

University of Groningen

Preeclampsia As Modulator of Offspring Health

Stojanovska, Violeta; Scherjon, Sicco A; Plosch, Torsten

Published in:
Biology of Reproduction

DOI:
[10.1095/biolreprod.115.135780](https://doi.org/10.1095/biolreprod.115.135780)

IMPORTANT NOTE: You are advised to consult the publisher's version (publisher's PDF) if you wish to cite from it. Please check the document version below.

Document Version
Publisher's PDF, also known as Version of record

Publication date:
2016

[Link to publication in University of Groningen/UMCG research database](#)

Citation for published version (APA):
Stojanovska, V., Scherjon, S. A., & Plösch, T. (2016). Preeclampsia As Modulator of Offspring Health. *Biology of Reproduction*, 94(3), 1-10. [53]. DOI: 10.1095/biolreprod.115.135780

Copyright

Other than for strictly personal use, it is not permitted to download or to forward/distribute the text or part of it without the consent of the author(s) and/or copyright holder(s), unless the work is under an open content license (like Creative Commons).

Take-down policy

If you believe that this document breaches copyright please contact us providing details, and we will remove access to the work immediately and investigate your claim.

Downloaded from the University of Groningen/UMCG research database (Pure): <http://www.rug.nl/research/portal>. For technical reasons the number of authors shown on this cover page is limited to 10 maximum.

Minireview

Preeclampsia As Modulator of Offspring Health¹

Violeta Stojanovska,² Sicco A. Scherjon, and Torsten Plösch

Department of Obstetrics and Gynecology, University of Groningen, University Medical Center Groningen, The Netherlands

ABSTRACT

A balanced intrauterine homeostasis during pregnancy is crucial for optimal growth and development of the fetus. The intrauterine environment is extremely vulnerable to multisystem pregnancy disorders such as preeclampsia, which can be triggered by various pathophysiological factors, such as angiogenic imbalance, immune responses, and inflammation. The fetus adapts to these conditions by a mechanism known as developmental programming that can lead to increased risk of chronic noncommunicable diseases in later life. This is shown in a substantial number of epidemiological studies that associate preeclampsia with increased onset of cardiovascular and metabolic diseases in the later life of the offspring. Furthermore, animal models based predominantly on one of the pathophysiological mechanism of preeclampsia, for example, angiogenic imbalance, immune response, or inflammation, do address the susceptibility of the preeclamptic offspring to increased maternal blood pressure and disrupted metabolic homeostasis. Accordingly, we extensively reviewed the latest research on the role of preeclampsia on the offspring's metabolism and cardiovascular phenotype. We conclude that future research on the pathophysiological changes during preeclampsia and methods to intervene in the harsh intrauterine environment will be essential for effective therapies.

early development, epigenetics, metabolism, preeclampsia

INTRODUCTION

The global prevalence of chronic cardiometabolic noncommunicable diseases (NCDs) diseases such as hypertension, cardiovascular disease, diabetes mellitus type 2, and metabolic syndrome has markedly increased during the past decades [1]. A number of genes and behavioral changes have been identified as initiators and mediators of these complex cardiometabolic diseases [2–4]. However, the increasing prevalence of NCDs cannot be accounted for by only these

determinants. Biological factors already present during early development can lead to immediate cardiometabolic fetal responses that might have long-term effects.

The developmental origins of health and disease, or the Barker hypothesis, attempts to explain the high incidence of chronic NCDs by unfavorable in utero conditions. Depending on the severity of the insult during specific critical windows of fetal development, permanent tissue adjustments can occur, leading to long-term changes in organ function [5]. During pregnancy, the key regulatory organ of the intrauterine environment is the placenta, which serves as a metabolic, immune, and endocrine organ. It enables and regulates transport of gasses, nutrients, hormones, immunoglobulins, and waste products between the mother and the fetus in order to maintain a favorable developmental homeostasis [6]. Hostile environmental factors present during early life, when rapid growth and differentiation is expected, can have a powerful impact on physiological health for a lifetime.

Preeclampsia is a pregnancy-associated syndrome, characterized by hypertension and proteinuria, affecting 2%–8% of the population worldwide [7]. It remains a major obstetric problem due to the high prevalence of maternal and fetal mortality and morbidity. Although the etiology is puzzling, several pathophysiological mechanisms combined have proven to be involved at least in the clinical course of preeclampsia. Antiangiogenic imbalance, excessive inflammation, hypoxia, and/or autoantibodies targeting the renin-angiotensin system make up the harsh intrauterine environment during preeclampsia [8, 9]. All these factors may interact with the genome of the mother and the fetus in terms of gene expression modulation, ultimately affecting the expressed phenotype.

In this review, first we address epidemiological and human studies that show a contribution of preeclampsia to cardiometabolic alterations in the offspring. Further, we focus on animal studies in this research area, approaching three different mechanistic scenarios of preeclampsia. Finally, we discuss possible mechanisms that may explain relevance of preeclampsia in developmental programming of metabolic and cardiovascular diseases in the offspring.

EVIDENCE FROM HUMAN STUDIES: OFFSPRING STATUS AFTER PREECLAMPSIA

Birth weight screening is still an important assessment of optimal in utero nutrition and development. During preeclampsia, 13%–60% of the pregnancies are complicated by decreased birth weight depending on the region, maternal age, and the severity of the disease [10, 11]. Therefore, preeclampsia is one of the leading factors of fetal growth restriction [12, 13]. Low birth weight per se is already an established risk factor for

¹This work was supported by the Netherlands Organization for Health Research and Development (ZonMw, grant no. 91211053).

²Correspondence: Violeta Stojanovska, University of Groningen, University Medical Center Groningen, PB 30 001 9700RB Groningen, The Netherlands. E-mail: v.stojanovska@umcg.nl

Received: 26 September 2015.
First decision: 3 November 2015.
Accepted: 15 January 2016.

© 2016 by the Society for the Study of Reproduction, Inc. This article is available under a Creative Commons License 4.0 (Attribution-Non-Commercial), as described at <http://creativecommons.org/licenses/by-nc/4.0>

eISSN: 1529-7268 <http://www.biolreprod.org>
ISSN: 0006-3363

cardiovascular and metabolic diseases in later life, although the causal mechanisms are still speculative [14, 15].

Preeclampsia is characterized by new-onset hypertension during pregnancy ($\geq 140/90$ mmHg) along with proteinuria. However, little is known about neonatal blood pressure after this complication of pregnancy. An early report indicated that term neonates from preeclamptic mothers have a transient hypertension [16]. A more recent study showed that premature neonates from preeclamptic mothers, compared to controls, have early neonatal hypotension [17]. As indicated, blood pressure levels are also altered in these children, which appears to be associated with the gestational age. Furthermore, observation of blood pressure in school-age children previously exposed to preeclampsia showed higher systolic and diastolic blood pressure already at 8 yr of age [18–23]. Additionally, it was reported that these children have smaller hearts, increased heart rate, and features of cardiac diastolic dysfunction [24] as well as an increased risk of congenital heart defects, namely septal defects [25, 26]. However, in a cohort study, a 65-yr follow-up of preeclamptic offspring did not show an increased risk of coronary heart disease, but increased stroke incidence was reported [27].

Evaluation of endothelial functionality with noninvasive assessment can provide considerable insight into blood pressure risk stratification. School-age children previously exposed to preeclampsia showed increased vascular stiffness in the pulmonary and peripheral vascular system [24, 28]. Moreover, intact endothelial morphology is a potent vascular tone regulator. Analysis of endothelial cord cells showed a decreased number of endothelial colony-forming cells in contrast to increased senescent progenitor cells [29, 30]. This is indicative for at least advanced endothelial cord cell aging in the preeclamptic neonates.

The body mass index (BMI), plasma glucose, and lipid concentrations serve as strong indicators of optimal metabolic functioning and, when increased, are risk factors for cardiovascular and metabolic diseases. Preeclampsia shares many features with the metabolic syndrome, including increased maternal concentrations of proinflammatory cytokines, insulin, leptin, triglycerides, free fatty acid, and low-density cholesterol, usually in absence of diabetes [31]. Children from preeclamptic mothers show an increased risk of hospitalizations for endocrine and metabolic diseases in the first 5 yr of life [32]. In adolescence, premature-born preeclamptic males have an increased BMI in comparison to premature males born from normotensive pregnancies [33]. Cord blood samples from preeclamptic children show altered lipid profiles and increased tumor necrosis factor alpha (TNF- α) when studied for metabolic and inflammatory parameters [31, 34, 35], but in adolescence, these changes in glucose and lipid profiles are not prominent anymore [19, 21]. These effects may be influenced to some extent by maternal metabolic blood parameters and placental insufficiency. However, in school-age children previously exposed to preeclampsia, the metabolic phenotype shows changes only after subclustering of this group. The quantitative insulin sensitivity check index (QUICKI) serves as a predictive marker for diabetes onset based on fasting plasma glucose and insulin levels, and low values correspond to increased insulin resistance. Subdivision of groups based on QUICKI did show increased leptin and triglycerides levels in preeclampsia-exposed children that had independently low QUICKI values [36]. This suggests that insulin resistance independently, superimposed on earlier preeclampsia exposure, can serve as a strong predictor of the metabolic syndrome. These clinical observations reflect a transiently affected neonatal metabo-

lism, which is not continuous through adolescence, but possibly can lead to increased susceptibility to the metabolic syndrome after a second environmental stressor, such as a metabolic stress.

