroblasts from macrophages</li> <li>• Exports haem from mitochondria (FLVCR1b) in haem-producing cells</li> </ul>	<ul style="list-style-type: none"> <li>• Enhanced alternative splicing of FLVCR1 has been observed in patients with Diamond Blackfan anaemia, a rare form of anaemia characterized by a reduction of erythroid progenitors</li> <li>• Similar anaemia is observed in FLVCR1-deficient mice, which also have impaired T-cell development</li> </ul>	53,341-345
Ferroportin	<i>See Table 1</i>				
Hephaestin (Hp)	<i>HEPH</i>	<ul style="list-style-type: none"> <li>• Transmembrane protein that contains 6 copper ions per molecule; member of the multi-copper oxidase family</li> <li>• Mainly expressed in the intestine, but also in placenta, heart, brain and pancreatic <math>\beta</math>-cells</li> </ul>	<ul style="list-style-type: none"> <li>• Oxidizes Fe(II) to Fe(III) via concurrent reduction of dioxygen to water</li> <li>• Enables binding of Fe(III) to transferrin to facilitate cellular iron export by ferroportin</li> </ul>	<ul style="list-style-type: none"> <li>• No known mutations in humans</li> <li>• Hephaestin-deficient mice have iron-deficiency anaemia with enterocytic iron overload</li> </ul>	346-348

Table 3 – Therapeutic agents targeting iron metabolism under commercial development

Target	Name	Company	Type	Mechanism	Indication	Development status	Trial identifiers <sup>a</sup>
BMP6 receptor pathway	LY3113593	Eli Lilly and Company	Humanized antibody	Capture of BMP6 to limit hepcidin expression	Anaemia	Phase 2 (lost through attrition 2016; see Further information)	NCT02604160 NCT02144285
	FMX-8	FerruMax Pharmaceuticals, Inc	HJV-Fc fusion protein	Capture of BMP6 to limit hepcidin expression	Anaemia	Phase 2 (terminated trials 2016)	NCT02228655 NCT01873534
Ferroportin	LY2928057	Eli Lilly and Company	Humanized IgG4 antibody	Binding of ferroportin to prevent its degradation by hepcidin, thereby stabilizing ferroportin expression	Anaemia	Phase 2 (removed from pipeline 2016)	NCT01991483 NCT01330953
Hepcidin	M012	Merganser Biotech Australia Pty Ltd.	Peptide	Hepcidin mimetic peptide that functions as a hepcidin agonist	$\beta$ -thalassemia, low risk myelodysplastic syndrome, polycythaemia vera	Phase 1	ACTRN1261600093482
	LJPC-401	La Jolla Pharmaceutical Company	Peptide	Hepcidin mimetic peptide that functions as a hepcidin agonist	Iron overload	Phase 1	ACTRN12616000132448

	PTG-300	Protagonist Therapeutics	Peptide	Hepcidin mimetic peptide that functions as a hepcidin agonist	Iron overload	preclinical	<i>n.a.</i>
	PRS-080	Pieris Pharmaceuticals GmbH	Anticalin	Bioengineered lipocalin that captures hepcidin to prevent ferroportin degradation	Anaemia, functional iron deficiency	Phase 1b	NCT02754167 NCT02340572
	NOX-H94 (lexaptetid pegol)	NOXXON Pharma AG	L-RNA spiegelmer	L-stereoisomeric RNA aptamer that captures hepcidin to prevent ferroportin degradation	Anaemia	Phase 2	NCT02079896 NCT01691040 NCT01522794 NCT01372137
	12B9m	Amgen Inc	Human IgG2 antibody	Capture of hepcidin to prevent ferroportin degradation	Anaemia	Preclinical	<i>n.a.</i>
Iron	Deferoxamine (Desferal)	Novartis	small molecule	Chelation of iron	Iron intoxication and overload	Marketed	
	Deferasirox (Exjade)	Novartis	Small molecule	Chelation of iron	Iron overload	Marketed	
	Deferiprone (Ferriprox)	ApoPharma Inc.	Small molecule	Chelation of iron	Iron overload	Marketed	
	DpC	Oncochel Therapeutics	Small molecule	Chelation of iron	Advanced solid tumours	Phase 1	NCT02688101

Matriptase-2	IONIS-TMPRSS6-LRx	Ionis Pharmaceuticals	Antisense DNA oligo-nucleotide	Inhibition of MT-2 expression to promote hepcidin transcription	$\beta$ -thalassemia	Preclinical	<i>n.a.</i>
	ALN-TMP	Alnylam	siRNA	Inhibition of MT-2 expression to promote hepcidin transcription	$\beta$ -thalassemia, iron overload	Preclinical	<i>n.a.</i>
Prolyl hydroxylase	FG-4592 / ASP1517 (Roxadustat)	FibroGen Astellas Pharma AstraZeneca	Small molecule	Inhibition of prolyl hydroxylases to stabilize HIFs	Anaemia in chronic kidney disease and end-stage renal disease	Phase 3	NCT02805374 NCT02780726 NCT02779764 NCT02780141
	GSK1278863 / GSK1278863A (Daprodustat)	GlaxoSmithKline	Small molecule	Inhibition of prolyl hydroxylases to stabilize HIFs	Anaemia in chronic kidney disease Wound healing	Phase 3	NCT02879305 NCT02876835 NCT02829320 NCT02791763
	AKB-6548 (Vadadustat)	Procter & Gamble Akebia Therapeutics Mitsubishi Tanabe Pharma	Small molecule	Inhibition of prolyl hydroxylases to stabilize HIFs	Anaemia in chronic kidney disease	Phase 3	NCT02892149 NCT02865850 NCT02680574 NCT02648347
	BAY85-3934 (Molidustat)	Bayer HealthCare AG	Small molecule	Inhibition of prolyl hydroxylases to stabilize HIFs	Anaemia in chronic kidney disease	Phase 2	NCT02312973 NCT02064426 NCT01975818 NCT02021409

	JTZ-951	Japan Tobacco Inc.	Small molecule	Inhibition of prolyl hydroxylases to stabilize HIFs	Anaemia in chronic kidney disease	Phase 2	NCT02805244 JPRN-JapicCTI-152892 JPRN-JapicCTI-152891 JPRN-JapicCTI-152881
	ZYAN1	Cadila Healthcare Ltd	Small molecule	Inhibition of prolyl hydroxylases to stabilize HIFs	Anaemia in chronic kidney disease	Phase 1	CTRI/2016/02/006665
	JNJ-42905343	Johnson & Johnson	Small molecule	Inhibition of prolyl hydroxylases to stabilize HIFs	Anaemia, functional iron deficiency	Preclinical	<i>n.a.</i>
Transferrin receptor 1	MBP-426	Mebiopharm Co., Ltd	Liposomal oxaliplatin functionalized with transferrin	Drug targeting to highly TfR1-expressing tumours	Gastric, gastro-oesophageal, and oesophageal adenocarcinoma	Phase 2 (last trial initiated 2009)	NCT00964080 NCT00355888
	SGT-53	SynerGene Therapeutics, Inc.	Liposomal plasmid DNA encoding p53 functionalized with TfR1-targeting single-chain variable fragment	Gene therapy to highly TfR1-expressing tumours and that crosses the blood-brain barrier	Solid tumours	Phase 2	NCT02354547 NCT02340156 NCT02340117 NCT00470613

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SGT-94	SynerGene Therapeutics, Inc.	Liposomal plasmid DNA encoding RB94, functionalized with TfR1- targeting single- chain variable fragment	Gene therapy to highly TfR1-expressing tumours	Solid tumours	Phase 1	NCT01517464
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<sup>a</sup> Most recent clinical trials (4), ordered by date of registration on the WHO International Clinical Trials Registry Platform (<http://apps.who.int/trialsearch/Default.aspx>)

## References

1. Rouault, T. A. Iron-sulfur proteins hiding in plain sight. *Nat. Chem. Biol.* **11**, 442–445 (2015).
2. Hohenberger, J., Ray, K. & Meyer, K. The biology and chemistry of high-valent iron-oxo and iron-nitrido complexes. *Nat. Commun.* **3**, 720 (2012).
3. Ganz, T. Systemic Iron Homeostasis. *Physiol. Rev.* **93**, 1721–1741 (2013).
4. Camaschella, C. Iron-Deficiency Anemia. *N. Engl. J. Med.* **372**, 1832–1843 (2015).
5. Rivella, S.  $\beta$ -thalassemias: paradigmatic diseases for scientific discoveries and development of innovative therapies. *Haematologica* **100**, 418–430 (2015).
6. Stadtman, E. R. Protein oxidation and aging. *Science* **257**, 1220–1224 (1992).
7. Fenton, H. J. H. Oxidation of tartaric acid in presence of iron. *J. Chem. Soc. Trans.* **65**, 899–910 (1894).
8. Haber, F. & Weiss, J. Über die Katalyse des Hydroperoxydes. *Naturwissenschaften* **20**, 948–950 (1932).
9. Park, C. H., Valore, E. V., Waring, A. J. & Ganz, T. Hepcidin, a Urinary Antimicrobial Peptide Synthesized in the Liver. *J. Biol. Chem.* **276**, 7806–7810 (2001).
10. Krause, A. *et al.* LEAP-1, a novel highly disulfide-bonded human peptide, exhibits antimicrobial activity<sup>1</sup>. *FEBS Lett.* **480**, 147–150 (2000).
11. Nicolas, G. *et al.* Lack of hepcidin gene expression and severe tissue iron overload in upstream stimulatory factor 2 (USF2) knockout mice. *Proc. Natl. Acad. Sci.* **98**, 8780–8785 (2001).



12. Donovan, A. *et al.* Positional cloning of zebrafish ferroportin1 identifies a conserved vertebrate iron exporter. *Nature* **403**, 776–781 (2000).
13. Nemeth, E. *et al.* Heparin regulates cellular iron efflux by binding to ferroportin and inducing its internalization. *Science* **306**, 2090–2093 (2004).
14. McKie, A. T. *et al.* A Novel Duodenal Iron-Regulated Transporter, IREG1, Implicated in the Basolateral Transfer of Iron to the Circulation. *Mol. Cell* **5**, 299–309 (2000).
15. Donovan, A. *et al.* The iron exporter ferroportin/Slc40a1 is essential for iron homeostasis. *Cell Metab.* **1**, 191–200 (2005).
16. Abboud, S. & Haile, D. J. A Novel Mammalian Iron-regulated Protein Involved in Intracellular Iron Metabolism. *J. Biol. Chem.* **275**, 19906–19912 (2000).
17. Aisen, P., Leibman, A. & Zweier, J. Stoichiometric and site characteristics of the binding of iron to human transferrin. *J. Biol. Chem.* **253**, 1930–1937 (1978).
18. Fotticchia, I. *et al.* Energetics of ligand-receptor binding affinity on endothelial cells: An in vitro model. *Colloids Surf. B Biointerfaces* **144**, 250–256 (2016).
19. Dautry-Varsat, A., Ciechanover, A. & Lodish, H. F. pH and the recycling of transferrin during receptor-mediated endocytosis. *Proc. Natl. Acad. Sci. U. S. A.* **80**, 2258–2262 (1983).
20. Trinder, D., Zak, O. & Aisen, P. Transferrin receptor-independent uptake of different transferrin by human hepatoma cells with antisense inhibition of receptor expression. *Hepatology* **23**, 1512–1520 (1996).
21. Gunshin, H. *et al.* Cloning and characterization of a mammalian proton-coupled metal-ion transporter. *Nature* **388**, 482–488 (1997).

