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1 **Normoglycemic ketonemia as biochemical presentation in ketotic glycogen storage disease**

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25 **Abbreviations:** FI, fasting intolerance; GSD, glycogen storage disease; KB, ketone bodies.
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37 **Abstract**

38 **Background:** According to the textbooks, the ketotic glycogen storage disease (GSD) types O,
39 III, VI, IX and XI are associated with fasting ketotic hypoglycemia and considered milder as
40 gluconeogenesis is intact.

41 **Methods:** Retrospective cohort study of biochemical profiles from supervised clinical fasting
42 studies performed in ketotic GSD patients in our metabolic center. For data analysis,
43 hypoglycemia was defined as plasma glucose concentration <2.6 mmol/L. Total KB was defined
44 as the sum of blood acetoacetate and β -hydroxybutyrate concentrations. If the product of glucose
45 and KB concentrations was greater than 10, a ketolysis defect was suspected.

46 **Results:** Data could be collected from 13 fasting studies in 12 patients with GSD III (n=4), GSD
47 VI (n=3) and GSD IX (n=5). Six patients remained normoglycemic with median glucose
48 concentration of 3.9 mmol/L [range: 2.8-4.6 mmol/L] and median total KB concentration of 1.9
49 mmol/L [range: 0.6-5.1 mmol/L]. The normoglycemic patients included type VI (3 out of 3) and
50 type IX (3 out of 5) patients. All type III patients developed ketotic hypoglycemia. Interestingly,
51 in five patients (1 GSD III, 1 GSD VI and 3 GSD IX), the biochemical profile suggested a
52 ketolysis defect.

53 **Conclusion:** Normoglycemic ketonemia is a common biochemical presentation in patients with
54 GSD types VI and IX and ketonemia can precede hypoglycemia in all studied GSD types.
55 Therefore, GSD VI and IX should be added to the differential diagnosis of ketotic
56 normoglycemia and KB concentrations should be routinely measured in ketotic GSD patients.

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60 **Compliance with Ethics Guidelines**

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62 **Conflict of Interest:** Francjan J van Spronsen has received research grants and consultancy and
63 speaker's fees from Merck Serono and Danone Nutricia. He is a member of the scientific
64 advisory board of Merck Serono and chair of the scientific advisory board of Danone Nutricia. In
65 the last 5 years, Terry GJ Derks has received speaker's fees from Danone Nutricia, Vitaflo and
66 Recordati and research fees from Sigma Tau and Vitaflo. Irene J Hoogeveen, Rixt M van der
67 Ende, Foekje de Boer, and M Rebecca Heiner-Fokkema declare that they have no conflict of
68 interest to disclose.

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70

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72

73 **Contributions of individual authors:** Irene J Hoogeveen and Rixt M van der Ende collected
74 and analyzed data from supervised clinical fasting studies, performed the data analysis, drafted
75 the first version of the manuscript, and wrote the final manuscript.

76 Francjan J van Spronsen was involved in clinical management and monitoring and critically
77 reviewed and revised the manuscript, and approved the final manuscript as submitted.

78 Foekje de Boer was involved in dietary management, critically reviewed and revised the
79 manuscript, and approved the final manuscript as submitted.

80 M Rebecca Heiner-Fokkema supervised the data analysis of the fasting studies, critically
81 reviewed and revised the manuscript, and approved the final manuscript as submitted.

82 Terry G J Derks initiated this study, was involved in clinical management and monitoring,
83 drafted the first version of the manuscript, critically reviewed and revised the manuscript, and
84 wrote the final manuscript.

85

86 All authors approved the final manuscript as submitted and agree to be accountable for all
87 aspects of the work. All authors confirm the absence of previous similar or simultaneous
88 publications.

89 **Informed Consent:** All procedures followed were in accordance with the ethical standards of
90 the institutional responsible committee on human experimentation and with the Helsinki
91 Declaration of 1975, as revised in 2000. Since all data were retrieved retrospectively and
92 analyses anonymously, no informed consent was needed.

93

94 **Introduction**

95 Fasting intolerance (FI) is biochemically associated with hypoglycemia and/or metabolic
96 acidosis, the latter often caused by increased concentrations of lactate and/or ketones. The
97 differential diagnosis of childhood FI includes many endocrine disorders and inborn errors of
98 metabolism, among which several types of glycogen storage disease (GSD).