INTRAUTERINE ADVERSE ENVIRONMENT DURING PREECLAMPSIA AND OFFSPRING OUTCOME: ANIMAL STUDIES

Animal models of preeclampsia can provide a unique possibility for understanding the causal relationship and the molecular networks of preeclampsia-induced offspring pathology. Unfortunately, there is currently no perfect animal model of preeclampsia due to the complex and poorly understood pathophysiology of this disease (see Table 1 for an overview). Most of the presented models are based only on one pathophysiological feature, failing to reproduce the whole spectrum of preeclampsia characteristics. It is important to unravel whether all these experimental pathophysiological changes, which appear during preeclampsia, contribute to partial or complete cardiovascular and metabolic changes in the offspring. The use of several animal models of preeclampsia could help to distinguish the independent and/or dependent contribution of each of these factors to the developmental programming of offspring health. Below, we will discuss the animal studies that involve offspring follow-up after induction of major pathophysiological conditions of preeclampsia, excluding the genetic or surgically induced animal models of preeclampsia.

Angiogenic Disparity

Angiogenic dysbalance is a well-known feature of preeclampsia. The antiangiogenic factors soluble fms-like tyrosine kinase-1 (sFlt-1) and soluble endoglin (sEng) are increased in preeclamptic patients, after the 30th week of pregnancy [37]. Both sFlt-1 and sEng promote vascular dysfunction and capillary permeability, liver dysfunction, and neurological abnormalities via antagonization of proangiogenic factors such as VEGF and PlGF or TGF β signaling [38–40].

Adenoviral overexpression of sFlt-1 mimics the clinical course of preeclampsia in rodents [38]. Fetuses in this model show restricted growth that can be maintained until adulthood or can show catch-up growth until the age of 6 mo (Table 1) [41, 42]. Solely, sFlt-1 exposure during pregnancy imposes sex-specific glucose and/or insulin responses (to a glucose bolus) in the offspring, suggesting sex-specific differences in developmental programming of glucose metabolism. In addition, hypertension was observed only in the male offspring [41–43]. Sex-specific offspring outcomes are poorly understood, but one possible reason can be sexually dimorphic adaptations of the placenta [44].

When another environmental stressor, such as maternal obesity, is introduced during sFlt-1-induced preeclampsia, the offspring's birth weight is not compromised. On the contrary, several biochemical parameters such as blood glucose, cholesterol, triglycerides, and leptin are increased in combination with increased fat tissue depositions and aberrant carotid vascular reactivity in both sexes [43, 45]. This may indicate that a single antiangiogenic intrauterine insult can influence sexual dimorphic changes in placenta by priming the males towards hypertensive phenotype, but this is not sufficient for profound metabolic alterations without additional trigger factors.

Unfortunately, the effects of increased sEng on offspring health are still largely unknown. In vivo studies in mice have

TABLE 1. Spectrum of cardiometabolic alterations in offspring from pre-eclamptic mothers (animal models).^a

Model	Species/strain	Offspring outcome	Offspring age	Reference
sFlt-1 overexpression	CD-1 mice	Catch up growth IPGTT variations	24 wk	[42]
sFlt-1 overexpression	CD-1 mice	Hypertension in males Low BW in comparison to controls	9 wk	[41]
sFlt-1 overexpression	CD-1 mice	Hypertension in male offspring Metabolic changes: hypercholesterolemia, hyperleptinemia in females, and hypertriglyceridemia in males	24 wk	[45]
Second impact: prepregnancy obesity		+ Second impact: more detrimental effect on weight gain and metabolic effects		
Prepregnancy obesity and sFlt-1 overexpression	CD-1 mice	Fasting glucose increased in males Altered vascular responsiveness in both sexes	12 wk	[43]
AT1 AA immunization Second impact: high sugar diet 20% sucrose	Wistar rats	Insulin resistance + Second impact: altered lipid and glucose profile without hypertension	40 wk	[54]
AT1 AA passive immunization	Wistar rats	Myocardial remodelling	3 wk	[143]
AT1 AA passive immunization	C67Bl/6J mice	Abnormal kidney and liver development	GD 18	[53]
LPS injections	Sprague Dawley rats	Increased BW and fat deposits, hyperleptinemia, hypertension (no major sex-specific effects)	24 wk	[60]
LPS injections	Sprague Dawley rats	Hypertension Proteinuria Decreased glomeruli	25 wk	[61]
LPS injections	Sprague Dawley rats	Hypertension Left ventricle hypertrophy	32 wk	[62]
LPS injections	CD-1 mice	Decreased body weight Impaired spermatogenesis	35 wk	[64]
LPS injections + HF diet during pregnancy until 3 mo of age	Sprague Dawley rats	Hypertension Insulin resistance	12 wk	[65]
LPS injections Second impact: HF diet	ICR mice	Metabolic phenotype altered only due to the HF diet	No data	[66]

^a IPGTT, intraperitoneal glucose tolerance test; BW, body weight; AT1 AA, angiotensin II type I receptor autoantibodies; LPS, lipopolysaccharide; HF, high fat; GD, gestational day.

shown that direct administration of sEng increases the vascular resistance and subsequently the blood pressure [46]. In patients with diabetes and hypertension, sEng is positively correlated with the basal glucose levels, suggesting a potential role in glucose metabolism [47]. In accordance with previous findings and the known synergistic effect of sEng and sFlt-1 on preeclampsia outcome, we can speculate on the effects on offspring health in a similar or superimposed manner.

Angiotensin II Type I Receptor Antibodies

Angiotensin II type I receptor autoantibodies (AT1 AA) are found in 70%–95% of women diagnosed with preeclampsia, compared to 30% of healthy controls. A higher antibody titer is proportionally correlated to the severity of the disease [48, 49]. In addition, AT1 AA display an agonistic effect on the AT1 receptor, promoting vasoconstriction and aldosterone secretion, in a manner similar to angiotensin II [50–52].

Passive immunization with AT1 AA in rodents is associated with the development of proteinuria and hypertension at the end of pregnancy [53, 54]. The fetuses show growth restriction and remodeling in several organs, such as the liver, heart, and kidney. At the histopathological level, glomerular loss, myocardial apoptosis, and immature cell liver infiltration are observed in the offspring, suggesting an adaptive decline in fetal growth and organogenesis, possibly due to maternal-fetal transfer of AT1 AA. Irani et al. [53] reported unaffected

functionality of these transported antibodies, and successful activation of fetal AT1 receptors may contribute to systemic vasoconstriction and hypoxia that can predispose the offspring to organ maladaptation.

Zhang et al. [54] did long-term follow-up on offspring derived from dams actively immunized against AT1 receptor antigen. Middle-age checkup at 10 mo of age showed elevated fasting insulin levels and an increased homeostasis model assessment index, suggesting the development of insulin resistance (Table 1). This was expected, especially because AT1 receptors are involved in insulin signaling of beta cells [55]. An additional 2 mo feeding with a high sugar diet of these adult offspring leads to even more pronounced metabolic alterations such as increased triglycerides, decreased high-density cholesterol, impaired glucose tolerance, and enlarged visceral fat depositions [54]. All these alterations are contributors to the progression of the metabolic syndrome. Surprisingly, blood pressure was normal in these animals, although they had been exposed to the AT1 antibodies in utero and in the weaning period via the maternal milk. One possible interpretation is that intrarenal angiotensin II, contrary to plasma angiotensin II, may be positively involved in blood pressure regulation. Another important comment is that vascular endothelium has relatively large regeneration capacities, and if there is no constant provocation with AT1 antibodies, no endothelial-related rise in blood pressure will occur.

In sum, AT1 antibody exposure does not affect the fetal blood pressure but can have detrimental effects on organ formation and insulin resistance, which can be potentiated with an unhealthy diet. Nevertheless, more studies are needed in order to elucidate the underlying mechanisms of AT1 AA-induced fetal metabolic programming.

Inflammatory Milieu

Mild inflammation is generally considered a normal feature of pregnancy, whereas more exaggerated systemic inflammatory responses are characteristic of preeclampsia [9]. In accordance, proinflammatory cytokine concentrations are increased (TNF α , IL-6, IL-1 β) in preeclamptic patients [56–58]. The association between inflammation and preeclampsia served as the basis for an experimental animal model of preeclampsia by low-dose intravenous infusion of bacterial endotoxin [59]. Nowadays, most of the developmental studies that involve exposure to lipopolysaccharide (LPS) during pregnancy are focused on the immunological consequences without concentrating on the possible preeclamptic symptoms in the dam.