22. Liuzzi, J. P., Aydemir, F., Nam, H., Knutson, M. D. & Cousins, R. J. Zip14 (Slc39a14) mediates non-transferrin-bound iron uptake into cells. *Proc. Natl. Acad. Sci.* **103**, 13612–13617 (2006).
23. Wang, C.-Y. *et al.* ZIP8 Is an Iron and Zinc Transporter Whose Cell-surface Expression Is Up-regulated by Cellular Iron Loading. *J. Biol. Chem.* **287**, 34032–34043 (2012).
24. Fleming, M. D. *et al.* Microcytic anaemia mice have a mutation in Nramp2, a candidate iron transporter gene. *Nat. Genet.* **16**, 383–386 (1997).
25. Fleming, M. D. *et al.* Nramp2 is mutated in the anemic Belgrade (b) rat: Evidence of a role for Nramp2 in endosomal iron transport. *Proc. Natl. Acad. Sci.* **95**, 1148–1153 (1998).
26. Shawki, A. *et al.* Intestinal DMT1 is critical for iron absorption in the mouse but is not required for the absorption of copper or manganese. *Am. J. Physiol. - Gastrointest. Liver Physiol.* **309**, G635–G647 (2015).
27. Mims, M. P. *et al.* Identification of a human mutation of DMT1 in a patient with microcytic anemia and iron overload. *Blood* **105**, 1337–1342 (2005).
28. Canonne-Hergaux, F., Zhang, A.-S., Ponka, P. & Gros, P. Characterization of the iron transporter DMT1 (NRAMP2/DCT1) in red blood cells of normal and anemic mk/mk mice. *Blood* **98**, 3823–3830 (2001).
29. Jenkitkasemwong, S. *et al.* SLC39A14 Is Required for the Development of Hepatocellular Iron Overload in Murine Models of Hereditary Hemochromatosis. *Cell Metab.* **22**, 138–150 (2015).
30. McKie, A. T. *et al.* An Iron-Regulated Ferric Reductase Associated with the Absorption of Dietary Iron. *Science* **291**, 1755–1759 (2001).

31. Ohgami, R. S. *et al.* Identification of a ferrireductase required for efficient transferrin-dependent iron uptake in erythroid cells. *Nat Genet* **37**, 1264–1269 (2005).
32. Ohgami, R. S., Campagna, D. R., McDonald, A. & Fleming, M. D. The Steap proteins are metalloredutases. *Blood* **108**, 1388–1394 (2006).
33. Tripathi, A. K. *et al.* Prion protein functions as a ferrireductase partner for ZIP14 and DMT1. *Free Radic. Biol. Med.* **84**, 322–330 (2015).
34. Singh, A. *et al.* Prion protein regulates iron transport by functioning as a ferrireductase. *J. Alzheimers Dis. JAD* **35**, 541–552 (2013).
35. Kristiansen, M. *et al.* Identification of the haemoglobin scavenger receptor. *Nature* **409**, 198–201 (2001).
36. Andersen, C. B. F. *et al.* Structure of the haptoglobin-haemoglobin complex. *Nature* **489**, 456–459 (2012).
37. Hvidberg, V. *et al.* Identification of the receptor scavenging hemopexin-heme complexes. *Blood* **106**, 2572–2579 (2005).
38. Rajagopal, A. *et al.* Haem homeostasis is regulated by the conserved and concerted functions of HRG-1 proteins. *Nature* **453**, 1127–1131 (2008).
39. Duffy, S. P. *et al.* The Fowler Syndrome-Associated Protein FLVCR2 Is an Importer of Heme. *Mol. Cell. Biol.* **30**, 5318–5324 (2010).
40. White, C. *et al.* HRG1 Is Essential for Heme Transport from the Phagolysosome of Macrophages during Erythrophagocytosis. *Cell Metab.* **17**, 261–270 (2013).
41. Li, J. Y. *et al.* Scara5 Is a Ferritin Receptor Mediating Non-Transferrin Iron Delivery. *Dev. Cell* **16**, 35–46 (2009).

42. Li, L. *et al.* Binding and uptake of H-ferritin are mediated by human transferrin receptor-1. *Proc. Natl. Acad. Sci. U. S. A.* **107**, 3505–3510 (2010).
43. Moss, D., Powell, L. W., Arosio, P. & Halliday, J. W. Characterization of the ferritin receptors of human T lymphoid (MOLT-4) cells. *J. Lab. Clin. Med.* **119**, 273–279 (1992).
44. Chen, T. T. *et al.* TIM-2 is expressed on B cells and in liver and kidney and is a receptor for H-ferritin endocytosis. *J. Exp. Med.* **202**, 955–965 (2005).
45. Shaw, G. C. *et al.* Mitoferrin is essential for erythroid iron assimilation. *Nature* **440**, 96–100 (2006).
46. Bou-Abdallah, F. The iron redox and hydrolysis chemistry of the ferritins. *Biochim. Biophys. Acta BBA - Gen. Subj.* **1800**, 719–731 (2010).
47. Leibold, E. A. & Munro, H. N. Cytoplasmic protein binds in vitro to a highly conserved sequence in the 5' untranslated region of ferritin heavy- and light-subunit mRNAs. *Proc. Natl. Acad. Sci.* **85**, 2171–2175 (1988).
48. Rouault, T. A., Hentze, M. W., Caughman, S. W., Harford, J. B. & Klausner, R. D. Binding of a cytosolic protein to the iron-responsive element of human ferritin messenger RNA. *Science* **241**, 1207–1210 (1988).
49. Vashchenko, G. & MacGillivray, R. T. A. Multi-Copper Oxidases and Human Iron Metabolism. *Nutrients* **5**, 2289–2313 (2013).
50. Ghosh, S., Hevi, S. & Chuck, S. L. Regulated secretion of glycosylated human ferritin from hepatocytes. *Blood* **103**, 2369–2376 (2004).
51. Cohen, L. A. *et al.* Serum ferritin is derived primarily from macrophages through a nonclassical secretory pathway. *Blood* **116**, 1574–1584 (2010).

52. Mancias, J. D., Wang, X., Gygi, S. P., Harper, J. W. & Kimmelman, A. C. Quantitative proteomics identifies NCOA4 as the cargo receptor mediating ferritinophagy. *Nature* **509**, 105–109 (2014).
53. Keel, S. B. *et al.* A Heme Export Protein Is Required for Red Blood Cell Differentiation and Iron Homeostasis. *Science* **319**, 825–828 (2008).
54. Taher, A. T. *et al.* Overview on practices in thalassemia intermedia management aiming for lowering complication rates across a region of endemicity: the OPTIMAL CARE study. *Blood* **115**, 1886–1892 (2010).
55. Finberg, K. E. *et al.* Mutations in TMPRSS6 cause iron-refractory iron deficiency anemia (IRIDA). *Nat. Genet.* **40**, 569–571 (2008).
56. Gardenghi, S. *et al.* Distinct roles for hepcidin and interleukin-6 in the recovery from anemia in mice injected with heat-killed *Brucella abortus*. *Blood* **123**, 1137–1145 (2014).
57. Canali, S. *et al.* Activin B Induces Noncanonical SMAD1/5/8 Signaling via BMP Type I Receptors in Hepatocytes: Evidence for a Role in Hepcidin Induction by Inflammation in Male Mice. *Endocrinology* **157**, 1146–1162 (2016).
58. Weiss, G. & Schett, G. Anaemia in inflammatory rheumatic diseases. *Nat. Rev. Rheumatol.* **9**, 205–215 (2013).
59. Eaton, J. W. & Qian, M. Molecular bases of cellular iron toxicity. *Free Radic. Biol. Med.* **32**, 833–840 (2002).
60. Dixon, S. J. *et al.* Ferroptosis: An Iron-Dependent Form of Nonapoptotic Cell Death. *Cell* **149**, 1060–1072 (2012).
61. Skouta, R. *et al.* Ferrostatins Inhibit Oxidative Lipid Damage and Cell Death in Diverse Disease Models. *J. Am. Chem. Soc.* **136**, 4551–4556 (2014).

62. Kang, Y., Tiziani, S., Park, G., Kaul, M. & Paternostro, G. Cellular protection using Flt3 and PI3K $\alpha$  inhibitors demonstrates multiple mechanisms of oxidative glutamate toxicity. *Nat. Commun.* **5**, 3672 (2014).
63. Friedmann Angeli, J. P. *et al.* Inactivation of the ferroptosis regulator Gpx4 triggers acute renal failure in mice. *Nat. Cell Biol.* **16**, 1180–1191 (2014).
64. Linkermann, A. *et al.* Synchronized renal tubular cell death involves ferroptosis. *Proc. Natl. Acad. Sci.* **111**, 16836–16841 (2014).
65. Jiang, L. *et al.* Ferroptosis as a p53-mediated activity during tumour suppression. *Nature* **520**, 57–62 (2015).
66. Sun, X. *et al.* HSPB1 as a novel regulator of ferroptotic cancer cell death. *Oncogene* **34**, 5617–5625 (2015).
67. Yang, W. S. *et al.* Regulation of Ferroptotic Cancer Cell Death by GPX4. *Cell* **156**, 317–331 (2014).
68. Dixon, S. J. & Stockwell, B. R. The role of iron and reactive oxygen species in cell death. *Nat. Chem. Biol.* **10**, 9–17 (2014).
69. Sabharwal, S. S. & Schumacker, P. T. Mitochondrial ROS in cancer: initiators, amplifiers or an Achilles' heel? *Nat. Rev. Cancer* **14**, 709–721 (2014).
70. Conrad, M., Angeli, J. P. F., Vandenabeele, P. & Stockwell, B. R. Regulated necrosis: disease relevance and therapeutic opportunities. *Nat. Rev. Drug Discov.* **15**, 348–366 (2016).
71. Chow, A. *et al.* CD169+ macrophages provide a niche promoting erythropoiesis under homeostasis and stress. *Nat. Med.* **19**, 429–436 (2013).
72. Ramos, P. *et al.* Macrophages support pathological erythropoiesis in polycythemia vera and  $\beta$ -thalassemia. *Nat. Med.* **19**, 437–445 (2013).