99 There are at least 13 types of GSD, which are classified according to the protein defect and organ
100 distribution (Laforet et al 2012). The ketotic GSD types 0, III, VI, IX and XI are associated with
101 fasting ketotic hypoglycemia and considered relatively mild compared to GSD type I because
102 gluconeogenesis is intact. Traditionally ketotic *hypoglycemia* is considered the common
103 diagnostic biochemical phenotype upon fasting in patients with ketotic GSD types (Laforet et al
104 2012), although cohort studies have demonstrated that this is not always the case (Beauchamp et
105 al 2007a; Beauchamp et al 2007b). It has recently been reported that ketotic GSD types can be
106 easily misinterpreted as idiopathic ketotic hypoglycemia (Brown et al 2014).

107 Regular monitoring of ketone bodies (KB) is recommended to titrate dietary management in
108 ketotic GSD patients (Dagli et al 2010; Dagli and Weinstein 2009, Goldstein et al 2011), but
109 experimental data are lacking. Therefore, we have performed this retrospective study of
110 supervised clinical fasting studies in patients with ketotic GSD.

111 **Patients and Methods**

112 Subjects - The Section of Metabolic Diseases, Beatrix Children's Hospital, University Medical
113 Center Groningen is a tertiary metabolic center and a reference center for hepatic GSD patients.
114 In the period 1993-2012, 539 supervised clinical fasting studies have been performed in 476
115 patients. From this cohort all patients with ketotic GSD have been identified to perform a
116 retrospective study of the biochemical profiles of their supervised clinical fasting studies. Data
117 were anonymously retrieved from both the paper and electronic medical files.

118 Fasting studies - Supervised clinical fasting studies were performed as described elsewhere
119 (Bonfont et al 1990; Van Veen et al 2011), for either diagnostic or therapeutic reasons, i.e. to
120 titrate dietary management. Most diagnostic studies were performed before 2003, when plasma
121 acylcarnitine profiling became available in our laboratory. Fasting studies were only performed
122 in healthy patients in good nutritional condition. Subjects were admitted one day before the
123 fasting test when they were <8 years of age. There was no limitation in water intake. After the
124 last meal and an individually tailored period of fasting, an intravenous catheter was inserted for
125 blood sampling at hourly intervals. Carefully supervised fasting was continued until glucose
126 concentrations dropped below 2.6 mmol/L or until development of symptoms or signs of
127 hypoglycemia.

128 Statistical analysis - SPSS Statistics version 22.0 (IBM Corp., Armonk New York, USA) was
129 used to calculate median and range for concentrations of glucose and KB. For data analysis,
130 hypoglycemia was defined as plasma glucose concentration <2.6 mmol/L (Koh et al 1988). Total
131 KB was defined as the sum of blood acetoacetate and β -hydroxybutyrate concentrations. If the
132 product of glucose and KB concentrations was greater than 10, a ketolysis defect was suspected
133 (Touati et al 2012).

134 **Results**

135 Table 1 presents patients characteristics from 12 patients with GSD type III (n=4), type VI (n=3)
136 and type IX (n=5), in whom a total of 13 supervised clinical fasting studies were performed. A
137 fasting study was performed twice in patient 2 due to therapeutic reasons. For patient 2 and 12 no
138 confirmatory molecular studies were available, however diagnosis was confirmed enzymatically,
139 in leukocytes and erythrocytes, respectively. Patient 2 died at the age of 27 years by a car
140 accident, his sister was homozygous for the c.2039G>A AGL founder mutation from the island
141 of Aruba.

142 Table 2 presents biochemical data during the fasting studies in the individual patients. Six
143 patients showed normoglycemia during fasting, i.e. median blood glucose concentration in these
144 patients were 3.9 mmol/L [range: 2.8-4.6 mmol/L] with median total KB concentration of 1.9
145 mmol/L [range: 0.6-5.1 mmol/L]. The *normoglycemic* patients included type VI (3 out of 3) and
146 type IX (3 out of 5) patients. All type III patients developed ketotic hypoglycemia, but
147 interestingly, a remarkable increase in KB preceded hypoglycemia in these patients. Patient 4
148 displayed hypoglycemia at the end of the fasting study, although her glucose concentration was
149 not below 2.6 mmol/L during the last combined KB and glucose measurement. Patient 9
150 developed hypoglycemia very quickly without clinical manifestations. In patient 11 the fasting
151 study was terminated because of abdominal pain, nausea and vomiting, although plasma blood
152 glucose concentrations were normal. At the moment of terminating the blood KB concentration
153 was 3.3 mmol/L. For patient 12 only one combined measurement of blood glucose and KB
154 concentration could be obtained, however it showed an elevated KB concentration of 0.6
155 mmol/L already after three hours of fasting. Interestingly, the product of glucose and KB

156 suggested a ketolysis defect in 5 patients. One of these 5 patients underwent a fasting study for
157 diagnostic purposes.