Midgestational LPS exposure is characterized by a large range of cardiovascular events such as increased blood pressure, aortal vascular impairment, left ventricular hypertrophy, diastolic dysfunction, and myocardial apoptosis in adult offspring, without specific sex differences [60–64]. This implies striking endothelial and cardiac sensitivity of the fetus for inflammation that is maintained until adulthood, programming the offspring's health toward cardiovascular functional decline. This, in part, can be explained by upregulation of the NF- κ B signaling pathway, an increase of reactive oxygen species (ROS), and downregulation of the renal dopaminergic system leading to hypertension and vascular instability [64].

Combined effects of LPS and high-fat diet exposure during pregnancy have differential effects on offspring's glucose and lipid metabolism (Table 1). It was shown that midgestation exposure to LPS and high-fat feeding until 3 mo of age can lead to impaired liver function and insulin resistance [65]. On the contrary, exposure to LPS in late gestation with additional high-fat diet stress after the lactation period did not result in an impaired metabolic phenotype in the offspring [66]. This suggests that timing of LPS exposure is crucial for fetal metabolic programming and in part can be explained by changes in maturational properties of the placenta, which in the last term of pregnancy are fully developed, possibly resulting in placental impermeability for the intermediate metabolic effectors of LPS [67]. Another important observation is that LPS and high-fat diet combined have a beneficial effect on blood pressure and the inflammatory response in the offspring, but not on the insulin resistance progression and liver dysfunction. Midgestation exposure to LPS seems to attenuate the offspring sensitivity to high-fat diet-induced inflammation [65]. In contrast, an aberrant inflammatory response on its own is not sufficient for a systemic breakdown in the regulation of insulin resistance.

Altogether, the data indicate that the developmental programming of offspring health via preeclampsia is caused by a two-hit combination of, first, systemic immunomodulatory and antiangiogenic signals during mid to late gestation and, second, a later host susceptibility marked by unhealthy lifestyle (e.g., a Western diet). These animal data have important translational consequences because the first hit is needed to affect the offspring's development, and the presence of the second hit explains why only a minority of human fetuses

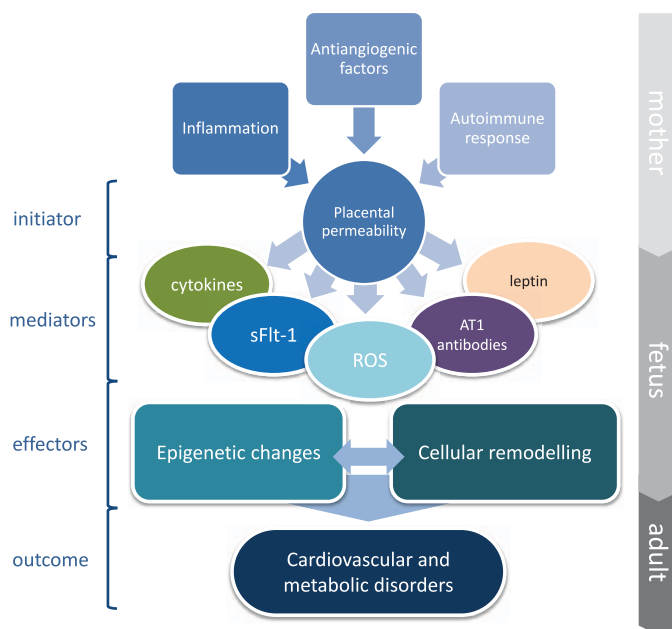


FIG. 1. The impact of preeclampsia on offspring/adult health. Schematic diagram of how possible pre-eclamptic scenarios are shaping the intrauterine environment, influencing the placental structure (initiator), and imposing unfavorable signaling network (mediators) in the offspring. The latter can program several aspects of the metabolic and cardiovascular system, mainly via organ remodeling and epigenetic modulation of gene expression (effectors).

exposed to preeclampsia develop detrimental cardiovascular and metabolic diseases later.

UNDERLYING MECHANISMS OF DEVELOPMENTAL PROGRAMMING

In order to interpret the developmental programming of cardiometabolic health via preeclampsia, we underline below the conserved mechanisms of chronic disease development, their interaction with the preeclamptic environment, and their effects on embryonic growth and epigenetic status (Fig. 1). Understanding the specific mechanisms by which preeclampsia impacts offspring welfare is crucial for developing appropriate strategies to improve the negative effects of the harsh intrauterine environment.

PLACENTAL PERMEABILITY: THE INITIATOR

The placental blood barrier serves as a protector and nutrient sensor between the mother and the child. In preeclampsia, placental morphology is perturbed showing superficial trophoblast invasion and insufficient remodeling of spiral arteries in the myometrium [68, 69]. Thus, with this defective placentation, two separate factors can influence its permeability: the placental composition and the exchange surface area.

Several factors influence placental composition, including an intact syncytiotrophoblast layer and cellular junctional assembly. The syncytiotrophoblast, a continuous membrane layer of the placenta, serves as a checkpoint for placental transport [70–72]. During preeclampsia, this layer is highly apoptotic [73], suggesting a dysfunctional adaptation of the placenta to increased fetal nutrient demand or an effect of the increased proinflammatory cytokines during preeclampsia. Consistent with this idea, tight junctions integral membrane proteins, important for paracellular transport of water and

nutrients, are extremely susceptible to TGF β and IL-1 β destruction, which can be reversed *in vitro* by specific cytokine inhibitors [74, 75]. Given that discontinuous placental membranes are accompanied with increased porosity, it is probable that loss of syncytiotrophoblast integrity underlines the defective nutrient transport of preeclamptic placenta.

An altered placental surface area in preeclampsia has been reported along with decreased placental weight, changes in the shape, and increased thickness, probably due to compensatory mechanisms [76]. The growth of the placenta was reported to be compromised only on the minor axis and corresponded to the severity of the preeclampsia. This axis is speculated to coincide with mediolateral development of the fetus, suggesting that this area is not spared during preeclampsia. In accordance, fetal length is less compromised in offspring in comparison with a severely affected abdominal circumference [34, 77].

A central question is whether these structural changes in the preeclamptic placenta are determinants for the transport of pathological signaling molecules to the fetus. It is known that inflammatory cytokines and AT1 AA can cross the placenta, but no data on the transfer of antiangiogenic molecules to the fetus is available [53, 78]. These molecules can act as potent signaling modifiers of glucose and lipid metabolism, but a definitive description of their mechanisms of action is lacking. Currently, we rely solely on animal data, for example, being challenged with an inflammatory cytokine such as TNF α induces insulin receptor downregulation in the liver that promotes the development of liver insulin resistance [79, 80]. An IL-6 challenge in rodents showed increased mobilization of acyl-CoA, a metabolic active form of fatty acid, in the skeletal muscles that has been strongly associated with lipid accumulation in muscles and peripheral insulin resistance [79–81]. AT1 AA exposure has detrimental effects on function of the liver, by NF- κ B and NADPH oxidase dependent release of ROS [82, 83]. Ubiquitous exposure to sFlt-1 leads to hypovascularization in several organs, including pancreas and adipose tissue, that in turn can affect the beta cell mass and energy expenditure of the adipose tissue [84, 85]. In addition, prolonged exposure to sFlt-1 is involved in the development of diastolic dysfunction and heart failure [86]. Taken together, increased exposure of the fetal organism to these molecules may have a detrimental effect on proper metabolic and cardiovascular functioning.

ROS: THE MEDIATORS

Multiple lines of evidence suggests that oxidative damage is one of the underlying mechanisms of many chronic diseases such as type 2 diabetes, obesity, hypertension, atherosclerosis, and metabolic syndrome [87, 88]. Oxidative stress occurs as soon as the production and consumption of ROS are imbalanced.

During preeclampsia, inflammatory cytokines and AT1 AA promote increased ROS production by up to 40% when compared to control placentas [89–92]. Whether oxidative distress is a cause or consequence of placental dysfunction and/or fetal nutrient demand is a matter of ongoing debate, and most studies simply describe an association of ROS overflow with metabolic consequences rather than mechanistic connections.

Normal fetal development is dependent on tightly controlled oxidative stress exposure for optimal cellular signaling, differentiation, and proliferation [93]. However, during preeclampsia the functionality of the placenta is reduced and the antioxidant capacity is diminished, suggesting increased

oxidative stress transfer to the fetus. Cord blood analyzed from preeclamptic mothers showed either decreased antioxidant activity [94] or increased oxidative stress markers [95–98], but not for all [99], suggesting a possibility of lipid peroxidation and protein inactivation in the fetus [100]. Several tissues are extremely susceptible to oxidative damage, including beta cells and vascular endothelium mainly due to low cytoprotective mechanisms [101, 102]. Furthermore, treatment of hypoxic dams with antioxidants during gestation ameliorates the vascular dysfunction in the offspring, indicating that antioxidant treatment may indeed be an interventional treatment [103]. On the contrary, clinical trials that involved routine antioxidant supplementation during pregnancy contradict the idea of preventive effect towards preeclampsia [104, 105]. Another trial that included only high-risk preeclampsia patients reported protective effect of antioxidant vitamins in combination with L-arginine [106], suggesting that exclusive antioxidant treatment is not sufficient to combat preeclampsia.