73. She, H. *et al.* Iron activates NF- $\kappa$ B in Kupffer cells. *Am. J. Physiol. - Gastrointest. Liver Physiol.* **283**, G719–G726 (2002).
74. Zhang, Z. *et al.* Ferroportin1 deficiency in mouse macrophages impairs iron homeostasis and inflammatory responses. *Blood* **118**, 1912–1922 (2011).
75. Zanganeh, S. *et al.* Iron oxide nanoparticles inhibit tumour growth by inducing pro-inflammatory macrophage polarization in tumour tissues. *Nat. Nanotechnol.* (2016). doi:10.1038/nnano.2016.168
76. Sindrilaru, A. *et al.* An unrestrained proinflammatory M1 macrophage population induced by iron impairs wound healing in humans and mice. *J. Clin. Invest.* **121**, 985–997 (2011).
77. Vinchi, F. *et al.* Hemopexin therapy reverts heme-induced proinflammatory phenotypic switching of macrophages in a mouse model of sickle cell disease. *Blood* **127**, 473–486 (2016).
78. Jabara, H. H. *et al.* A missense mutation in TFRC, encoding transferrin receptor 1, causes combined immunodeficiency. *Nat. Genet.* **48**, 74–78 (2016).
79. Wandersman, C. & Delepelaire, P. Bacterial Iron Sources: From Siderophores to Hemophores. *Annu. Rev. Microbiol.* **58**, 611–647 (2004).
80. Bao, G. *et al.* Iron traffics in circulation bound to a siderocalin (Ngal)–catechol complex. *Nat. Chem. Biol.* **6**, 602–609 (2010).
81. Flo, T. H. *et al.* Lipocalin 2 mediates an innate immune response to bacterial infection by sequestering iron. *Nature* **432**, 917–921 (2004).
82. Arezes, J. *et al.* Hepcidin-Induced Hypoferremia Is a Critical Host Defense Mechanism against the Siderophilic Bacterium *Vibrio vulnificus*. *Cell Host Microbe* **17**, 47–57 (2015).

83. Chen, S. *et al.* Transforming Growth Factor  $\beta$ 1 (TGF- $\beta$ 1) Activates Heparin mRNA Expression in Hepatocytes. *J. Biol. Chem.* **291**, 13160–13174 (2016).
84. Guida, C. *et al.* A novel inflammatory pathway mediating rapid hepcidin-independent hypoferrremia. *Blood* **125**, 2265–2275 (2015).
85. Piskounova, E. *et al.* Oxidative stress inhibits distant metastasis by human melanoma cells. *Nature* **527**, 186–191 (2015).
86. Torti, S. V. & Torti, F. M. Iron and cancer: more ore to be mined. *Nat. Rev. Cancer* **13**, 342–355 (2013).
87. Bystrom, L. M. & Rivella, S. Cancer cells with irons in the fire. *Free Radic. Biol. Med.* **79**, 337–342 (2015).
88. Kim, K.-S., Son, H.-G., Hong, N.-S. & Lee, D.-H. Associations of Serum Ferritin and Transferrin Saturation With All-cause, Cancer, and Cardiovascular Disease Mortality: Third National Health and Nutrition Examination Survey Follow-up Study. *J. Prev. Med. Pub. Health* **45**, 196–203 (2012).
89. Ashmore, J. H., Rogers, C. J., Kelleher, S. L., Lesko, S. M. & Hartman, T. J. Dietary Iron and Colorectal Cancer Risk: A Review of Human Population Studies. *Crit. Rev. Food Sci. Nutr.* **56**, 1012–1020 (2016).
90. Miller, L. D. *et al.* An Iron Regulatory Gene Signature Predicts Outcome in Breast Cancer. *Cancer Res.* **71**, 6728–6737 (2011).
91. Pinnix, Z. K. *et al.* Ferroportin and Iron Regulation in Breast Cancer Progression and Prognosis. *Sci. Transl. Med.* **2**, 43ra56 (2010).
92. Habashy, H. O. *et al.* Transferrin receptor (CD71) is a marker of poor prognosis in breast cancer and can predict response to tamoxifen. *Breast Cancer Res. Treat.* **119**, 283–293 (2009).



93. Jeong, S. M., Hwang, S. & Seong, R. H. Transferrin receptor regulates pancreatic cancer growth by modulating mitochondrial respiration and ROS generation. *Biochem. Biophys. Res. Commun.* **471**, 373–379 (2016).
94. Schonberg, D. L. *et al.* Preferential Iron Trafficking Characterizes Glioblastoma Stem-like Cells. *Cancer Cell* **28**, 441–455 (2015).
95. Högemann-Savellano, D. *et al.* The Transferrin Receptor: A Potential Molecular Imaging Marker for Human Cancer. *Neoplasia* **5**, 495–506 (2003).
96. Isobe, T. *et al.* Human STEAP3 maintains tumor growth under hypoferric condition. *Exp. Cell Res.* **317**, 2582–2591 (2011).
97. Savci-Heijink, C. D., Halfwerk, H., Koster, J. & Vijver, M. J. A novel gene expression signature for bone metastasis in breast carcinomas. *Breast Cancer Res. Treat.* **156**, 249–259 (2016).
98. Tesfay, L. *et al.* Heparin regulation in prostate and its disruption in prostate cancer. *Cancer Res.* **75**, 2254–2263 (2015).
99. Chen, Y. *et al.* Disordered signaling governing ferroportin transcription favors breast cancer growth. *Cell. Signal.* **27**, 168–176 (2015).
100. Zhang, S. *et al.* Disordered hepcidin–ferroportin signaling promotes breast cancer growth. *Cell. Signal.* **26**, 2539–2550 (2014).
101. Marques, O. *et al.* Local iron homeostasis in the breast ductal carcinoma microenvironment. *BMC Cancer* **16**, 187 (2016).
102. Sanders, A. J. *et al.* Genetic upregulation of matriptase-2 reduces the aggressiveness of prostate cancer cells in vitro and in vivo and affects FAK and paxillin localisation. *J. Cell. Physiol.* **216**, 780–789 (2008).

103. Chen, Y. *et al.* Myeloid zinc-finger 1 (MZF-1) suppresses prostate tumor growth through enforcing ferroportin-conducted iron egress. *Oncogene* **34**, 3839–3847 (2015).
104. Calzolari, A. *et al.* Transferrin Receptor 2 Is Frequently and Highly Expressed in Glioblastomas. *Transl. Oncol.* **3**, 123–134 (2010).
105. Ward, R. J., Zucca, F. A., Duyn, J. H., Crichton, R. R. & Zecca, L. The role of iron in brain ageing and neurodegenerative disorders. *Lancet Neurol.* **13**, 1045–1060 (2014).
106. Simpson, I. A. *et al.* A Novel Model for Brain Iron Uptake: Introducing the Concept of Regulation. *J. Cereb. Blood Flow Metab.* **35**, 48–57 (2015).
107. McCarthy, R. C. & Kosman, D. J. Mechanisms and regulation of iron trafficking across the capillary endothelial cells of the blood-brain barrier. *Front. Mol. Neurosci.* **8**, 31 (2015).
108. Nielsen, J. E., Jensen, L. N. & Krabbe, K. Hereditary haemochromatosis: a case of iron accumulation in the basal ganglia associated with a parkinsonian syndrome. *J. Neurol. Neurosurg. Psychiatry* **59**, 318–321 (1995).
109. Dekker, M. C. J. *et al.* Mutations in the hemochromatosis gene (HFE), Parkinson's disease and parkinsonism. *Neurosci. Lett.* **348**, 117–119 (2003).
110. Wang, X.-S. *et al.* Increased incidence of the Hfe mutation in amyotrophic lateral sclerosis and related cellular consequences. *J. Neurol. Sci.* **227**, 27–33 (2004).
111. Bucossi, S. *et al.* Association of K832R and R952K SNPs of Wilson's disease gene with Alzheimer's disease. *J. Alzheimers Dis. JAD* **29**, 913–919 (2012).

112. Kong, S. M. Y. *et al.* Parkinson's disease-linked human PARK9/ATP13A2 maintains zinc homeostasis and promotes  $\alpha$ -Synuclein externalization via exosomes. *Hum. Mol. Genet.* **23**, 2816–2833 (2014).
113. Kell, D. B. Towards a unifying, systems biology understanding of large-scale cellular death and destruction caused by poorly liganded iron: Parkinson's, Huntington's, Alzheimer's, prions, bactericides, chemical toxicology and others as examples. *Arch. Toxicol.* **84**, 825–889 (2010).
114. Rouault, T. A. Iron metabolism in the CNS: implications for neurodegenerative diseases. *Nat. Rev. Neurosci.* **14**, 551–564 (2013).
115. Singh, N., Das, D., Singh, A. & Mohan, M. L. Prion protein and metal interaction: physiological and pathological implications. *Curr. Issues Mol. Biol.* **12**, 99–107 (2010).
116. Singh, A. *et al.* Abnormal Brain Iron Homeostasis in Human and Animal Prion Disorders. *PLOS Pathog* **5**, e1000336 (2009).
117. Dringen, R., Bishop, G. M., Koeppe, M., Dang, T. N. & Robinson, S. R. The Pivotal Role of Astrocytes in the Metabolism of Iron in the Brain. *Neurochem. Res.* **32**, 1884–1890 (2007).
118. Urrutia, P. *et al.* Inflammation alters the expression of DMT1, FPN1 and hepcidin, and it causes iron accumulation in central nervous system cells. *J. Neurochem.* **126**, 541–549 (2013).
119. Raha-Chowdhury, R. *et al.* Expression and cellular localization of hepcidin mRNA and protein in normal rat brain. *BMC Neurosci.* **16**, 24 (2015).
120. Raha, A. A., Vaishnav, R. A., Friedland, R. P., Bomford, A. & Raha-Chowdhury, R. The systemic iron-regulatory proteins hepcidin and

- ferroportin are reduced in the brain in Alzheimer's disease. *Acta Neuropathol. Commun.* **1**, 55 (2013).
121. Wang, J., Jiang, H. & Xie, J.-X. Ferroportin1 and hephaestin are involved in the nigral iron accumulation of 6-OHDA-lesioned rats. *Eur. J. Neurosci.* **25**, 2766–2772 (2007).
122. Halon, M. *et al.* Changes in skeletal muscle iron metabolism outpace amyotrophic lateral sclerosis onset in transgenic rats bearing the G93A hmSOD1 gene mutation. *Free Radic. Res.* **48**, 1363–1370 (2014).
123. Zarruk, J. G. *et al.* Expression of iron homeostasis proteins in the spinal cord in experimental autoimmune encephalomyelitis and their implications for iron accumulation. *Neurobiol. Dis.* **81**, 93–107 (2015).
124. Kroner, A. *et al.* TNF and Increased Intracellular Iron Alter Macrophage Polarization to a Detrimental M1 Phenotype in the Injured Spinal Cord. *Neuron* **83**, 1098–1116 (2014).
125. Mehta, V. *et al.* Iron Is a Sensitive Biomarker for Inflammation in Multiple Sclerosis Lesions. *PLOS ONE* **8**, e57573 (2013).
126. Jeong, S. Y. & David, S. Glycosylphosphatidylinositol-anchored Ceruloplasmin Is Required for Iron Efflux from Cells in the Central Nervous System. *J. Biol. Chem.* **278**, 27144–27148 (2003).
127. Ayton, S., Faux, N. G., Bush, A. I. & Alzheimer's Disease Neuroimaging Initiative. Ferritin levels in the cerebrospinal fluid predict Alzheimer's disease outcomes and are regulated by APOE. *Nat. Commun.* **6**, 6760 (2015).
128. Baraibar, M. A., Barbeito, A. G., Muhoberac, B. B. & Vidal, R. A mutant light-chain ferritin that causes neurodegeneration has enhanced propensity toward oxidative damage. *Free Radic. Biol. Med.* **52**, 1692–1697 (2012).