158 Figure 1 presents concentrations of glucose and KB longitudinally in time for all fasted GSD
159 type VI patients, demonstrating normoglycemia despite remarkable increase of KB
160 concentrations.

161 **Discussion**

162 This study demonstrates that normoglycemic ketonemia is a common biochemical phenotype in
163 GSD type VI and IX and that ketonemia can precede hypoglycemia in all studied GSD types.
164 This is important from both a diagnostic and management point of view.

165 In this study normoglycemic ketonemia was presented by half of the GSD patients. Five out of
166 twelve patients displayed a biochemical phenotype suggestive of a ketolysis defect (Bonnetfont et
167 al 1990). It was recently reported that especially GSD IX is an unappreciated cause of idiopathic
168 ketotic hypoglycemia (Brown et al 2014). As this study also included diagnostic fasting studies,
169 to our opinion it emphasizes the potential risk of underdiagnosing ketotic GSD. Ketotic GSD
170 should therefore be included in the differential diagnosis of childhood FI associated
171 normoglycemic ketonemia.

172 Previously, supervised clinical fasting studies have played a central diagnostic role as an
173 informative functional *in vivo* test (Bonnetfont et al 1990), but nowadays these studies are
174 considered obsolete. Moreover, fasting studies are relatively time-consuming, expensive,
175 invasive and potentially dangerous. These fasting studies have merely been replaced after the
176 introduction of new laboratory techniques, like acylcarnitine profiling (Millington et al 1990).
177 More recently next generation sequencing and/or exome sequencing have developed into
178 powerful diagnostic confirmatory tests (Wang et al 2012). In our experience there are few
179 indications for the traditional clinical fasting studies, under exceptional circumstances and well-
180 controlled conditions, to characterize the clinical *in vivo* implications for patients with unknown
181 variations in the metabolome or genome.

182 Several factors complicate the recognition of patients with ketotic GSD. During ‘quick’ physical
183 examination at an emergency room, both the soft hepatomegaly (like in GSD types VI and IX)

184 and failure to thrive may be easily overlooked. Simple laboratory tests in blood are not routinely
185 requested in stress samples from patients with FI. In untreated GSD patients, (a specific
186 combination of) plasma concentrations of lactate, transaminases, uric acid, triglycerides and
187 cholesterol is usually abnormal. In contrast, the traditional hormonal and secondary metabolic
188 tests (like analysis of plasma acylcarnitines and urinary organic acids) are usually normal, even
189 when samples are obtained under critical conditions. The above-mentioned investigations are
190 important first-line tests in patients with FI to select candidates for confirmatory molecular
191 and/or enzymatic testing for GSD.

192 It is not known why some ketotic GSD patients display hypoglycemia and some do not. This
193 variation is especially observed in GSD VI; hepatic phosphorylase deficiency, encoded by the
194 *PYGL* gene (OMIM #232700) and GSD IX; hepatic phosphorylase b kinase deficiency, encoded
195 by the *PHKA2* gene (OMIM #300798; X-linked GSD IX), the *PHKB* gene (OMIM #172490),
196 and *PHKG2* gene (OMIM #172471) respectively. Beauchamp et al reported hypoglycemia in 5
197 out of 13 GSD VI patients on either fasting or glucose loading tests (Beauchamp et al 2007a),
198 while in GSD IX Beauchamp et al reported hypoglycemia as a presenting sign in 5 out of 15
199 GSD IX patients (Beauchamp et al 2007b). The hypoglycemia in GSD IX patients included those
200 with mutations in the *PHKG2* gene, which is in line with Bali et al, who reported fasting
201 hypoglycemia in all 5 patients with *PHKG2* mutations (Bali et al 2014). This finding may be
202 very well explained by the fact that mutations of the *PHKG2* gene contains the catalytic site of
203 hepatic phosphorylase b kinase.