LEPTIN SIGNALING AS MEDIATOR

Leptin is a satiety hormone, and acting via JAK2/STAT3 and PI3K-Akt signaling pathways, leptin has a major impact on energy homeostasis, body composition, and appetite in early fetal and later adult life [107]. Moreover, leptin expression is responsive to the intrauterine and fetal environment, showing overexpression in monochorionic twin placenta only on the side of the growth restricted fetus [108]. The extent to which leptin signaling is implicated in overall fetal metabolism is unknown, but there is evidence that it stimulates fatty acid oxidation in muscles, increases the glucose turnover in brain, heart, and brown adipose tissue, and inhibits global lipid accumulation [109]. By contrast, reduction of leptin concentrations and the state of leptin resistance share similar effects on metabolism, promoting hyperinsulinemia and hyperglycemia. During preeclampsia, maternal leptin concentrations show a 2-fold increase in comparison to control subjects, irrespective of the BMI [110–114], which is usually combined with hyperinsulinemia, altered lipid profile, and decreased 2-methoxyestradiol, which serves as an important vasoprotector and vasodilator [115, 116]. There is a dichotomy, therefore, between the protective metabolic effects of leptin and apparently deleterious effects of hyperleptinemia on maternal health. In part, this can be explained by a development of leptin resistance, mainly due to inactivation of STAT3 intracellular activity that is also decreased in preeclamptic pregnancies [117].

Fetal cord leptin concentrations are increased [110, 118] in preeclamptic pregnancies, possibly due to increased nutrient demand of the fetus and/or increased placental permeability and consequent leptin flow to the fetus. Hyperleptinemia *in utero* can alter the adrenal responsiveness [56, 119] in the fetus and together with increased inflammatory markers can develop a defense mechanism of leptin resistance, which ultimately can lead to deleterious effects on cardiovascular and metabolic health.

CELLULAR ADAPTATIONS AS EFFECTORS

Optimal organ functioning is dependent on the quantity, morphology, and functionality of relevant cell types due to appropriate differentiation of pluripotent embryonic stem cells. Although the mechanisms of embryonic cell fate decisions are obscure, the presence of low energy levels and prominent signaling networks are strongly correlated with disturbed metabolic stem cell fate [120–122]. Importantly, all these

adverse conditions are also present in utero during preeclampsia.

Moreover, several reports showed changes in the number of nephrons, beta cells, and/or cardiomyocytes in offspring exposed to a harsh intrauterine environment [123–126]. A decreased number of nephrons contributes to low rates of renal ultrafiltration that affects blood circulating volume, which ultimately can lead to increased blood pressure [126]. Decreased beta-cell mass adaptation due to early life stressors, such as undernutrition and placental insufficiency, possibly can have an influence on later disease development, for example, diabetes mellitus [127]. Initial heart size has an influence on the end diastolic volume and serves as a predictive index for myocardial disease [24].

These (mal)adaptive changes are observed mainly in organs constructed from long-lived postmitotic cells [123, 128]. Because these cells are not—or rarely—dividing cells, their development during intrauterine life is extremely important in order to prepare them for long-term functionality. Tightly controlled processes regulate cell number while their functioning is dependent on specific signaling molecules and energy sources. During preeclampsia, the increased concentration of inflammatory markers and improper vascular signaling molecules might perturb these regulatory processes essential in organ formation [129]. Combined with poor nutrient supply via the placenta, this can lead to detrimental effects on offspring health.

EPIGENETIC CHANGES AS EFFECTORS

Exposure to different environmental stimuli, especially during critical windows of development, results in the formation of adaptive epigenetic marks as part of the adaptive stress response [5]. The epigenetic marking system includes changes in DNA methylation, histone modifications, and noncoding RNA (ncRNA) expression. Usually, they are established early in development and act as regulators of developmental, tissue, and sex-specific gene expression [130–133].

DNA methylation is a unique form of gene regulation because it involves direct covalent modification within the genome and can provide long-term stability in a heritable transgenerational way [134]. Methylation of important regulatory sites, for example, gene promoters or enhancers, is mostly connected to gene repression, resulting in gene expression downregulation [135].

DNA methylation analysis of cord blood cells is a valuable target for studying the early epigenetic consequences of preeclampsia on the fetus. Several studies analyzed DNA methylation of genes involved in fetal growth and development that are also highly sensitive to environmental perturbations. Hypomethylation has been observed in the promoter region of the 11 β -hydroxysteroid dehydrogenase type 2 (HSD11B2) in cord blood samples from neonates exposed to preeclampsia [136]. Decreased methylation was also reported for insulin-like growth factor 2 (IGF2) in the differentially methylated regions, important for gene regulation of imprinted genes [137]. By contrast, in preeclamptic placentas, HSD11B2 and IGF2 gene expression levels are decreased [138, 139]. Therefore, there is a discrepancy between the reported hypomethylated status and the observed downregulated activity of these genes in other studies. It is tempting to speculate that this is a compensatory change in methylation to ensure favorable offspring functioning, but on the other side, it can be an atypical decrease in gene expression that can lead to metabolic maladaptation.

A recent study used a genomewide methylation analysis in which neonatal cord blood DNA from mothers diagnosed with early onset preeclampsia showed promoter hypo- or hypermethylation for different subsets of genes. Prominent DNA modifications were primarily discovered in genes involved in lipid metabolism and inflammation, pointing out that early epigenetic disruptions can be seen in preeclamptic children [140]. Altogether, these findings support an effect of preeclampsia on the methylation status of the neonates cord blood, but it is unclear whether this is a protective or maladaptive effect. Although this does not prove any causal relationship with long-term health effects, it can be used as an initial proof of principle for conduction of new cohort studies.

To our knowledge, there are no data concerning histone modifications and/or ncRNAs in offspring from preeclamptic mothers. Communication between DNA methylation and chromatin modifiers or promoter regions of ncRNAs has been established [141, 142], and abnormal methylation either solely or via other epigenetic marks can be an important mediator of fetal metabolism. It is becoming clear that these molecules are implicated in several diseases, and successful unclosing of their role in developmental programming can lead to possible biological biomarkers or targets for therapy.

CONCLUDING REMARKS

Taking into consideration the great amount of evidence, it is reasonable to suggest that preeclampsia constrains the cardiometabolic health of the offspring. Still, it remains difficult to estimate the degree of involvement of preeclampsia into the cardiovascular and metabolic health programming of the offspring. The major obstacle is the presence of multiple pathophysiological pathways implicated in the development and the clinical course of preeclampsia that may influence each other or act independently all at once or in series. Depending on which mechanism is dominantly involved, and which secondary environmental stressors are present, different aspects of the metabolism and the cardiovascular system can be affected. The role of the placenta as a central initiator of long-term preeclamptic consequences in the offspring is just the beginning of what needs to be explored (Fig. 1). However, understanding of all mechanisms by which preeclampsia alters fetal growth and development and later on programs it toward chronic disorders is crucial for identification of individuals at risk and for development of future clinical interventions or prevention strategies.

REFERENCES

1. World Health Organization. WHO Noncommunicable Diseases Country Profiles. Geneva, Switzerland: WHO Press; 2014: 2014.
2. Pollex RL, Hegele RA. Genetic determinants of the metabolic syndrome. *Nat Clin Pract Cardiovasc Med* 2006; 3:482–489.
3. Mitchell BD, Imumorin IG. Genetic determinants of diabetes and atherosclerosis. *Curr Atheroscler Rep* 2002; 4:193–198.
4. Norman RE, Carpenter DO, Scott J, Brune MN, Sly PD. Environmental exposures: an underrecognized contribution to noncommunicable diseases. *Rev Environ Health* 2013; 28:59–65.
5. Jiménez-Chillarón JC, Díaz R, Martínez D, Pentinat T, Ramón-Krauel M, Ribó S, Plösch T. The role of nutrition on epigenetic modifications and their implications on health. *Biochimie* 2012; 94:2242–2263.
6. Carter AM. Evolution of placental function in mammals: the molecular basis of gas and nutrient transfer, hormone secretion, and immune responses. *Physiol Rev* 2012; 92:1543–1576.
7. Duley L. The global impact of pre-eclampsia and eclampsia. *Semin Perinatol* 2009; 33:130–137.
8. Laresgoiti-Servitje E, Gomez-Lopez N. The pathophysiology of preeclampsia involves altered levels of angiogenic factors promoted by