129. Biasiotto, G., Lorenzo, D. D., Archetti, S. & Zanella, I. Iron and Neurodegeneration: Is Ferritinophagy the Link? *Mol. Neurobiol.* **53**, 5542–5574 (2016).
130. Stadler, N., Lindner, R. A. & Davies, M. J. Direct Detection and Quantification of Transition Metal Ions in Human Atherosclerotic Plaques: Evidence for the Presence of Elevated Levels of Iron and Copper. *Arterioscler. Thromb. Vasc. Biol.* **24**, 949–954 (2004).
131. Satchell, L. & Leake, D. S. Oxidation of Low-Density Lipoprotein by Iron at Lysosomal pH: Implications for Atherosclerosis. *Biochemistry (Mosc.)* **51**, 3767–3775 (2012).
132. Sullivan, J. L. Do Hemochromatosis Mutations Protect Against Iron-Mediated Atherogenesis? *Circ. Cardiovasc. Genet.* **2**, 652–657 (2009).
133. Robbins, C. S. *et al.* Local proliferation dominates lesional macrophage accumulation in atherosclerosis. *Nat. Med.* **19**, 1166–1172 (2013).
134. Chinetti-Gbaguidi, G., Colin, S. & Staels, B. Macrophage subsets in atherosclerosis. *Nat. Rev. Cardiol.* **12**, 10–17 (2015).
135. Gursel, O. *et al.* Premature Atherosclerosis in Children With  $\beta$ -Thalassemia Major: *J. Pediatr. Hematol. Oncol.* **34**, 630–634 (2012).
136. Cheung, Y. F., Chow, P. C., Chan, G. C. & Ha, S. Y. Carotid intima-media thickness is increased and related to arterial stiffening in patients with beta-thalassaemia major. *Br. J. Haematol.* **135**, 732–734 (2006).
137. Li, J. J. *et al.* Hpcidin Destabilizes Atherosclerotic Plaque Via Overactivating Macrophages After Erythrophagocytosis. *Arterioscler. Thromb. Vasc. Biol.* **32**, 1158–1166 (2012).

138. Kautz, L. *et al.* Testing the Iron Hypothesis in a Mouse Model of Atherosclerosis. *Cell Rep.* **5**, 1436–1442 (2013).
139. Mehta, N. U. *et al.* Apolipoprotein E<sup>-/-</sup> Mice Lacking Hemopexin Develop Increased Atherosclerosis via Mechanisms That Include Oxidative Stress and Altered Macrophage Function. *Arterioscler. Thromb. Vasc. Biol.* **36**, 1152–1163 (2016).
140. Boyle, J. J. *et al.* Activating Transcription Factor 1 Directs Mhem Atheroprotective Macrophages Through Coordinated Iron Handling and Foam Cell Protection. *Circ. Res.* **110**, 20–33 (2012).
141. Bories, G. *et al.* Liver X Receptor Activation Stimulates Iron Export in Human Alternative Macrophages. *Circ. Res.* **113**, 1196–1205 (2013).
142. Fernández-Real, J. M., McClain, D. & Manco, M. Mechanisms Linking Glucose Homeostasis and Iron Metabolism Toward the Onset and Progression of Type 2 Diabetes. *Diabetes Care* **38**, 2169–2176 (2015).
143. Kim, B.-J. *et al.* Iron overload accelerates bone loss in healthy postmenopausal women and middle-aged men: A 3-year retrospective longitudinal study. *J. Bone Miner. Res.* **27**, 2279–2290 (2012).
144. Camacho, A. *et al.* Iron overload in a murine model of hereditary hemochromatosis is associated with accelerated progression of osteoarthritis under mechanical stress. *Osteoarthritis Cartilage* **24**, 494–502 (2016).
145. Dunaief, J. L. *et al.* Macular Degeneration in a Patient with Aceruloplasminemia, a Disease Associated with Retinal Iron Overload. *Ophthalmology* **112**, 1062–1065 (2005).

146. Fargion, S., Valenti, L. & Fracanzani, A. L. Beyond hereditary hemochromatosis: New insights into the relationship between iron overload and chronic liver diseases. *Dig. Liver Dis.* **43**, 89–95 (2011).
147. Keberle, H. The Biochemistry of Desferrioxamine and its Relation to Iron Metabolism. *Ann. N. Y. Acad. Sci.* **119**, 758–768 (1964).
148. Borgna-Pignatti, C. *et al.* Survival and complications in patients with thalassemia major treated with transfusion and deferoxamine. *Haematologica* **89**, 1187–1193 (2004).
149. Olivieri, N. F. *et al.* Iron-Chelation Therapy with Oral Deferiprone in Patients with Thalassemia Major. *N. Engl. J. Med.* **332**, 918–922 (1995).
150. Nick, H. *et al.* Development of Tridentate Iron Chelators: From Desferrithiocin to ICL670. *Curr. Med. Chem.* **10**, 1065–1076 (2003).
151. Porter, J. *et al.* Health-Related Quality of Life, Treatment Satisfaction, Adherence and Persistence in  $\beta$ -Thalassemia and Myelodysplastic Syndrome Patients with Iron Overload Receiving Deferasirox: Results from the EPIC Clinical Trial. *Anemia* **2012**, e297641 (2012).
152. Borgna-Pignatti, C. & Marsella, M. Iron Chelation in Thalassemia Major. *Clin. Ther.* **37**, 2866–2877 (2015).
153. Li, H. *et al.* Transferrin therapy ameliorates disease in  $\beta$ -thalassemic mice. *Nat. Med.* **16**, 177–182 (2010).
154. Donfrancesco, A. *et al.* Effects of a Single Course of Deferoxamine in Neuroblastoma Patients. *Cancer Res.* **50**, 4929–4930 (1990).
155. Donfrancesco, A. *et al.* Deferoxamine followed by cyclophosphamide, etoposide, carboplatin, thiotepa, induction regimen in advanced neuroblastoma: Preliminary results. *Eur. J. Cancer* **31**, 612–615 (1995).

156. Kovacevic, Z., Chikhani, S., Lovejoy, D. B. & Richardson, D. R. Novel Thiosemicarbazone Iron Chelators Induce Up-Regulation and Phosphorylation of the Metastasis Suppressor N-myc Down-Stream Regulated Gene 1: A New Strategy for the Treatment of Pancreatic Cancer. *Mol. Pharmacol.* **80**, 598–609 (2011).
157. Lynch, S. G., Peters, K. & LeVine, S. M. Desferrioxamine in chronic progressive Multiple Sclerosis: a pilot study. *Mult. Scler.* **2**, 157–160 (1996).
158. McLachlan, D. R. C. *et al.* Intramuscular desferrioxamine in patients with Alzheimer's disease. *The Lancet* **337**, 1304–1308 (1991).
159. Devos, D. *et al.* Targeting Chelatable Iron as a Therapeutic Modality in Parkinson's Disease. *Antioxid. Redox Signal.* **21**, 195–210 (2013).
160. Preza, G. C. *et al.* Minihepcidins are rationally designed small peptides that mimic hepcidin activity in mice and may be useful for the treatment of iron overload. *J. Clin. Invest.* **121**, 4880–4888 (2011).
161. Casu, C. *et al.* Minihepcidin peptides as disease modifiers in mice affected by  $\beta$ -thalassemia and polycythemia vera. *Blood* **128**, 265–276 (2016).
162. Ramos, E. *et al.* Minihepcidins prevent iron overload in a hepcidin-deficient mouse model of severe hemochromatosis. *Blood* **120**, 3829–3836 (2012).
163. Vadhan-Raj, S. *et al.* Phase 1 Study of a Hepcidin Antagonist, LY2787106, in Cancer-Associated Anemia. *Blood* **126**, 537–537 (2015).
164. Cooke, K. S. *et al.* A fully human anti-hepcidin antibody modulates iron metabolism in both mice and nonhuman primates. *Blood* **122**, 3054–3061 (2013).



165. Hohlbaum, A. *et al.* Iron mobilization and pharmacodynamic marker measurements in non-human primates following administration of PRS-080, a novel and highly specific anti-hepcidin therapeutic. *Am. J. Hematol.* **88**, E41 (2013).
166. van Eijk, L. T. *et al.* Effect of the antihepcidin Spiegelmer lexaptetid on inflammation-induced decrease in serum iron in humans. *Blood* **124**, 2643–2646 (2014).
167. Boyce, M. *et al.* Safety, pharmacokinetics and pharmacodynamics of the anti-hepcidin Spiegelmer lexaptetid pegol in healthy subjects. *Br. J. Pharmacol.* **173**, 1580–1588 (2016).
168. Le Gac, G. *et al.* Structure-Function Analysis of the Human Ferroportin Iron Exporter (SLC40A1): Effect of Hemochromatosis Type 4 Disease Mutations and Identification of Critical Residues. *Hum. Mutat.* **34**, 1371–1380 (2013).
169. Taniguchi, R. *et al.* Outward- and inward-facing structures of a putative bacterial transition-metal transporter with homology to ferroportin. *Nat. Commun.* **6**, 8545 (2015).
170. Fernandes, A. *et al.* The molecular basis of hepcidin-resistant hereditary hemochromatosis. *Blood* **114**, 437–443 (2009).
171. Fung, E. *et al.* High-Throughput Screening of Small Molecules Identifies Heparin Antagonists. *Mol. Pharmacol.* **83**, 681–690 (2013).
172. Leung, D. *et al.* LY2928057, An Antibody Targeting Ferroportin, Is a Potent Inhibitor Of Heparin Activity and Increases Iron Mobilization In Normal Cynomolgus Monkeys. *Blood* **122**, 3433–3433 (2013).