204 Uncooked cornstarch and protein are the keystones of dietary management in ketotic GSD, the
205 latter serving as an alternative source for gluconeogenesis to maintain normoglycemia (Derks
206 and Smit 2015). In ketotic GSD types, increased KB concentrations reflect increased

207 mitochondrial fatty acid oxidation, which is associated with activation of gluconeogenesis and
208 secondary endogenous proteolysis from muscle tissue. Instead of maintenance of
209 normoglycemia, prevention of increasing KB concentrations could therefore be regarded as a
210 more relevant aim in optimizing metabolic control.

211 At a relatively young age, one GSD III patient (patient 4) displayed a decrease in both KB and
212 glucose concentrations with prolonged fasting. Hypoketosis has been reported before in GSD III
213 patients (Seigel et al 2008; Clemente et al 2010), in whom exogenous carbohydrate requirements
214 are still relatively high (Derks and Van Rijn 2015). We speculate that, as a consequence of
215 dietary management with frequent high carbohydrate meals, there may have been a relatively
216 high plasma insulin state together with high intracellular malonyl-CoA levels, physiologically
217 inhibiting long-chain mitochondrial fatty acid oxidation at the level of carnitine
218 palmitoyltransferase type I.

219 This study has several limitations. First, data have retrospectively been retrieved from electronic
220 and paper files, from fasting studies that have mostly been performed at least ten years ago.
221 Second, fasting studies have been conducted in only a subset of our GSD patients, which could
222 have introduced a selection bias. Third, these fasting studies originate from a period, in which the
223 general opinion on dietary management and outcome parameters for ketotic GSD types was
224 different. Last, the definition of hypoglycemia is debatable in several ways. We have defined
225 hypoglycemia as a *plasma* glucose concentration <2.6 mmol/L, measured by calibrated meters
226 with a constant factor of 1.11 for conversion between blood glucose and plasma glucose
227 concentrations (D'Orazio et al 2005). Therefore, the plasma glucose concentrations are on
228 average 11% higher compared to blood concentrations, depending on the hematocrit and the
229 water component in blood. Also, hypoglycemia defined by a single number does not distinguish

230 the difference values at which an individual starts to compensate for inadequate glucose supply
231 to the brain (Cornblath et al 2000).

232 To date, in contrast with GSD III (Derks and Smit 2015; Kishnani et al 2010), there are no
233 formal diagnostic and management guidelines for GSD VI and IX. Based on expert opinion,
234 caregivers are advised to titrate dietary management, aiming at normoglycemia and maintenance
235 of blood β -hydroxybutyrate concentrations lower than 0.3 mmol/L, measured by a portable blood
236 ketone meter (Dagli et al 2010; Dagli and Weinstein 2009; Goldstein et al 2011). This study
237 provides short-term, indirect biochemical evidence substantiating these management advices, but
238 there is a lack of data on long-term clinical outcome parameters, like growth, liver size,
239 laboratory studies, hepatic complications and bone density.

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264 **Conclusion**

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266 This is the first study that critically analyzed blood glucose and KB concentrations during fasting

267 in ketotic GSD patients. Normoglycemic ketonemia is a common biochemical presentation in

268 patients with GSD types VI and IX and ketonemia can precede hypoglycemia in all studied GSD

269 types. Therefore, GSD VI and IX should be added to the differential diagnosis of ketotic

270 normoglycemia and KB concentrations should be routinely measured in ketotic GSD patients.

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275 **References**

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277 Bali D, Goldstein J, Fredrickson K, Rehder C, Boney A, Austin S (2014) Variability of disease
278 spectrum in children with liver phosphorylase kinase deficiency caused by mutations in the
279 PHKG2 gene. *Mol Genet Metab.* **111**: 309–313.

280 Beauchamp NJ, Taybert J, Champion MP, Layet V, Heinz-Erian P, Dalton A, et al (2007a) High
281 frequency of missense mutations in glycogen storage disease type VI. *J Inherit Metab Dis* **30**:
282 722–734.

283 Beauchamp NJ, Dalton A, Ramaswami U, Niinikoski H, Mention K, Kenny P, et al (2007b)
284 Glycogen storage disease type IX: High variability in clinical phenotype. *Mol Genet Metab* **92**:
285 88–99.

286 Bonnefont JP, Specola NB, Vassault A, Lombes A, Ogier H, de Klerk JBC, et al (1990) The
287 fasting test in paediatrics: Application to the diagnosis of pathological hypo- and hyperketotic
288 states. *Eur J Pediatr* **150**: 80–85.