- hypoxia and autoantibody-mediated mechanisms. *Biol Reprod* 2012; 87: 36–36.
9. Redman CWG, Sargent IL. Immunology of pre-eclampsia. *Am J Reprod Immunol* 2010; 63:534–543.
 10. Srinivas SK, Edlow AG, Neff PM, Sammel MD, Andrela CM, Elovitz MA. Rethinking IUGR in preeclampsia: dependent or independent of maternal hypertension? *J Perinatol* 2009; 29:680–684.
 11. Weiler J, Tong S, Palmer KR. Is fetal growth restriction associated with a more severe maternal phenotype in the setting of early onset pre-eclampsia? A retrospective study. *PLoS One* 2011; 6:e26937.
 12. Xiao R, Sorensen TK, Williams MA, Luthy DA. Influence of pre-eclampsia on fetal growth. *J Matern Fetal Neonatal Med* 2003; 13: 157–162.
 13. Ødegård RA, Vatten LJ, Nilsen ST, Salvesen KÅ, Austgulen R. Preeclampsia and fetal growth. *Obstet Gynecol* 2000; 96:950–955.
 14. Eriksson JG, Forse T, Tuomilehto J, Osmond C. Early growth, adult income, and risk of stroke. *Stroke* 2000; 31:869–875.
 15. Jong M, Lafeber HN, Cranendonk A, van Weissenbruch MM. Components of the metabolic syndrome in early childhood in very-low-birth-weight infants. *Horm Res Paediatr* 2014; 81:43–49.
 16. Miller FC, Read JA, Cabal L, Siassi B. Heart rate and blood pressure in infants of pre-eclamptic mothers during the first hour of life. *Crit Care Med* 1983; 11:532–535.
 17. Teng RJ, Wu TJ, Sharma R, Garrison RD, Hudak ML. Early neonatal hypotension in premature infants born to preeclamptic mothers. *J Perinatol* 2006; 26:471–475.
 18. Davis EF, Newton L, Lewandowski AJ, Lazdam M, Kelly BA, Kyriakou T, Leeson P. Pre-eclampsia and offspring cardiovascular health: mechanistic insights from experimental studies. *Clin Sci* 2012; 123: 53–72.
 19. Tenhola S, Rahiala E, Martikainen A, Halonen P, Voutilainen R. Blood pressure, serum lipids, fasting insulin, and adrenal hormones in 12-year-old children born with maternal preeclampsia. *J Clin Endocrinol Metab* 2003; 88:1217–1222.
 20. Miettola S, Hartikainen AL, Väärasmäki M, Bloigu A, Ruokonen A, Järvelin MR, Pouta A. Offspring's blood pressure and metabolic phenotype after exposure to gestational hypertension in utero. *Eur J Epidemiol* 2013; 28:87–98.
 21. Fraser A, Nelson SM, MacDonald-Wallis C, Sattar N, Lawlor DA. Hypertensive disorders of pregnancy and cardiometabolic health in adolescent offspring. *Hypertension* 2013; 62:614–620.
 22. Tenhola S, Rahiala E, Halonen P, Vanninen E, Voutilainen R. Maternal preeclampsia predicts elevated blood pressure in 12-year-old children: evaluation by ambulatory blood pressure monitoring. *Pediatr Res* 2006; 59:320–324.
 23. Lim W, Lee Y, Yap FK, Aris IM, Ngee L, Meaney M, Gluckman PD, Godfrey KM, Kwek K, Chong Y, Saw S. Maternal blood pressure during pregnancy and early childhood blood pressures in the offspring: the GUSTO birth cohort study. *Med* 2015; 94:1–9.
 24. Fugelseth D, Ramstad HB, Kvehaugen AS, NESTAAS E, Støylen A, Staff AC. Myocardial function in offspring 5-8 years after pregnancy complicated by preeclampsia. *Early Hum Dev* 2011; 87:531–535.
 25. Brodwall K, Leirgul E, Greve G, Vollset SE, Holmstrøm H, Tell GS, Øyen N. Possible common aetiology behind maternal preeclampsia and congenital heart defects in the child: a cardiovascular diseases in Norway project study. *Paediatr Perinat Epidemiol* 2015; 30:76–85.
 26. Auger N, Fraser WD, Healy-Profítos J, Arbour L. Association between preeclampsia and congenital heart defects. *JAMA* 2015; 314:1588.
 27. Kajantie E, Eriksson JG, Osmond C, Thornburg K, Barker DJP. Preeclampsia is associated with increased risk of stroke in the adult offspring the Helsinki birth cohort study. *Stroke* 2009; 40:1176–1180.
 28. Jayet PY, Rimoldi SF, Stuber T, Salinas Salmòn C, Hutter D, Rexhaj E, Thalmann S, Schwab M, Turini P, Sartori-Cucchia C, Nicod P, Villena M, et al. Pulmonary and systemic vascular dysfunction in young offspring of mothers with preeclampsia. *Circulation* 2010; 122:488–494.
 29. Hwang HS, Maeng YS, Park YW, Koos BJ, Kwon YG, Kim YH. Increased senescence and reduced functional ability of fetal endothelial progenitor cells in pregnancies complicated by preeclampsia without intrauterine growth restriction. *Am J Obstet Gynecol* 2008; 199:1–7.
 30. Muñoz-Hernandez R, Miranda ML, Stiefel P, Lin RZ, Praena-Fernández JM, Dominguez-Simeon MJ, Villar J, Moreno-Luna R, Melero-Martín JM. Decreased level of cord blood circulating endothelial colony-forming cells in preeclampsia. *Hypertension* 2014; 64:165–171.
 31. Rodie VA, Caslake MJ, Stewart F, Sattar N, Ramsay JE, Greer IA, Freeman DJ. Fetal cord plasma lipoprotein status in uncomplicated human pregnancies and in pregnancies complicated by pre-eclampsia and intrauterine growth restriction. *Atherosclerosis* 2004; 176:181–187.
 32. Wu CS, Nohr EA, Bech BH, Vestergaard M, Catov JM, Olsen J. Health of children born to mothers who had preeclampsia: a population-based cohort study. *Am J Obstet Gynecol* 2009; 201:269.
 33. Washburn L, Nixon P, Russell G, Snively BM, O'Shea TM. Adiposity in adolescent offspring born prematurely to mothers with preeclampsia. *J Pediatr* 2013; 162:912–917.
 34. Ophir E, Dourleshter G, Hirsh Y, Fait V, German L, Bornstein J. Newborns of pre-eclamptic women: a biochemical difference present in utero. *Acta Obstet Gynecol Scand* 2006; 85:1172–1178.
 35. Guillemette L, Lacroix M, Allard C, Patenaude J, Battista M-C, Doyon M, Moreau J, Ménard J, Ardilouze J-L, Perron P, Côté A-M, Hivert M-F. Preeclampsia is associated with an increased pro-inflammatory profile in newborns. *J Reprod Immunol* 2015; 112:111–114.
 36. Seppä S, Voutilainen R, Tenhola S. Markers of insulin sensitivity in 12-year-old children born from preeclamptic pregnancies. *J Pediatr* 2015; 167:125–130.
 37. Powers RW, Jeyabalan A, Clifton RG, van Dorsten P, Hauth JC, Klebanoff MA, Lindheimer MD, Sibai B, Landon M, Miodovnik M. Soluble fms-like tyrosine kinase 1 (sFlt1), endoglin and placental growth factor (PlGF) in preeclampsia among high risk pregnancies. *PLoS One* 2010; 5:e13263.
 38. Maynard SE, Min J, Merchan J, Lim K, Li J, Mondal S, Libermann TA, Morgon JP, Sellke FW, Stillman IE, Epstein FH, Sukhatme VP, et al. Excess placental soluble fms-like tyrosine kinase 1 (sFlt1) may contribute to endothelial dysfunction, hypertension, and proteinuria in preeclampsia. *J Clin Invest* 2003; 111:649–658.
 39. Chaiworapongsa T, Romero R, Espinoza J, Bujold E, Mee Kim Y, Gonçalves LF, Gomez R, Edwin S. Evidence supporting a role for blockade of the vascular endothelial growth factor system in the pathophysiology of preeclampsia: Young Investigator Award. *Am J Obstet Gynecol* 2004; 190:1541–1550.
 40. Levine RJ, Lam C, Qian C, Yu KF, Maynard SE, Sachs BP, Sibai BM, Epstein FH, Romero R, Thadhani R, Karumanchi SA. Soluble endoglin and other circulating antiangiogenic factors in preeclampsia. *N Engl J Med* 2006; 355:992–1005.
 41. Lu F, Bytautiene E, Tamayo E, Gamble P, Anderson GD, Hankins GDV, Longo M, Saade GR. Gender-specific effect of overexpression of sFlt-1 in pregnant mice on fetal programming of blood pressure in the offspring later in life. *Am J Obstet Gynecol* 2007; 197:1–5.
 42. McDonnold M, Tamayo E, Kechichian T, Gamble P, Longo M, Hankins GDV, Saade GR, Costantine MM. The effect of prenatal pravastatin treatment on altered fetal programming of postnatal growth and metabolic function in a preeclampsia-like murine model. *Am J Obstet Gynecol* 2014; 210:542.
 43. Byers BD, Betancourt A, Lu F, Hankins GDV, Longo M, Saade GR, Bytautiene E. The effect of prepregnancy obesity and sFlt-1-induced preeclampsia-like syndrome on fetal programming of adult vascular function in a mouse model. *Am J Obstet Gynecol* 2009; 200:432.
 44. Pruis MG, Gellhaus A, Kühnel E, Lendvai Á, Bloks VW, Groen AK, Plösch T. Sex-specific placental differences as a contributor to sex-specific metabolic programming? *Acta Physiol* 2015; 215:127–129.
 45. Bytautiene E, Tamayo E, Kechichian T, Drever N, Gamble P, Hankins GDV, Saade GR. Prepregnancy obesity and sFlt1-induced preeclampsia in mice: developmental programming model of metabolic syndrome. *Am J Obstet Gynecol* 2011; 204:398.
 46. Venkatesha S, Toporsian M, Lam C, Hanai J, Mammoto T, Kim YM, Bdolah Y, Lim K-H, Yuan H-T, Libermann TA, Stillman IE, Roberts D, et al. Soluble endoglin contributes to the pathogenesis of preeclampsia. *Nat Med* 2006; 12:642–649.
 47. Blázquez-Medela AM, García-Ortiz L, Gómez-Marcos MA, Recio-Rodríguez JI, Sánchez-Rodríguez A, López-Novoa JM, Martínez-Salgado C. Increased plasma soluble endoglin levels as an indicator of cardiovascular alterations in hypertensive and diabetic patients. *BMC Med* 2010; 8:86.
 48. Siddiqui AH, Irani RA, Blackwell SC, Ramin SM, Kellems RE, Xia Y. Angiotensin receptor agonistic autoantibody is highly prevalent in preeclampsia: correlation with disease severity. *Hypertension* 2010; 55: 386–393.
 49. Herse F, Lamarca B. Angiotensin II type 1 receptor autoantibody (AT1-AA)-mediated pregnancy hypertension. *Am J Reprod Immunol* 2013; 69: 413–418.
 50. Wallukat G, Homuth V, Fischer T, Lindschau C, Horstkamp B, Büpner A, Baur E, Nissen E, Vetter K, Neichel D, Dudenhausen JW, Haller H, et al.