173. Andriopoulos Jr, B. *et al.* BMP6 is a key endogenous regulator of hepcidin expression and iron metabolism. *Nat. Genet.* **41**, 482–487 (2009).
174. Poli, M. *et al.* Glycol-split nonanticoagulant heparins are inhibitors of hepcidin expression in vitro and in vivo. *Blood* **123**, 1564–1573 (2014).
175. Yu, P. B. *et al.* Dorsomorphin inhibits BMP signals required for embryogenesis and iron metabolism. *Nat. Chem. Biol.* **4**, 33–41 (2008).
176. Cuny, G. D. *et al.* Structure–activity relationship study of bone morphogenetic protein (BMP) signaling inhibitors. *Bioorg. Med. Chem. Lett.* **18**, 4388–4392 (2008).
177. Akinc, A. *et al.* Targeting the Hepcidin Pathway with RNAi Therapeutics for the Treatment of Anemia. *Blood* **118**, 688–688 (2011).
178. Babitt, J. L. *et al.* Bone morphogenetic protein signaling by hemojuvelin regulates hepcidin expression. *Nat. Genet.* **38**, 531–539 (2006).
179. Silvestri, L. *et al.* The Serine Protease Matriptase-2 (TMPRSS6) Inhibits Hepcidin Activation by Cleaving Membrane Hemojuvelin. *Cell Metab.* **8**, 502–511 (2008).
180. Du, X. *et al.* The Serine Protease TMPRSS6 Is Required to Sense Iron Deficiency. *Science* **320**, 1088–1092 (2008).
181. Maurer, E. *et al.* Insights into Matriptase-2 Substrate Binding and Inhibition Mechanisms by Analyzing Active-Site-Mutated Variants. *ChemMedChem* **7**, 68–72 (2012).
182. Sisay, M. T. *et al.* Identification of the First Low-Molecular-Weight Inhibitors of Matriptase-2. *J. Med. Chem.* **53**, 5523–5535 (2010).
183. Guo, S. *et al.* Reducing TMPRSS6 ameliorates hemochromatosis and  $\beta$ -thalassemia in mice. *J. Clin. Invest.* **123**, 1531–1541 (2013).

184. Casu, C. *et al.* Combination of Tmprss6- ASO and the iron chelator deferiprone improves erythropoiesis and reduces iron overload in a mouse model of beta-thalassemia intermedia. *Haematologica* **101**, e8–e11 (2016).
185. Schmidt, P. J. *et al.* An RNAi therapeutic targeting Tmprss6 decreases iron overload in Hfe<sup>-/-</sup> mice and ameliorates anemia and iron overload in murine  $\beta$ -thalassemia intermedia. *Blood* **121**, 1200–1208 (2013).
186. Béliveau, F., Désilets, A. & Leduc, R. Probing the substrate specificities of matriptase, matriptase-2, hepsin and DESC1 with internally quenched fluorescent peptides. *FEBS J.* **276**, 2213–2226 (2009).
187. Beckmann, A.-M. *et al.* En Route to New Therapeutic Options for Iron Overload Diseases: Matriptase-2 as a Target for Kunitz-Type Inhibitors. *ChemBioChem* **17**, 595–604 (2016).
188. Zhen, A. W. *et al.* The small molecule, genistein, increases hepcidin expression in human hepatocytes. *Hepatology* **58**, 1315–1325 (2013).
189. Chung, B., Matak, P., McKie, A. T. & Sharp, P. Leptin Increases the Expression of the Iron Regulatory Hormone Hepcidin in HuH7 Human Hepatoma Cells. *J. Nutr.* **137**, 2366–2370 (2007).
190. Zhang, S.-P. AG490: An inhibitor of hepcidin expression *in vivo*. *World J. Gastroenterol.* **17**, 5032 (2011).
191. Fatih, N. *et al.* Natural and synthetic STAT3 inhibitors reduce hepcidin expression in differentiated mouse hepatocytes expressing the active phosphorylated STAT3 form. *J. Mol. Med.* **88**, 477–486 (2010).
192. Czachorowski, M., Lam-Yuk-Tseung, S., Cellier, M. & Gros, P. Transmembrane Topology of the Mammalian Slc11a2 Iron Transporter. *Biochemistry (Mosc.)* **48**, 8422–8434 (2009).

193. Wang, D., Song, Y., Li, J., Wang, C. & Li, F. Structure and metal ion binding of the first transmembrane domain of DMT1. *Biochim. Biophys. Acta BBA - Biomembr.* **1808**, 1639–1644 (2011).
194. Ehrnstorfer, I. A., Geertsma, E. R., Pardon, E., Steyaert, J. & Dutzler, R. Crystal structure of a SLC11 (NRAMP) transporter reveals the basis for transition-metal ion transport. *Nat. Struct. Mol. Biol.* **21**, 990–996 (2014).
195. Buckett, P. D. & Wessling-Resnick, M. Small molecule inhibitors of divalent metal transporter-1. *Am. J. Physiol. - Gastrointest. Liver Physiol.* **296**, G798–G804 (2009).
196. Horonchik, L. & Wessling-Resnick, M. The Small-Molecule Iron Transport Inhibitor Ferristatin/NSC306711 Promotes Degradation of the Transferrin Receptor. *Chem. Biol.* **15**, 647–653 (2008).
197. Byrne, S. L. *et al.* Ferristatin II Promotes Degradation of Transferrin Receptor-1 In Vitro and In Vivo. *PLOS ONE* **8**, e70199 (2013).
198. Yanatori, I., Yasui, Y., Noguchi, Y. & Kishi, F. Inhibition of iron uptake by ferristatin II is exerted through internalization of DMT1 at the plasma membrane. *Cell Biol. Int.* **39**, 427–434 (2015).
199. Alkhateeb, A. A. *et al.* The small molecule ferristatin II induces hepatic hepcidin expression in vivo and in vitro. *Am. J. Physiol. - Gastrointest. Liver Physiol.* **308**, G1019–G1026 (2015).
200. IARC Working Group on the Evaluation of Carcinogenic Risks to Humans. *In A review of human carcinogens. Part F: Chemical agents and related occupations* **100F**, 53–64 (World Health Organization, 2012).

201. Wetli, H. A., Buckett, P. D. & Wessling-Resnick, M. Small-Molecule Screening Identifies the Selenazal Drug Ebselen as a Potent Inhibitor of DMT1-Mediated Iron Uptake. *Chem. Biol.* **13**, 965–972 (2006).
202. Davis, M. T. & Bartfay, W. J. Ebselen Decreases Oxygen Free Radical Production and Iron Concentrations in the Hearts of Chronically Iron-Overloaded Mice. *Biol. Res. Nurs.* **6**, 37–45 (2004).
203. Cadieux, J. A. *et al.* Synthesis and biological evaluation of substituted pyrazoles as blockers of divalent metal transporter 1 (DMT1). *Bioorg. Med. Chem. Lett.* **22**, 90–95 (2012).
204. Zhang, Z. *et al.* Discovery of benzylisothioureas as potent divalent metal transporter 1 (DMT1) inhibitors. *Bioorg. Med. Chem. Lett.* **22**, 5108–5113 (2012).
205. Montalbetti, N. *et al.* Discovery and characterization of a novel non-competitive inhibitor of the divalent metal transporter DMT1/SLC11A2. *Biochem. Pharmacol.* **96**, 216–224 (2015).
206. Span, K. *et al.* A novel oral iron-complex formulation: Encapsulation of hemin in polymeric micelles and its in vitro absorption. *Eur. J. Pharm. Biopharm.* **108**, 226–234 (2016).
207. Latunde-Dada, G. O. *et al.* A Nanoparticulate Ferritin-Core Mimetic Is Well Taken Up by HuTu 80 Duodenal Cells and Its Absorption in Mice Is Regulated by Body Iron. *J. Nutr.* **144**, 1896–1902 (2014).
208. Taylor, K. M., Morgan, H. E., Johnson, A. & Nicholson, R. I. Structure–function analysis of a novel member of the LIV-1 subfamily of zinc transporters, ZIP14. *FEBS Lett.* **579**, 427–432 (2005).

209. Peyssonnaud, C. *et al.* Regulation of iron homeostasis by the hypoxia-inducible transcription factors (HIFs). *J. Clin. Invest.* **117**, 1926–1932 (2007).
210. Qian, Z.-M. *et al.* Divalent metal transporter 1 is a hypoxia-inducible gene. *J. Cell. Physiol.* **226**, 1596–1603 (2011).
211. Shah, Y. M., Matsubara, T., Ito, S., Yim, S.-H. & Gonzalez, F. J. Intestinal Hypoxia-Inducible Transcription Factors Are Essential for Iron Absorption following Iron Deficiency. *Cell Metab.* **9**, 152–164 (2009).
212. Tacchini, L., Bianchi, L., Bernelli-Zazzera, A. & Cairo, G. Transferrin receptor induction by hypoxia. HIF-1-mediated transcriptional activation and cell-specific post-transcriptional regulation. *J. Biol. Chem.* **274**, 24142–24146 (1999).
213. Taylor, M. *et al.* Hypoxia-Inducible Factor-2 $\alpha$  Mediates the Adaptive Increase of Intestinal Ferroportin During Iron Deficiency in Mice. *Gastroenterology* **140**, 2044–2055 (2011).
214. Scheuermann, T. H. *et al.* Allosteric inhibition of hypoxia inducible factor-2 with small molecules. *Nat. Chem. Biol.* **9**, 271–276 (2013).
215. Scheuermann, T. H. *et al.* Isoform-Selective and Stereoselective Inhibition of Hypoxia Inducible Factor-2. *J. Med. Chem.* **58**, 5930–5941 (2015).
216. Bruick, R. K. & McKnight, S. L. A Conserved Family of Prolyl-4-Hydroxylases That Modify HIF. *Science* **294**, 1337–1340 (2001).
217. Epstein, A. C. R. *et al.* *C. elegans* EGL-9 and Mammalian Homologs Define a Family of Dioxygenases that Regulate HIF by Prolyl Hydroxylation. *Cell* **107**, 43–54 (2001).