289 Brown LM, Corrado MM, van der Ende RM, Derks TGJ, Chen M, Siegel S, et al (2014)
290 Evaluation of glycogen storage disease as a cause of ketotic hypoglycemia in children. *J Inherit*
291 *Metab Dis* **38**: 489–493.

292 Clemente M, Gussinyer M, Arranz JA, Riudor E, Yeste D, Albisa M, Carrascosa A (2010)
293 Glycogen Storage Disease Type III with Hypoketosis. *J Pediatr Endocrinol & Metab* **23**: 833-
294 836

295 Cornblath M, Hawdon JM, Williams a F, Aynsley-Green a, Ward-Platt MP, Schwartz R, et al
296 (2000) Controversies regarding definition of neonatal hypoglycemia: suggested operational
297 thresholds. *Pediatrics* **105**: 1141–1145.

298 Dagli A, Sentner C, Weinstein D (2010) Glycogen storage disease type III. In Pagon RA, Adam
299 MP, Ardinger HH, et al., editors. *GeneReviews*. University of Washington, Seattle.

300 Dagli A, Weinstein D (2009) Glycogen storage disease type VI. In Pagon RA, Adam MP,
301 Ardinger HH, et al., editors. *GeneReviews*. University of Washington, Seattle.

302 Derks TGJ, van Rijn M (2015) Lipids in hepatic glycogen storage diseases: pathophysiology
303 monitoring of dietary management and future directions. *J Inherit Metab Dis* **38**: 537-543

304 Derks TGJ, Smit GPA (2015) Dietary management in glycogen storage disease type III: what is
305 the evidence ? *J Inherit Metab Dis* **38**: 545–550.

306 D’Orazio P, Burnett R, Fogh-Andersen N, Jacobs E, Kuwa K, Kulpman W, et al (2005)
307 Approved IFCC Recommendation on Reporting Results for Blood Glucose. *Clin Chem* **51**:
308 1573–1576.

- 309 Goldstein J, Austin S, Kishnani P et al (2011) Phosphorylase Kinase Deficiency. In Pagon RA,
310 Adam MP, Ardinger HH, et al., editors. *GeneReviews*. University of Washington, Seattle.
- 311 Kishnani PS, Austin SL, Arn P, Bali DS, Boney A, Case LE, et al (2010) Glycogen Storage
312 Disease Type III diagnosis and management guidelines. *Genet Med* **12**: 446–463.
- 313 Koh TH, Aynsley-Green a, Tarbit M, Eyre J (1988). Neural dysfunction during hypoglycaemia.
314 *Arch Dis Child* **63**: 1353–1358.
- 315 Laforêt P, Weinstein DA, Smit GPA (2012) The Glycogen Storage Diseases and Related
316 Disorders, chapter 6. In Saudubray JM, van de Berghe G, Walter J, editors. *Inborn metabolic*
317 *diseases: diagnosis and treatment*. Springer, Berlin.
- 318 Millington DS, Kodo N, Norwood DL, Roe CR (1990) Tandem Mass-Spectrometry - a New
319 Method for Acylcarnitine Profiling With Potential for Neonatal Screening for Inborn-Errors of
320 Metabolism. *J Inherit Metab Dis* **13**: 321–324.
- 321 Seigel J, Weinstein DA, Hillman R, Colbert B, Matthews B, Bachrab B (2008) Glycogen storage
322 disease type IIIa presenting as non-ketotic hypoglycemia: use of a newly approved commercially
323 available mutation analysis to non-invasively confirm the diagnosis. *J Pediatr Endocrinol Metab*
324 **6**: 587-590.
- 325 Touati G, Mochel F, Rabier D (2012) Diagnostic Procedures: Functional Tests and Post-mortem
326 Protocol, chapter 4. In Saudubray JM, van den Berghe G, Walter J, editors. *Inborn Metabolic*
327 *Diseases: diagnosis and treatment*. Springer, Berlin.
- 328 Van Veen MR, van Hasselt PM, de Sain-van der Velden MGM, Verhoeven N, Hofstede FC, de
329 Koning TJ, et al (2011). Metabolic profiles in children during fasting. *Pediatrics* **127**: 1021–
330 1027.
- 331 Wang J, Cui H, Lee N-C, Hwu W-L, Chien Y-H, Craigen WJ, et al (2012) Clinical application of
332 massively parallel sequencing in the molecular diagnosis of glycogen storage diseases of
333 genetically heterogeneous origin. *Genet Med*. **15**: 106-114
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336 **Legends Figures**

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339 **Figure 1. Longitudinal course of fasting in GSD type VI**

340 Legend: •□ Glucose concentration, ◆□ Ketone Bodies concentration, - - - Cut off

341 point hypoglycemia.