- Patients with preeclampsia develop agonistic autoantibodies against the angiotensin AT1 receptor. *J Clin Invest* 1999; 103:945–952.
51. Wenzel K, Rajakumar A, Haase H, Geusens N, Hubner N, Schulz H, Brewer J, Roberts L, Hubel CA, Herse F, Hering L, Qadri F, et al. Angiotensin II type 1 receptor antibodies and increased angiotensin II sensitivity in pregnant rats. *Hypertension* 2011; 58:77–84.
 52. Hubel CA, Wallukat G, Wolf M, Herse F, Rajakumar A, Roberts JM, Markovic N, Thadhani R, Luft FC, Dechend R. Agonistic angiotensin II type 1 receptor autoantibodies in postpartum women with a history of preeclampsia. *Hypertension* 2007; 49:612–617.
 53. Irani RA, Zhang Y, Blackwell SC, Zhou CC, Ramin SM, Kellems RE, Xia Y. The detrimental role of angiotensin receptor agonistic autoantibodies in intrauterine growth restriction seen in preeclampsia. *J Exp Med* 2009; 206:2809–2822.
 54. Zhang S, Zhang X, Yang L, Yan Z, Yan L, Tian J, Li X, Song L, Wang L, Yang X, Zheng R, Lau WB, et al. Increased susceptibility to metabolic syndrome in adult offspring of angiotensin type 1 receptor autoantibody-positive rats. *Antioxid Redox Signal* 2012; 17:733–743.
 55. Chu KY, Tung L, Carlsson P-O, Leung PS. Angiotensin II. Type 1 receptor blockade improves β -cell function and glucose tolerance in a mouse model of type 2 diabetes. *Diabetes* 2006; 55:367–374.
 56. Lockwood CJ, Yen C-F, Basar M, Kayisli UA, Martel M, Buhimschi I, Buhimschi C, Huang SJ, Krikun G, Schatz F. Preeclampsia-related inflammatory cytokines regulate interleukin-6 expression in human decidual cells. *Am J Pathol* 2008; 172:1571–1579.
 57. Sharma A, Satyam A, Sharma JB. Leptin, IL-10 and inflammatory markers (TNF- α , IL-6 and IL-8) in pre-eclamptic, normotensive pregnant and healthy non-pregnant women. *Am J Reprod Immunol* 2007; 58:21–30.
 58. Lau SY, Guild SJ, Barrett CJ, Chen Q, Mccowan L, Jordan V, Chamley LW. Tumor necrosis factor- α , interleukin-6, and interleukin-10 levels are altered in preeclampsia: a systematic review and meta-analysis. *Am J Reprod Immunol* 2013; 70:412–427.
 59. Faas M, Schuiling G, Baller J, Visscher C, Bakker W. A new model for human preeclampsia: ultra-low-dose endotoxin infusion in pregnant rats. *Am J Obstet Gynecol* 1994; 171:158–164.
 60. Wei Y, Li X, Zhou J. Prenatal exposure to lipopolysaccharide results in increases in blood pressure and body weight in rats. *Acta Pharmacol Sin* 2007; 28:651–656.
 61. Hao X-Q, Zhang H-G, Yuan Z-B, Yang D-L, Hao L-Y, Li X-H. Prenatal exposure to lipopolysaccharide alters the intrarenal renin-angiotensin system and renal damage in offspring rats. *Hypertens Res* 2010; 33: 76–82.
 62. Wei Y, Du W, Xiong X, He X, Yi P, Deng Y, Chen D. Prenatal exposure to lipopolysaccharide results in myocardial remodeling in adult murine offspring. *J Inflamm* 2013; 10:35.
 63. Zhao S, Zhang H, Cao D, Liu Y, Li X. Lipopolysaccharide exposure during pregnancy leads to aortic dysfunction in offspring rats. *PLoS One* 2014; 9:e102273.
 64. Wang X, Luo H, Chen C, Chen K, Wang J, Cai Y, Zheng S, Yang X, Zhou L, Jose PA, Zeng C. Prenatal lipopolysaccharide exposure results in dysfunction of the renal dopamine D 1 receptor in offspring. *Free Radic Biol Med* 2014; 76:242–250.
 65. Hao XQ, Du JX, Li Y, Li M, Zhang SY. Prenatal exposure to lipopolysaccharide combined with pre- and postnatal high-fat diet result in lowered blood pressure and insulin resistance in offspring rats. *PLoS One* 2014; 9:e88127.
 66. Liu X, Wang B, Zhao M, Zhang C, Chen Y. Effects of maternal LPS exposure during pregnancy on metabolic phenotypes in female offspring. *PLoS One* 2014; 9:e114780.
 67. Malassiné A, Frenzo JL, Evain-Brion D. A comparison of placental development and endocrine functions between the human and mouse model. *Hum Reprod Update* 2003; 9:531–539.
 68. Noris M, Perico N, Remuzzi G. Mechanisms of disease: pre-eclampsia. *Nat Clin Pract Nephrol* 2005; 1:98–114.
 69. Lyall F, Robson SC, Bulmer JN. Spiral artery remodeling and trophoblast invasion in preeclampsia and fetal growth restriction relationship to clinical outcome. *Hypertension* 2013; 62:1046–1054.
 70. Sibley CP, Brownbill P, Dilworth M, Glazier JD. Review: adaptation in placental nutrient supply to meet fetal growth demand: Implications for programming. *Placenta* 2010; 31:S70–S74.
 71. Gude NM, Roberts CT, Kalonis B, King RG. Growth and function of the normal human placenta. *Thromb Res* 2004; 114:397–407.
 72. Powell TL, Jansson T, Illsley NP, Wennergren M, Korotkova M, Strandvik B. Composition and permeability of syncytiotrophoblast plasma membranes in pregnancies complicated by intrauterine growth restriction. *Biochim Biophys Acta Biomembr* 1999; 1420:86–94.
 73. Ishihara N, Matsuo H, Murakoshi H, Laoag-Fernandez JB, Samoto T, Maruo T. Increased apoptosis in the syncytiotrophoblast in human term placentas complicated by either preeclampsia or intrauterine growth retardation. *Am J Obstet Gynecol* 2002; 186:158–166.
 74. Liévano S, Alarcón L, Chávez-Munguía B, González-Mariscal L. Endothelia of term human placentae display diminished expression of tight junction proteins during preeclampsia. *Cell Tissue Res* 2006; 324: 433–448.
 75. Tossetta G, Paolinelli F, Avellini C, Salvolini E, Ciarmela P, Lorenzi T, Emanuelli M, Toti P, Giulianti R, Gesuita R, Crescimanno C, Voltolini C, et al. IL-1 β and TGF- β weaken the placental barrier through destruction of tight junctions: an in vivo and in vitro study. *Placenta* 2014; 35:509–516.
 76. Kajantie E, Thornburg KL, Eriksson JG, Osmond C, Barker DJP. In preeclampsia, the placenta grows slowly along its minor axis. *Int J Dev Biol* 2010; 54:469–473.
 77. Rasmussen S, Irgens LM. Fetal growth and body proportion in preeclampsia. *Obstet Gynecol* 2003; 101:575–583.
 78. Dahlgren J, Samuelsson AM, Jansson T, Holmäng A. Interleukin-6 in the maternal circulation reaches the rat fetus in mid-gestation. *Pediatr Res* 2006; 60:147–151.
 79. Miles PDG, Romeo OM, Higo K, Cohen A, Rafaat K, Olefsky JM. TNF α -induced insulin resistance in vivo and its prevention by troglitazone. *Diabetes* 1997; 46:1678–1683.
 80. Cheung AT, Ree D, Kolls JK, Fuselier J, Coy DH, Bryer-Ash M. An in vivo model for elucidation of the mechanism of tumor necrosis factor- α (TNF- α)-induced insulin resistance: evidence for differential regulation of insulin signaling by TNF- α . *Endocrinology* 1998; 139: 4928–4935.
 81. Kim HJ, Higashimori T, Park SY, Choi H, Dong JY, Kim YJ, Noh HL, Cho YR, Cline G, Kim YB, Kim JK. Differential effects of interleukin-6 and -10 on skeletal muscle and liver insulin action in vivo. *Diabetes* 2004; 53:1060–1067.
 82. Dechend R, Gieffers J, Dietz R, Joerres A, Rupp J, Luft FC, Maass M. Hydroxymethylglutaryl coenzyme A reductase inhibition reduces Chlamydia pneumoniae-induced cell interaction and activation. *Circulation* 2003; 108:261–265.
 83. Jin Z, Zhang W, Chai W, Zheng Y, Zhi J. Antibodies against AT1 receptors are associated with vascular endothelial and smooth muscle function impairment: protective effects of hydroxysafflor yellow A. *PLoS One* 2013; 8:e67020.
 84. D'Hoker J, De Leu N, Heremans Y, Baeyens L, Minami K, Ying C, Lavens A, Chintinne M, Stangé G, Magenheimer J, Swisa A, Martens G, et al. Conditional hypovascularization and hypoxia in islets do not overtly influence adult β -cell mass or function. *Diabetes* 2013; 62:4165–4173.
 85. Herse F, Fain JN, Janke J, Engeli S, Kuhn C, Frey N, Weich HA, Bergmann A, Kappert K, Karumanchi SA, Luft FC, Muller DN, et al. Adipose tissue-derived soluble fms-like tyrosine kinase 1 is an obesity-relevant endogenous paracrine adipokine. *Hypertension* 2011; 58:37–42.
 86. Patten IS, Rana S, Shahul S, Rowe GC, Jang C, Liu L, Hacker MR, Rhee JS, Mitchell J, Mahmood F, Hess P, Farrell C, et al. Cardiac angiogenic imbalance leads to peripartum cardiomyopathy. *Nature* 2012; 485: 333–338.
 87. Furukawa S, Fujita T, Shimabukuro M, Iwaki M, Yamada Y, Nakajima Y, Nakayama O, Makishima M, Matsuda M, Shimomura I. Increased oxidative stress in obesity and its impact on metabolic syndrome. *J Clin Invest* 2004; 114:1752–1761.
 88. Houstis N, Rosen ED, Lander ES. Reactive oxygen species have a causal role in multiple forms of insulin resistance. *Nature* 2006; 440:944–948.
 89. Tsukimori K, Fukushima K, Tsushima A, Nakano H. Generation of reactive oxygen species by neutrophils and endothelial cell injury in normal and preeclamptic pregnancies. *Hypertension* 2005; 46:696–700.
 90. Matsubara K, Matsubara Y, Hyodo S, Katayama T, Ito M. Role of nitric oxide and reactive oxygen species in the pathogenesis of preeclampsia. *J Obstet Gynaecol Res* 2010; 36:239–247.
 91. Walsh SW, Vaughan JE, Wang Y, Roberts LJ. Placental isoprostane is significantly increased in preeclampsia. *FASEB J* 2000; 14:1289–1296.
 92. Aris A, Benali S, Ouellet A, Moutquin J, Leblanc S. Potential biomarkers of preeclampsia: inverse correlation between hydrogen peroxide and nitric oxide early in maternal circulation and at term in placenta of women with preeclampsia. *Placenta* 2009; 30:342–347.
 93. Denery PA. Oxidative stress in development: nature or nurture? *Free Radic Biol Med* 2010; 49:1147–1151.