218. McDonough, M. A. *et al.* Cellular oxygen sensing: Crystal structure of hypoxia-inducible factor prolyl hydroxylase (PHD2). *Proc. Natl. Acad. Sci.* **103**, 9814–9819 (2006).
219. Maxwell, P. H. & Eckardt, K.-U. HIF prolyl hydroxylase inhibitors for the treatment of renal anaemia and beyond. *Nat. Rev. Nephrol.* **12**, 157–168 (2016).
220. Barrett, T. D. *et al.* Prolyl hydroxylase inhibition corrects functional iron deficiency and inflammation-induced anaemia in rats. *Br. J. Pharmacol.* **172**, 4078–4088 (2015).
221. Rajagopalan, S., Rane, A., Chinta, S. J. & Andersen, J. K. Regulation of ATP13A2 via PHD2-HIF1 $\alpha$  Signaling Is Critical for Cellular Iron Homeostasis: Implications for Parkinson's Disease. *J. Neurosci.* **36**, 1086–1095 (2016).
222. Provenzano, R. *et al.* Roxadustat (FG-4592) Versus Epoetin Alfa for Anemia in Patients Receiving Maintenance Hemodialysis: A Phase 2, Randomized, 6- to 19-Week, Open-Label, Active-Comparator, Dose-Ranging, Safety and Exploratory Efficacy Study. *Am. J. Kidney Dis.* **67**, 912–924 (2016).
223. Brigandi, R. A. *et al.* A Novel Hypoxia-Inducible Factor–Prolyl Hydroxylase Inhibitor (GSK1278863) for Anemia in CKD: A 28-Day, Phase 2A Randomized Trial. *Am. J. Kidney Dis.* **67**, 861–871 (2016).
224. Hartman, C. S. *et al.* AKB-6548, a new hypoxia-inducible factor prolyl hydroxylase inhibitor increases hemoglobin while decreasing ferritin in a 28-day, phase 2a dose escalation study in stage 3 and 4 chronic kidney disease patients with anemia. in *Proceedings of the American Society of Nephrology Kidney Week* (2011).

225. Boettcher, M. F. *et al.* First-in-man study with BAY 85-3934—a new oral selective HIF-PH inhibitor for the treatment of renal anemia. *J Am Soc Nephrol* **24**, 347A (2013).
226. Akizawa, T., Hanaki, K. & Arai, M. JTZ-951, an Oral Novel Hif-Phd Inhibitor, Elevates Hemoglobin in Japanese Anemic Patients with Chronic Kidney Disease Not on Dialysis. *Nephrol. Dial. Transplant.* **30**, iii196 (2015).
227. Jain, M. *et al.* Pharmacological Characterization of ZYAN1, a Novel Prolyl Hydroxylase Inhibitor for the Treatment of Anemia. *Drug Res.* **66**, 107–112 (2015).
228. Salvati, A. *et al.* Transferrin-functionalized nanoparticles lose their targeting capabilities when a biomolecule corona adsorbs on the surface. *Nat. Nanotechnol.* **8**, 137–143 (2013).
229. Singh, R. *et al.* Dose-Dependent Therapeutic Distinction between Active and Passive Targeting Revealed Using Transferrin-Coated PGMA Nanoparticles. *Small* **12**, 351–359 (2016).
230. van der Meel, R., Vehmeijer, L. J. C., Kok, R. J., Storm, G. & van Gaal, E. V. B. Ligand-targeted particulate nanomedicines undergoing clinical evaluation: Current status. *Adv. Drug Deliv. Rev.* **65**, 1284–1298 (2013).
231. Senzer, N. *et al.* Phase I Study of a Systemically Delivered p53 Nanoparticle in Advanced Solid Tumors. *Mol. Ther.* **21**, 1096–1103 (2013).
232. Zhang, H. *et al.* Transferrin-mediated fullerenes nanoparticles as Fe<sup>2+</sup>-dependent drug vehicles for synergistic anti-tumor efficacy. *Biomaterials* **37**, 353–366 (2015).



233. O'Neill, P. M., Barton, V. E. & Ward, S. A. The Molecular Mechanism of Action of Artemisinin—The Debate Continues. *Molecules* **15**, 1705–1721 (2010).
234. Wiley, D. T., Webster, P., Gale, A. & Davis, M. E. Transcytosis and brain uptake of transferrin-containing nanoparticles by tuning avidity to transferrin receptor. *Proc. Natl. Acad. Sci.* **110**, 8662–8667 (2013).
235. Sehlin, D. *et al.* Antibody-based PET imaging of amyloid beta in mouse models of Alzheimer's disease. *Nat. Commun.* **7**, 10759 (2016).
236. Jutz, G., van Rijn, P., Santos Miranda, B. & Böker, A. Ferritin: A Versatile Building Block for Bionanotechnology. *Chem. Rev.* **115**, 1653–1701 (2015).
237. Crich, S. G. *et al.* Targeting ferritin receptors for the selective delivery of imaging and therapeutic agents to breast cancer cells. *Nanoscale* **7**, 6527–6533 (2015).
238. Lei, Y. *et al.* Targeted tumor delivery and controlled release of neuronal drugs with ferritin nanoparticles to regulate pancreatic cancer progression. *J. Controlled Release* **232**, 131–142 (2016).
239. Hu, H. *et al.* The M2 phenotype of tumor-associated macrophages in the stroma confers a poor prognosis in pancreatic cancer. *Tumor Biol.* **37**, 8657–8664 (2016).
240. Van Gorp, H., Delputte, P. L. & Nauwynck, H. J. Scavenger receptor CD163, a Jack-of-all-trades and potential target for cell-directed therapy. *Mol. Immunol.* **47**, 1650–1660 (2010).
241. Etzerodt, A. *et al.* Efficient intracellular drug-targeting of macrophages using stealth liposomes directed to the hemoglobin scavenger receptor CD163. *J. Controlled Release* **160**, 72–80 (2012).

242. Amin, M. L., Kim, D. & Kim, S. Development of hematin conjugated PLGA nanoparticle for selective cancer targeting. *Eur. J. Pharm. Sci.* **91**, 138–143 (2016).
243. Takle, G. B. *et al.* Delivery of Oligoribonucleotides to Human Hepatoma Cells Using Cationic Lipid Particles Conjugated to Ferric Protoporphyrin IX (Heme). *Antisense Nucleic Acid Drug Dev.* **7**, 177–185 (1997).
244. Kell, D. B. Iron behaving badly: inappropriate iron chelation as a major contributor to the aetiology of vascular and other progressive inflammatory and degenerative diseases. *BMC Med. Genomics* **2**, 2 (2009).
245. Nairz, M. *et al.* ‘Ride on the ferrous wheel’ – The cycle of iron in macrophages in health and disease. *Immunobiology* **220**, 280–294 (2015).
246. Ganz, T. & Nemeth, E. Iron homeostasis in host defence and inflammation. *Nat. Rev. Immunol.* **15**, 500–510 (2015).
247. Miron, V. E. *et al.* M2 microglia and macrophages drive oligodendrocyte differentiation during CNS remyelination. *Nat. Neurosci.* **16**, 1211–1218 (2013).
248. Biswas, S. K., Chittechath, M., Shalova, I. N. & Lim, J.-Y. Macrophage polarization and plasticity in health and disease. *Immunol. Res.* **53**, 11–24 (2012).
249. Bronte, V. & Murray, P. J. Understanding Local Macrophage Phenotypes In Disease: Modulating macrophage function to treat cancer. *Nat. Med.* **21**, 117–119 (2015).
250. Crielaard, B. J. *et al.* Macrophages and liposomes in inflammatory disease: Friends or foes? *Int. J. Pharm.* **416**, 499–506 (2011).

251. Kiessling, F., Mertens, M. E., Grimm, J. & Lammers, T. Nanoparticles for Imaging: Top or Flop? *Radiology* **273**, 10–28 (2014).
252. Lartigue, L. *et al.* Biodegradation of Iron Oxide Nanocubes: High-Resolution In Situ Monitoring. *ACS Nano* **7**, 3939–3952 (2013).
253. Daldrup-Link, H. E. *et al.* MRI of tumor-associated macrophages with clinically applicable iron oxide nanoparticles. *Clin. Cancer Res. Off. J. Am. Assoc. Cancer Res.* **17**, 5695–5704 (2011).
254. Hervault, A. *et al.* Doxorubicin loaded dual pH- and thermo-responsive magnetic nanocarrier for combined magnetic hyperthermia and targeted controlled drug delivery applications. *Nanoscale* **8**, 12152–12161 (2016).
255. Xian-hui, D. *et al.* Age-related changes of brain iron load changes in the frontal cortex in APP<sup>swe</sup>/PS1 $\Delta$ E9 transgenic mouse model of Alzheimer's disease. *J. Trace Elem. Med. Biol.* **30**, 118–123 (2015).
256. Salazar, J. *et al.* Divalent metal transporter 1 (DMT1) contributes to neurodegeneration in animal models of Parkinson's disease. *Proc. Natl. Acad. Sci.* **105**, 18578–18583 (2008).
257. Brookes, M. J. *et al.* Modulation of iron transport proteins in human colorectal carcinogenesis. *Gut* **55**, 1449–1460 (2006).
258. Hulet, S. W., Powers, S. & Connor, J. R. Distribution of transferrin and ferritin binding in normal and multiple sclerotic human brains. *J. Neurol. Sci.* **165**, 48–55 (1999).
259. Li, W., Xu, L.-H., Forssell, C., Sullivan, J. L. & Yuan, X.-M. Overexpression of Transferrin Receptor and Ferritin Related to Clinical Symptoms and Destabilization of Human Carotid Plaques. *Exp. Biol. Med.* **233**, 818–826 (2008).

260. Jefferies, W. A. *et al.* Reactive microglia specifically associated with amyloid plaques in Alzheimer's disease brain tissue express melanotransferrin. *Brain Res.* **712**, 122–126 (1996).
261. Kalaria, R. N., Sromek, S. M., Grahovac, I. & Harik, S. I. Transferrin receptors of rat and human brain and cerebral microvessels and their status in Alzheimer's disease. *Brain Res.* **585**, 87–93 (1992).
262. Faucheux, B. A. *et al.* Distribution of 125I-Ferrotransferrin Binding Sites in the Mesencephalon of Control Subjects and Patients with Parkinson's Disease. *J. Neurochem.* **60**, 2338–2341 (1993).
263. Morris, C. M., Candy, J. M., Omar, S., Bloxham, C. A. & Edwardson, J. A. Transferrin receptors in the Parkinsonian midbrain. *Neuropathol. Appl. Neurobiol.* **20**, 468–472 (1994).
264. Visanji, N. P. *et al.* Iron deficiency in parkinsonism: region-specific iron dysregulation in Parkinson's disease and multiple system atrophy. *J. Park. Dis.* **3**, 523–537 (2013).
265. Zhang, Z. *et al.* Parenchymal accumulation of CD163+ macrophages/microglia in multiple sclerosis brains. *J. Neuroimmunol.* **237**, 73–79 (2011).
266. Pey, P., Pearce, R. K., Kalaitzakis, M. E., Griffin, W. S. T. & Gentleman, S. M. Phenotypic profile of alternative activation marker CD163 is different in Alzheimer's and Parkinson's disease. *Acta Neuropathol. Commun.* **2**, 21 (2014).
267. Boyle, J. J. *et al.* Coronary Intraplaque Hemorrhage Evokes a Novel Atheroprotective Macrophage Phenotype. *Am. J. Pathol.* **174**, 1097–1108 (2009).