342 **Table 1. Patient characteristics**

Case	Age ^a (Y ^m)	Sex ^b	GSD type	Molecular defect						
				Gene	Exon nr	Nucleotide change allele 1	Coding effect allele 1	Exon nr	Nucleotide change allele 2	Coding effect allele 2
1	18 ^{0/12}	F	IIIa	AGL	17	c.2039G>A	p.Trp680X	17	c.2039G>A	p.Trp680X
2 (1)*	11 ^{0/12}	M	IIIa	-	-	-	-	-	-	-
2 (2)*	16 ^{6/12}	M	IIIa	-	-	-	-	-	-	-
3*	22 ^{5/12}	F	IIIa	AGL	17	c.2039G>A	p.Trp680X	17	c.2039G>A	p.Trp680X
4	1 ^{5/12}	M	III	AGL	13	c.1571G>A	p.Arg524His	-	-	-
5	3 ^{3/12}	M	VI	PYGL	3	c.385G>A	p.Asp129Asn	20	c.2446C>T	p.Arg816*
6	4 ^{10/12}	M	VI	PYGL	3	c.418C>G	p.Leu140Val	11	c.1366G>A	p.Val456Met
7	1 ^{10/12}	M	VI	PYGL	1	c.131G>A	p.Arg44His	16	c.1900G>C	p.Asp634His
8	4 ^{4/12}	M	IX	PHKA2	33	c.3614C>T	p.Pro1205Leu			
9	1 ^{7/12}	F	IX	PHKA2	33	c.3614C>T	p.Pro1205Leu			
10	2 ^{6/12}	M	IX	PHKA2	-	DelXp22.13	-			
11	7 ^{10/12}	M	IX	PHKB	14	c.1265dup	-	27	c.2316-2A>C	p.Asn422fs
12	2 ^{3/12}	M	IX	-	-	-	-	-	-	-

343 Legend: ^a: age during the fasting study in years and months, ^b: M=male, F=female, -:

344 mutation unknown, *: patients are siblings.

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352 **Table 2. Biochemical data of the fasting studies**

Case	Purpose test ^a	Duration of Fasting	Glucose	KB ^b	FFA	FFA/KB	KBxGlucose ^c
	(D/T)	(hh:mm)	(mmol/L)	(mmol/L)	(mmol/L)		
1	T	11:30 18:30	3.5 2.4	1.4 4.0	0.9 1.2	0.6 0.3	4.9 9.6
2	T	10:30 11:30	2.1 2.0	4.8 6.9	1.8 1.3	0.4 0.2	10.1 13.8
2	T	11:00 17:00	3.3 1.7	3.8 4.8	1.2 1.6	0.3 0.3	12.5 8.2
3	T	11:00 16:30	3.2 1.7	2.6 3.2	0.9 0.8	0.3 0.3	8.3 5.4
4	D	04:00 05:00	4.8 4.4	2.0 1.5	1.2 0.8	0.6 0.5	9.6 6.6
5	D	12:30 15:15	5.1 3.0	1.5 3.5	1.0 1.1	0.7 0.3	7.7 10.5
6	D	08:45 14:45	4.1 3.5	0.7 0.8	0.6 0.8	0.9 1.0	2.9 2.8
7	D	09:00 12:00	2.7 3.0	1.4 1.9	- -	- -	3.8 5.7
8	T	02:00 08:15	4.6 3.8	1.6 5.1	0.9 1.2	0.6 0.2	7.4 19.4
9	T	03:00 07:00	3.7 2.3	0.6 3.3	- 1.1	- 0.3	2.2 7.6
10	T	08:30 14:15	3.5 2.5	1.8 6.1	1.4 1.6	0.8 0.3	6.3 15.3
11	T	07:50 14:50	4.1 3.8	1.4 3.3	0.9 0.9	0.6 0.3	5.7 12.5
12	D/T	03:10	4.3 -	0.6 -	0.6 -	1.0 -	2.6 -

353 Legend: ^a:D=diagnostic, T=therapeutic, ^b:KB is the sum of acetoacetate and β -
354 hydroxybutyrate, ^c:suspect ketolysis defect is defined as a product of glucose and KB
355 greater than 10 (Touati et al 2012).

356