94. Mistry HD, Wilson V, Ramsay MM, Symonds ME, Pipkin FB. Reduced selenium concentrations and glutathione peroxidase activity in pre-eclamptic pregnancies. *Hypertension* 2008; 52:881–888.
95. Tsukahara H, Ohta N, Sato S, Hiraoka M, Shukunami K-I, Uchiyama M, Kawakami H, Sekine K, Mayumi M. Concentrations of pentosidine, an advanced glycation end-product, in umbilical cord blood. *Free Radic Res* 2004; 38:691–695.
96. Torrance HL, Krediet TG, Vreman HJ, Visser GH, van Bel F. Oxidative stress and proinflammatory cytokine levels are increased in premature neonates of preeclamptic mothers with HELLP syndrome. *Neonatology* 2008; 94:138–142.
97. Hilali N, Kocyigit A, Demir M, Camuzcuoglu A, Incebiyik A, Camuzcuoglu H, Vural M, Taskin A. DNA damage and oxidative stress in patients with mild preeclampsia and offspring. *Eur J Obstet Gynecol Reprod Biol* 2013; 170:377–380.
98. Erdem M, Harma M, Harma IM, Arikan I, Barut A. Comparative study of oxidative stress in maternal blood with that of cord blood and maternal milk. *Arch Gynecol Obstet* 2012; 285:371–375.
99. Braekke K, Harsem NK, Staff AC. Oxidative stress and antioxidant status in fetal circulation in preeclampsia. *Pediatr Res* 2006; 60:560–564.
100. Yaacobi N, Ohel G, Hochman A. Reactive oxygen species in the process of labor. *Arch Gynecol Obs* 1999; 263:23–24.
101. Lenzen S, Drinkgem J. Low antioxidant enzyme gene expression in pancreatic islets compared with various other mouse tissues. *Free Radic Biol Med* 1996; 20:463–466.
102. Kajiji M, Hirota M, Inai Y, Kiyooka T, Morimoto T, Iwasaki T, Endo K, Mohri S, Shimizu J, Yada T, Ogasawara Y, Naruse K, et al. Impaired NO-mediated vasodilation with increased superoxide but robust EDHF function in right ventricular arterial microvessels of pulmonary hypertensive rats. *Am J Physiol Heart Circ Physiol* 2007; 292: H2737–H2744.
103. Richter HG, Camm EJ, Modi BN, Naeem F, Cross CM, Cindrova-Davies T, Spasic-Boskovic O, Dunster C, Mudway IS, Kelly FJ, Burton GJ, Poston L, et al. Ascorbate prevents placental oxidative stress and enhances birth weight in hypoxic pregnancy in rats. *J Physiol* 2012; 590: 1377–1387.
104. Roberts JM, Myatt L, Spong CY, Thom EA, Hauth JC, Leveno KJ, Pearson GD, Wapner RJ, Varner MW, Thorp JM, Mercer BM, Peaceman AM, et al. Vitamins C and E to prevent complications of pregnancy-associated hypertension. *N Engl J Med* 2010; 362:1282–1291.
105. Parrish RM, Martin NJ, Lamarca BB, Ellis B, Parrish AS, Owens YM, May LW. Randomized, placebo controlled, double blind trial evaluating early pregnancy phytonutrient supplementation in the prevention of preeclampsia. *J Perinatol* 2013; 33:593–599.
106. Vadillo-Ortega F, Perchart-Perera O, Espino S, Avila-Vergara MA, Ibarra I, Ahued R, Godines M, Parry S, Macones G, Strauss JF. Effect of supplementation during pregnancy with L-arginine and antioxidant vitamins in medical food on pre-eclampsia in high risk population: randomised controlled trial. *BMJ* 2011; 342:d2901.
107. Yang R, Barouch LA. Leptin signaling and obesity: cardiovascular consequences. *Circ Res* 2007; 101:545–559.
108. Schrey S, Kingdom J, Baczyk D, Fitzgerald B, Keating S, Ryan G, Drewlo S. Leptin is differentially expressed and epigenetically regulated across monozygotic twin placenta with discordant fetal growth. *Mol Hum Reprod* 2013; 19:764–772.
109. Huynh FK, Neumann UH, Wang Y, Rodrigues B, Kieffer TJ, Covey SD. A role for hepatic leptin signaling in lipid metabolism via altered very low density lipoprotein composition and liver lipase activity in mice. *Hepatology* 2013; 57:543–554.
110. McCarthy JF, Misra DN, Roberts JM. Maternal plasma leptin is increased in preeclampsia and positively correlates with fetal cord concentration. *Am J Obstet Gynecol* 1999; 180:731–736.
111. Laivuori H, Kaaja R, Koistinen H, Karonen SL, Andersson S, Koivisto V, Ylikorkala O. Leptin during and after preeclamptic or normal pregnancy: its relation to serum insulin and insulin sensitivity. *Metabolism* 2000; 49:259–263.
112. Mise H, Yura S, Itoh H, Nuamah MA, Takemura M, Sagawa N, Fujii S. The relationship between maternal plasma leptin levels and fetal growth restriction. *Endocr J* 2007; 54:945–951.
113. Stepan H, Richter J, Kley K, Kralisch S, Jank A, Schaarschmidt W, Ebert T, Lössner U, Jessnitzner B, Kratzsch J, Blüher M, Stumvoll M, et al. Serum levels of growth arrest specific protein 6 are increased in preeclampsia. *Regul Pept* 2013; 182:7–11.
114. Eleuterio NM, Palei ACT, Rangel Machado JS, Tanus-Santos JE, Cavalli RC, Sandrim VC. Correlations between circulating levels of adipokines and anti-angiogenic factors in women with BMI <30 and a late-onset preeclampsia. *Hypertens Pregnancy* 2014; 33:72–80.
115. Hauth JC, Clifton RG, Roberts JM, Myatt L, Spong CY, Leveno KJ, Varner MW, Wapner RJ, Thorp JM, Mercer BM, Peaceman AM, Ramin SM, et al. Maternal insulin resistance and preeclampsia. *Am J Obstet Gynecol* 2011; 204:327.
116. Vrijkotte TGM, Krukziener N, Hutten BA, Vollebregt KC, Van Eijsden M, Twickler MB. Maternal lipid profile during early pregnancy and pregnancy complications and outcomes: the ABCD study. *J Clin Endocrinol Metab* 2012; 97:3917–3925.
117. Zhang Z, Yang X, Zhang L, Duan Z, Jia L, Wang P, Shi Y, Li Y, Gao J. Decreased expression and activation of Stat3 in severe preeclampsia. *J Mol Histol* 2015; 46:205–219.
118. Ødegård RA, Vatten LJ, Nilsen ST, Salvesen KÅ, Austgulen R. Umbilical cord plasma leptin is increased in preeclampsia. *Am J Obstet Gynecol* 2002; 186:427–432.
119. Yuen BSJ, Owens PC, Symonds ME, Keisler DH, McFarlane JR, Kauter KG, McMillen IC. Effects of leptin on fetal plasma adrenocorticotrophic hormone and cortisol concentrations and the timing of parturition in the sheep. *Biol Reprod* 2004; 70:1650–1657.
120. Kobayashi CI, Suda T. Regulation of reactive oxygen species in stem cells and cancer stem cells. *J Cell Physiol* 2012; 227:421–430.
121. Bauer S. Cytokine control of adult neural stem cells. *Ann N Y Acad Sci* 2009; 1153:48–56.
122. Chaudhari P, Ye Z, Jang Y-Y. Roles of reactive oxygen species in the fate of stem cells. *Antioxid Redox Signal* 2014; 20:1881–1890.
123. Poladia DP, Kish K, Kutay B, Bauer J, Baum M, Bates CM. Link between reduced nephron number and hypertension: studies in a mutant mouse model. *Pediatr Res* 2006; 59:489–493.
124. Corstius HB, Zimanyi MA, Maka N, Herath T, Thomas W, Van Der Laarse A, Wreford NG, Black MJ. Effect of intrauterine growth restriction on the number of cardiomyocytes in rat hearts. *Pediatr Res* 2005; 57:796–800.
125. Hinchliffe SA, Lynch MR, Sargent PH, Howard CV, Van Velzen D. The effect of intrauterine growth retardation on the development of renal nephrons. *Br J Obstet Gynaecol* 1992; 99:296–301.
126. Cebrian C, Asai N, D'Agati V, Costantini F. The number of fetal nephron progenitor cells limits ureteric branching and adult nephron endowment. *Cell Rep* 2014; 7:127–137.
127. Stanger BZ, Tanaka AJ, Melton DA. Organ size is limited by the number of embryonic progenitor cells in the pancreas but not the liver. *Nature* 2007; 445:886–891.
128. Terman A, Kurz T, Navratil M, Arriaga EA, Brunk UT. Mitochondrial turnover and aging of long-lived postmitotic cells: the mitochondrial-lysosomal axis theory of aging. *Antioxid Redox Signal* 2010; 12: 503–535.
129. Rafalski VA, Mancini E, Brunet A. Energy metabolism and energy-sensing pathways in mammalian embryonic and adult stem cell fate. *J Cell Sci* 2012; 125:5597–5608.
130. Jaenisch R, Young R. Stem cells, the molecular circuitry of pluripotency and nuclear reprogramming. *Cell* 2008; 132:567–582.
131. Ellis HL, Shioda K, Rosenthal NF, Coser KR, Shioda T. Masculine epigenetic sex marks of the CYP19A1/aromatase promoter in genetically male chicken embryonic gonads are resistant to estrogen-induced phenotypic sex conversion. *Biol Reprod* 2012; 87:23.
132. Marcho C, Bevilacqua A, Tremblay KD, Mager J. Tissue-specific regulation of Igf2r/Airn imprinting during gastrulation. *Epigenetics Chromatin* 2015; 8:10.
133. Cantone I, Fisher AG. Epigenetic programming and reprogramming during development. *Nat Struct Mol Biol* 2013; 20:282–9.
134. Orozco LD, Rubbi L, Martin LJ, Fang F, Hormozdiari F, Che N, Smith AD, Lusk AJ, Pellegrini M. Intergenerational genomic DNA methylation patterns in mouse hybrid strains. *Genome Biol* 2014; 15:R68.
135. Nissenbaum J, Bar-Nur O, Ben-David E, Benvenisty N. Global indiscriminate methylation in cell-specific gene promoters following reprogramming into human induced pluripotent stem cells. *Stem Cell Reports* 2013; 1:509–517.
136. Hu W, Weng X, Dong M, Liu Y, Li W, Huang H. Alteration in methylation level at 11β-hydroxysteroid dehydrogenase type 2 gene promoter in infants born to preeclamptic women. *BMC Genet* 2014; 15: 96.
137. He J, Zhang A, Fang M, Fang R, Ge J, Jiang Y, Zhang H, Han C, Ye X, Yu D, Huang H, Liu Y, et al. Methylation levels at IGF2 and GNAS DMRs in infants born to preeclamptic pregnancies. *BMC Genomics* 2013; 14:472.