268. Tiainen, S. *et al.* High numbers of macrophages, especially M2-like (CD163-positive), correlate with hyaluronan accumulation and poor outcome in breast cancer. *Histopathology* **66**, 873–883 (2015).
269. Lundholm, M. *et al.* Secreted Factors from Colorectal and Prostate Cancer Cells Skew the Immune Response in Opposite Directions. *Sci. Rep.* **5**, 15651 (2015).
270. Mignogna, C. *et al.* A reappraisal of macrophage polarization in glioblastoma: Histopathological and immunohistochemical findings and review of the literature. *Pathol. - Res. Pract.* **212**, 491–499 (2016).
271. LeVine, S. M. Iron deposits in multiple sclerosis and Alzheimer's disease brains. *Brain Res.* **760**, 298–303 (1997).
272. Connor, J. R., Menzies, S. L., St. Martin, S. M. & Mufson, E. J. A histochemical study of iron, transferrin, and ferritin in Alzheimer's diseased brains. *J. Neurosci. Res.* **31**, 75–83 (1992).
273. Ji, J. *et al.* Low expression of ferroxidases is implicated in the iron retention in human atherosclerotic plaques. *Biochem. Biophys. Res. Commun.* **464**, 1134–1138 (2015).
274. Loeffler, D. A. *et al.* Transferrin and Iron in Normal, Alzheimer's Disease, and Parkinson's Disease Brain Regions. *J. Neurochem.* **65**, 710–716 (1995).
275. Galesloot, T. E. *et al.* Serum hepcidin is associated with presence of plaque in postmenopausal women of a general population. *Arterioscler. Thromb. Vasc. Biol.* **34**, 446–456 (2014).
276. Orlandi, R. *et al.* Heparin and ferritin blood level as noninvasive tools for predicting breast cancer. *Ann. Oncol.* **25**, 352–357 (2014).

277. LeVine, S. M. *et al.* Ferritin, transferrin and iron concentrations in the cerebrospinal fluid of multiple sclerosis patients. *Brain Res.* **821**, 511–515 (1999).
278. Sato, Y. *et al.* Cerebrospinal fluid ferritin in glioblastoma: Evidence for tumor synthesis. *J. Neurooncol.* **40**, 47–50 (1998).
279. Sung, K.-C. *et al.* Ferritin Is Independently Associated With the Presence of Coronary Artery Calcium in 12 033 Men. *Arterioscler. Thromb. Vasc. Biol.* **32**, 2525–2530 (2012).
280. Chua, A. C., Knuiman, M. W., Trinder, D., Divitini, M. L. & Olynyk, J. K. Higher concentrations of serum iron and transferrin saturation but not serum ferritin are associated with cancer outcomes. *Am. J. Clin. Nutr.* **104**, 736–742 (2016).
281. Wang, R., Liu, Z. & Yan, L. Serum iron levels and Parkinson’s disease risk: evidence from a meta-analysis. *Int. J. Clin. Exp. Med.* **9**, 3167–3172 (2016).
282. Valberg, L. S., Flanagan, P. R., Kertesz, A. & Ebers, G. C. Abnormalities in Iron Metabolism in Multiple Sclerosis. *Can. J. Neurol. Sci.* **16**, 184–186 (1989).
283. Faux, N. G. *et al.* An anemia of Alzheimer’s disease. *Mol. Psychiatry* **19**, 1227–1234 (2014).
284. Hare, D. J. *et al.* Decreased Plasma Iron in Alzheimer’s Disease Is Due to Transferrin Desaturation. *ACS Chem. Neurosci.* **6**, 398–402 (2015).
285. Meynard, D. *et al.* Lack of the bone morphogenetic protein BMP6 induces massive iron overload. *Nat. Genet.* **41**, 478–481 (2009).
286. Feng, Q., Migas, M. C., Waheed, A., Britton, R. S. & Fleming, R. E. Ferritin upregulates hepatic expression of bone morphogenetic protein 6 and

- hepcidin in mice. *Am. J. Physiol. - Gastrointest. Liver Physiol.* **302**, G1397–G1404 (2012).
287. Rausa, M. *et al.* Bmp6 Expression in Murine Liver Non Parenchymal Cells: A Mechanism to Control their High Iron Exporter Activity and Protect Hepatocytes from Iron Overload? *PLOS ONE* **10**, e0122696 (2015).
288. Daher, R. *et al.* Heterozygous Mutations in BMP6 Pro-peptide Lead to Inappropriate Heparin Synthesis and Moderate Iron Overload in Humans. *Gastroenterology* **150**, 672–683.e4 (2016).
289. Kautz, L. *et al.* Identification of erythroferrone as an erythroid regulator of iron metabolism. *Nat. Genet.* **46**, 678–684 (2014).
290. Kautz, L., Jung, G., Nemeth, E. & Ganz, T. Erythroferrone contributes to recovery from anemia of inflammation. *Blood* **124**, 2569–2574 (2014).
291. Montosi, G. *et al.* Autosomal-dominant hemochromatosis is associated with a mutation in the ferroportin (SLC11A3) gene. *J. Clin. Invest.* **108**, 619–623 (2001).
292. Njajou, O. T. *et al.* A mutation in SLC11A3 is associated with autosomal dominant hemochromatosis. *Nat. Genet.* **28**, 213–214 (2001).
293. Wallace, D. F., Clark, R. M., Harley, H. A. J. & Subramaniam, V. N. Autosomal dominant iron overload due to a novel mutation of ferroportin1 associated with parenchymal iron loading and cirrhosis. *J. Hepatol.* **40**, 710–713 (2004).
294. Healey, E. G. *et al.* Repulsive guidance molecule is a structural bridge between neogenin and bone morphogenetic protein. *Nat. Struct. Mol. Biol.* **22**, 458–465 (2015).

295. Papanikolaou, G. *et al.* Mutations in HFE2 cause iron overload in chromosome 1q-linked juvenile hemochromatosis. *Nat. Genet.* **36**, 77–82 (2004).
296. D'Alessio, F., Hentze, M. W. & Muckenthaler, M. U. The hemochromatosis proteins HFE, Tfr2, and HJV form a membrane-associated protein complex for hepcidin regulation. *J. Hepatol.* **57**, 1052–1060 (2012).
297. Roetto, A. *et al.* Mutant antimicrobial peptide hepcidin is associated with severe juvenile hemochromatosis. *Nat. Genet.* **33**, 21 (2003).
298. Vaiopoulos, G. *et al.* Arthropathy in juvenile hemochromatosis. *Arthritis Rheum.* **48**, 227–230 (2003).
299. Feder, J. N. *et al.* A novel MHC class I-like gene is mutated in patients with hereditary haemochromatosis. *Nat. Genet.* **13**, 399–408 (1996).
300. Wallace, D. F. *et al.* Combined deletion of Hfe and transferrin receptor 2 in mice leads to marked dysregulation of hepcidin and iron overload. *Hepatology* **50**, 1992–2000 (2009).
301. Latour, C. *et al.* Differing impact of the deletion of hemochromatosis-associated molecules HFE and transferrin receptor-2 on the iron phenotype of mice lacking bone morphogenetic protein 6 or hemojuvelin. *Hepatology* **63**, 126–137 (2016).
302. Schmidt, P. J., Toran, P. T., Giannetti, A. M., Bjorkman, P. J. & Andrews, N. C. The Transferrin Receptor Modulates Hfe-Dependent Regulation of Hepcidin Expression. *Cell Metab.* **7**, 205–214 (2008).
303. Nemeth, E. *et al.* IL-6 mediates hypoferremia of inflammation by inducing the synthesis of the iron regulatory hormone hepcidin. *J. Clin. Invest.* **113**, 1271–1276 (2004).



304. Boulanger, M. J., Chow, D., Brevnova, E. E. & Garcia, K. C. Hexameric Structure and Assembly of the Interleukin-6/IL-6  $\alpha$ -Receptor/gp130 Complex. *Science* **300**, 2101–2104 (2003).
305. Meynard, D. *et al.* Regulation of TMPRSS6 by BMP6 and iron in human cells and mice. *Blood* **118**, 747–756 (2011).
306. Nai, A. *et al.* Limiting hepatic Bmp-Smad signaling by matriptase-2 is required for erythropoietin-mediated hepcidin suppression in mice. *Blood* blood-2015-11-681494 (2016). doi:10.1182/blood-2015-11-681494
307. Bell, C. H. *et al.* Structure of the Repulsive Guidance Molecule (RGM)–Neogenin Signaling Hub. *Science* **341**, 77–80 (2013).
308. Zhao, N. *et al.* Neogenin Facilitates the Induction of Hepcidin Expression by Hemojuvelin in the Liver. *J. Biol. Chem.* **291**, 12322–12335 (2016).
309. Kawabata, H. *et al.* Molecular Cloning of Transferrin Receptor 2. A new member of the transferrin receptor-like family. *J. Biol. Chem.* **274**, 20826–20832 (1999).
310. Camaschella, C. *et al.* The gene TFR2 is mutated in a new type of haemochromatosis mapping to 7q22. *Nat. Genet.* **25**, 14–15 (2000).
311. Pagani, A. *et al.* Regulation of cell surface transferrin receptor-2 by iron-dependent cleavage and release of a soluble form. *Haematologica* **100**, 458–465 (2015).
312. Jiang, Y., Oliver, P., Davies, K. E. & Platt, N. Identification and Characterization of Murine SCARA5, a Novel Class A Scavenger Receptor That Is Expressed by Populations of Epithelial Cells. *J. Biol. Chem.* **281**, 11834–11845 (2006).

313. Ojala, J. R. M., Pikkarainen, T., Elmberger, G. & Tryggvason, K. Progressive Reactive Lymphoid Connective Tissue Disease and Development of Autoantibodies in Scavenger Receptor A5-Deficient Mice. *Am. J. Pathol.* **182**, 1681–1695 (2013).
314. Herz, J., Clouthier, D. E. & Hammer, R. E. LDL receptor-related protein internalizes and degrades uPA-PAI-1 complexes and is essential for embryo implantation. *Cell* **71**, 411–421 (1992).
315. Overton, C. D., Yancey, P. G., Major, A. S., Linton, M. F. & Fazio, S. Deletion of Macrophage LDL Receptor-Related Protein Increases Atherogenesis in the Mouse. *Circ. Res.* **100**, 670–677 (2007).
316. Philippidis, P. *et al.* Hemoglobin Scavenger Receptor CD163 Mediates Interleukin-10 Release and Heme Oxygenase-1 Synthesis Antiinflammatory Monocyte-Macrophage Responses In Vitro, in Resolving Skin Blisters In Vivo, and After Cardiopulmonary Bypass Surgery. *Circ. Res.* **94**, 119–126 (2004).
317. Vargas, J. D. *et al.* Stromal cell-derived receptor 2 and cytochrome b561 are functional ferric reductases. *Biochim. Biophys. Acta* **1651**, 116–123 (2003).
318. Gunshin, H. *et al.* Cybrd1 (duodenal cytochrome b) is not necessary for dietary iron absorption in mice. *Blood* **106**, 2879–2883 (2005).
319. Choi, J. *et al.* Duodenal Reductase Activity and Spleen Iron Stores Are Reduced and Erythropoiesis Is Abnormal in Dcytb Knockout Mice Exposed to Hypoxic Conditions. *J. Nutr.* **142**, 1929–1934 (2012).
320. Gunshin, H. *et al.* Slc11a2 is required for intestinal iron absorption and erythropoiesis but dispensable in placenta and liver. *J. Clin. Invest.* **115**, 1258–1266 (2005).

321. Kvarnung, M. *et al.* Mutations in FLVCR2 associated with Fowler syndrome and survival beyond infancy. *Clin. Genet.* **89**, 99–103 (2016).
322. Delaby, C. *et al.* Subcellular Localization of Iron and Heme Metabolism Related Proteins at Early Stages of Erythrophagocytosis. *PLOS ONE* **7**, e42199 (2012).
323. Gomes, I. M., Maia, C. J. & Santos, C. R. STEAP Proteins: From Structure to Applications in Cancer Therapy. *Mol. Cancer Res.* **10**, 573–587 (2012).
324. Grandchamp, B. *et al.* A novel type of congenital hypochromic anemia associated with a nonsense mutation in the STEAP3/TSAP6 gene. *Blood* **118**, 6660–6666 (2011).
325. Liu, D. *et al.* Human STEAP3 mutations with no phenotypic red cell changes. *Blood* **127**, 1067–1071 (2016).
326. Knisely, A. S., Gelbart, T. & Beutler, E. Molecular characterization of a third case of human atransferrinemia. *Blood* **104**, 2607–2607 (2004).
327. Levy, J. E., Jin, O., Fujiwara, Y., Kuo, F. & Andrews, N. Transferrin receptor is necessary for development of erythrocytes and the nervous system. *Nat. Genet.* **21**, 396–399 (1999).
328. Hojyo, S. *et al.* The Zinc Transporter SLC39A14/ZIP14 Controls G-Protein Coupled Receptor-Mediated Signaling Required for Systemic Growth. *PLOS ONE* **6**, e18059 (2011).
329. Ji, C. & Kosman, D. J. Molecular mechanisms of non-transferrin-bound and transferrin-bound iron uptake in primary hippocampal neurons. *J. Neurochem.* **133**, 668–683 (2015).

330. Cozzi, A. *et al.* Human L-ferritin deficiency is characterized by idiopathic generalized seizures and atypical restless leg syndrome. *J. Exp. Med.* **210**, 1779–1791 (2013).
331. Poss, K. D. & Tonegawa, S. Heme oxygenase 1 is required for mammalian iron reutilization. *Proc. Natl. Acad. Sci.* **94**, 10919–10924 (1997).
332. Kovtunovych, G., Eckhaus, M. A., Ghosh, M. C., Ollivierre-Wilson, H. & Rouault, T. A. Dysfunction of the heme recycling system in heme oxygenase 1-deficient mice: effects on macrophage viability and tissue iron distribution. *Blood* **116**, 6054–6062 (2010).
333. Kawashima, A., Oda, Y., Yachie, A., Koizumi, S. & Nakanishi, I. Heme oxygenase-1 deficiency: The first autopsy case. *Hum. Pathol.* **33**, 125–130 (2002).
334. Scortegagna, M. *et al.* Multiple organ pathology, metabolic abnormalities and impaired homeostasis of reactive oxygen species in *Epas1*<sup>-/-</sup> mice. *Nat. Genet.* **35**, 331–340 (2003).
335. Zhuang, Z. *et al.* Somatic HIF2A Gain-of-Function Mutations in Paraganglioma with Polycythemia. *N. Engl. J. Med.* **367**, 922–930 (2012).
336. Sanchez, M., Galy, B., Muckenthaler, M. U. & Hentze, M. W. Iron-regulatory proteins limit hypoxia-inducible factor-2 $\alpha$  expression in iron deficiency. *Nat. Struct. Mol. Biol.* **14**, 420–426 (2007).
337. Troadec, M.-B. *et al.* Targeted deletion of the mouse Mitoferrin1 gene: from anemia to protoporphyria. *Blood* **117**, 5494–5502 (2011).
338. Percy, M. J. *et al.* A family with erythrocytosis establishes a role for prolyl hydroxylase domain protein 2 in oxygen homeostasis. *Proc. Natl. Acad. Sci. U. S. A.* **103**, 654–659 (2006).

339. Takeda, K. *et al.* Placental but Not Heart Defects Are Associated with Elevated Hypoxia-Inducible Factor  $\alpha$  Levels in Mice Lacking Prolyl Hydroxylase Domain Protein 2. *Mol. Cell. Biol.* **26**, 8336–8346 (2006).
340. Kono, S. Aceruloplasminemia. *Curr. Drug Targets* **13**, 1190–1199 (2012).
341. Quigley, J. G. *et al.* Identification of a Human Heme Exporter that Is Essential for Erythropoiesis. *Cell* **118**, 757–766 (2004).
342. Byon, J. C. H., Chen, J., Doty, R. T. & Abkowitz, J. L. FLVCR is necessary for erythroid maturation, may contribute to platelet maturation, but is dispensable for normal hematopoietic stem cell function. *Blood* **122**, 2903–2910 (2013).
343. Chiabrando, D. *et al.* The mitochondrial heme exporter FLVCR1b mediates erythroid differentiation. *J. Clin. Invest.* **122**, 4569–4579 (2012).
344. Rey, M. A. *et al.* Enhanced alternative splicing of the FLVCR1 gene in Diamond Blackfan anemia disrupts FLVCR1 expression and function that are critical for erythropoiesis. *Haematologica* **93**, 1617–1626 (2008).
345. Philip, M. *et al.* Heme Exporter FLVCR Is Required for T Cell Development and Peripheral Survival. *J. Immunol.* **194**, 1677–1685 (2015).
346. Vulpe, C. D. *et al.* Hephaestin, a ceruloplasmin homologue implicated in intestinal iron transport, is defective in the sla mouse. *Nat. Genet.* **21**, 195–199 (1999).
347. Griffiths, T. A. M., Mauk, A. G. & MacGillivray, R. T. A. Recombinant Expression and Functional Characterization of Human Hephaestin: A Multicopper Oxidase with Ferroxidase Activity. *Biochemistry (Mosc.)* **44**, 14725–14731 (2005).

348. Hudson, D. M. *et al.* Human hephaestin expression is not limited to enterocytes of the gastrointestinal tract but is also found in the antrum, the enteric nervous system, and pancreatic  $\beta$ -cells. *Am. J. Physiol. - Gastrointest. Liver Physiol.* **298**, G425–G432 (2010).

### **Author biographies**

#### *Bart J. Crielaard*

Bart Crielaard is an assistant professor at the Zernike Institute for Advanced Materials, University of Groningen, The Netherlands. After finishing his medical training, he further specialized in Biomedical Engineering and obtained a PhD degree in Pharmaceutics from Utrecht University, The Netherlands. During his PhD and his postdoctoral fellowship at Weill Cornell Medical College, U.S.A., he developed a particular interest in macrophage biology and iron metabolism, which could be exploited for therapeutic purposes. In his current role as a group leader at the University of Groningen, he is currently investigating this and other strategies for the development of new therapeutics.

#### *Twan Lammers*

Twan Lammers obtained a DSc degree in Radiation Oncology from Heidelberg University, Germany, in 2008 and a PhD degree in Pharmaceutics from Utrecht University, The Netherlands, in 2009. In the same year, he started the Nanomedicine and Theranostics group at the Institute for Experimental Molecular Imaging RWTH Aachen University, Germany. In 2014, he was promoted to full professor at RWTH Aachen. Since 2012, he has also worked as a part-time assistant professor at the Department of Targeted Therapeutics at the University of Twente. His primary research interests include drug targeting to tumors, image-guided drug delivery and tumor-targeted combination therapies.

#### *Stefano Rivella*

Stefano Rivella is Professor of Pediatrics with tenure at the Children's Hospital of Philadelphia and University of Pennsylvania, U.S.A., and holds the Kwame Ohene-Frempong Chair on Sickle Cell Anemia. He characterized the role of seminal

factors contributing to morbidity and mortality in various haematological disorders, including hepcidin and ferroportin, which control iron absorption and organ iron distribution; and macrophages, which play an important role in iron recycling, inflammation as well as erythroid proliferation and maturation. Based on these studies, he contributed to the development of novel therapeutics presently in clinical and preclinical trials.

### **Key points**

- Iron metabolism is a tightly regulated physiological process that has relatively low redundancy and its deregulation often leads to iron deficiency or iron overload.
- Iron deficiency and iron overload are historically associated with erythroid disorders, but a deregulated iron metabolism has also been implicated in numerous ageing-related, non-haematological disorders including neurodegenerative disorders, atherosclerosis and cancer.
- Intracellular iron is directly involved in the formation of reactive oxygen species that can cause cellular oxidative damage, and which are important for ferroptosis, a form of non-apoptotic cell death.
- The internalization of iron by macrophages can modulate macrophage activity towards a pro-inflammatory phenotype, which is likely also dependent on the pathway of iron intake.
- Agents that interfere with key regulators of iron metabolism and cellular iron trafficking represent a promising new class of therapeutic agents for various diseases, as they exploit pathological pathways that are complementary to those targeted by existing treatments.
- Targeting therapeutics to diseased tissues that express high levels of transferrin receptor is a strategy that is used by several agents under clinical development, and extension of this strategy towards other iron metabolism-associated cellular transporters may be advantageous.