138. Aufdenblatten M, Baumann M, Raio L, Dick B, Frey BM, Schneider H, Surbek D, Hocher B, Mohaupt MG. Prematurity is related to high placental cortisol in preeclampsia. *Pediatr Res* 2009; 65:198–202.
139. Bourque DK, Avila L, Peñaherrera M, von Dadelszen P, Robinson WP. Decreased placental methylation at the H19/IGF2 imprinting control region is associated with normotensive intrauterine growth restriction but not preeclampsia. *Placenta* 2010; 31:197–202.
140. Ching T, Ha J, Song M-A, Tiirikainen M, Molnar J, Berry MJ, Towner D, Garmire LX. Genome-scale hypomethylation in the cord blood DNAs associated with early onset preeclampsia. *Clin Epigenetics* 2015; 7.
141. Reddington JP, Perricone SM, Nestor CE, Reichmann J, Youngson NA, Suzuki M, Reinhardt D, Dunican DS, Prendergast JG, Mjoseng H, Ramsahoye BH, Whitelaw E, et al. Redistribution of H3K27me3 upon DNA hypomethylation results in de-repression of Polycomb target genes. *Genome Biol* 2013; 14:R25.
142. Li Y, Zhang Y, Li S, Lu J, Chen J, Zhao Z, Bai J, Xu J, Li X. Genome-wide DNA methylome analysis reveals novel epigenetically dysregulated non-coding RNAs in human breast cancer. *Sci Rep* 2014; 1–12.
143. Jin Z, Zhang W, Yang H, Wang X, Zheng Y, Zhang Q, Zhi J. Maternal treatment with agonistic autoantibodies against type-1 angiotensin ii receptor in late pregnancy increases apoptosis of myocardial cells and myocardial susceptibility to ischemia-reperfusion injury in offspring rats. *PLoS One* 2013; 8:e80709.